

Fig. S1. The addition of a Flag tag to the C-terminus of ACE3 and XYR1 and deletion of *ace3*. (A), Construction of the ACE3-Flag, XYR1-Flag, and $\Delta ace3$ mutants. (B), *T. reesei* strains cultured on MA medium, using lactose as the sole source of carbon. LML 2.1 is the erasable hygromycin selection marker in *T. reesei*.



Fig. S2. The addition of a Flag tag to the C-terminus of ACE3 inhibits cellulase production. Biomass production from *T. reesei* strains on MA medium, using 2% glucose (A), xylan (B), or Avicel (C) as the sole source of carbon. (D), Activities of *p*NPCase and xylanase in *T. reesei* strains, in the presence of Avicel and xylan, respectively. Values are the mean \pm SD of the results from three independent experiments. Asterisks indicate a significant difference compared to the untreated strain (**p* < 0.05, Student's *t* test). n.s. indicates no significant difference.



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Fig. S3. Adding a Flag tag to the N-terminus of ACE3 or XYR1. (A) Construction of Flag-ACE3 and Flag-XYR1. (B) *p*NPCase and xylanase activities of *T. reesei* strains in Avicel and xylan, respectively. LML 2.12 is the erasable hygromycin selection marker in *T. reesei*. Values are the mean \pm SD of the results from three independent experiments. Asterisks indicate a significant difference compared to the untreated strain (*p < 0.05, Student's *t* test). n.s. indicates no significant difference.



Fig. S4. Differential transcription pattern of the WT and $\Delta ace3$ strain in response to lactose, as measured by RNA-Seq. (A), Volcano plot analysis of up and downregulated genes in the absence of *ace3* versus that observed for the WT in the presence of lactose. (B), GO annotations analysis of differential genes for the $\Delta ace3$ and QM6a strain in response to lactose. (C), number of differential genes in each molecular functional category.



Fig. S5. Phenotypic differences of yellow pigment production in *T. reesei* QM6a, Δ*ace3*, 6aACE3-Flag, and 6aA7 strains.

Coils output for ACE3



Fig. S6. Predicted coiled-coil domains in the complete ACE3 sequence using COILS.



Fig. S7. Phylogenetic analysis of ACE3 and its homologs. Sequence alignments were performed with ClustalW, and the neighbor-joining tree was generated with MEGA 6.0. Numbers on the tree branches represent the bootstrap support calculated per 1000 bootstrap replicates. ACE3 sequences belonging to the *Trichoderma* clade are indicated by boxes.



Fig. S8. *p*NPCase activities (A) and relative transcriptional levels of *cbh1* (B) of *T. reesei* strains QM6a and 6aDcbh1p1AB in Avicel. Values are the mean \pm SD of the results from three independent experiments. Asterisks indicate a significant difference compared to the untreated strain (**p* < 0.05, Student's *t* test).



Fig. S9. EMSAs of ACE3 bound to the promoter regions of selected genes.



Fig. S10. Truncation of the C-terminus of ACE3 terminates its function. (A), Construction of *T. reesei* 6aA8 and 6aA7. The native ACE3 cassette named A8 that contains all 734 amino acids of ACE3, is used as the positive control. The A7 cassette (amino acids 1-700 of ACE3) is constructed by the deletion of the C-terminal 34 amino acids of ACE3. (B), *p*NPCase and xylanase activities of *T. reesei* strains 6aA8 and 6aA7 in Avicel and xylan, respectively. (C), Relative transcriptional levels of *cbh1* in *T. reesei* strains 6aA8 and 6aA7, in Avicel. LML 2.1 is the erasable hygromycin selection marker in *T. reesei*. Values are the mean \pm SD of the results from three independent experiments. Asterisks indicate a significant difference compared to the untreated strain (**p* < 0.05, Student's *t* test).



Fig. S11. Truncation of the C-terminus of XYR1 terminates its function. (A), Construction of *T. reesei* TRX8, TRX7, TRX6, TRX5, TRX3, and TRX2. The native XYR1 cassette named X8, which contains all 940 amino acids of XYR1, is used as the positive control. XYR1 truncated analogues X7 (amino acids 1-909 of XYR1), X6 (amino acids 1-865 of XYR1), X5 (amino acids 1-775 of XYR1), X3 (amino acids 1-700 of XYR1), and X2 (amino acids 1-368 of XYR1) were constructed by deleting the corresponding amino acids. (B), *p*NPCase activities of *T. reesei* strains TRX8, TRX7, TRX6, TRX5, TRX3, and TRX2 in Avicel. (C), Relative transcriptional levels of *cbh1* in *T. reesei* strains TRX8, TRX7, TRX6, TRX5, TRX3, and TRX2 in Avicel. (D), Xylanase activities of *T. reesei* strains TRX8, TRX7, TRX6, TRX5, TRX3, and TRX2 in xylan. LML 2.1 is the erasable hygromycin selection marker in *T. reesei*. Values are the mean \pm SD of the results from three independent experiments. Asterisks indicate a significant difference compared to the untreated strain (**p* < 0.05, Student's *t* test).



Fig. S12. Relative transcriptional levels of xyn1 and xyn2 in *T. reesei* QM6a, TRX3, TRX5, TRX6, TRX8, and 6aXYR1-Flag in Xylan. (A) xyn1. (B) xyn2. Values are represented as mean \pm SD of the results from three independent experiments. n.s. indicates no significant difference.



Fig. S13. Relative transcriptional levels of *cbh1*, *cbh2*, *egl1*, and *egl2* in *T. reesei* strains TRX6G4 and TRX6 in Avicel. Values are the mean \pm SD of the results from three independent experiments. Asterisks indicate a significant difference compared to the untreated strain (*p < 0.05, Student's *t* test).



respectively. The two domains of mCherry (mN, N-terminal aa 1–159 of mCherry; mC, C-terminal aa 160–237 of mCherry) were fused with the N-terminus of XYR1 or ACE3. The mCherry fused XYR1 and ACE3 were under the control of the *pdc* promoter and were transformed successively into *T. reesei*. Positive interactions in the co-expressed strains TRmC-XYR1/mN-ACE3 and TRmC-ACE3/mN-XYR1 resulted in red fluorescence. TRmC-XYR1, TRmN-ACE3, TRmC-ACE3, TRmN-XYR1, and QM6a are used as negative controls. The BiFC assay results revealed the protein-protein interactions of the XYR1c and ACE3a domains in *E. coli*. Strains expressing protein partners fused with mN or mC are indicated on each panel. Negative controls for the BiFC assay of the interaction of the XYR1c-ACE3a domains were performed (XYR1c-mN + mC and mN + mC-ACE3a). Positive interactions resulted in fluorescence.