



Supporting Information

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DNA-Encoded Libraries: Aryl Fluorosulfonates as Versatile Electrophiles Enabling Facile On-DNA Suzuki, Sonogashira, and Buchwald Reactions

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Electronic Supporting Information

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Hongtao Xu ^{a#}, Fei Ma ^{a#}, Nan Wang ^a, Wei Hou ^b, Huan Xiong ^a, Fengping Lu ^a, Jie Li ^a, Shuyue Wang ^a, Peixiang Ma ^a, Guang Yang ^{a,*}, Richard A. Lerner ^{c*}

^aShanghai Institute for Advanced Immunochemical Studies, ShanghaiTech University, 201210 Shanghai, China

^bCollege of Pharmaceutical Science, and Institute of Drug Development & Chemical Biology (IDD & CB), Zhejiang University of Technology, 310014 Hangzhou, China.

^cDepartment of Chemistry, Scripps Research Institute, La Jolla, CA 92037

Contributed equally

* To whom correspondence may be addressed.

Email: yangguang@shanghaitech.edu.cn, or rlerner@scripps.edu.

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General Methods

All reagents and DNA headpiece HP-NH₂ (5'- / 5phos / GAGTCA / iSp9 / iUniAmM / iSp9 / TGACTCCC-3', Figure 1) were obtained from commercial sources unless

otherwise noted and used as received. All on-DNA reactions were performed in 1.5 mL or 5 mL Eppendorf tubes. On-DNA reactions in the studies of reaction condition optimization and substrate scope extension were analyzed by LC-MS. Typically, samples were dissolved in an appropriate amount of distilled and deionized water (ddH₂O) and injected into a reverse-phase chromatography column (Xbridge Oligonucleotide BEH C18 column, 1.7 μm, 2.1×50 mm). The elution was carried out as followings: 5–95% solvent B over 4.5 min, 0.4 mL/min, λ = 260 nm; solvent A: 0.75% v/v hexafluoroisopropanol/ 0.038% v/v triethylamine in methanol/water = 5/95; solvent B: 0.75% v/v hexafluoroisopropanol/ 0.038% v/v triethylamine in methanol/water = 90/10. The effluents were analyzed by an ABSCIEX 4600 TOF electrospray mass spectrometer in negative ion mode. Mass deconvolution was carried out using Peakview software. Percent conversion for DNA conjugated reactions were based on the areas of TIC peaks of LC-MS.

The Structure of DNA Headpiece

DNA headpiece HP-NH₂(5'/5Phos/GAGTCA/iSp9/iUniAmM/iSp9/TGACTCCC-3')

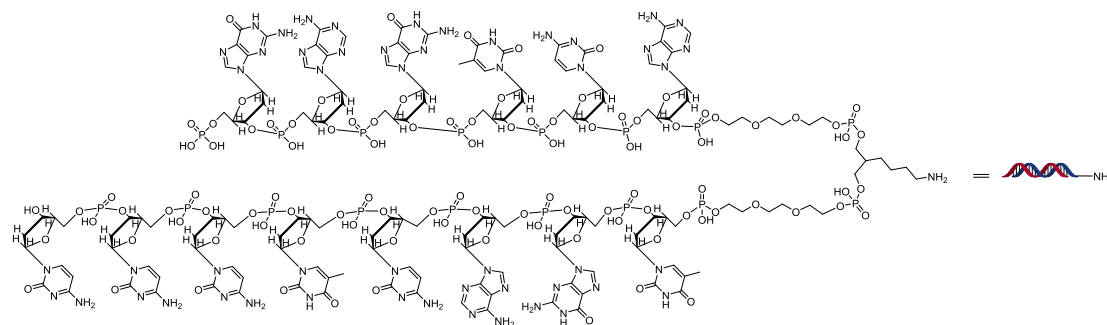
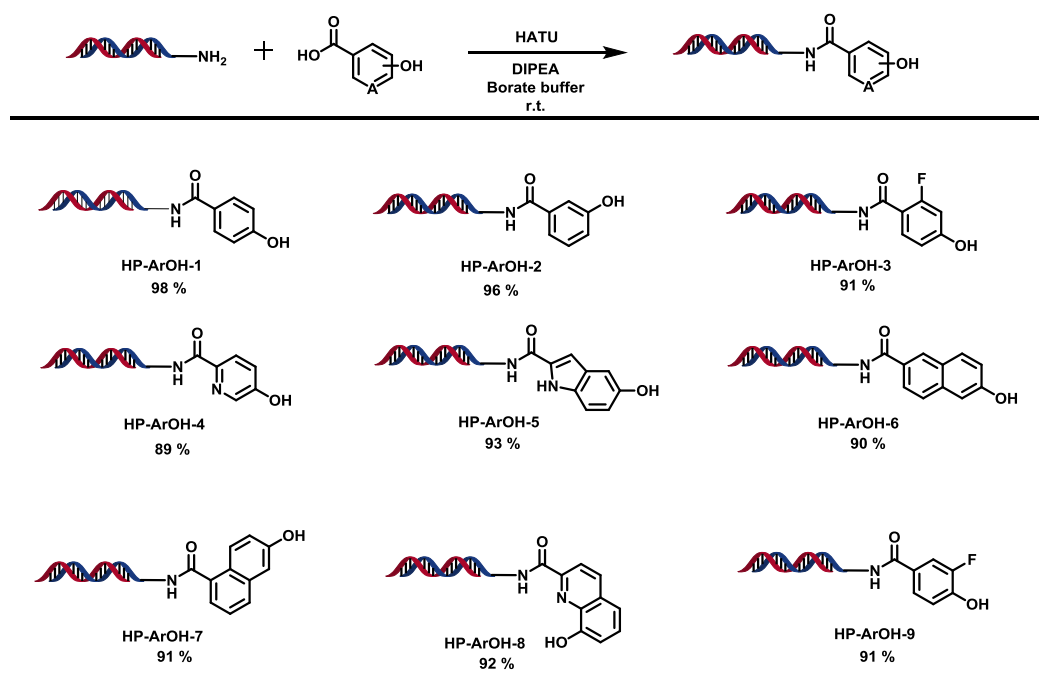


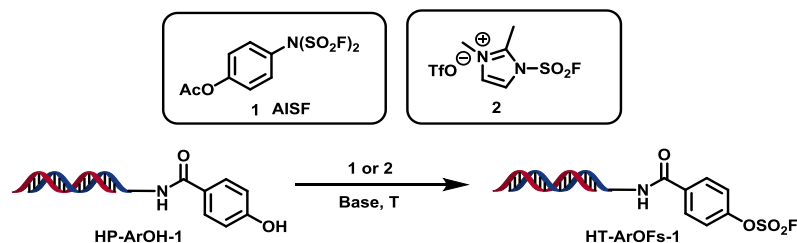
Figure 1. The Structure of DNA Headpiece

Scheme S1. Synthesis of DNA conjugated phenols



To a solution of DNA headpiece (600 μ L, 300 nmol) in borate buffer (250 mM, pH = 9.4), was added a mixture of DMA solution of HATU (60 μ L, 200 mM), DIPEA (60 μ L, 200 mM) and acids (60 μ L, 200 mM). The resultant mixture was vortexed and stood at 25 $^{\circ}$ C for 8 hours. 5M NaCl (78 μ L) and cold ethanol (2.34 mL) were sequentially added, and the resultant mixture was stored at -80 $^{\circ}$ C for 30 min. The mixture was centrifuged at 4 $^{\circ}$ C for 30 min at 12000 rpm to remove the supernatant. The resulting pellet was re-dissolved in ddH₂O (300 μ L), which was used in following reaction without further purification.

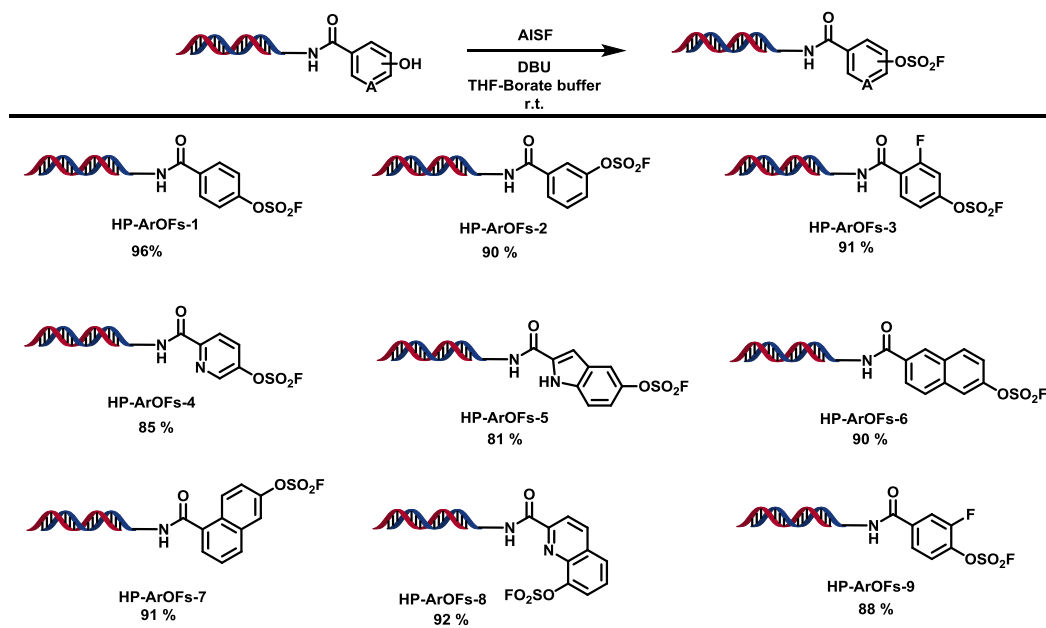
Table S1. Optimization of reaction conditions for HP-DNA-Aryl fluorosulfonates synthesis^a



Entry	SO ₂ F	base	T(h)	Conversion (%) ^b
1	1	DBU	0.5	54
2	1	DBU	1	73
3	1	DBU	1.5	86
4	1	DBU	2	96
5	1	DBU	4	90
6	2	Et ₃ N	0.5	71
7	2	Et ₃ N	1	86
8	2	Et ₃ N	1.5	84
9	2	Et ₃ N	2	82
10	2	Et ₃ N	4	80

^aReaction conditions: 1 equiv of **HP-Ar-OH-1** (1 mM in borate buffer), 100 equiv of **1** (100 mM in THF) or **2** (100 mM in CH₃CN), 200 equiv of DBU (200 mM in THF) or Et₃N (200 mM in CH₃CN), borate buffer (pH 9.4); ^b Conversion of **HP-Ar-OH-1** determined by LC-MS.

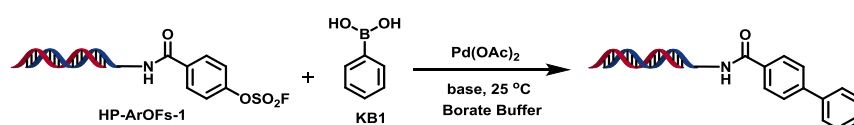
Scheme S2. Preparation of DNA conjugated (hetero)aryl fluorosulfonates



To the DNA conjugated phenols (300 μ L, 1 mM), were added sequentially 300 μ L AISF (100 mM), 300 μ L borate buffer (250 mM, pH = 9.4), and 300 μ L DBU (200

mM). The resultant mixture was vortexed and stood at 25 °C for 8 hours. 5.0M NaCl (120 μ L) and cold neat ethanol (3.6 mL) were sequentially added, and the resultant mixture was incubated at -80 °C for at least 30 min. The mixture was centrifuged at 4 °C for 30 min at 12,000 rpm to remove the supernatant. The resulting pellet was re-dissolved in ddH₂O (300 μ L), which was used in following reaction without further purification.

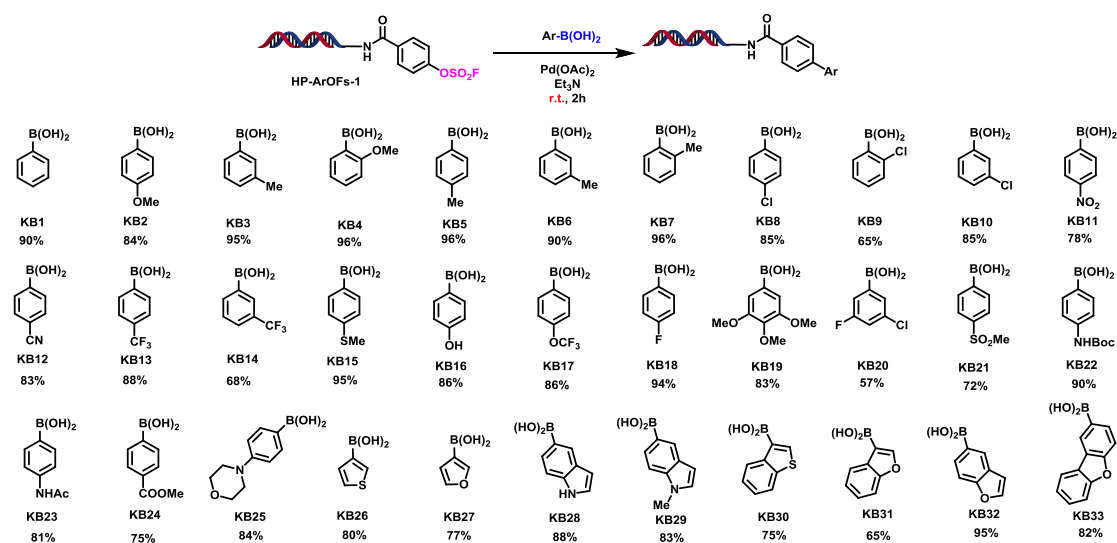
Table S2. Optimization of on-DNA Suzuki-Miyaura coupling reaction conditions^a



Entry	Base	Conversion (%) ^b
1	Et ₃ N	90
2	DIPEA	10
3	Na ₂ CO ₃	0
4	K ₂ CO ₃	0
5	NaOH	0

^aReaction conditions: 1 equiv of **HP-ArOFs-1** (1 mM in borate buffer), 400 equiv of **KB1** (400 mM in 1,4-dioxane), 20 equiv of Pd(OAc)₂ (20 mM in 1,4-dioxane), base (1000 mM in 1,4-dioxane), water; ^b Conversion of **HP-ArOFs-1** was determined by LC-MS.

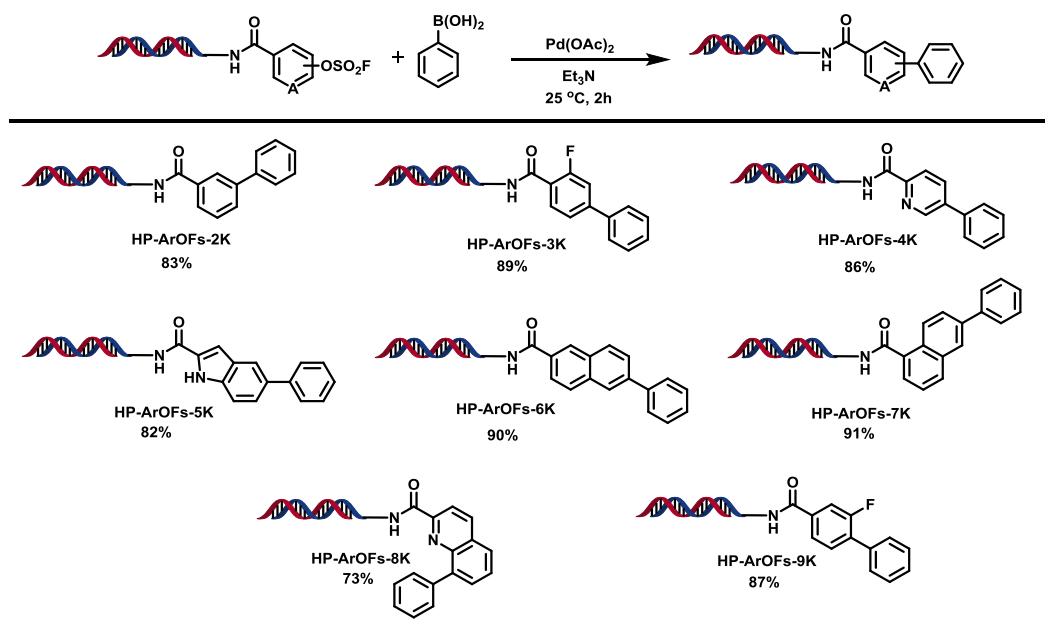
General Procedures for on-DNA Suzuki-Miyaura Coupling of Representative Boronic Acids with HP-ArOFs-1



To **HP-ArOFs-1** (10 nmol, 10 μ L, 1 mM in water), was added 400 equiv. of aromatic

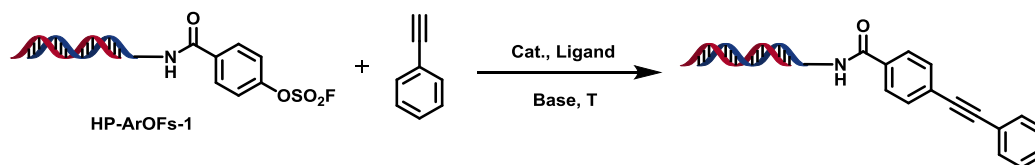
boronic acids (10 μ L, 400 mM in 1,4-dioxane) followed by 1000 equiv. of Et₃N (2 μ L, 5 M in 1,4-dioxane), 20 equiv. of Pd(OAc)₂ (10 μ L, 20 mM in 1,4-dioxane) and 20 μ L H₂O. The mixture was vortexed and stood at 25 °C for 2 hours. After reaction, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with Pd(OAc)₂, 6 μ L 1 M in ddH₂O) were added to the mixture, and the reaction mixture was heated at 60 °C for at least 30 minutes. The mixture was centrifuged at 4 °C for 30 min at 12,000 rpm, and the resultant supernatant was collected. Add 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20 °C) to the resultant supernatant. The mixture was vortexed and incubated at -80 °C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4°C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μ L) for LC-MS detection.

