Rare protein-altering telomere-related gene variants in patients with chronic hypersensitivity pneumonitis

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ONLINE DATA SUPPLEMENT

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SUPPLEMENTAL METHODS

Clinical Data Collection:

Clinical data collection included demographics, pulmonary function tests, and radiologic, and histopathologic studies. Race and ethnicity were self-reported on enrollment questionnaires. Family history of interstitial lung disease was self-reported on enrollment questionnaires and verified by review of ILD clinical notes. The date of death was recorded clinically and confirmed using the United States Social Security Death Index. Lung transplants and date of lung transplants were recorded clinically. HRCTs were scored by expert chest radiologists (B.M.E. or T.S.H.) using a structured data collection form that included assessment of 2011 ATS/ERS/JRS/ALAT criteria for UIP patterns, semi-quantitative extent of fibrosis, and individual morphologic features (Figure E1). Surgical lung biopsies were scored prospectively in the UCSF cases only, at the time of multidisciplinary team discussion, by an expert lung pathologist (K.J.D.) using a structured pathology data collection form (Figure E2).

DNA Sequencing:

Sequencing was performed on DNA isolated from peripheral blood in both cohorts. Whole genome sequencing data was generated in the discovery cohort to a read depth of 30X using the Illumina HiSeq X Ten sequencer (Human Longevity Inc, San Diego, CA, USA) and KAPA Hyper Library Kit (Roche Sequencing, Pleasanton, CA, USA). Whole exome sequencing data was generated in the replication cohort to a read depth of 50X using the Illumina Novaseq 6000 S4 Flowcell sequencer (Novogene, Sacramento, CA, USA) and the SureSelect Human All Exome V6 capture kit (Agilent, Santa Clara, CA, USA).

Variant discovery

Discovery cohort: Detailed variant discovery methods for this cohort are as described in [E1]. *Replication cohort:* Raw Illumina paired-end 150bp reads were first subjected to quality control. Adapters were removed from the sequencing reads using scythe (https://github.com/vsbuffalo/scythe, version 0.994 beta). Base quality was controlled using a window-based method, sickle (https://github.com/najoshi/sickle, version 1.33), with the cutoff set at 30. Reads that are less than 30bp in length were discarded. Next, reads that have passed the quality control were mapped to GRCh38 reference genome using BWA-MEM[E2] with parameter -M for downstream analysis compatibility. PCR duplicates were removed using Picard-tools (http://broadinstitute.github.io/picard/, version 2.18.4). Variants were identified using HaplotypeCaller function in GATK[E3] (version 4.0.5.2), followed by variant recalibration using the recommendations from GATK developers.

Variants were annotated for genomic effect using snpEff (v4.3q) and identifiers using dbSNP build 150 (release July 10, 2017).

Samples in both cohorts were excluded for call rates less than 90% (no samples were excluded), identity by descent analysis (patient relatedness), $\pi 0.125$ or greater (i.e. third-degree relatives), or sex mismatch. Variant sites were excluded for call rates less than 95%.

Principle Component Analysis

Principal component analysis (PCA) was performed in Plink version 1.9 (see Figures E3-E6). A European subgroup was selected using the first two principal components corresponding to the cluster of self-reported non-Hispanic white patients (discovery cohort: PC1 range -0.05 to -0.022, PC2 range -0.025 to 0.025; replication cohort: PC1 and range 0.0-0.05). PCA was then repeated in this subgroup and outliers greater than 3 standard deviations of the mean of PC1 or in the restricted PCA were further excluded.

Case-Control Analysis

Discovery Cohort: Discovery cohort cases were age- and sex-matched to age-related macular degeneration (AMD) and asthma samples that were sequenced on the same platform during the same calendar years as described in reference [E1]. Cases and controls of European ancestry (EUR admixture >=0.8) were included in the analysis:

Variable	CHP cases	Controls
Ν	117	351

Age in years, mean (sd)	63.5 (11.2)	63.6 (10.8)
Female, N (%)	68 (58.1)	180 (51.3)
N AMD	NA	124
N Asthma	NA	227

The frequency of qualifying variants in cases and controls (per individual) were compared using the odds ratio and Fischer's exact p-values.

Replication Cohort: Qualifying variants were identified in the entire Exome Aggregation Consortium (v0.3.1) (ExAC) dataset (n=60,706) and the ExAC non-Finnish European (NFE) sub-population (n=33,370) using the same criteria as CHP cases. The frequency of qualifying variants in cases and controls (per individual) were compared using the odds ratio and Fischer's exact p-values. This was done for all CHP cases compared to the entire ExAC cohort and again restricted to the CHP cases of non-Hispanic white race/ethnicity compared to the ExAC NFE sub-population.

References

[E1] Dressen A, Abbas AR, Cabanski C, et al. Analysis of protein-altering variants in telomerase genes and their association with MUC5B common variant status in patients with idiopathic pulmonary fibrosis: a candidate gene sequencing study. The Lancet Respiratory medicine (2018).

[E2] L. H., Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM, arXiv:1303.3997 (2013).

[E3] R. Poplin, V. Ruano-Rubio, M.A. DePristo, T.J. Fennell, M.O. Carneiro, G.A. Van der Auwera, D.E. Kling, L.D. Gauthier, A. Levy-Moonshine, D. Roazen, K. Shakir, J. Thibault, S. Chandran, C. Whelan, M. Lek, S. Gabriel, M.J. Daly, B. Neale, D.G. MacArthur, E. Banks, Scaling accurate genetic variant discovery to tens of thousands of samples, bioRxiv (2018).

