Aryl hydrocarbon receptor (AhR) regulates adipocyte differentiation by assembling CRL4B ubiquitin ligase to target PPARγ for proteasomal degradation

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Table S1. Primers used in real-time PCR.

AhR fwd:	5'-GCCAGGACCAGTGTAGAGC-3'
AhR rev:	5'- ATTCAGCGCCTGTAACAAGAA -3'
PPARγ fwd:	5'- CCATTCTGGCCCACCAAC -3'
PPARγ rev:	5'- AATGCGAGTGGTCTTCCATCA -3'
CUL4B fwd:	5'- ACCTCGTCCTTCTGCCTTGGG -3'
CUL4B rev:	5'- CTGAGCCGGGCTTGCTGTTAG -3'
GAPDH fwd:	5'- TGTGTCCGTCGTGGATCTGA -3'
GAPDH rev:	5'- CCTGCTTCACCACCTTCTTGAT-3'
Fabp4 fwd:	5'- GATGCCTTTGTGGGAACCTG -3'
Fabp4 rev:	5'- TCCTGTCGTCTGCGGTGATT -3'
Adipsin fwd:	5'- CTGAACCCTACAAGCGAT -3'
Adipsin rev:	5'- GACCCAACGAGGCATTCT -3'
CD36 fwd:	5'- GGCCAACGTATTGCGACAT -3'
CD36 rev:	5'- CAGATCCGAACACAGCGTAGA -3'



Figure S1. 3T3-L1 differentiation. mRNA levels of *PPAR* γ and its related genes in 3T3-L1 cells. At different time points post-treatment, total RNA samples were extracted and subjected to qRT-PCR analysis of related genes (*Fabp4*, *CD36*, *Adipsin*, *Cul4b* and *Ahr*). Data were presented as mean \pm SD.; n=4 with ***P* < 0.01, ****P* < 0.001 by student's *t* test.



Figure S2. Efficiency of the overexpression and knockdown of AhR in 3T3-L1 cells. A and B, The mRNA levels of overexpression and knockdown of AhR were validated by qPCR.

C and D, The protein levels of overexpression and knockdown of AhR were validated by Western blots.





A, AhR did not affect PPARγ mRNA level in wild-type and AhR overexpression 3T3-L1 cells.
mRNA levels were measured by qRT-PCR analysis using primers targeted for AhR and PPARγ.
B, AhR did not affect PPARγ mRNA level in wild-type and AhR knockdown 3T3-L1 cells.
mRNA levels were measured by qRT-PCR analysis using primers targeted for AhR and PPARγ.



Figure S4. Defect in ubiquitination does not increase acetylation level of PPARy.

3T3-L1 cells were first transfected with siAhR or siControl. Cells were then transfected with plasmids expressing Flag-Cul4B, HA-Ub and Myc-PPARγ. Cells were treated with MG132 for 3 hours. Cells were then harvested for the following immunoprecipitation and Western blotting. Input (5%) was used for Western blotting.