Scheme S3. Reactions of DNA conjugated (hetero) aryl fluorosulfonates with phenylboronic acid



To the DNA conjugated (hetero) aryl fluorosulfates (10 nmol, 10 μ L, 1 mM in ddH₂O), were added 400 equiv. of phenylboronic acid (10 μ L, 400 mM in 1,4-dioxane) followed by 1000 equiv. of Et₃N (2 μ L, 5 M in 1,4-dioxane), 20 equiv.

of Pd(OAc)₂ (10 μL, 20 mM in 1,4-dioxane), and 20 μL H₂O. The mixture was vortexed and stood at 25 °C for 2 hours. After reaction, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with Pd(OAc)₂, 6 μL 1000 mM in water) were added to the mixture, and the reaction mixture was heated at 60 °C for at least 30 minutes. The mixture was centrifuged at 4 °C for 30 minutes at 12,000 rpm, and the resultant supernatant was collected. Add 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20 °C) to the resultant supernatant. The mixture was vortexed and incubated at -80 °C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4°C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μL) for LC-MS detection.

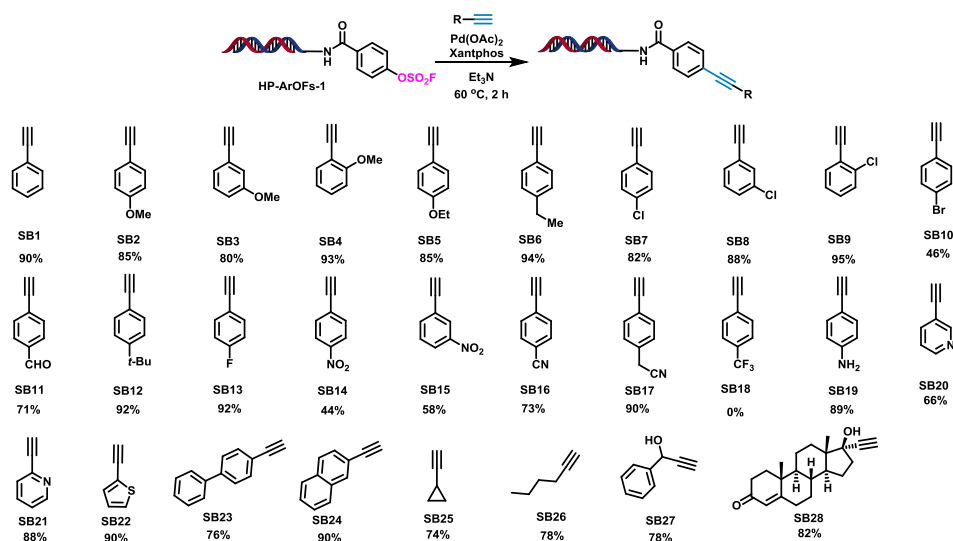
Table S3. Optimization of on-DNA Sonogoshira reaction conditions^a

Entry	Cat.	Ligand	Base	Conversion(%) ^b
1	Pd(OAc) ₂	Xphos	Et ₃ N	0
2	Pd(OAc) ₂	dppp	Et ₃ N	0
3	Pd(OAc) ₂	Dppf	Et ₃ N	0
4	Pd(OAc) ₂	P(t-Bu) ₃	Et ₃ N	0
5	Pd(OAc) ₂	PPh ₃	Et ₃ N	0
6	Pd(OAc) ₂	TPPS	Et ₃ N	0
7	Pd(OAc) ₂	Xantphos	Et ₃ N	77
8	Pd(PPh ₃) ₄	Xantphos	Et ₃ N	68
9	Pd ₂ (dba) ₃	Xantphos	Et ₃ N	48
10	Pd(PPh ₃) ₂ Cl ₂	Xantphos	Et ₃ N	46
11	[PdCl(C ₃ H ₅) ₂]	Xantphos	Et ₃ N	72
12	Pd(OAc) ₂	Xantphos	DBU	0
13	Pd(OAc) ₂	Xantphos	DIPEA	56
14	Pd(OAc) ₂	Xantphos	DABCO	72
15	Pd(OAc) ₂	Xantphos	NaHCO ₃	67
16	Pd(OAc) ₂	Xantphos	Na ₂ CO ₃	43
17	Pd(OAc) ₂	Xantphos	K ₂ CO ₃	32
18	Pd(OAc) ₂	Xantphos	Cs ₂ CO ₃	64
19	Pd(OAc) ₂	Xantphos	\	62
20 ^c	Pd(OAc) ₂	Xantphos	Et ₃ N	0
21 ^d	Pd(OAc) ₂	Xantphos	Et ₃ N	41
22 ^e	Pd(OAc) ₂	Xantphos	Et ₃ N	69
23 ^f	Pd(OAc)₂	Xantphos	Et₃N	85

^aReaction conditions: 1 equiv of **HP-ArOFs-1** (1 mM in ddH₂O), 200 equiv of 1-ethynyl-4-methoxybenzene (200 mM in DMA), 10 equiv of Cat. (10 mM in DMA), 20 equiv of ligand (20 mM in DMA), 1000 equiv of base (5 M in DMA), 60 °C, 2h; ^bConversion

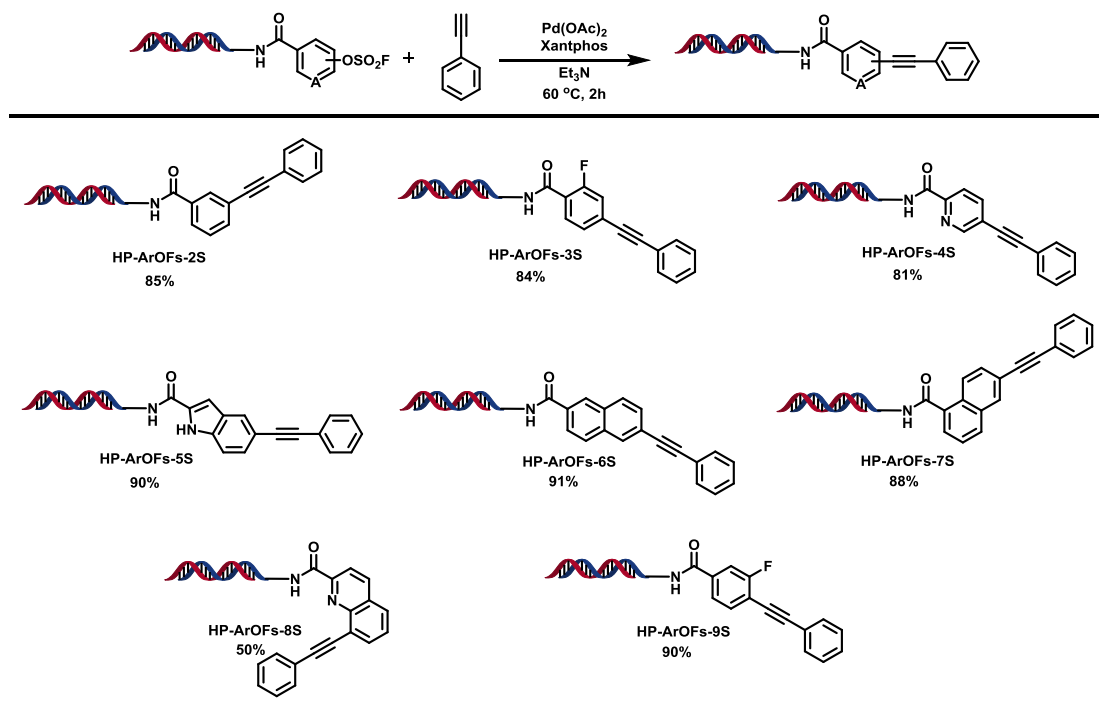
of **HP-ArOFs-1** was determined by LC-MS; ^c 25 °C was used; ^d 40 °C was used; ^e 80 °C was used; ^f 20 equiv of Pd(OAc)₂ (20 mM in DMA), 40 equiv of Xantphos (40 mM in DMA), 1000 equiv of Et₃N (5 M in DMA).

General Procedures for on-DNA Sonogashira Coupling of Representative Alkynes and HP-ArOFs-1

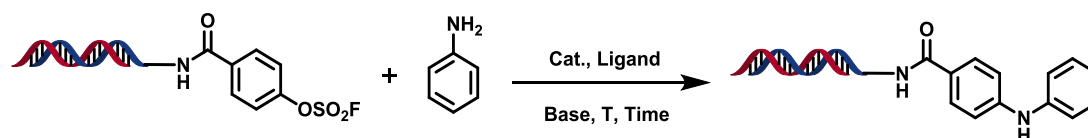


Pd(OAc)₂ (10 μL, 20 mM in DMA), Xantphos (10 μL, 40 mM in DMA) and alkynes (10 μL, 200 mM in DMA) were mixed to generate 30 μL 0.67 mM Pd / Xantphos / alkynes complex. To **HP-ArOFs-1** (10 nmol, 10 μL, 1 mM in ddH₂O), were added 1000 equiv. of Et₃N (2 μL, 5 M in DMA) and the above 30 μL 0.67 mM Pd / Xantphos / alkynes complex. The resulting mixture was vortexed and stood at 60 °C for 2 hours. After incubation, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with Pd(OAc)₂, 6 μL 1 M in ddH₂O) were added to the mixture, and heated at 60 °C for at least 30 minutes. The reaction mixture was next centrifuged at 4 °C for 30 min at 12,000 rpm, and the resultant supernatant was collected. To the supernatant, 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20°C) were added, vortexed, and incubated at -80°C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4°C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μL) for LC-MS detection.

Scheme S4. Reactions of DNA conjugated (hetero) aryl fluorosulfonates with 1-ethynyl-benzene



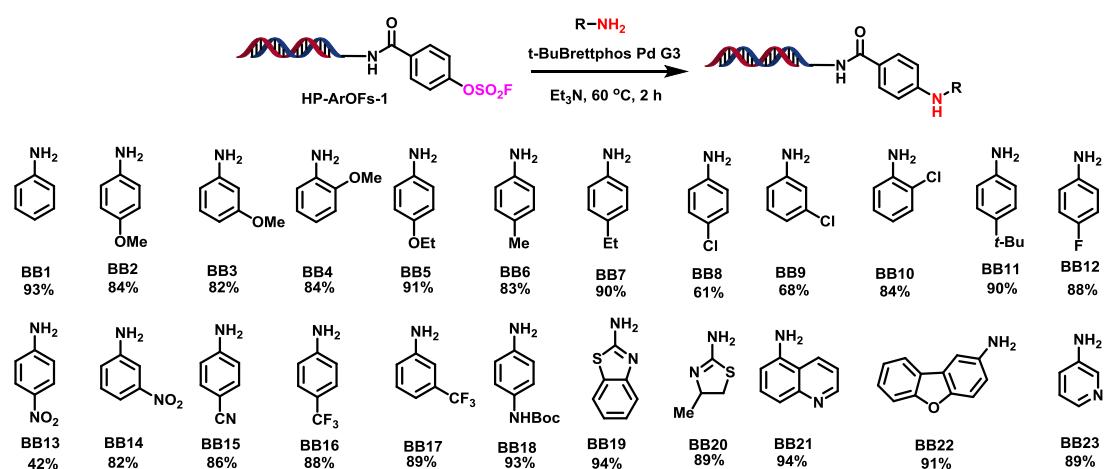
Pd(OAc)₂ (10 μL, 20 mM in DMA), Xantphos (10 μL, 40 mM in DMA) and Phenylacetylene (10 μL, 200 mM in DMA) were mixed to generate 30 μL 0.67 mM Pd / Xantphos / Phenylacetylene complex. To the DNA conjugated (hetero) aryl fluorosulfates (10 nmol, 10 μL, 1 mM in ddH₂O), was added 1000 equiv. of Et₃N (2 μL, 5 M in DMA) and the above 30μL 0.67 mM Pd / Xantphos / alkynes complex. The mixture was vortexed and stood at 60 °C for 2 hours. After incubation, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with Pd(OAc)₂, 6 μL 100 mM in ddH₂O) were added to the mixture, and heated at 60 °C for at least 30 minutes. The reaction mixture was centrifuged at 4 °C for 30 min at 12,000 rpm, and the resultant supernatant was collected. 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20°C) were added to the resultant supernatant. The resulting mixture was vortexed and incubated at -80°C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4°C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μL) for LC-MS detection.

Table S4. Optimization of on-DNA Buchwald reaction conditions^a

entry	Cat	ligand	Base	Conversion (%) ^b
1	Pd(OAc) ₂	Xphos	Et ₃ N	0
2	Pd(OAc) ₂	dppp	Et ₃ N	0
3	Pd(OAc) ₂	Dppf	Et ₃ N	0
4	Pd(OAc) ₂	P(t-Bu) ₃	Et ₃ N	0
5	Pd(OAc) ₂	PPh ₃	Et ₃ N	0
6	Pd(OAc) ₂	TPPS	Et ₃ N	0
7	Pd(OAc) ₂	Xantphos	Et ₃ N	0
8	Pd(PPh ₃) ₄	BINAP	Et ₃ N	0
9	Brettphos	\	Et ₃ N	89
10	t-BuBrettphos	\	Et ₃ N	91
11^c	t-BuBrettphos	\	Et ₃ N	92
12^d	t-BuBrettphos	\	Et₃N	93
13^e	t-BuBrettphos	\	Et ₃ N	87
14^f	t-BuBrettphos	\	Et ₃ N	83

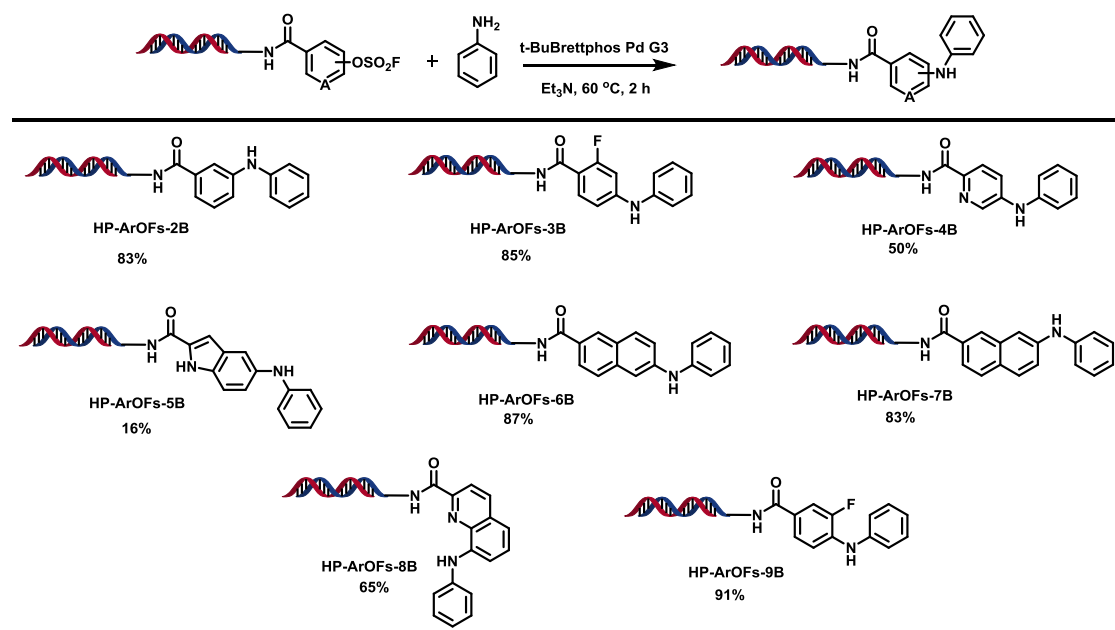
^a Reaction conditions: 1 equiv of **HP-ArOFs-1** (1 mM in water), 200 equiv of aniline (200 mM in DMA), 20 equiv of Cat. (10 mM in DMA), 40 equiv of ligand (20 mM in DMA), 1000 equiv of base (5000 mM in DMA), 60 °C, 2 hours; ^b Conversion of **HP-ArOFs-1** was determined by LC-MS; ^c 15 equiv of t-Brettphos Pd G3; ^d 10 equiv of t-Brettphos Pd G3; ^e 5 equiv of t-Brettphos Pd G3; ^f 3 equiv of t-Brettphos Pd G3.

General Procedures for on-DNA Buchwald Amination of Representative Amines with HP-ArOFs-1



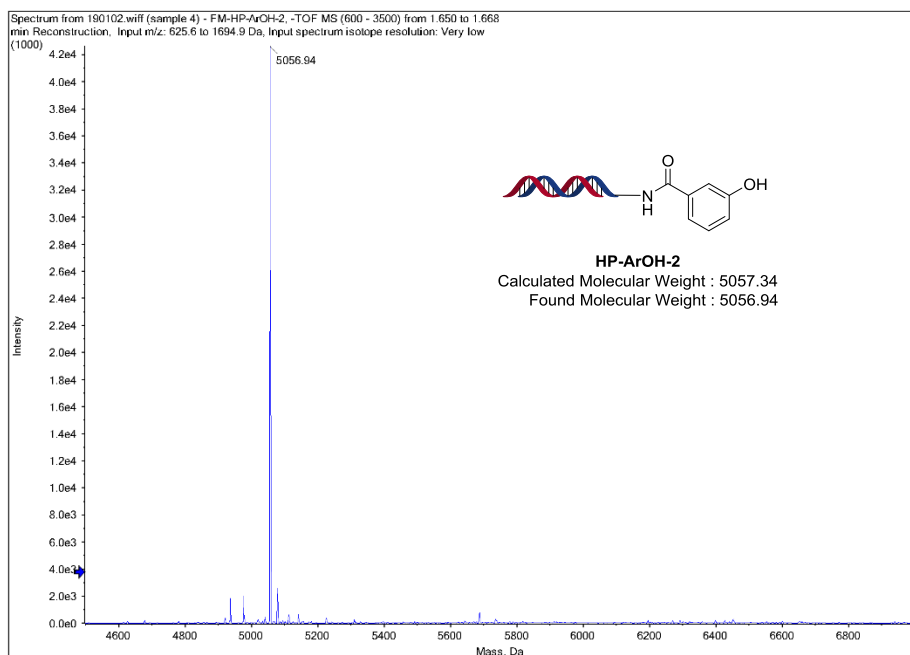
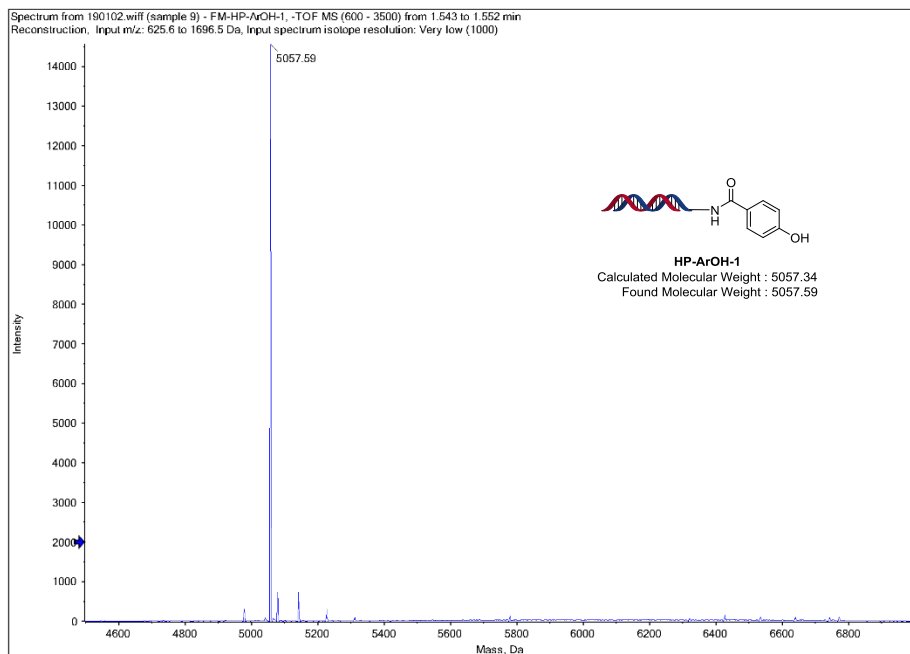
To **HP-ArOFs-1** (10 nmol, 10 μ L, 1 mM in ddH₂O), was added 200 equiv. of aromatic amines (10 μ L, 200 mM in DMA), followed by 1000 equiv. of Et₃N (2 μ L, 5 M in DMA) and 10 equiv. of t-BuBrettphos Pd G3 (10 μ L, 10 mM in DMA). The mixture was vortexed and stood at 60 °C for 2 hours. After the reaction, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with Pd(OAc)₂, 6 μ L 1 M in ddH₂O) were added to the mixture, and the resulting mixture was heated at 60 °C for at least 30 minutes followed by centrifugation at 4 °C for 30 min at 12,000 rpm. The resultant supernatant was collected, mixed with 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20 °C), vortexed, and incubated at -80°C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4°C in a micro centrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μ L) for LC-MS detection.