SUPPLEMENTAL TABLES AND FIGURES

Figure E1. Computed tomography of the chest data collection form

			Radiology Scorin	g and HR	CT Classification H	Form	
Nam	e	MRN	ILD #	Clinic Date	Biopsy Date	Review Date	Reviewer
Г	1. CT scan attributes				6. Nodules	No Yes	
	Date		No CT Availa	ble 🗌 Yes	Severity	Mild Moderate Severe	B
	Min Section Thickness				Definition	Ground glass Dense	
	Skipped or full chest	Skip	ped Full chest		Size	□<5 mm □ 5-10 mm □>10	-30 mm □>30 mm
	Supine images	No	□ Yes		Calcification	□No □Yes	
	Prone images	□ No	□ Yes		Craniocaudal distribution	□Lower □Middle □Upper	Diffuse
	Expiratory Images	□ No	□ Yes		Distribution	Perilymphatic Random	Centrilobular
L	Edge Enhanced	□ No	Yes		Tree-in-bud		
L	Scanned-in films	□ No	□Yes		7. Emphysema	⊠No □Yes	
					Centrilobular	None Mild Moderate	Severe
	2. Fibrosis	No	□ Yes		Paraseptal	None Mild Moderate	Severe
L	Irregular reticulation	None	e Mild Moderate Sever	e	Panlobular	None Mild Moderate	Severe
	Traction bronchiectasis	□ None	e Mild Moderate Sever	e			
L	Honeycombing	□ None	e Mild Moderate Sever	e	8. Misc. Lung Findings	⊠No ∐Yes	
	Craniocaudal distribution	Low	er 🗌 Middle 🗌 Upper 🗌 Diffu	se	Cysts	None Mild Moderate	Severe
- L	Axial distribution	Perip	heral Central Diffuse		Interiobular septal thick.	None Mild Moderate	Severe
	PMF-like opacities	□ No	☐ Yes		Crazy Paving		Severe
- H	PPFE-like opacities	L No	Yes Analysis not performed		Subpleural Sparing		
	Assoc. pulm. ossification		Yes Analysis not performed		Reversed Halo		
_					Perilobular Opacities		
	3. Ground glass opacity	No	□ Yes		9. Non-lung findings	⊠No □Yes □Analysis not	performed
_ L	Severity	☐ Mild	Moderate Severe		Extensive LAD	□ No □ Yes □ Analysis not	performed
_ L	Craniocaudal distribution	Low	er Middle Upper Diffu	ise	Pleural disease	□ No □ Yes □ Analysis not	performed
L	Axial distribution	Perip	bheral Central Diffuse		Esophageal dilation	No Yes Analysis not	performed
_					10.0		
L	4. Consolidation	No	□ Yes		10. Summary		
_ L	Severity	☐ Mild	Moderate Severe		UIP designation (new)		e for UIP
	Craniocaudal distribution	Low	er Middle Upper Diffu	se		Probable UIP Alternative of	liagnosis
L	Axial distribution	Perij	oheral Central Diffuse		UIP designation (old)		nsistent
_					Leading Diagnosis		
	5. Airways disease	No	□ Yes		Alt. diagnosis 1	1	
	MosaicPerfusion	Non	e □Mild □Moderate □Seve	re	Alt. diagnosis 2		
	AirTrapping	Non	e Mild Moderate Seve	re □N/A	Confidence in diagnosis		V
	Airways inflammation	Non	e Mild Moderate Seve	re	INDICES		
N	on-traction bronchiectasis	Non	e ☐Mild ☐Moderate ☐Seve	re	Empty Fields	13	
		-			LUP discremency?	15	
		D	EFINITIONS		>1 check selected	0	
	Ground glass onacity. Cont	fluent are	as of opacity that do not obscure th	e underlying	Actual Diagnosis	0	
	ressels. This is in the absend	ce of sign	ificant associated irregular reticula	tion.	Tetuar Diagnosis		
	Consolidation: Confluent an	eas of or	acity that do obscure the underlyin	g vessels.		OTHER REENITIONS	
1	rregular reticulation: Line	ar opacit	ies that do not represent smooth or	nodular	11	OTHER DEFINITIONS	
i	nterlobular septal thickening	g.			Severity:		
1	Fraction bronchiectasis: D	ilated and	l irregular bronchi due to adjacent	fibrosis with a	Mild: <10% lung invo	olvement	
1	ack of inflammatory thicker	ning or m	ucous impaction.		Moderate: 10-50% lui	ng involvement	
1	Ioneycombing: Air density	cysts wi	th relatively thick walls that are fir	st seen in the	Severe: >50% lung in	volvement	
S	ubpleural lung and then bec	ome stac	ked into multiple layers. At least 3	air density	For mosaic perfusion	and air trapping to be moderate or	severe they must also
1	ysis lined up in the subpleu	ie must h	a single row must be present. Al	so, at least	be present bilaterally and in	at least 3 lobes	
	Mosaic perfusion: Geograp	hic areas	of decreased lung attenuation with	sharn	Bronchiectasis (traction or	non-traction) severity:	
	nargins seen on inspiratory	CT.	or decreased rang automation with	or p	Calculate severity in e	each lobe as (0=none, 1=mild, 2=r	noderate, 3=severe)
	Air trapping: On expirtory	images, t	he presence of areas of lung that do	not	The sum of all lobes of	letermines overall severity:	
s	ignificantly increase in atter	nuation w	hen compared to the inspiratory in	nages.	Madarata 4.7		
1	Nodules: Multiple rounded	opacities.			Severe >8		
1	Emphysema: Centrilobular	emphyse	ma are focal, air density lucencies	without a wall	Confidence in diagnosis:		
	minally and aminating in th	a a a set to a 1	mentions of the unner labor Dame	antal			

typically predominating in the central portions of the upper lobes. <u>Paraseptal</u> <u>emphysema</u> are focal, air density lucencies with a thin wall located in the subpleural lung in a single layer. Panlobular emphysema is diffuse lung lucency with a lower

lung predominance. Cysts: Focal air attenuation lucencies with a thin, but perceptible wall that typically predominate in the central lung, however occasionally may be subpleural in location.