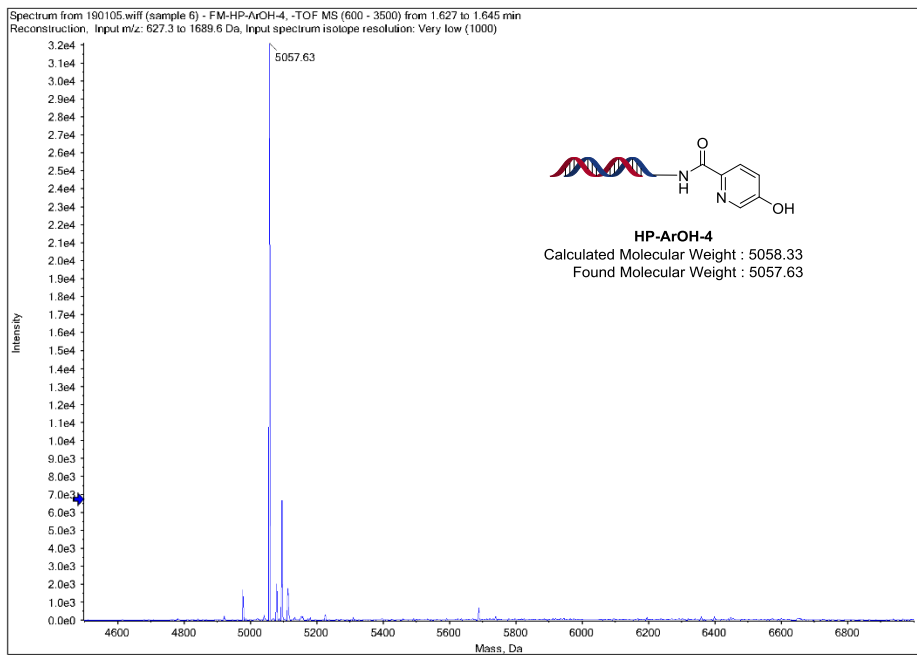
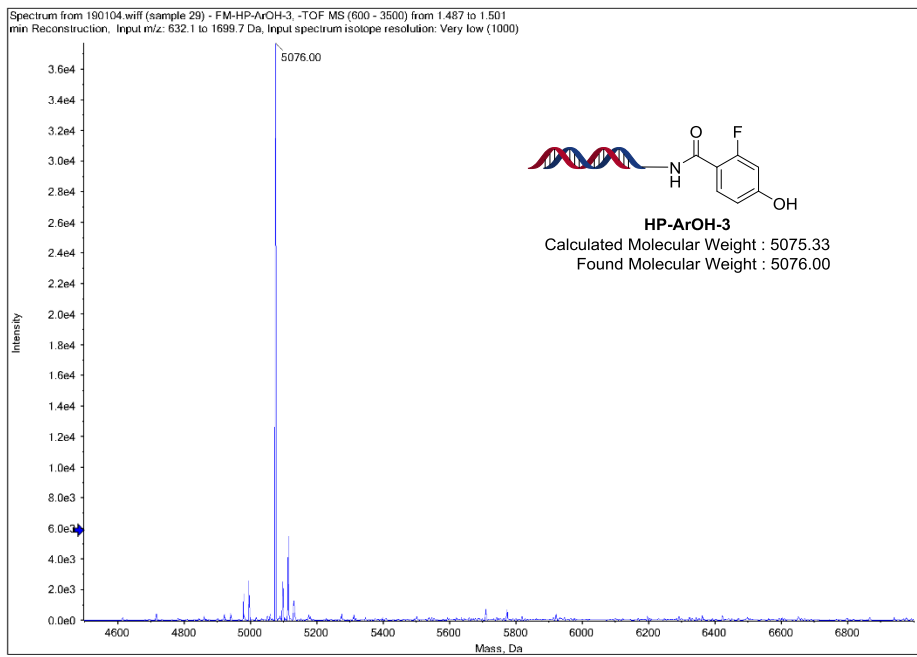
Scheme S5. Reactions of DNA conjugated (hetero) aryl fluorosulfonates with anilines

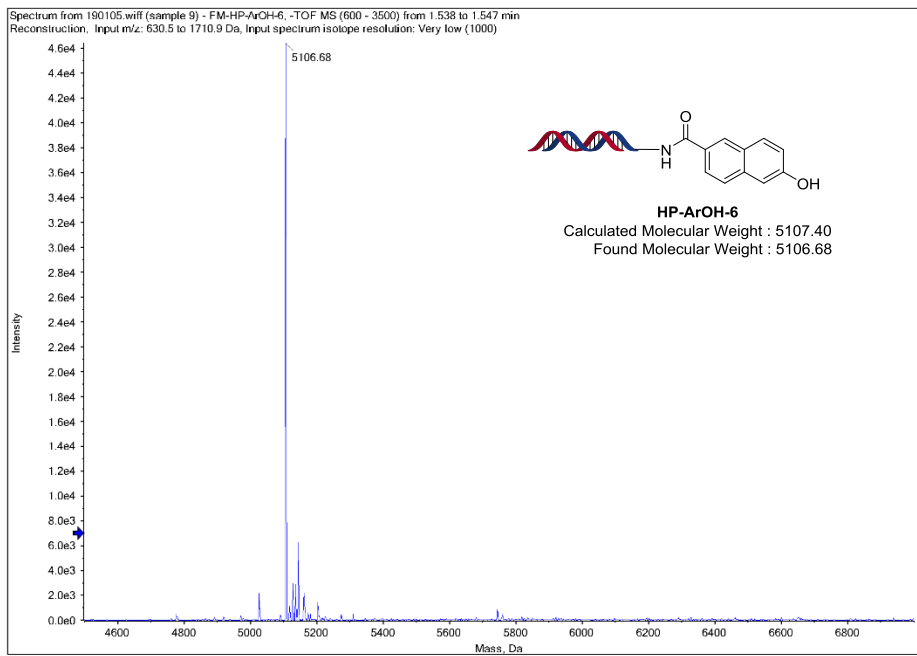
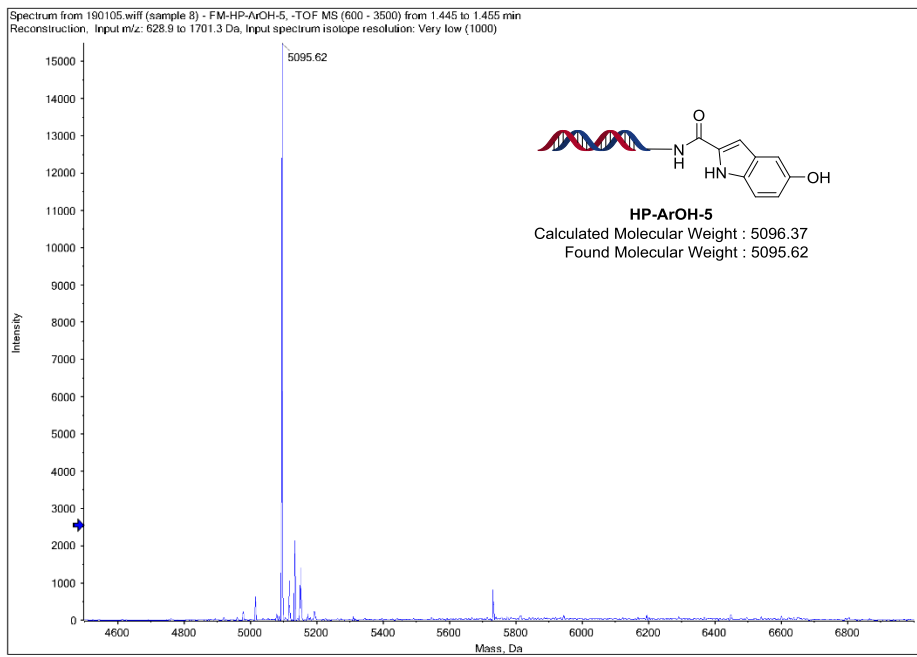


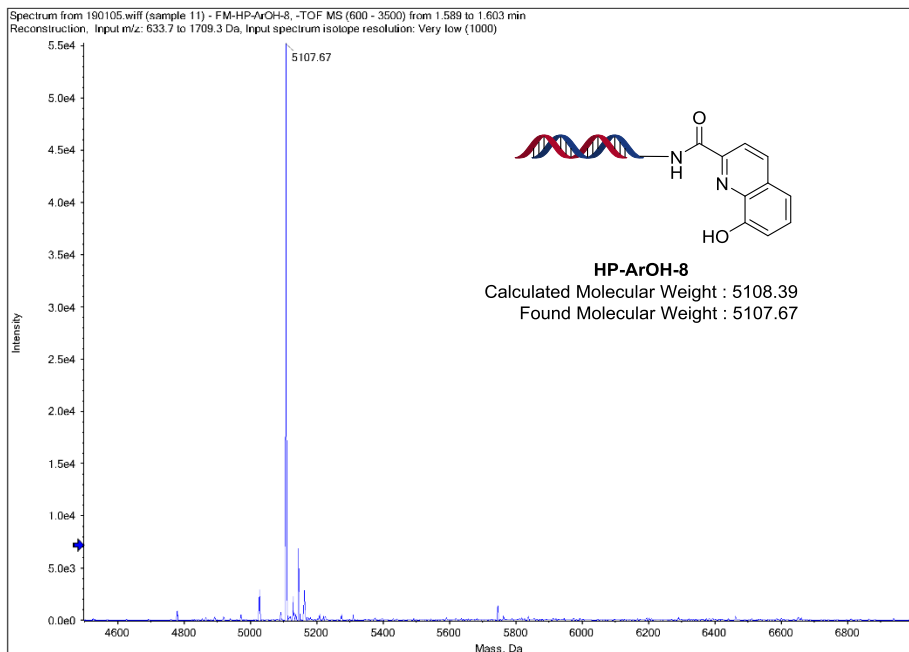
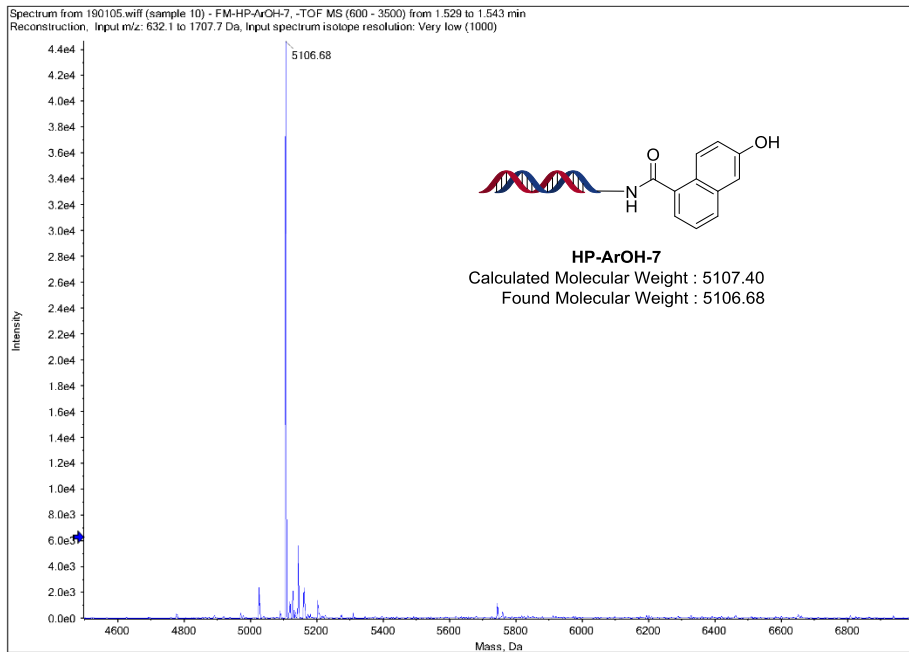
To DNA conjugated (hetero) aryl fluorosulfates (10 nmol, 10 μ L, 1 mM in water), were added 200 equiv. of aniline (10 μ L, 200 mM in DMA), followed by 1000 equiv. of Et₃N (2 μ L, 5 M in DMA) and 10 equiv. of *t*-BuBrettphos Pd G3 (10 μ L, 10 mM in DMA). The mixture was vortexed and stood at 60 °C for 2 hours. After the reaction, 30 equiv. of scavenger sodium diethyldithiocarbamic acid (compared with *t*-BuBrettphos Pd G3, 6 μ L 1 M in ddH₂O) were added to the mixture, and the reaction mixture was heated at 60 °C for at least 30 minutes. The resulting mixture was centrifuged at 4 °C for 30 min at 12,000 rpm, and the resultant supernatant was collected. 5 M NaCl solution (10% by volume) and cold ethanol (2.5 times by volume, ethanol stored at -20°C) were added to the resultant supernatant, and the mixture was vortexed and incubated at -80°C for at least 30 minutes. The sample was centrifuged for 30 minutes at 4°C in a microcentrifuge at 12,000 rpm to remove the supernatant. The resulting pellet (precipitate) was re-dissolved in ddH₂O (300 μ L) for LC-MS detection.

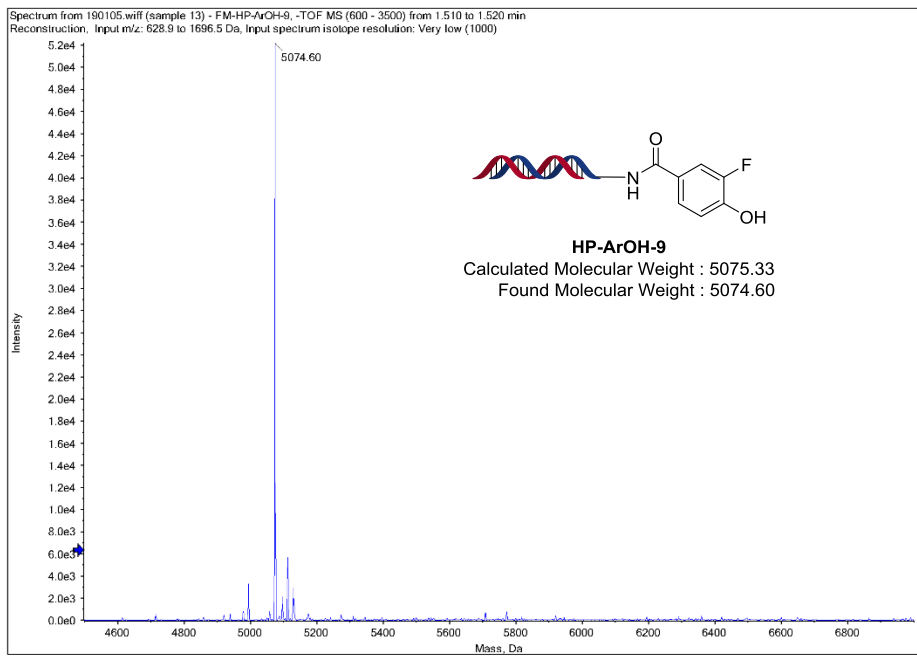
MS Spectra of DNA conjugated phenols



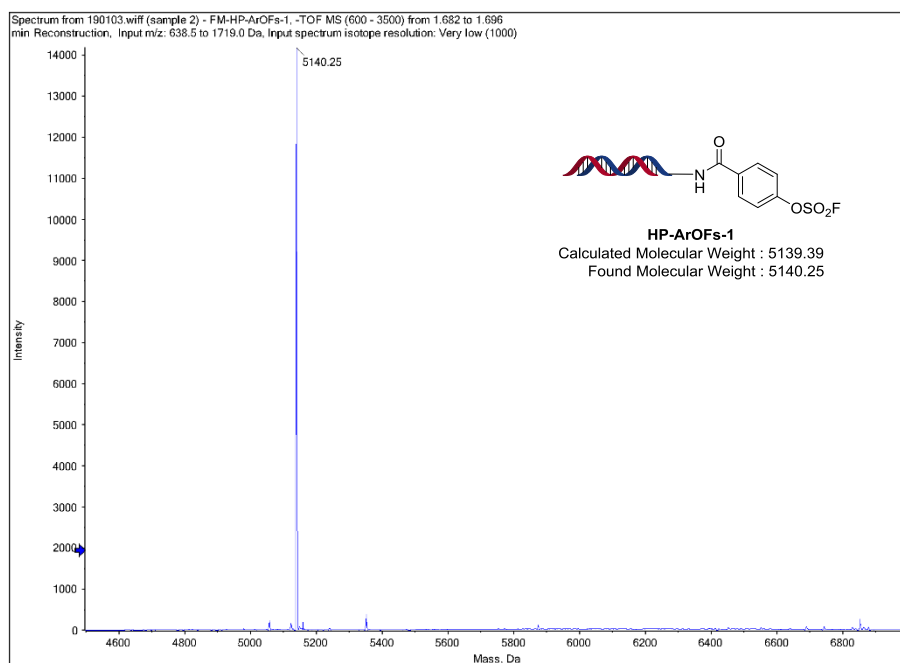


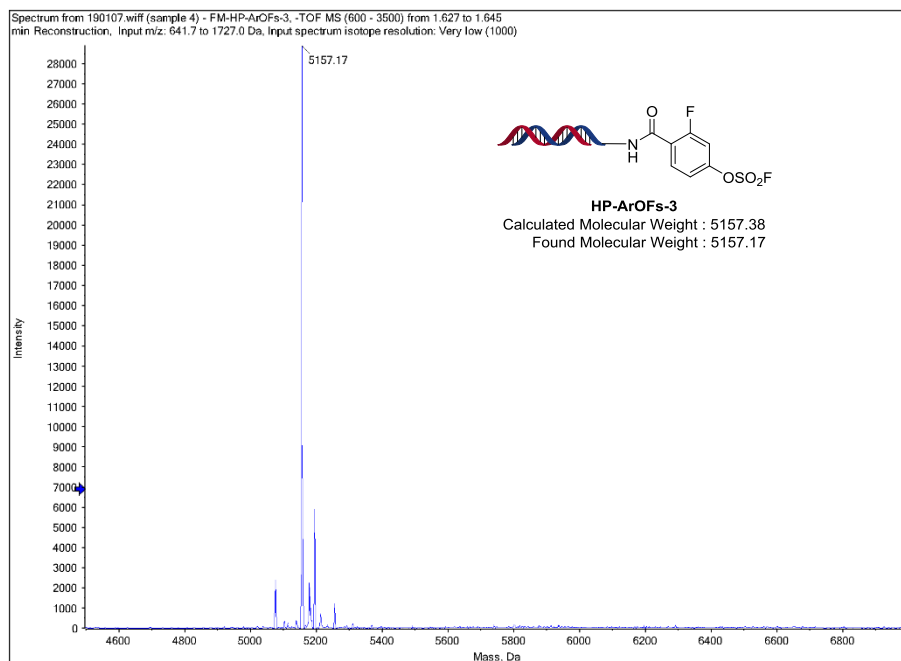
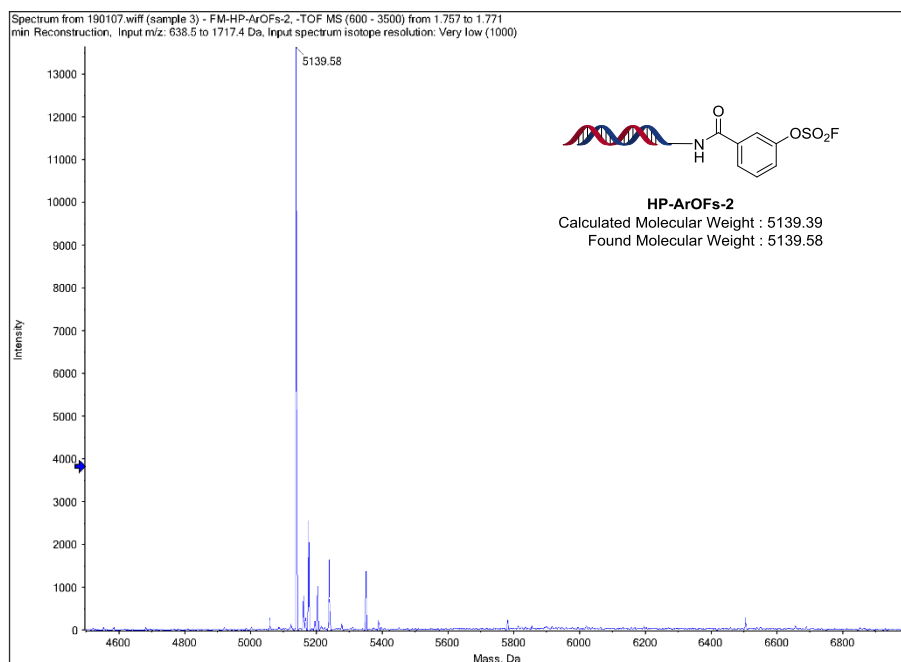


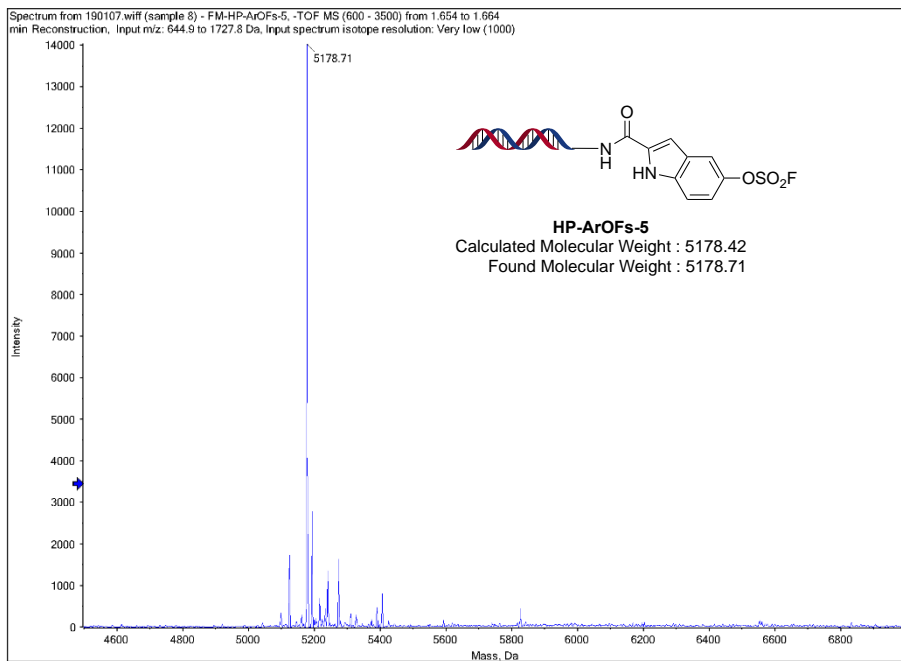
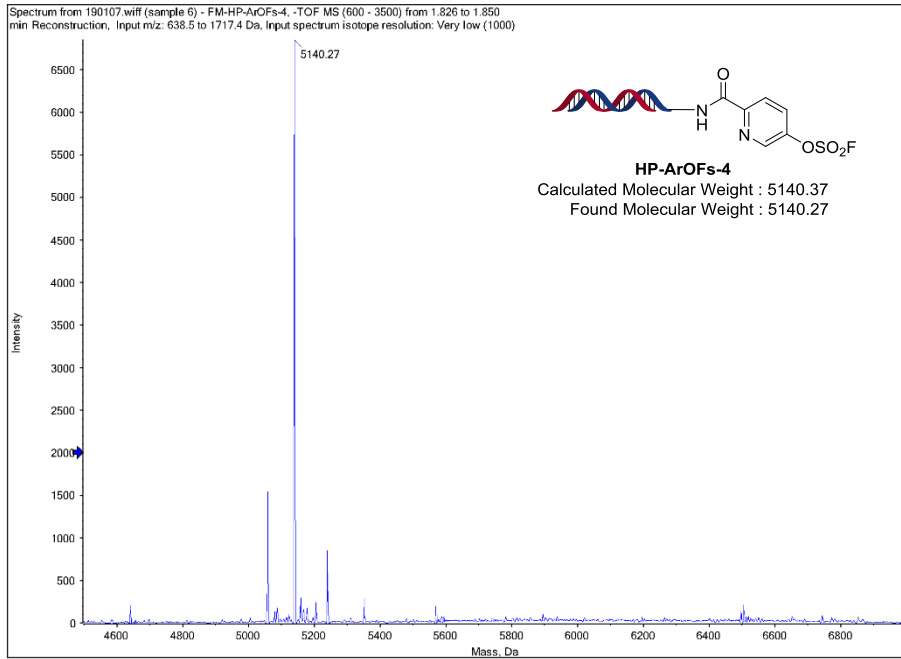


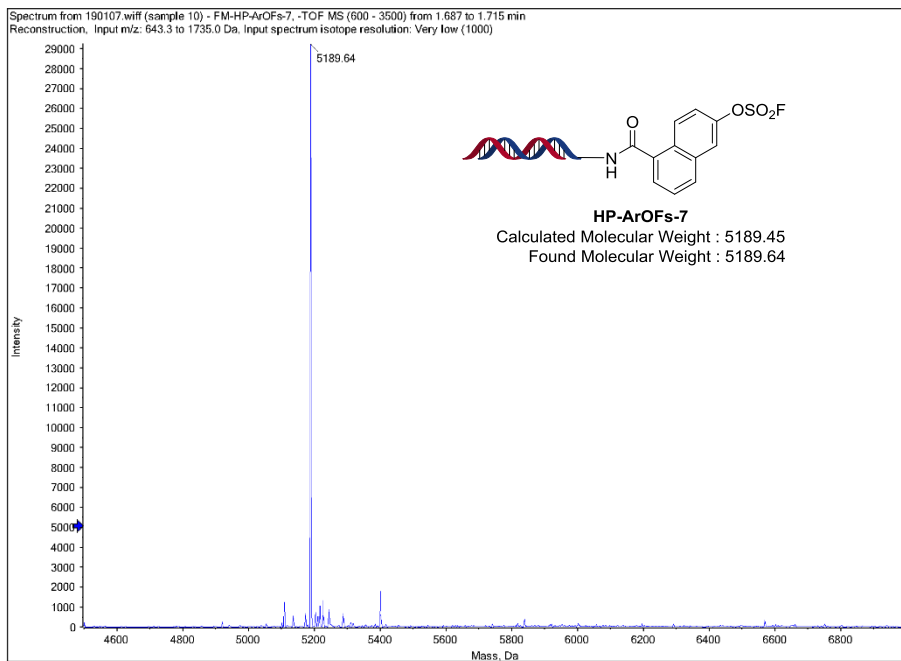
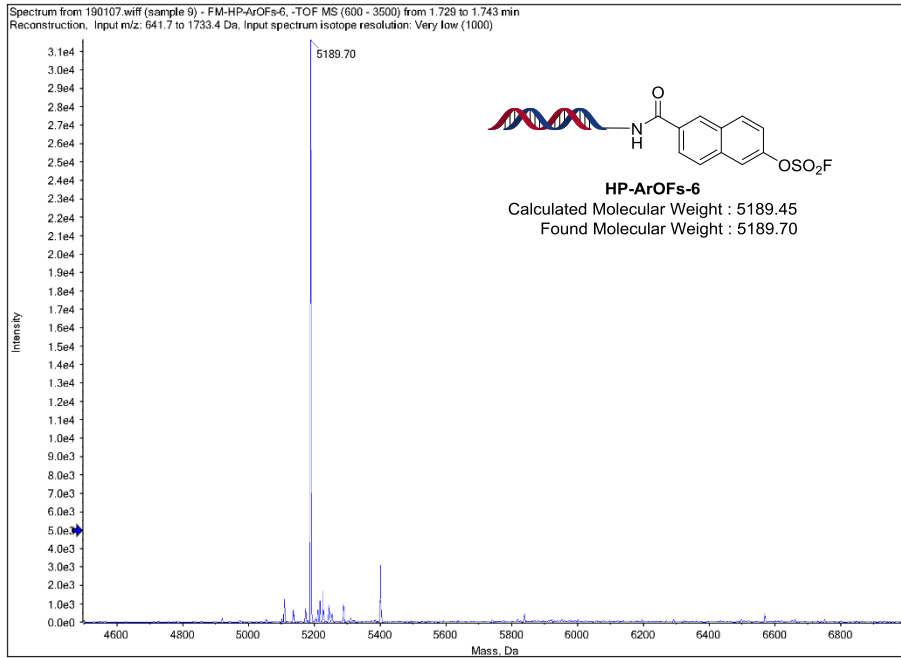


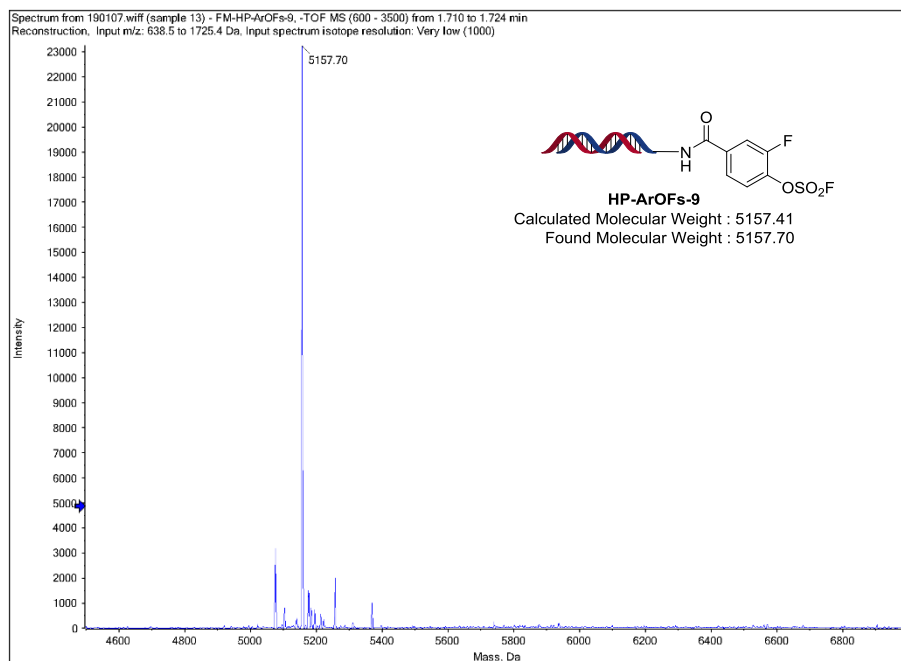
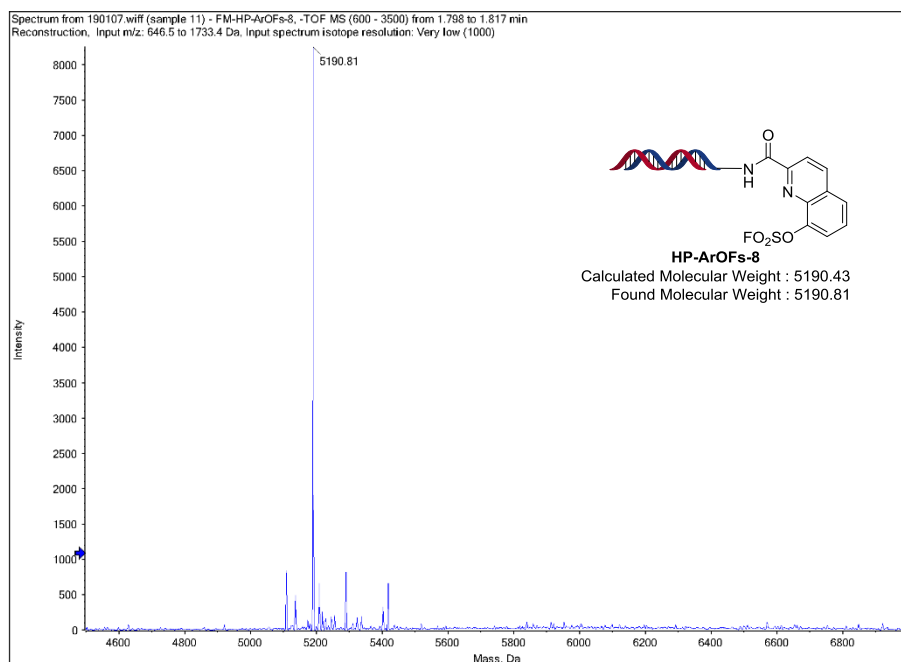
MS Spectra of DNA conjugated (hetero)aryl fluorosulfonates.