9. Non-lung findings	No ☐ Yes ☐ Analysis not performed
Extensive LAD	□ No □ Yes □ Analysis not performed
Pleural disease	□ No □ Yes □ Analysis not performed
Esophageal dilation	□ No □ Yes □ Analysis not performed
10. Summary	
UIP designation (new)	UIP Indeterminate for UIP
	Probable UIP Alternative diagnosis
UIP designation (old)	Definite Possible Inconsistent
Leading Diagnosis	
Alt. diagnosis 1	
Alt. diagnosis 2	
Confidence in diagnosis	High Intermediate Low
Notes	
Empty Fields	13
UIP discrepancy?	
>1 check selected	0
Actual Diagnosis	
	OTHER DEFINITIONS
everity:	
Mild: <10% lung inv	olvement
Source >50% lung in	ng involvement
Severe: >50% lung if	and air tranning to be moderate or severe they must also
present bilaterally and in	and an inapping to be moderate or severe they must also at least 3 lobes
ronchiectoric (treation of	a least 5 loves

tion or non-traction) severity: rity in each lobe as (0=none, 1=mild, 2=moderate, 3=severe) lobes determines overall severity: e 4-7 8 osis: High: ≥90% likelihood

Intermediate: 51-89% likelihood Low: ≤50% likelihood

Name:			
Last First			
Biopsy date: Biopsy site(s): RU		LUL Lingula	TBBx
Pathologist: Jones Other: RL		Lliguia LLL	Other
GENERAL OBSERVATIONS			
Low Magnification			
1. Heterogeneous distribution of abnormal areas	Yes	No	N/A
2. Accentuation of fibrosis at the pleural surface	Yes	No	N/A
3. Abnormalities airway centered	Yes	No	N/A
4. Diffuse alveolar septal thickening is present	Yes	No	N/A
High Magnification			
1. Temporal heterogeneity	Yes	No	N/A
2. Inflammation confined to areas of fibrosis	Yes	No	N/A
3. Microscopic honeycombing	Yes	No	N/A
4. Areas of normal lung	Yes	No	N/A
Specific Findings ($0 = ABSENT$, $1 = MILD$, $2 = MODER$	ATE, 3 =	MARKED)	Di t
1. Fibroblast foci (interstitial young connective tissue)	0	1	2 3
2. Organizing pneumonia (airway luminal granulation tissue	0	1	2 3
3. Lymphocytic interstitial infiltrate in areas on non-fibrotic lung	0	1	2 3
4. Alveolar macrophages	0	1	2 3
5. Dense collagen fibrosis	0	1	2 3
6. Granuloma/Giant cells	0	1	2 3
7. Germinal centers	0	1	2 3
8. Inorganic dust deposits	0	1	2 3
9. Hemosiderin laden macrophages and intra-alveolar blood	0	1	2 3
10. Eosinophilia/Eosinophilic abscess	0	1	2 3
11. Airway-centered inflammation	0	1	2 3
12. Small airways disease	0	1	2 3
13. Emphysema	0	1	2 3
14. Acute lung injury (hyaline membranes)	0	1	2 3

ILD HISTOPATHOLOGIC SCORING FORM



Figure E3. Principal component analysis, discovery cohort. Black lines define the European subgroup.

Figure E4. Principal component analysis, discovery cohort, restricted to the European subgroup, after excluding outliers.





Figure E5. Principal component analysis, replication cohort. Black lines define the European subgroup.

Figure E6. Principal component analysis, discovery cohort, restricted to the European subgroup, after excluding outliers.



	Cases	Cases	Controls	Controls		
	Count	Frequency	Count	Frequency	Odds Ratio	p-value
TERC	0	0	1	0.0028	NA	NA
TERT	3	0.0256	0	0	23.72	0.0441
TINF2	0	0	2	0.0057	0.38	0.5502
PARN	2	0.0171	1	0.0028	4.2	0.1643
RTEL1	4	0.0427	4	0.0114	3.38	0.0582
DKC1	1	0.0085	0	0	NA	NA
Combined	10	0.094	8	0.022	4.49	0.00173

 Table E1. Case-Control Analysis: Discovery Cohort

	Replication	(n=209)	ExAC (n=60,706)			
Gene	Count	Frequency	Count	Frequency	OR (95%CI)	p-value
TERC	0	0	120	0.0019767	NA	NA
TERT	9	0.0430622	131	0.0021579	20.8 (9.2-41.5)	1.65E-09
TINF2	0	0	158	0.0026027	NA	NA
PARN	2	0.00956938	126	0.0020756	4.65 (0.55-17.3)	0.719
RTEL1	6	0.02870813	536	0.0088294	3.32 (1.20-7.39)	0.0115
DKC1	0	0	47	0.0007742	NA	NA
Combined	17	0.08133971	1118	0.0184166	4.71 (2.68-7.79)	1.87E-11

 Table E2. Case-Control Analysis: Replication Cohort (all cases compared to all ExAC)

 Table E3. Case-Control Analysis: Replication Cohort (White/European individuals)

	Replication (n=169)		ExAC NFE (n=33,370)			
Gene	Count	Frequency	Count	Frequency	OR (95%CI)	p-value
TERC	0	0	59	0.00176806	NA	NA
TERT	6	0.035503	69	0.00206773	17.8 (6.2-41.5)	2.26E-06
TINF2	0	0	96	0.00287684	NA	NA
PARN	2	0.0118343	67	0.00200779	5.95 (0.70-22.7)	0.0475
RTEL1	5	0.0295858	294	0.00881031	3.43 (1.09-8.26)	0.0181
DKC1	0	0	27	0.00080911	NA	NA
Combined	13	0.0769231	612	0.01833983	4.46 (2.31-7.91)	1.94E-08

<u>Abbreviations:</u> ExAC = Exome Aggregation Consortium v0.3.1; NFE = non-Finnish European; OR = odds ratio; CI = confidence interval

Table E4. Clinical, Radiographic, and Pathologic Data for Individuals with Qualifying Variants