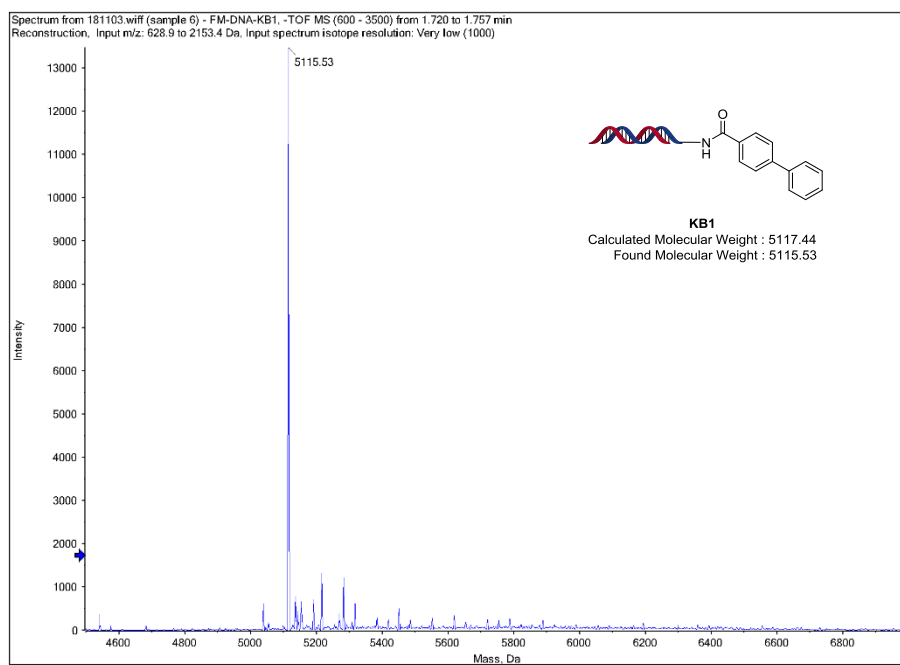


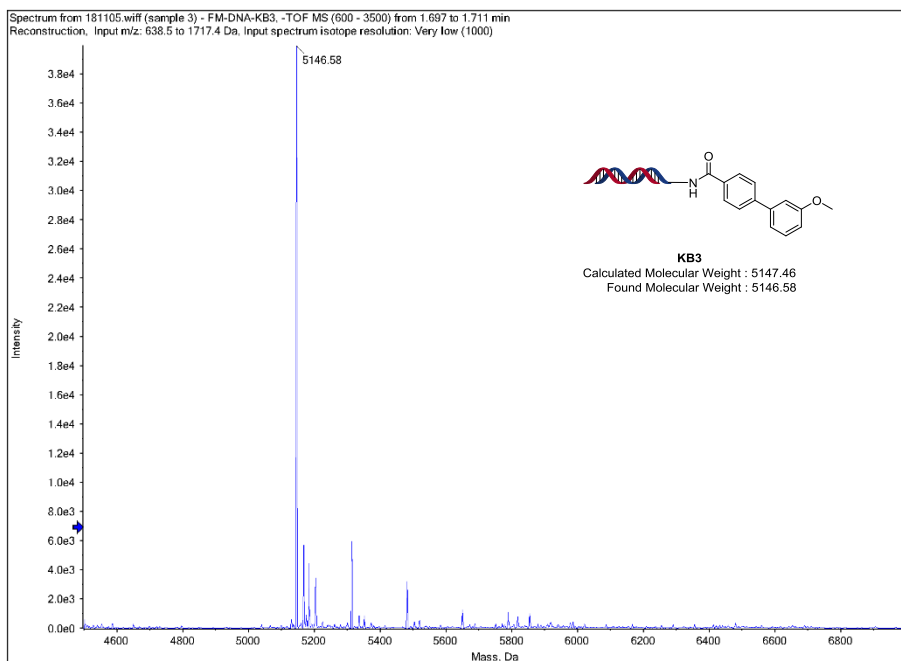
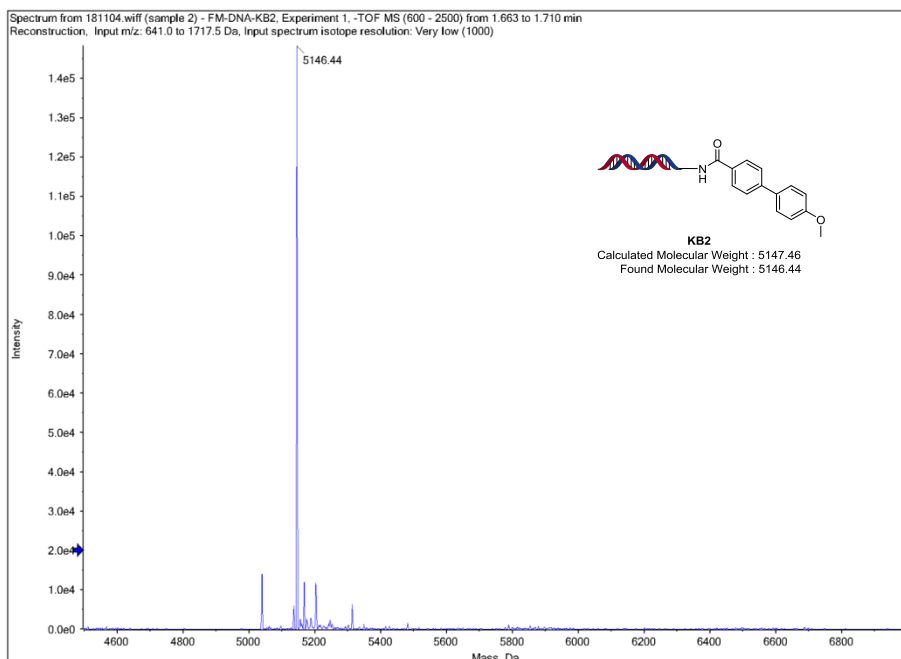


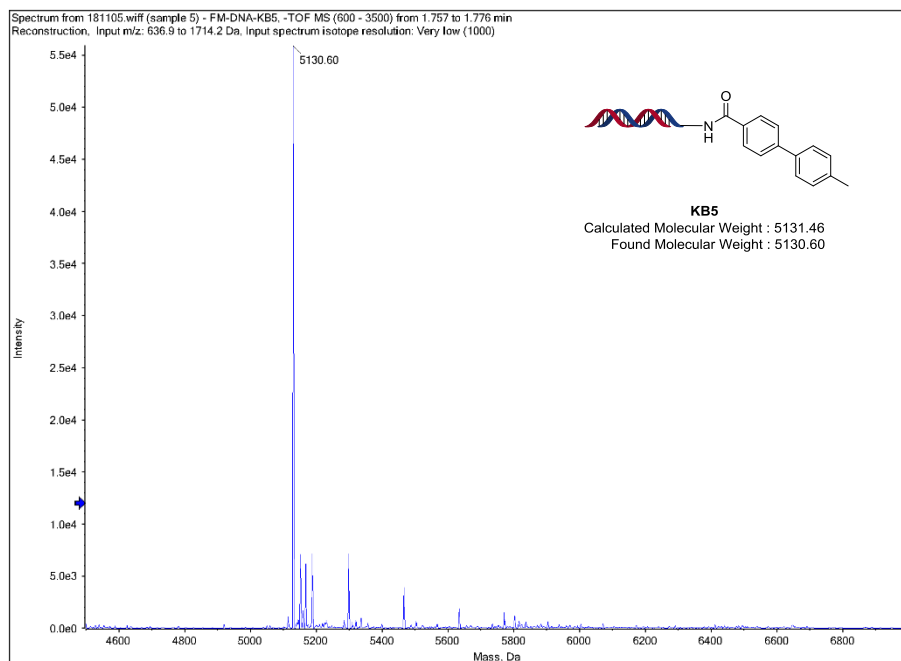
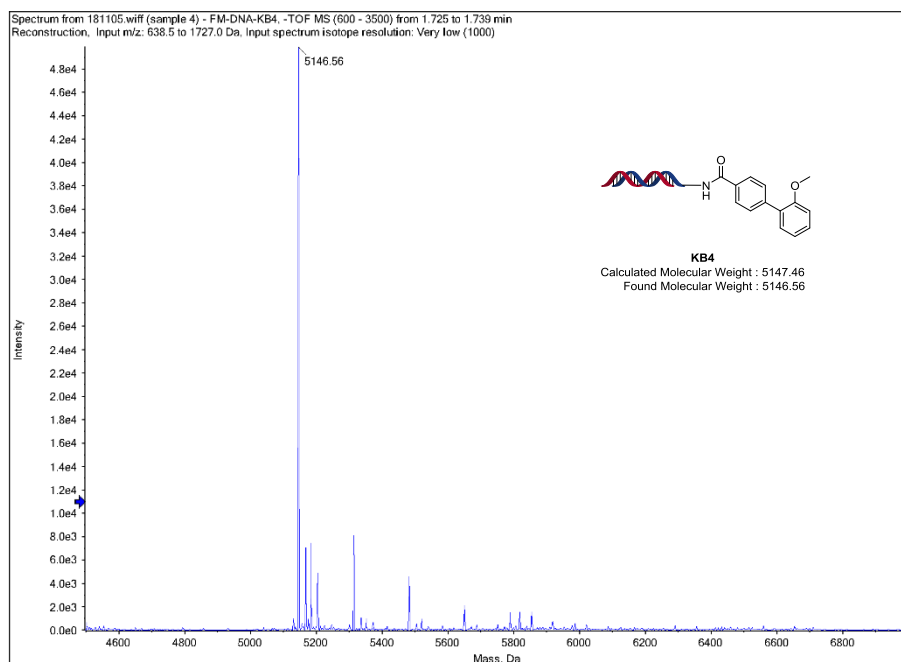


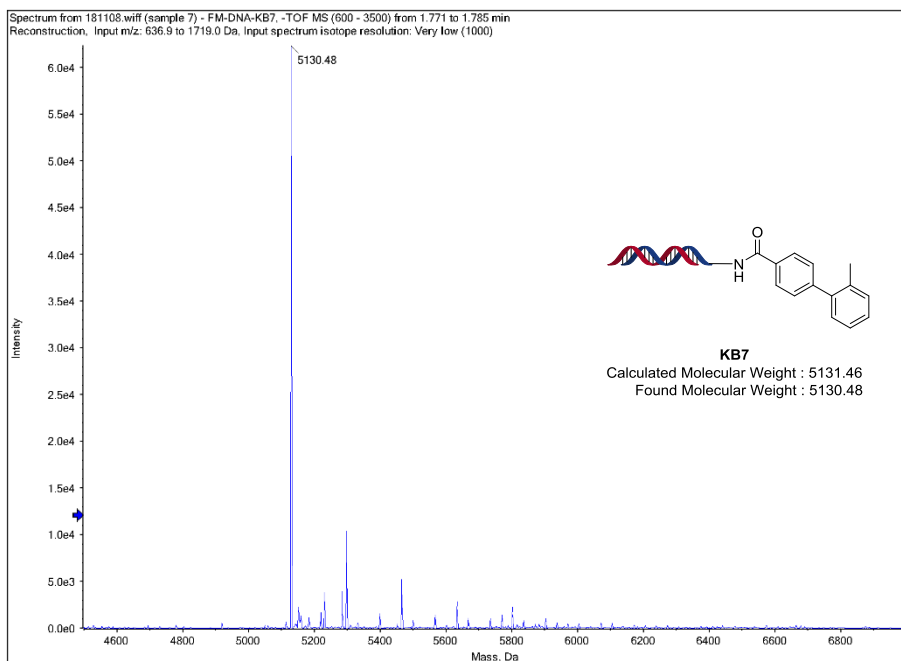
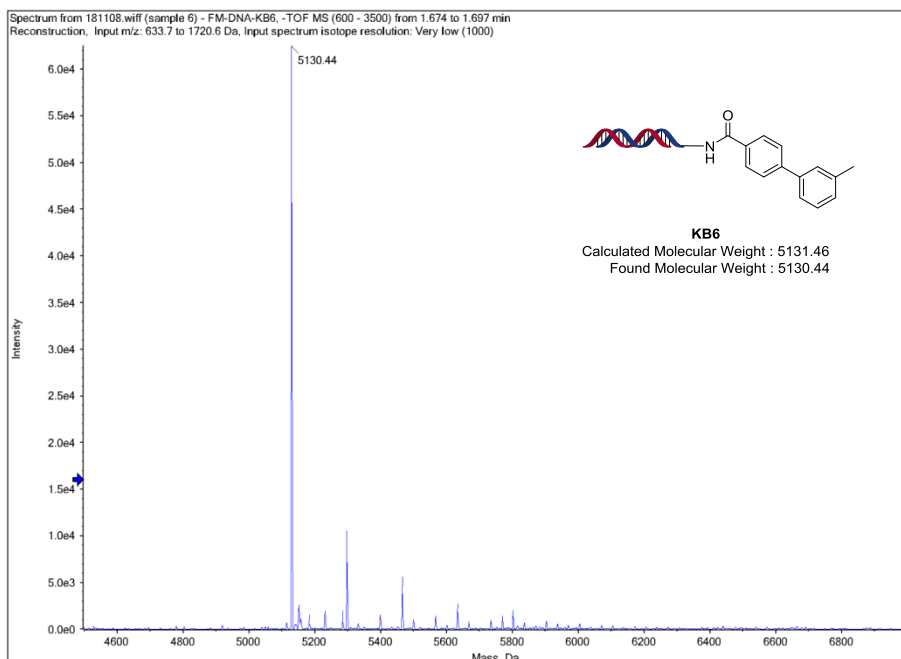


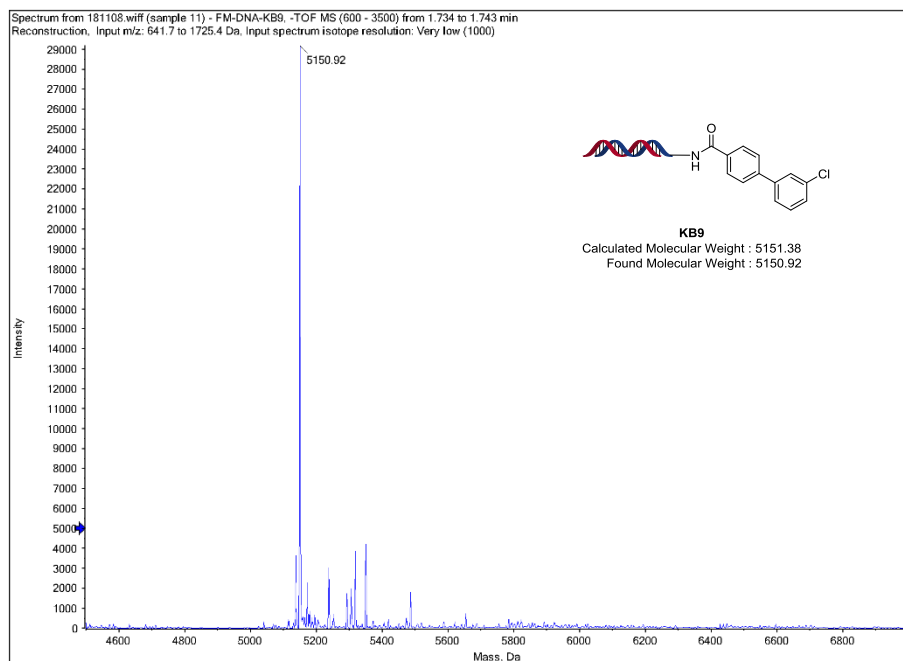
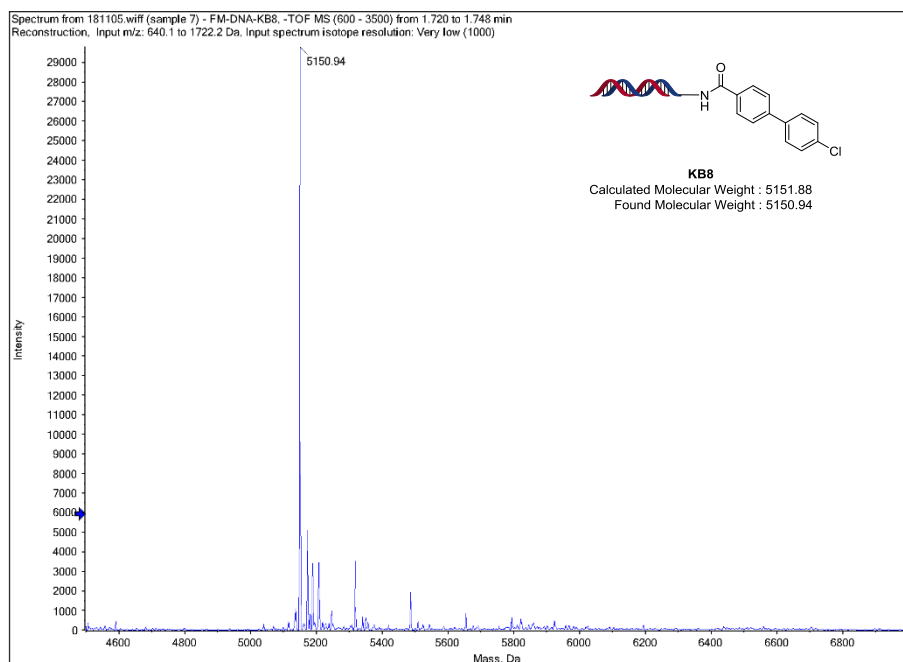
MS Spectra of On-DNA Suzuki-Miyaura coupling products

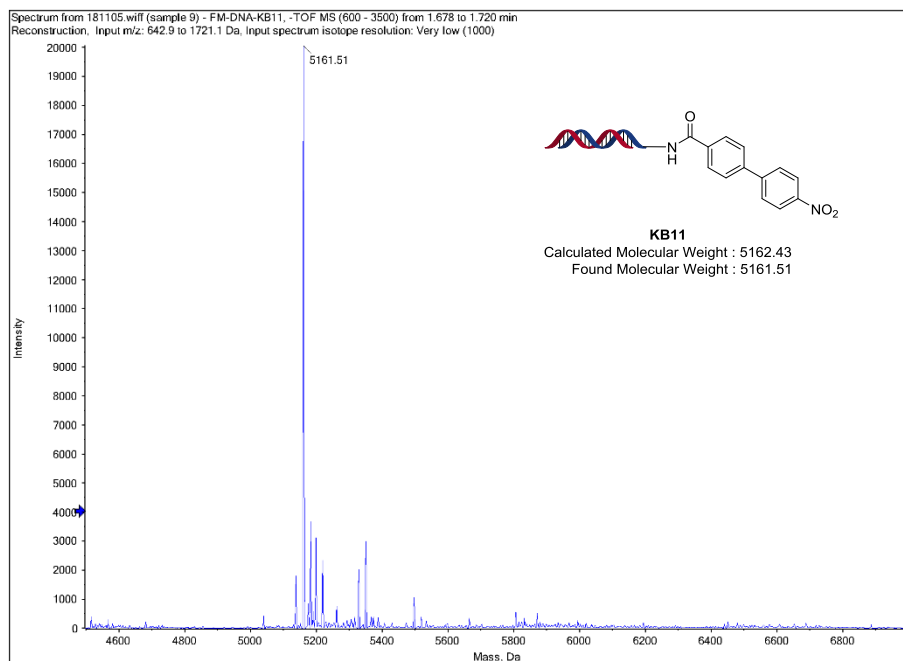
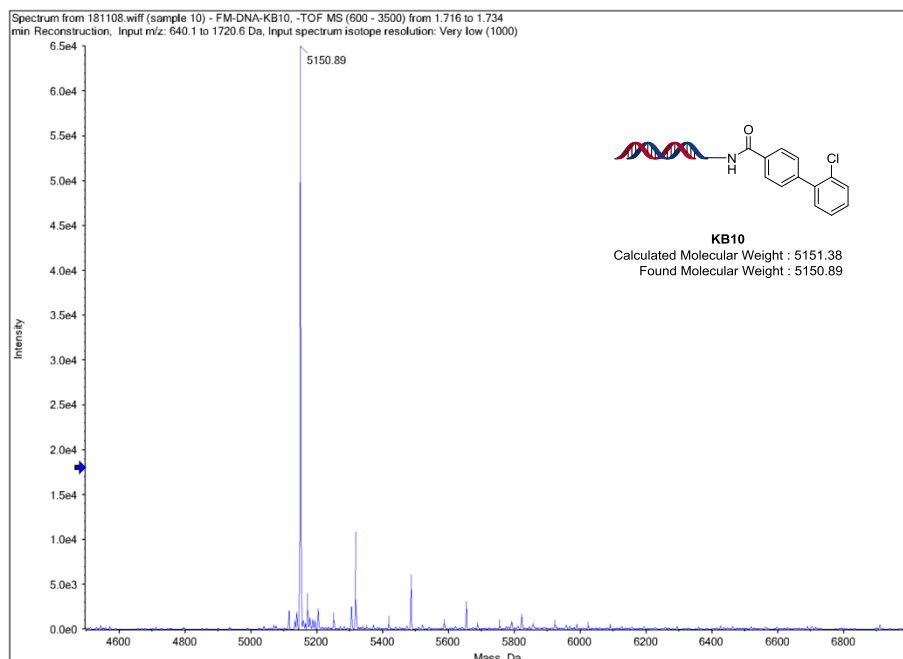


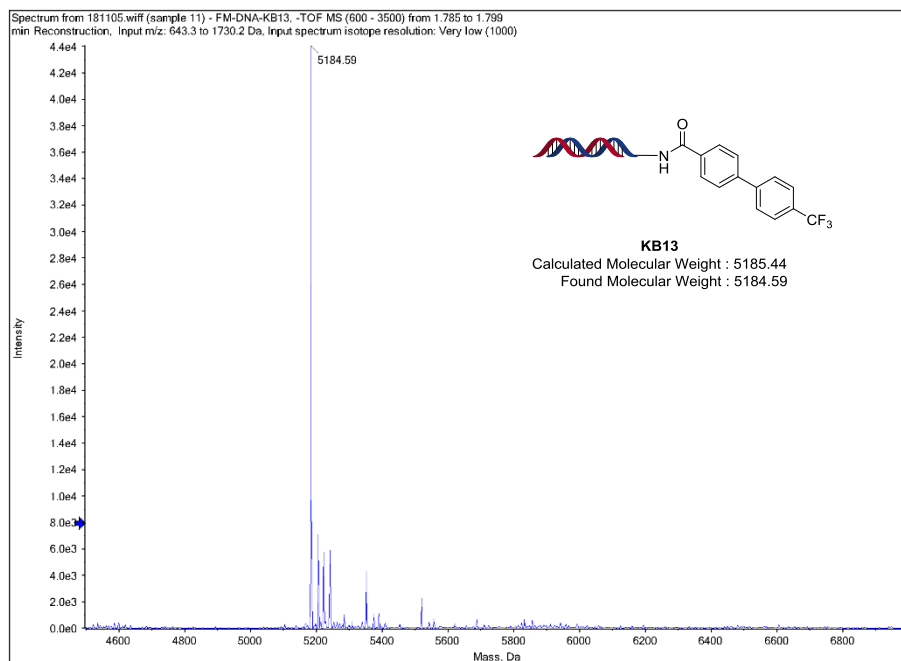
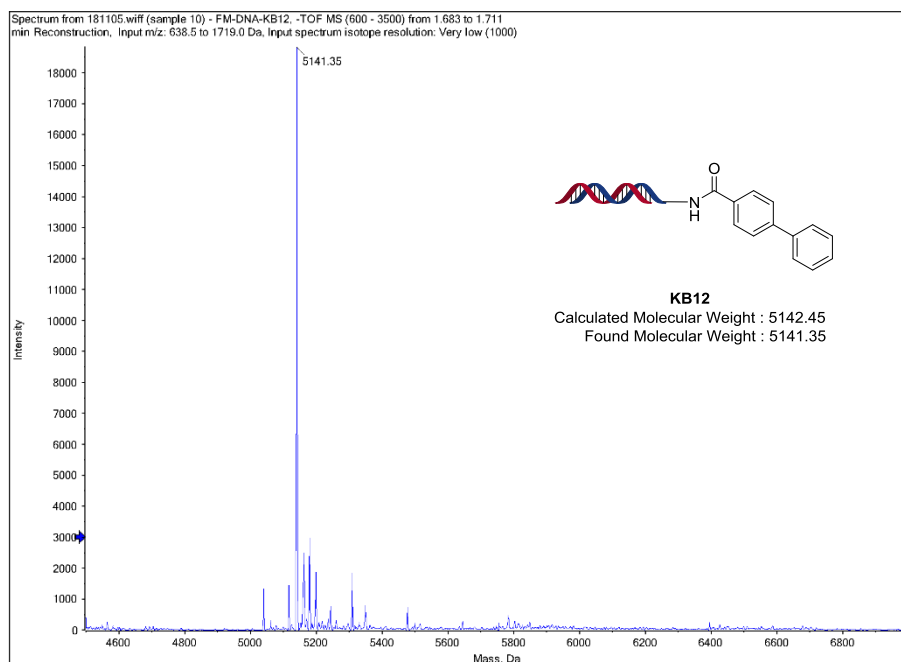


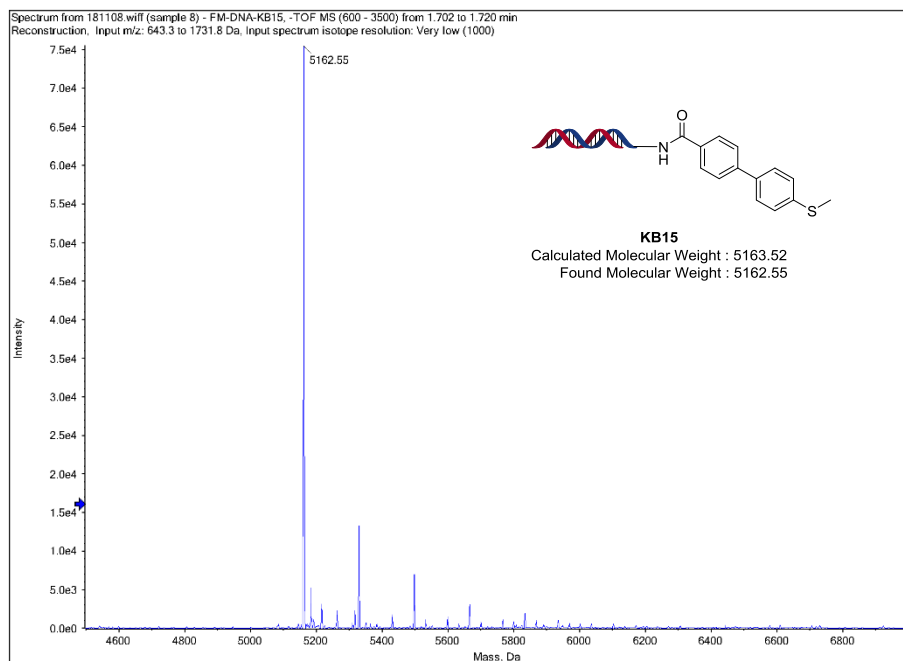
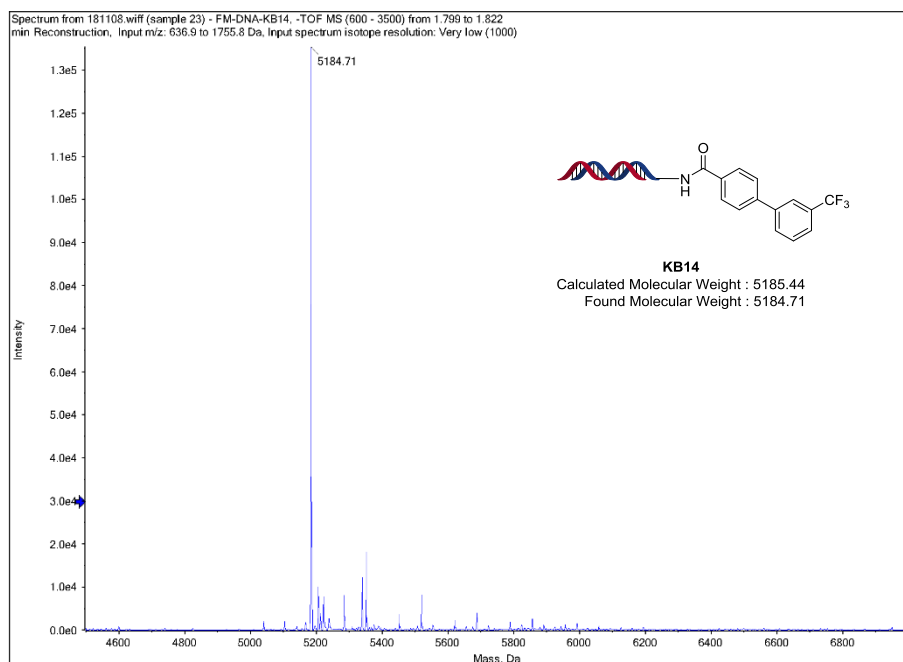


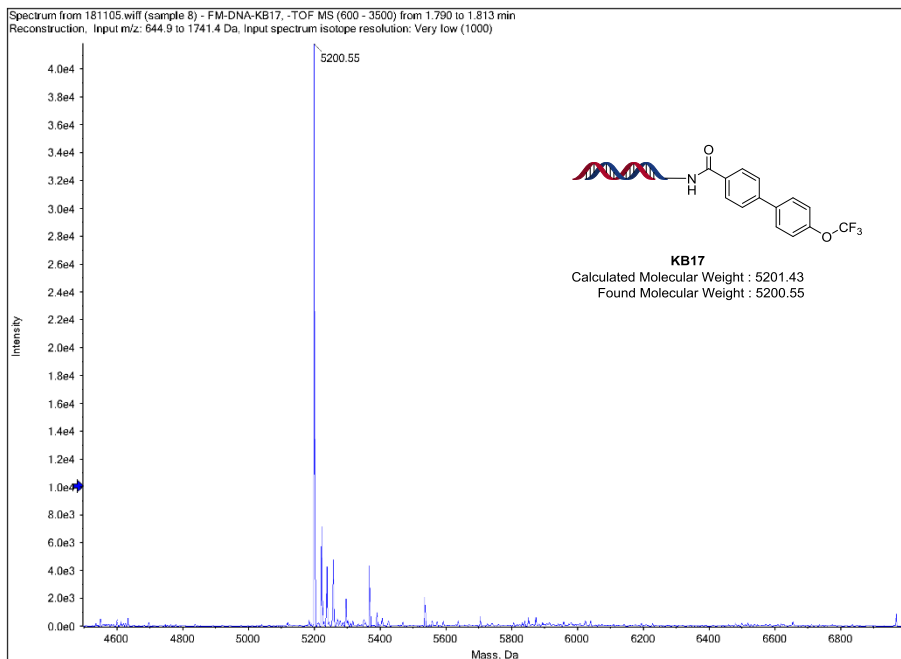
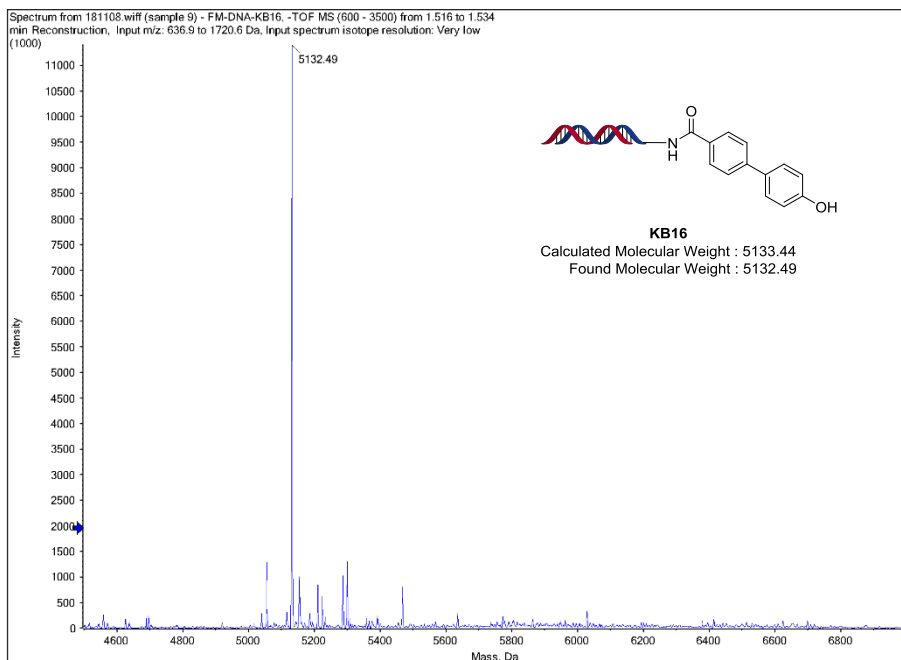


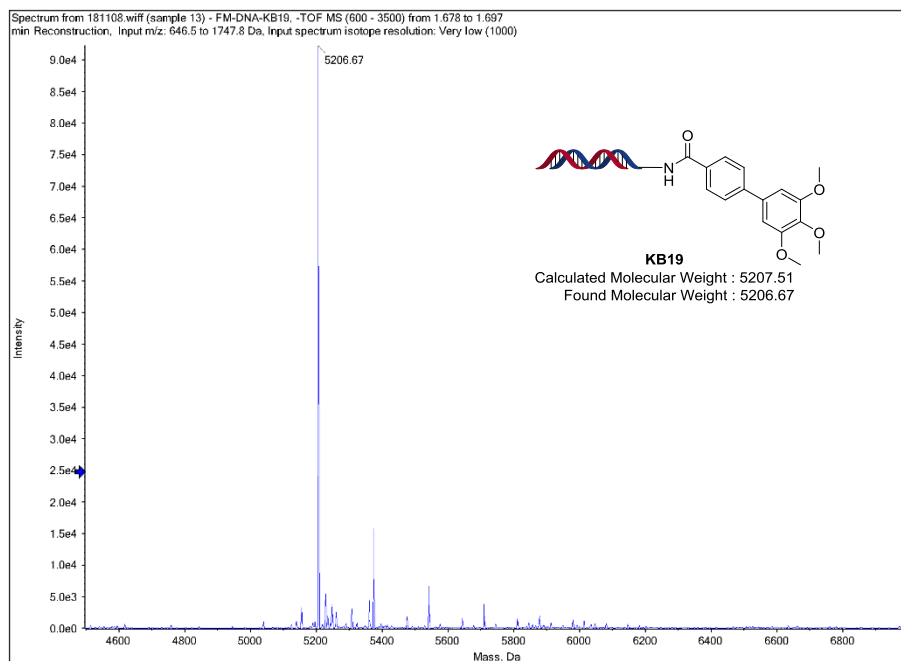
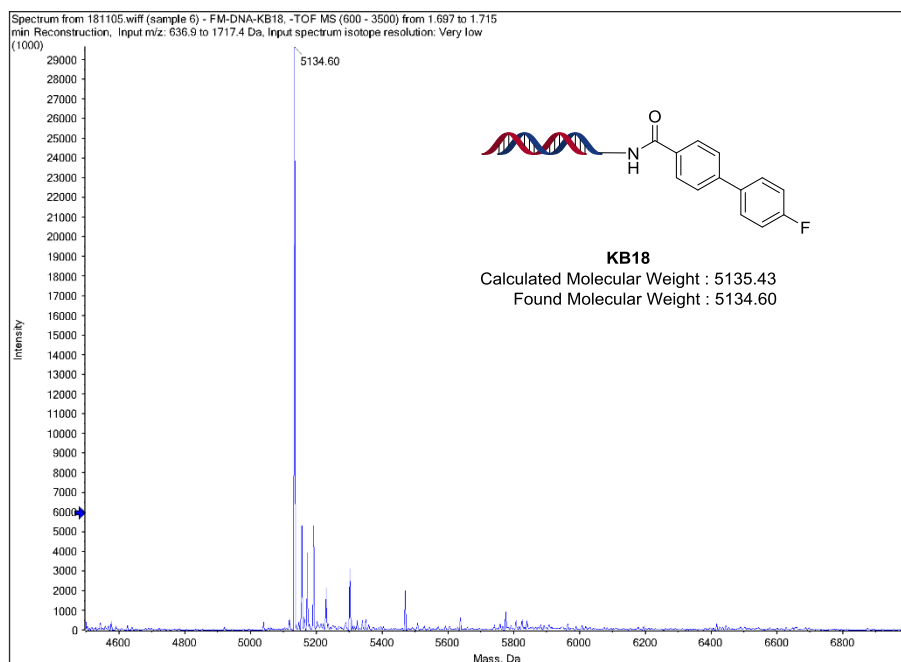


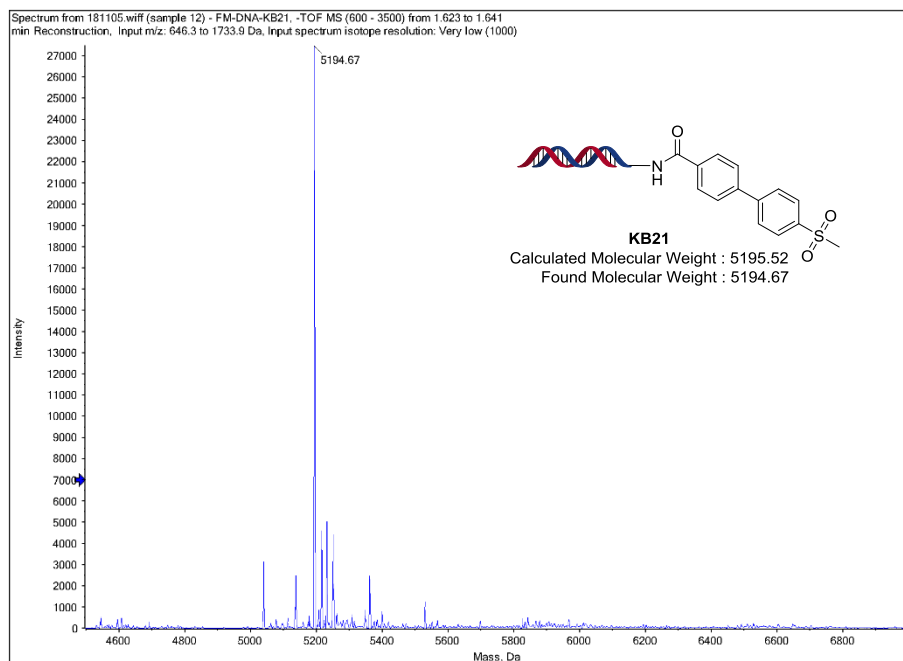
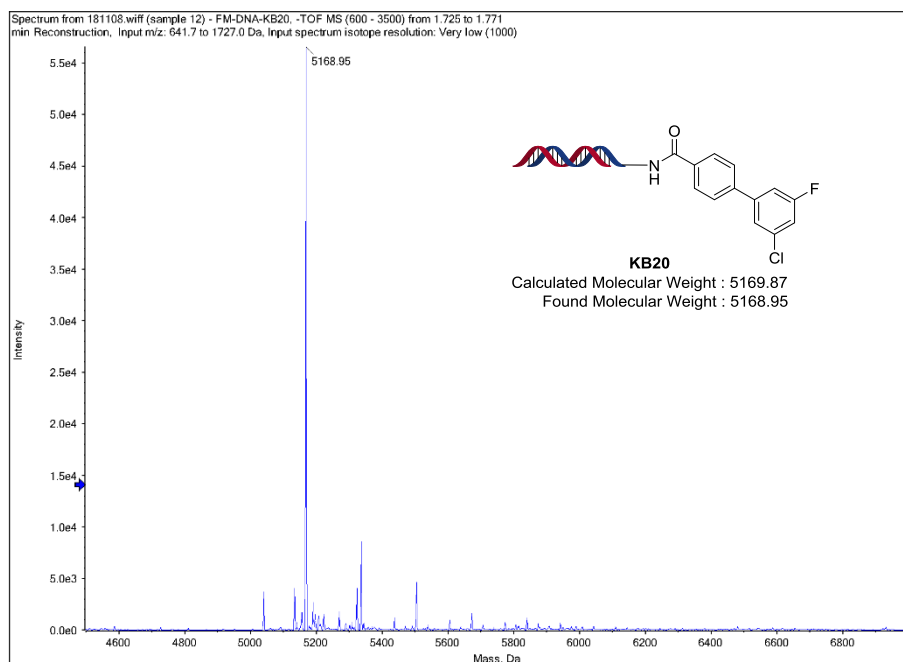