Gene	dbSNP	CHR	Position	REF	ALT	Effect	Exon	Protein	ExAC AF	ESP AF	PP2	SIFT	CADD	Cohort	Race/Ethnicity	Exposure	FH	TL	HRCT features	Biopsy findings
TERT	rs370445231	5	1260593	G	А	missense	12/16	R951W	8.26E-06	Absent	D (0.999)	T (0.106)	17.5	Replication	Hispanic	Mold	Yes	5559	Incon: Mod fib, PBV, GGO	cfNSIP
TERT	rs775014633	5	1268538	G	Α	missense	9/16	A855V	8.24E-06	Absent	P (0.522)	T (0.199)	7.4	Discovery	White	Down	No	5266	Incon: Mod fib, UM, AT	PBF, PFGs
TERT	rs387907249	5	1278781	G	Α	missense	6/16	A716T	Absent	Absent	D (0.998)	D (0.0)	23.1	Discovery	White	Bird	Yes	5055	Incon: UM	fNSIP, PFGs
TERT	rs866282352	5	1279329	G	Α	missense	5/16	R698W	Absent	Absent	D (0.981)	D (0.008)	23.1	Discovery	White	Mold	No	4248	Incon: Mild fib, GGO, AT	PBI, OP, PFGs
TERT	n/a	5	1280334	G	Т	missense	4/16	H592N	Absent	Absent	D (0.996)	D (0.005)	23.7	Replication	White	Unknown	No	4312	Incon: Mod fib, AT	Insufficient material
TERT	n/a	5	1282488	С	Α	missense	3/16	K570N	Absent	Absent	D (0.965)	D (0.0)	25.0	Discovery	White	Bird	Yes	4832	Possible	cfNSIP, PFGs
															African					
TERT	n/a	5	1282507	G	Α	missense	3/16	T564M	Absent	Absent	D (1.0)	D (0.0)	26.5	Replication	American	Unknown	No	5380	Incon: Mod fib, UM, PBV, AT	LII, NNGs
TERT	n/a	5	1293394	С	Т	missense	2/16	G498R	Absent	Absent	D (1.0)	D (0.0)	25.5	Replication	Hispanic	Down	No	4783	Incon: Mod fib, AT	No biopsy
TERT	rs34094720	5	1293652	G	А	missense	2/16	H412Y	0.0022	0.0043	D (0.998)	D (0.007)	20.2	Discovery	Hispanic	Unknown	No	6511	Incon: Mild fib, AT	cNSIP
														Discovery	White	Unknown	Yes	5782	Incon: Mod fib, GGO	cfNSIP, PFGs
														Discovery	White	Unknown	No	5486	Incon: No fib, GGO, AT	No biopsy
														Replication	White	Unknown	No	5516	Incon: Mild fib, UM	No biopsy
TERT	n/a	5	1294393	Т	С	missense	2/16	S165G	Absent	Absent	P (0.734)	D (0.005)	21.1	Replication	White	Mold	No	5415	Incon: Sev fib, PBV, AT, CLN	UIP+PFGs
TERT	rs1060502990	5	1294549	С	CG	frameshift	2/16	112-113X	Absent	Absent	n/a	n/a	15.9	Replication	White	Mold	No	4762	Incon: No fib, GGO, AT	No biopsy
TERT	n/a	5	1294656	Α	G	missense	2/16	L77P	Absent	Absent	D (0.0997)	D (0.028)	23.8	Replication	White	Unknown	No	4493	Possible	UIP+PFGs
TERT	n/a	5	1294829	С	Т	missense	1/16	C54Y	Absent	Absent	D (1.0)	D (0.011)	23.1	Replication	White	Bird	No	5010	Incon: Sev fib, UM, PBV, AT	fNSIP
TERC	rs552679780	3	169765027	С	Т	n/a	n/a	n/a	Absent	Absent	n/a	n/a	10.4	Discovery	Asian	Unknown	Yes	4850	Incon: Mild fib, UM, GGO	BCF, LII, PFGs
PARN	rs377199187	16	14432695	С	G	missense	22/24	R538P	8.28E-05	3.63E-04	P (0.928)	T (0.055)	4.1	Discovery	White	Unknown	No	5863	Incon: Mild fib, UM, PBV, GGO, AT	BCF, PFGs
PARN	n/a	16	14554142	Т	С	missense	20/24	K443R	Absent	Absent	P (0.755)	T (0.167)	25.5	Replication	White	Bird	No	5284	Incon: Mod fib, UM, PBV, GGO, AT	cNSIP
PARN	rs774170618	16	14604184	G	Α	missense	11/24	R249C	3.31E-05	Absent	D (0.998)	D (0.016)	31	Discovery	White	Mold	No	5683	Incon: Mod fib, UM, PBV, AT	No biopsy
																				UIP+BM, upper lobe
PARN	n/a	16	14610777	G	А	stop gained	7/24	Q141*	Absent	Absent	n/a	n/a	31	Replication	White	Bird	No	5788	Incon: Mod fib, UM	biopsy predominant
RTEL1	rs151214675	20	63661882	G	А	missense	4/35	A112T	2.22E-04	2.33E-04	D (0.998)	D (0.005)	27.3	Discovery	White	Down	No	5767	Incon: No fib, GGO, AT	No biopsy
														Replication	White	MAC	No	6216	Incon: No fib, GGO, AT, CLN	PBI, NNGs
													27.8							SLB: cfNSIP, OP, PFGs;
RTEL1	n/a	20	63672587	G	Т	missense	9/35	G244V	Absent	Absent	D (0.965)	D (0.003)		Discovery	White	Down	Yes	5386	Definite	Explant: UIP+PFGs
RTEL1	n/a	20	63688042	Т	Α	missense	18/35	F529L	Absent	Absent	P (0.85)	D (0.01)	22.6	Discovery	White	Mold	No	6630	Incon: Mod fib, UM, AT	No biopsy
RTEL1	rs1035926074	20	63689829	G	А	missense	24/35	R702H	Absent	Absent	D (1.0)	D (0.005)	26.0	Replication	White	Mold/Bird	No	6490	Possible	cfNSIP, PFGs
RTEL1	rs775121139	20	63690303	С	G	missense	26/35	P759A	1.66E-05	Absent	P (0.751)	D (0.021)	9.5	Replication	White	Unknown	No	6139	Incon: Mod fib, UM, GGO, AT	No biopsy
RTEL1	rs772872062	20	63691764	С	Т	missense	28/35	S860F	3.31E-05	Absent	P (0.848)	T (0.137)	19.8	Replication	White	Unknown	Yes	5273	Incon: Mild fib, GGO, AT, CLN	cfNSIP, PFGs
RTEL1	rs754727644	20	63692883	Α	Т	missense	23/35	N911Y	Absent	Absent	B (0.271)	D (0.013)	22.4	Discovery	White	Unknown	No	4839	Possible	fNSIP, OP, PFGs
RTEL1	rs753270617	20	63693161	G	А	missense	30/35	R957Q	8.28E-06	Absent	D (0.996	D (0.014)	25.9	Discovery	White	Down	No	5091	Incon: No fib, GGO, AT	cNSIP, PFGs
RTEL1	rs759564073	20	63693208	G	А	missense	30/35	G973R	8.29E-06	Absent	D (1.0)	D (0.003)	25.9	Replication	Unknown	Unknown	No	5478	Incon: Sev fib, PBV	UIP+LII+PFGs+BM
RTEL1	rs367598119	20	63695356	G	С	missense	34/35	K1176N	1.67E-05	Absent	P (0.501)	D (0.012)	17.0	Replication	White	Mold	No	6636	Incon: No fib, GGO, AT	cfNSIP, PFGs
DKC1	rs146700772	Х	154769233	Α	С	missense	9/15	S280R	2.80E-04	1.50E-04	P (0.519)	D (0.042)	23.3	Discovery	White	Unknown	No	5243	Incon: Mild fib, AT	cfNSIP, PFGs