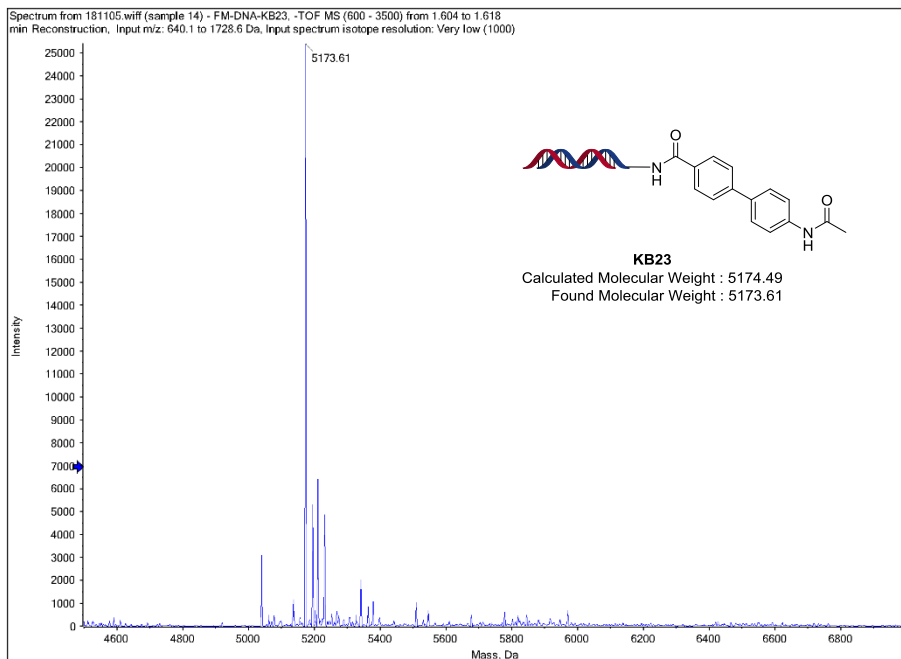
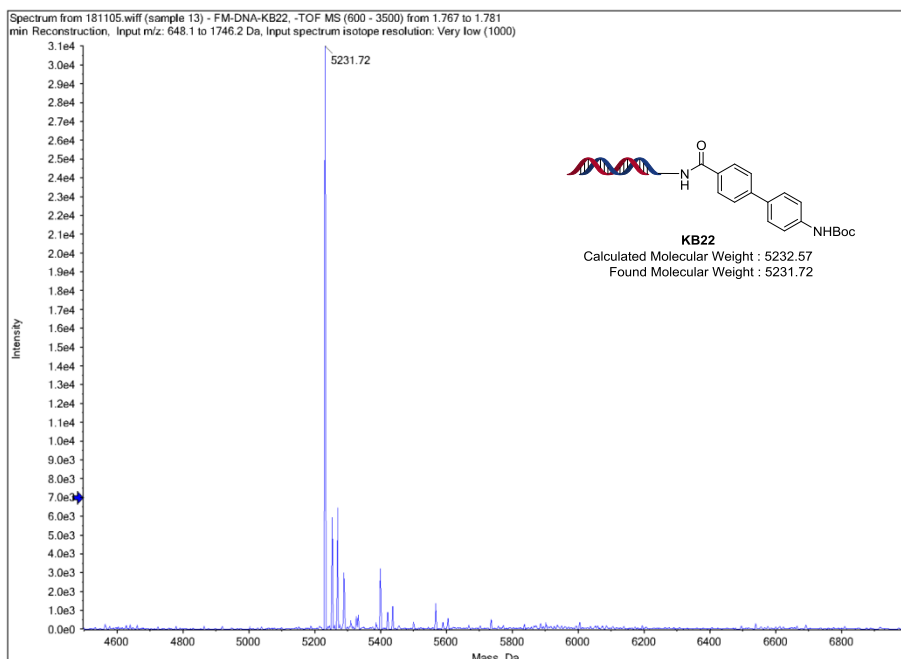


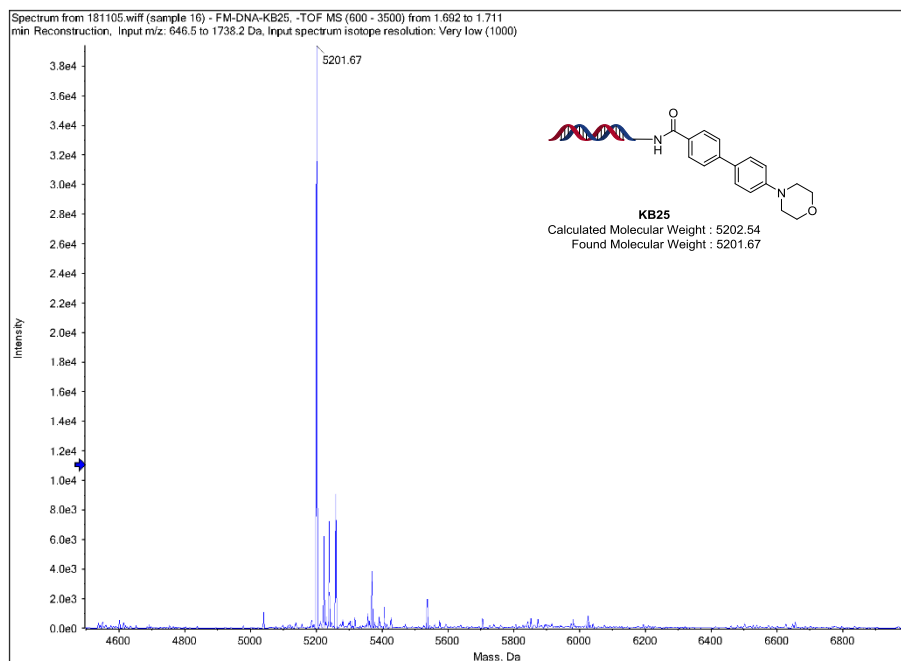
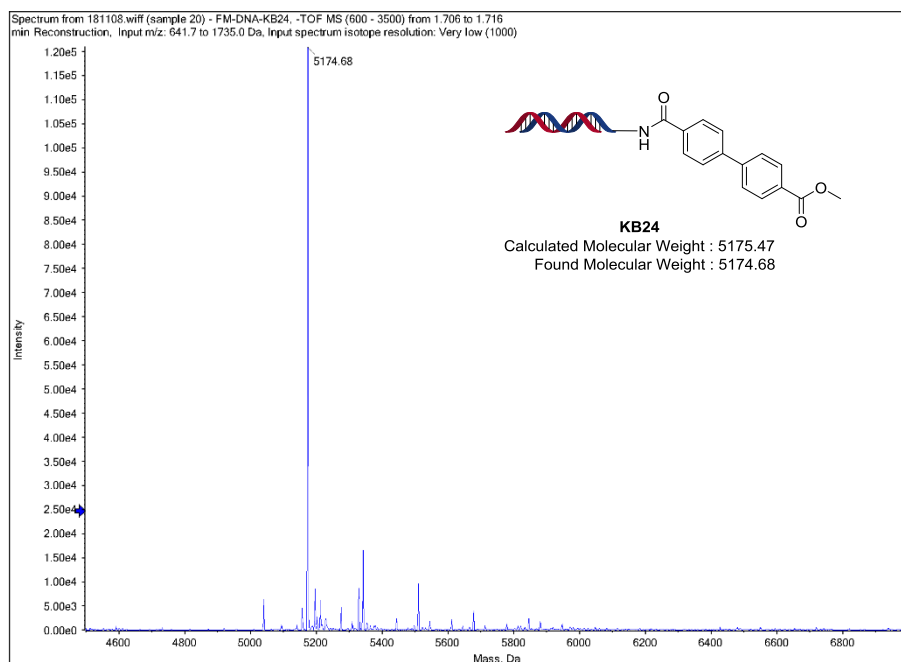


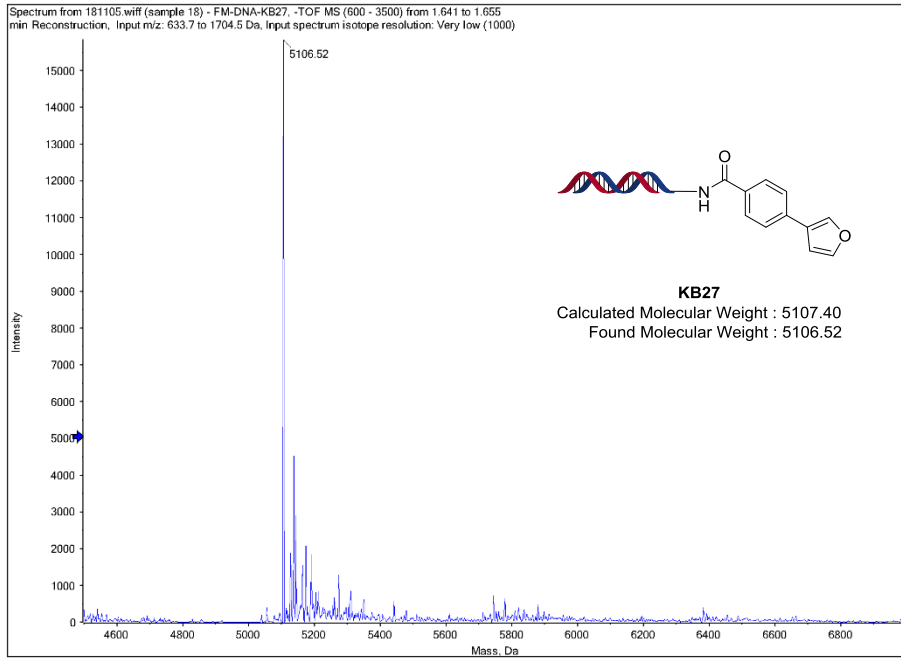
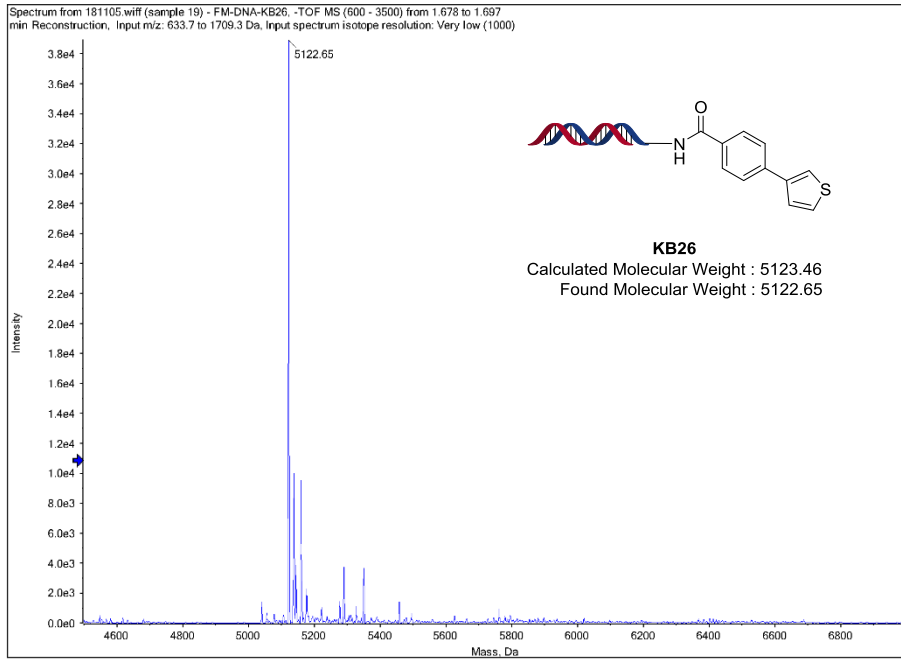


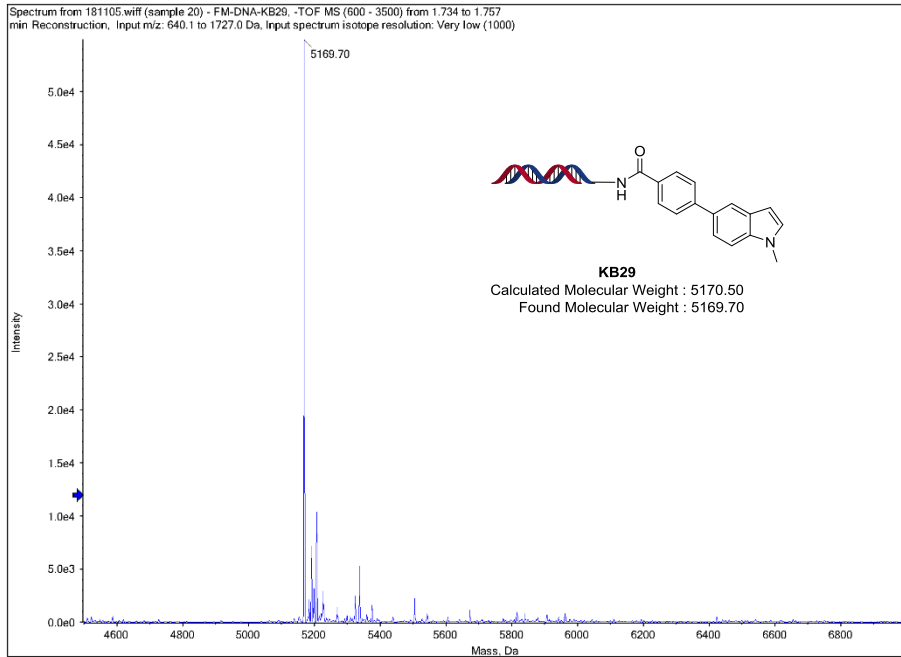
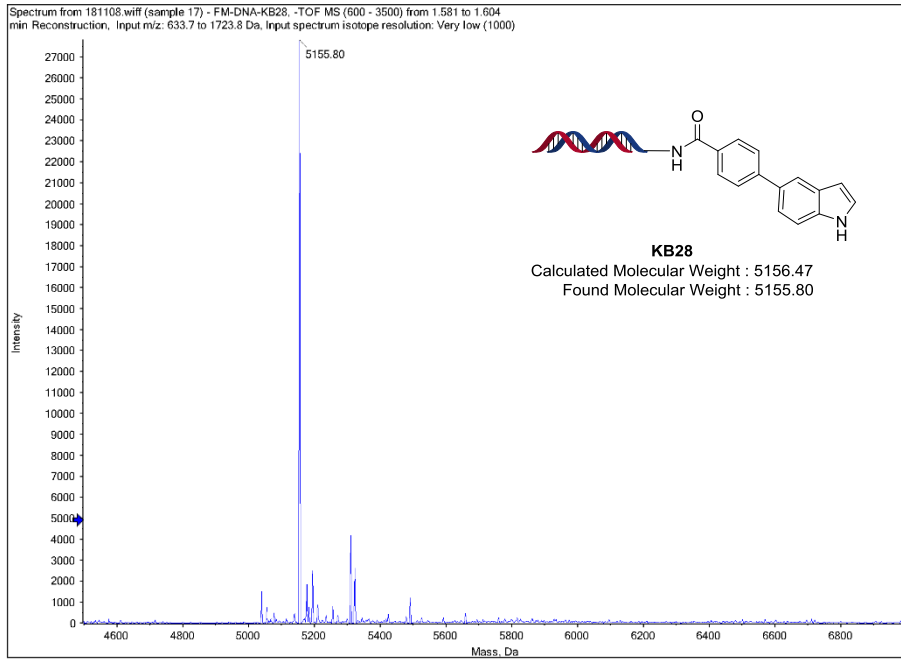