Footnote, by column:

- 1. Gene: TERT = telomerase reverse transcriptase; TERC = telomerase RNA component; PARN = poly(A)-specific ribonuclease; RTEL1 = regulator of telomere elongation helicase 1; DKC1 = dyskerin pseudouridine synthase 1
- 2. dbSNP = database of short genetic variations identifiers, build 150, release July 10, 2017
- 3. CHR = chromosome number
- 4. Position = genomic position, human genome version 38
- 5. REF = reference genome allele
- 6. ALT = alternate allele
- 7. Effect = genomic effect
- 8. Exon = exon number/total exons
- 9. Protein = effect on protein
- 10. ExAC AF = Exome Aggregation Consortium allele frequency
- 11. ESP AF = Exome sequencing project allele frequency
- 12. PP2 = Polyphen-2 prediction of effect on protein function, format: D = probably damaging, P = possibly damaging, B = benign (hdiv score)

- 13. SIFT = Sorting Intolerant From Tolerant prediction of effect on protein function, format: D = damaging, T = tolerated (score)
- 14. CADD = Combined Annotation Dependent Depletion score; score 10 or greater indicates top 10% of most deleterious in genome and score 20 or greater indicates top 1%
- 15. Cohort: cohort in which variant was identified
- 16. Race/Ethnicity: self-reported race/ethnicity
- 17. Exposure: exposure clinically suspected to be causing hypersensitivity pneumonitis
- 18. FH = family history of interstitial lung disease
- 19. TL = telomere length in base pairs

20. HRCT features: high-resolution computed tomography features; abbreviations: Incon = inconsistent with usual interstitial pneumonia pattern; Possible = possible usual interstitial pneumonia pattern; Definite = definite usual interstitial pneumonia pattern; Mod = moderate; Sev = severe; fib = fibrosis; PBV = peribronchovascular distribution; GGO = ground glass opacities; UM = upper-mid lung distribution; AT = air-trapping; CLN = centrilobular nodules

21. Biopsy findings: c = cellular, f = fibrotic, cf = cellular and fibrotic, NSIP = non-specific interstitial pneumonia; PBF = peribronchiolocentric fibrosis; PFGs = poorly formed granulomas; PBI = peribronchiolar inflammation; OP = organizing pneumonia; LII = lymphocytic interstitial inflammation; NNGs = non-necrotizing granulomas; UIP = usual interstitial pneumonia; BCF = bronchiolocentric fibrosis; BM = bronchiolar metaplasia; SLB = surgical lung biopsy, Explant = explanted lung

Table E5. Phenotypes associated with qualifying variants

Gene	dbSNP	CHR	Position	REF	ALT	Protein	ClinVar (OMIM); Clinical Significance
TERT	rs370445231	5	1260593	G	Α	R951W	None
							diopathic fibrosing alveolitis, chronic form (178500); Uncertain Significance
TERT	rs775014633	5	1268538	G	Α	A855V	Dyskeratosis congenita, autosomal dominant, 2 (613989); Uncertain Significance
TERT	rs387907249	5	1278781	G	Α	A716T	Pulmonary fibrosis and/or bone marrow failure, telomere-related, 1 (614742); Pathogenic
TERT	rs866282352	5	1279329	G	Α	R698W	None
TERT	n/a	5	1280334	G	Т	H592N	
TERT	n/a	5	1282488	С	Α	K570N	
TERT	n/a	5	1282507	G	Α	T564M	
TERT	n/a	5	1293394	С	Т	G498R	
TERT	rs34094720	5	1293652	G	А	H412Y	Conflicting interpretations for pathogenicity from Benign to Pathogenic for: Idiopathic fibrosing alveolitis, chronic form; Aplastic Anemia; Dyskeratosis congenita, autosomal dominant, 2; Dyskeratosis congenita, autosomal recessive, 4; Pulmonary fibrosis and/or bone marrow failure, telomere-related, 1; Dyskeratosis Congenita, Recessive
TERT	n/a	5	1294393	Т	С	\$165G	
							diopathic fibrosing alveolitis, chronic form (178500); Pathogenic
TERT	rs1060502990	5	1294549	С	CG	112-113X	Dyskeratosis congenita, autosomal dominant, 2 (613989); Pathogenic
TERT	n/a	5	1294656	A	G	L77P	
TERT	n/a	5	1294829	С	Т	C54Y	
TERC	rs552679780	3	169765027	С	Т	n/a	None
							Dyskeratosis congenita, autosomal recessive 6 (616353); Uncertain Significance
PARN	rs377199187	16	14432695	С	G	R538P	Pulmonary fibrosis and/or bone marrow failure, telomere-related, 4 (616371); Uncertain Significance
PARN	n/a	16	14554142	Т	С	K443R	
PARN	rs774170618	16	14604184	G	A	R249C	None
PARN	n/a	16	14610777	G	Α	Q141*	
RTEL1	rs151214675	20	63661882	G	Α	A112T	None
RTEL1	n/a	20	63672587	G	Т	G244V	
RTEL1	n/a	20	63688042	Т	Α	F529L	
							Dyskeratosis congenita, autosomal recessive, 5 (615190); Uncertain Significance
RTEL1	rs1035926074	20	63689829	G	Α	R702H	Pulmonary fibrosis and/or bone marrow failure, telomere-related, 3 (616373); Uncertain Significance
RTEL1	rs775121139	20	63690303	С	G	P759A	None
RTEL1	rs772872062	20	63691764	С	Т	S860F	None
RTEL1	rs754727644	20	63692883	Α	Т	N911Y	None
RTEL1	rs753270617	20	63693161	G	Α	R957Q	None
RTEL1	rs759564073	20	63693208	G	Α	G973R	None
RTEL1	rs367598119	20	63695356	G	С	K1176N	None
DKC1	rs146700772	X	154769233	А	С	S280R	Dyskeratosis congenita X-linked (305000); Pathogenic

ClinVar database, searched on 04/26/2019.
 OMIM = Online Mendelian Inheritance in Man database number

Table E6. Comparison of clinical features by telomere variant status

			Discovery			Replication	on Cohort					
		All Patients			European Ancestry			All Patients	-		European Ancestry	
Characteristic	No Variant	Variant	p-value	No Variant	Variant	p-value	No Variant	Variant	p-value	No Variant	Variant	p-value
Ν	128	16		93	12		192	17		148	11	
Age in Years, mean (SD)	63.6 (11.0)	62.4 (13.0)	0.70	64.3 (10.7)	60.1 (12.5)	0.21	65.1 (10.3)	65.4 (10.7)	0.92	65.7 (9.6)	67.7 (9.3)	0.53
Male sex, n (%)	53 (41.4%)	6 (37.5%)	1.00	39 (42%)	5 (42%)	1.00	85 (44.3%)	6 (35.3%)	0.61	69 (46.6%)	6 (54.5%)	0.76
Race, n (%)			0.81			1.00			0.39			0.44
African American	3 (2.3%)	0 (0.0%)					9 (4.7%)	1 (5.9%)				
Asian	3 (2.3%)	1 (6.2%)					4 (2.1%)	0 (0.0%)				
European	107 (83.6%)	14 (87.5%)		90 (97%)	12 (100%)		157 (81.8%)	12 (70.6%)		141 (95.3%)	10 (90.9%)	
Hispanic	12 (9.4%)	1 (6.2%)		1 (1%)	0 (0%)		15 (7.8%)	3 (17.6%)		3 (2.0%)	0 (0.0%)	
Other or Unknown	3 (2.3%)	0 (0.0%)		2 (2%)	0 (0%)		7 (3.6%)	1 (5.9%)		4 (2.7%)	1 (9.1%)	
Family history, n (%)	7 (5.5%)	5 (31.2%)	0.004	6 (6%)	4 (33%)	0.015	12 (6.2%)	2 (11.8%)	0.32	11 (7.4%)	0 (0.0%)	1.00
Ever-smoker, n (%)	66 (53.7%)	7 (43.8%)	0.60	49 (54%)	6 (50%)	1.00	102 (53.1%)	7 (41.2%)	0.45	80 (54.1%)	6 (54.5%)	1.00
Known exposure, n (%)	90 (70.3%)	9 (56.2%)	0.26	67 (72%)	8 (67%)	0.74	132 (68.8%)	10 (58.8%)	0.42	99 (66.9%)	6 (54.5%)	0.51
Exposure type, n (%)			0.71			0.83			0.17			0.48
Avian	63 (49.2%)	6 (37.5%)		47 (51%)	5 (42%)		84 (43.8%)	4 (23.5%)		64 (43.2%)	3 (27.3%)	
Mold	23 (18.0%)	3 (18.8%)		16 (17%)	3 (25%)		41 (21.4%)	4 (23.5%)		28 (18.9%)	2 (18.2%)	
Other	4 (3.1%)	0 (0.0%)		4 (4%)	0 (0%)		7 (3.6%)	2 (11.8%)		7 (4.7%)	1 (9.1%)	
Unknown	38 (29.7%)	7 (43.8%)		26 (28%)	4 (33%)		60 (31.2%)	7 (41.2%)		49 (33.1%)	5 (45.5%)	
FVC % predicted, mean	68 1 (19 1)	61.0(17.0)	0.16	697(205)	54 9 (14 2)	0.017	66 2 (17 9)	57 5 (20 1)	0.057	68.0 (18.0)	58.9 (19.1)	0.11
DLCO % predicted, mean		(17.0)	0.10	40.0 (15.0)	11.0 (20.2)	0.51	50.0 (21.5)	10 7 (14 0)	0.057	(0.0 (01.5)	50.5 (19.1)	0.10
(SD)	47.6 (16.4)	43.4 (17.9)	0.34	48.0 (15.9)	44.8 (20.2)	0.54	59.8 (21.5)	49.7 (14.6)	0.077	60.0 (21.5)	54.4 (13.3)	0.42
MUC5B genotype, n (%)			0.34		0.447-040	0.31			0.15	/ ///		0.61
G/G	70 (58.3%)	11 (68.8%)		49 (56%)	8 (67%)		104 (54.5%)	10 (58.8%)		77 (52.4%)	6 (54.5%)	
G/T	46 (38.3%)	4 (25.0%)		36 (41%)	3 (25%)		81 (42.4%)	5 (29.4%)		64 (43.5%)	4 (36.4%)	
T/T	4 (3.3%)	1 (6.2%)		3 (3%)	1 (8%)		6 (3.1%)	2 (11.8%)		6 (4.1%)	1 (9.1%)	
G/T or T/T Telomere length in bp	50 (42.4%) 6139	5 (31.2%) 5447	0.43	39 (45%) 6048	4 (33%) 5447	0.54	87 (45.5%) 6090	7 (41.2%) 5415	0.80	70 (47.6%) 6081	5 (45.5%) 5478	1.00
median (IQR)	(5639-6573)	(4996-5989)	0.004	(5615-6607)	(5141-5940)	0.012	(5716-6377)	(5010-5788)	< 0.001	(5716-6371)	(4763-6139)	0.007

Table E7. Comparison of radiologic features by telomere variant status

			Discov			Rep	lication							
		Α	Il Patients]	European			А		European			
Factor	Total	No Variant	Variant	p- value	No Variant	Variant	p- value	Total	No Variant	Variant	p- value	No Variant	Variant	p-value
Number with CT scored	134	119	15	n/a	86	11	n/a	204	187	17		143	11	
UIP pattern, n (%)				0.16			0.32				0.74			0.61
Definite	5 (3.7%)	4 (3.4%)	1 (6.7%)		3 (3%)	1 (9%)		13 (6.4%)	12 (6.5%)	1 (5.9%)		7 (4.9%)	1 (9.1%)	
Possible	12 (9.0%)	9 (7.6%)	3 (20.0%)		5 (6%) 78	1 (9%)		16 (7.9%)	14 (7.6%)	2 (11.8%)		13 (9.2%)	1 (9.1%)	
Inconsistent	117 (87.3%)	106 (89.1%)	11 (73.3%)		(91%)	9 (82%)		173 (85.6%)	159 (85.9%)	14 (82.4%)		122 (85.9%)	9 (81.8%)	
Definite/Possible	17 (12.7%)	13 (10.9%)	4 (26.7%)	0.100	8 (9%)	2 (18%)	0.32	29 (14.4%)	26 (14.1%)	3 (17.6%)	0.72	20 (14.1%)	2 (18.2%)	0.66
<u>Fibrosis, n (%)</u>				0.90	15		1.00				0.43			0.20
None	24 (17.9%)	21 (17.6%)	3 (20.0%)		(17%) 30	2 (18%)		30 (14.9%)	27 (14.6%)	3 (17.6%)		22 (15.5%)	3 (27.3%)	
Mild	48 (35.8%)	42 (35.3%)	6 (40.0%)		(35%) 35	4 (36%)		39 (19.3%)	38 (20.5%)	1 (5.9%)		28 (19.7%)	0 (0.0%)	
Moderate	54 (40.3%)	49 (41.2%)	5 (33.3%)		(41%)	4 (36%)		110 (54.5%)	100 (54.1%)	10 (58.8%)		77 (54.2%)	6 (54.5%)	
Severe	8 (6.0%)	7 (5.9%)	1 (6.7%)		6 (7%) 41	1 (9%)		23 (11.4%)	20 (10.8%)	3 (17.6%)		15 (10.6%)	2 (18.2%)	
Moderate/Severe	62 (43.1%)	56 (43.8%)	6 (37.5%)	0.79	(44%)	5 (42%)	1.00	133 (65.8%)	120 (64.9%)	13 (76.5%)	0.43	92 (64.8%)	8 (72.7%)	0.75
Honeycombing, n (%)	22 (16.4%)	18 (15.1%)	4 (26.7%)	0.27	16 (19%)	3 (27%)	0.45	40 (19.8%)	36 (19.5%)	4 (23.5%)	0.75	25 (17.6%)	3 (27.3%)	0.42
Traction bronchiectasis, n (%)	97 (72.4%)	87 (73.1%)	10 (66.7%)	0.56	64 (74%)	8 (73%)	1.00	158 (78.2%)	144 (77.8%)	14 (82.4%)	1.00	110 (77.5%)	8 (72.7%)	0.71
Upper-middle lung distribution, n (%)	47 (35.1%)	43 (36.1%)	4 (26.7%)	0.57	34 (40%)	3 (27%)	0.52	56 (32.0%)	51 (31.7%)	5 (35.7%)	0.77	35 (28.7%)	4 (50.0%)	0.24
Peribronchovacular distribution, n (%)	69 (51.5%)	62 (52.1%)	7 (46.7%)	0.79	43 (50%)	6 (55%)	1.00	93 (53.1%)	87 (54.0%)	6 (42.9%)	0.58	67 (54.9%)	3 (37.5%)	0.47
Ground glass opacities, n (%)	61 (45.5%)	57 (47.9%)	4 (26.7%)	0.17	37 (43%)	3 (27%)	0.52	108 (53.5%)	101 (54.6%)	7 (41.2%)	0.32	80 (56.3%)	5 (45.5%)	0.54
trapping, n (%)	76 (56.7%)	69 (58.0%)	7 (46.7%)	0.42	55 (64%)	6 (55%)	0.53	129 (63.9%)	117 (63.2%)	12 (70.6%)	0.61	94 (66.2%)	7 (63.6%)	1.00

Table E8. Comparison of pathologic features by telomere variant status

		All			European		
Feature	Total	No Variant	Variant	p-value	No Variant	Variant	p-value*
N	110	98	12		78	8	
Microscopic honeycombing	24 (22.0%)	20 (21%)	4 (33%)	0.30	16 (21%)	3 (38%)	0.37
Heterogenous distribution	75 (68.2%)	65 (66%)	10 (83%)	0.33	52 (67%)	8 (100%)	0.099
Subpleural accentuation	28 (25.9%)	23 (24%)	5 (42%)	0.29	19 (25%)	4 (50%)	0.21
Airway centered	80 (73.4%)	70 (72%)	10 (83%)	0.51	55 (71%)	7 (88%)	0.44
Dense collagen fibrosis				0.66			0.92
None	26 (23.6%)	24 (24%)	2 (17%)		19 (24%)	1 (12%)	
Mild	36 (32.7%)	33 (34%)	3 (25%)		28 (36%)	3 (38%)	
Moderate	20 (18.2%)	18 (18%)	2 (17%)		12 (15%)	1 (12%)	
Marked	28 (25.5%)	23 (23%)	5 (42%)		19 (24%)	3 (38%)	
Any	84 (76.4%)	74 (76%)	10 (83%)	0.73	59 (76%)	7 (88%)	0.67
Moderate-Marked	48 (43.6%)	41 (42%)	7 (58%)	0.36	31 (40%)	4 (50%)	0.71
Interstitial granulomas				0.97			0.84
None	32 (29.1%)	29 (30%)	3 (25%)		21 (27%)	1 (12%)	
Mild	19 (17.3%)	17 (17%)	2 (17%)		12 (15%)	1 (12%)	
Moderate	29 (26.4%)	26 (27%)	3 (25%)		23 (29%)	3 (38%)	
Marked	30 (27.3%)	26 (27%)	4 (33%)		22 (28%)	3 (38%)	
Any	78 (70.9%)	69 (70%)	9 (75%)	1.00	57 (73%)	7 (88%)	0.67
Moderate-Marked	59 (53.6%)	52 (53%)	7 (58%)	0.77	45 (58%)	6 (75%)	0.46
Lymphocytic infiltration				0.96			1.00
None	16 (14.8%)	15 (16%)	1 (8%)		9 (12%)	1 (12%)	
Mild	34 (31.5%)	30 (31%)	4 (33%)		24 (32%)	3 (38%)	
Moderate	52 (48.1%)	45 (47%)	7 (58%)		37 (49%)	4 (50%)	
Marked	6 (5.6%)	6 (6%)	0 (0%)		6 (8%)	0 (0%)	
Any	92 (85.2%)	81 (84%)	11 (92%)	0.69	67 (88%)	7 (88%)	1.00
Moderate-Marked	58 (53.7%)	51 (53%)	7 (58%)	0.77	43 (57%)	4 (50%)	0.73
Fibroblastic foci				0.004			0.019
None	69 (63.3%)	65 (67%)	4 (33%)		54 (70%)	3 (38%)	
Mild	31 (28.4%)	27 (28%)	4 (33%)		21 (27%)	3 (38%)	
Moderate	8 (7.3%)	5 (5%)	3 (25%)		2 (3%)	1 (12%)	
Marked	1 (0.9%)	0 (0%)	1 (8%)		0 (0%)	1 (12%)	
Any	40 (36.7%)	32 (33%)	8 (67%)	0.029	23 (30%)	5 (62%)	0.11
Moderate-Marked	9 (8.3%)	5 (5%)	4 (33%)	0.008	2 (3%)	2 (25%)	0.043
Small airway disease				0.75			0.22
None	15 (14.0%)	14 (15%)	1 (9%)		11 (14%)	1 (12%)	
Mild	50 (46.7%)	43 (45%)	7 (64%)		37 (49%)	6 (75%)	
Moderate	32 (29.9%)	30 (31%)	2 (18%)		22 (29%)	0 (0%)	
Marked	10 (9.3%)	9 (9%)	1 (9%)		6 (8%)	1 (12%)	
Any	92 (86.0%)	82 (85%)	10 (91%)	1.00	65 (86%)	7 (88%)	1.00
Moderate-Marked	42 (39.3%)	39 (41%)	3 (27%)	0.52	28 (37%)	1 (12%)	0.25

*Trends in associations between variant status and semi-quantitative categories were evaluated using a multinomial logistic regression model.

Table E9. Coverage statistics for candidate genes

	Discovery Cohort		Replication Cohort			
GENE	Median	Minimum	Median	Minimum		
DKC1	28	8	35	29		
PARN	28	20	29	25		
RTEL1	37	23	35	30		
TERC	32	22	51	15		
TERT	36	23	40	30		
TINF2	29	20	44	31		

<u>Footnote</u>:
1. Median = median of median coverage per exon across samples in read depth
2. Minimum = median of minimum coverage per exon across samples in read depth

Table E10. Comparison of variant inclusion criteria and variant frequencies between the current study and prior studies of telomere-related gene variants in idiopathic pulmonary fibrosis and rheumatoid arthritis-associated interstitial lung disease

Study	Genes included	MAF cut-off	Predicted effects	Frequency in Cases from Reference	Frequency in Controls from	Frequency in non- Hispanic White	Frequency in non- Hispanic White
				Study	Reference Study	Cases from	Cases from
						Discovery Cohort in	Replication Cohort in
						Current Study	Current Study
Ley et al	TERT, TERC,	0.005	1. LoF	n/a	n/a	n/a	n/a
(current study)	PARN, RTEL1,		2. PP2 – at least possibly				
	TINF2, DKC1		damaging				
			3. SIFT- deleterious				
Petrovski et al	Whole Exome	0.0005	1. LoF	TERT = 0.0496	TERT = 0.0014	TERT = 0.0248	TERT = 0.0237
(Reference 29)		cases/controls	2. PP2- at least probably	RTEL1 = 0.0229	RTEL1 = 0.0	RTEL1 = 0.0083	RTEL1 = 0.0059
		AND absent in	damaging	PARN = 0.0267	PARN = 0.0012	PARN = 0.0	PARN = 0.0059
		Exome Variant		Combined = 9.9%	Combined = 0.26%	Combined = 3.3%	Combined = 3.6%
		Server and ExAC					
Juge et al	ABCA3, PARN,	0.01	1. PP2 probably	TERT = 0.0741	TERT = 0.0168	TERT = 0.0413	TERT = 0.0355
(Reference 31)	RTEL1,		damaging or SIFT	RTEL1 = 0.0370	RTEL1 = 0.0139	RTEL1 = 0.0413	RTEL1 = 0.0296
	SFTPA2,		deleterious	PARN = 0.123	PARN = 0.0069	PARN = 0.0083	PARN = 0.0118
	SFTPC, TERT		2. GERP >=2	Combined = 12.3%	Combined = 3.8%	Combined = 8.3%	Combined = 7.7%
			3. CADD >=10				
Dressen et al	Whole	0.01	PP2 >=0.5 OR high-	TERT = 0.0424	TERT = 0.01	TERT = 0.0496	TERT = 0.0355
(Reference 30)	Genome		impact	RTEL1 = 0.0305	RTEL1 = 0.0176	RTEL1 = 0.0331	RTEL1 = 0.0355
				PARN = 0.0124	PARN = 0.001	PARN = 0.0165	PARN = 0.0118
				Combined = 8.5%	Combined = 2.9%	Combined = 9.9%	Combined = 8.3%

Abbreviations: MAF = minor allele frequency, ExAC = Exome Aggregation Consortium, LoF = loss of function, PP2 = polyphen-2, SIFT = Sorting Intolerant from Tolerant, GERP = Genomic Evolutionary Rate Profiling score, CADD = Combined Annotation Dependent Depletion score