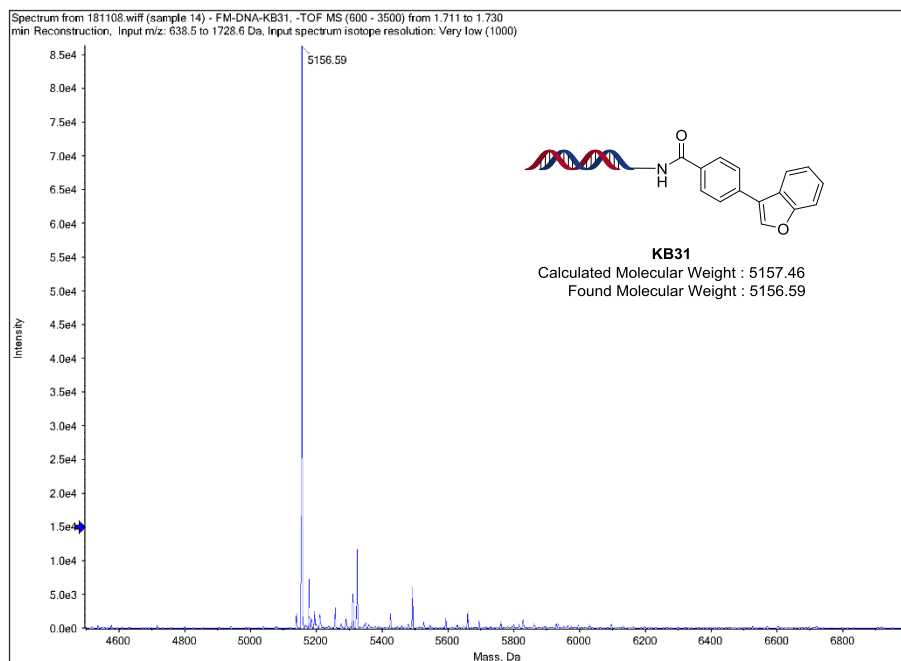
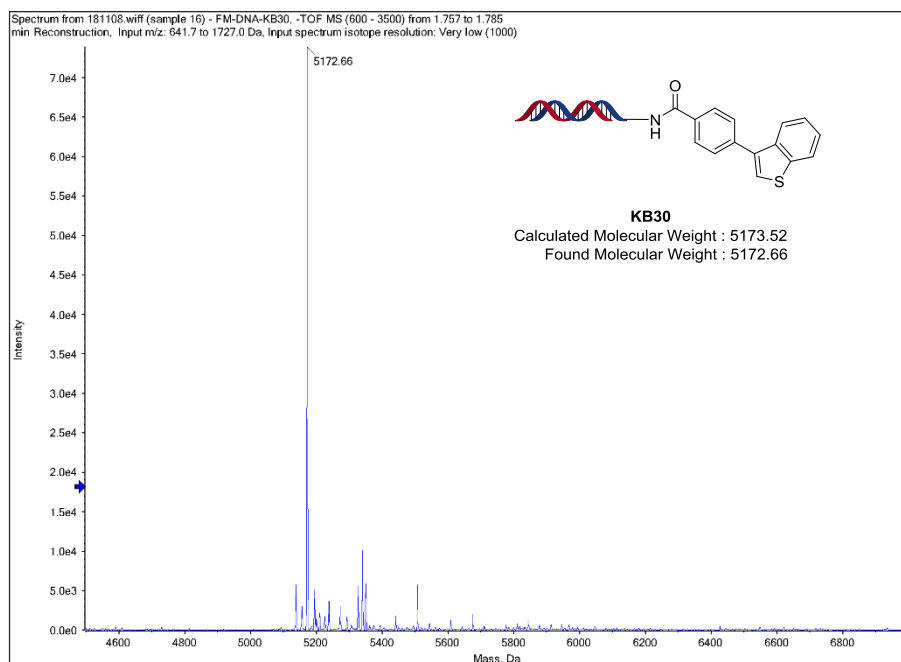


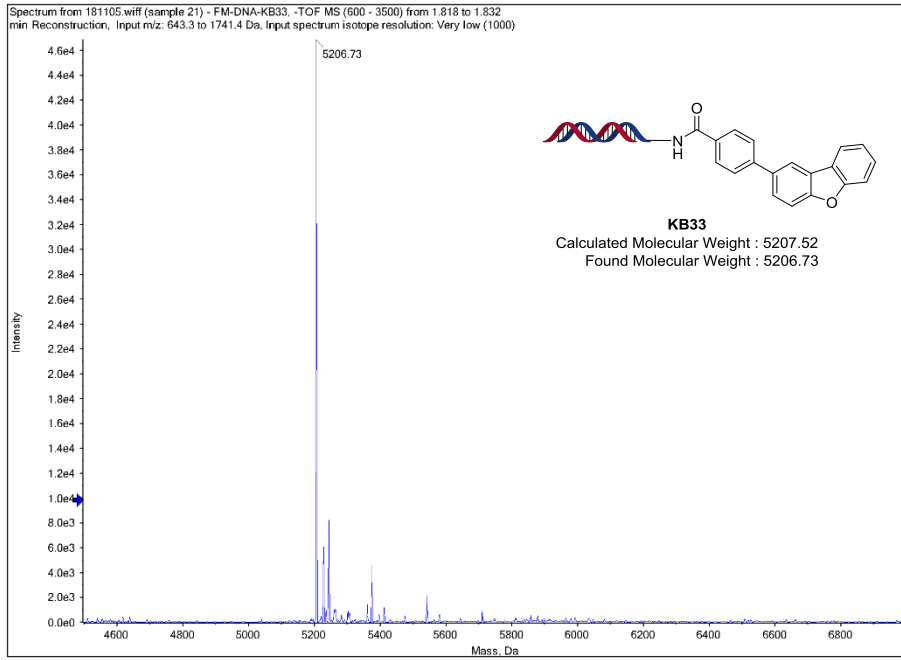
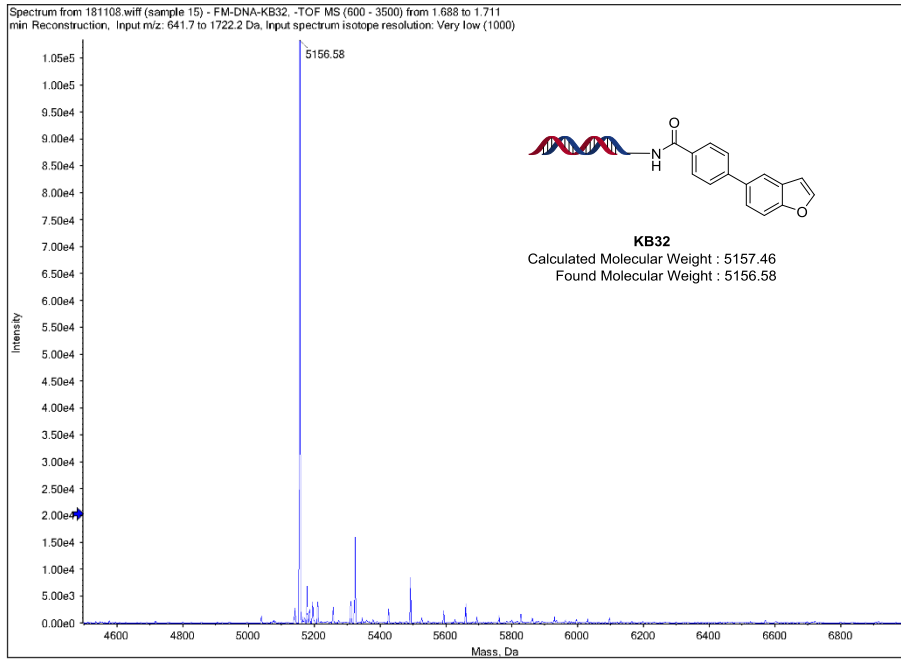


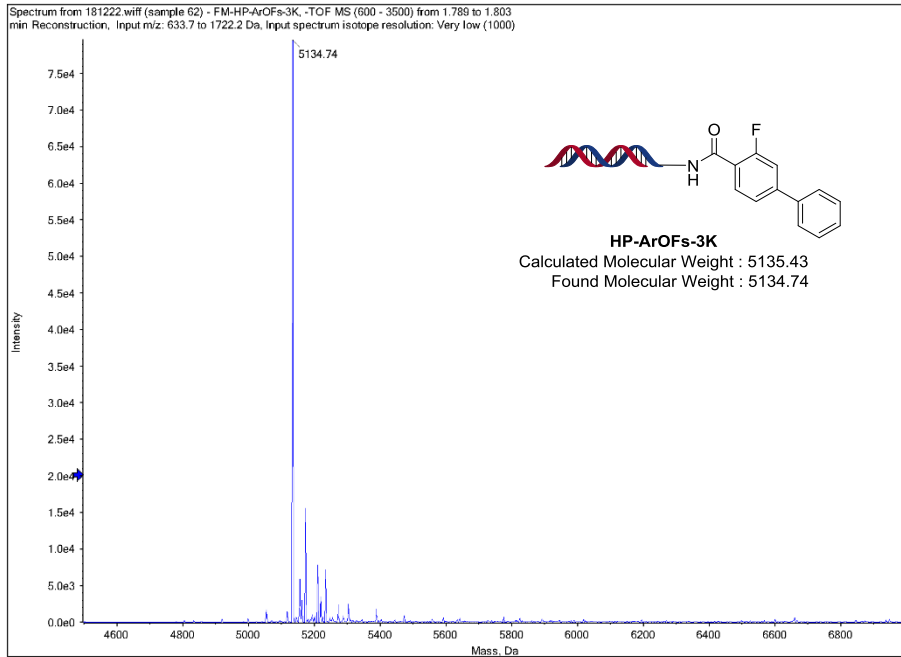
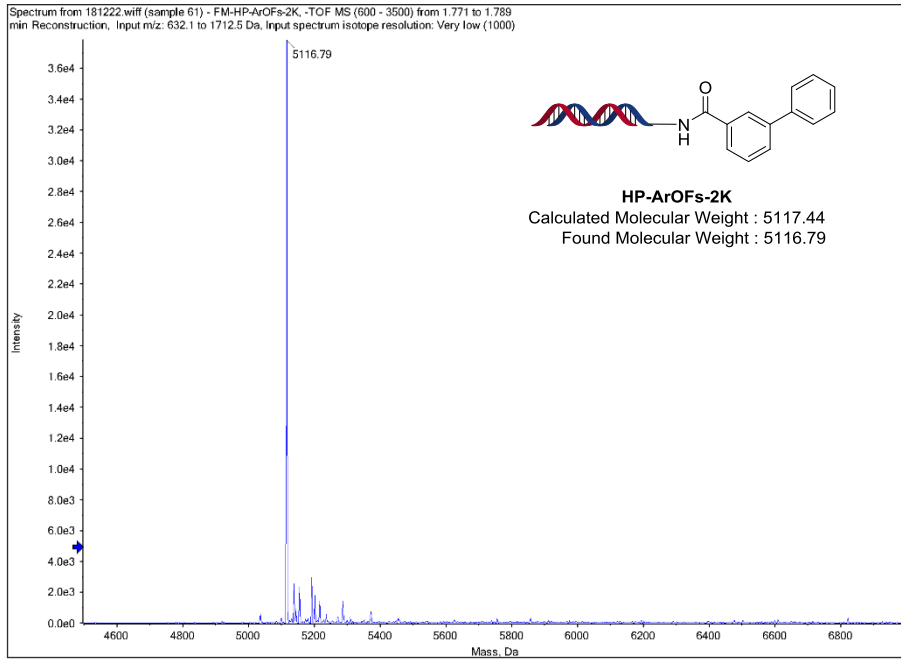


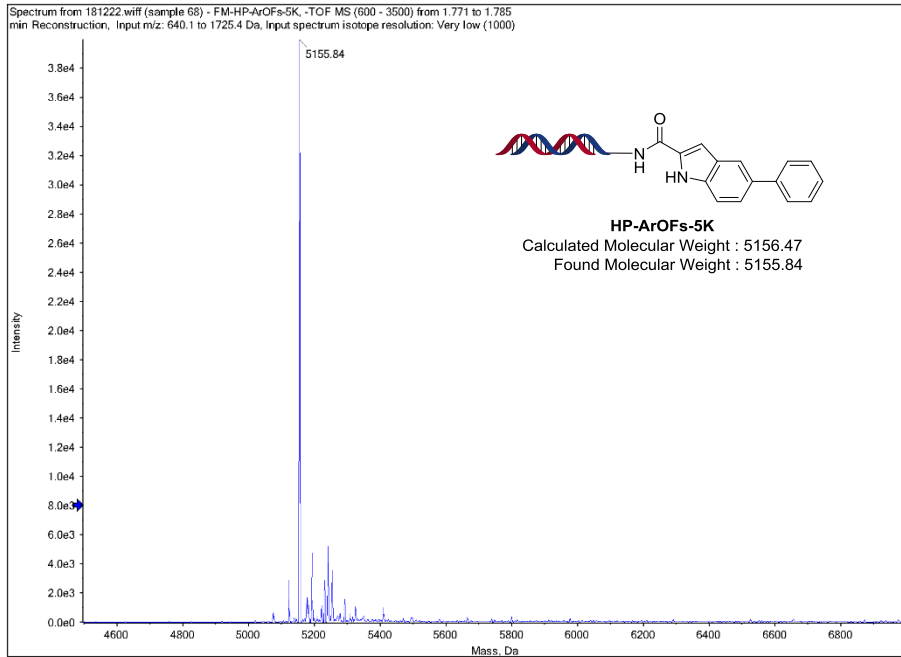
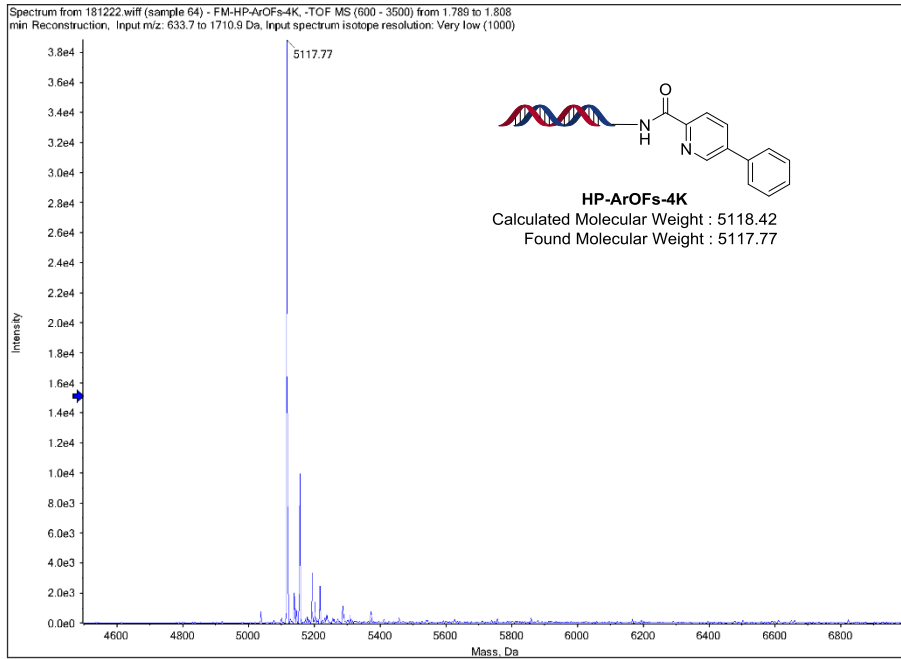


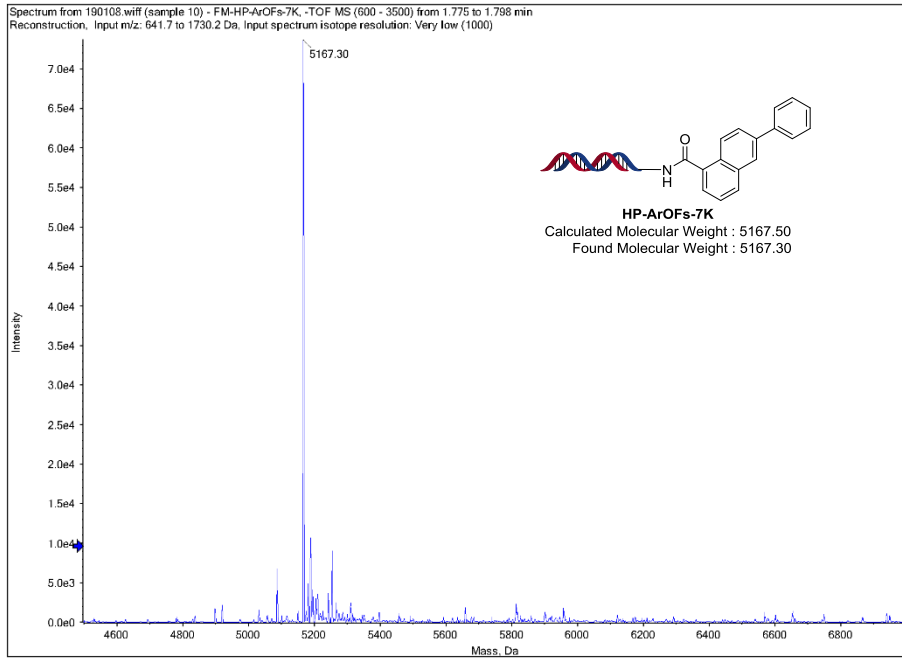
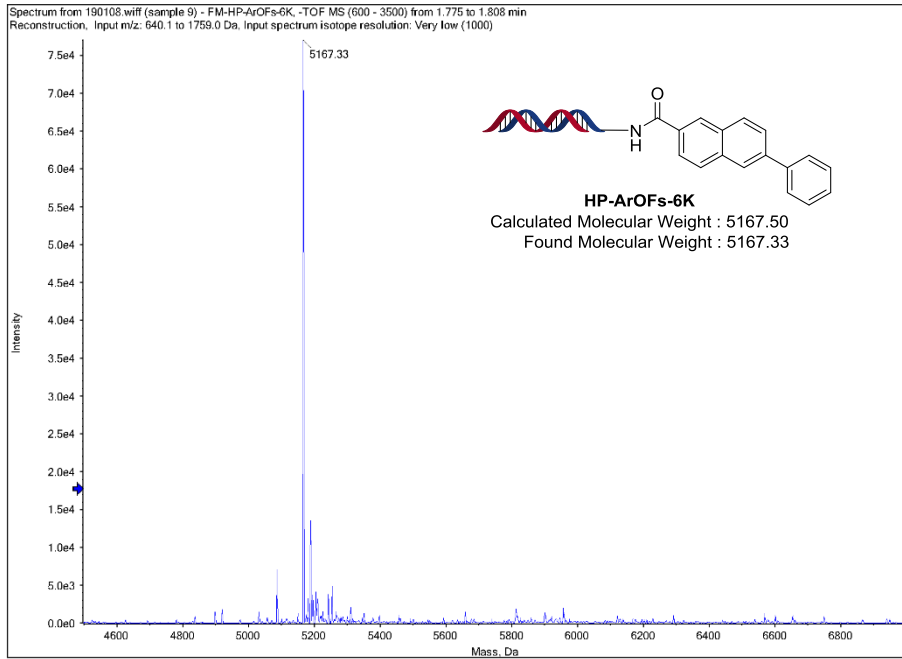


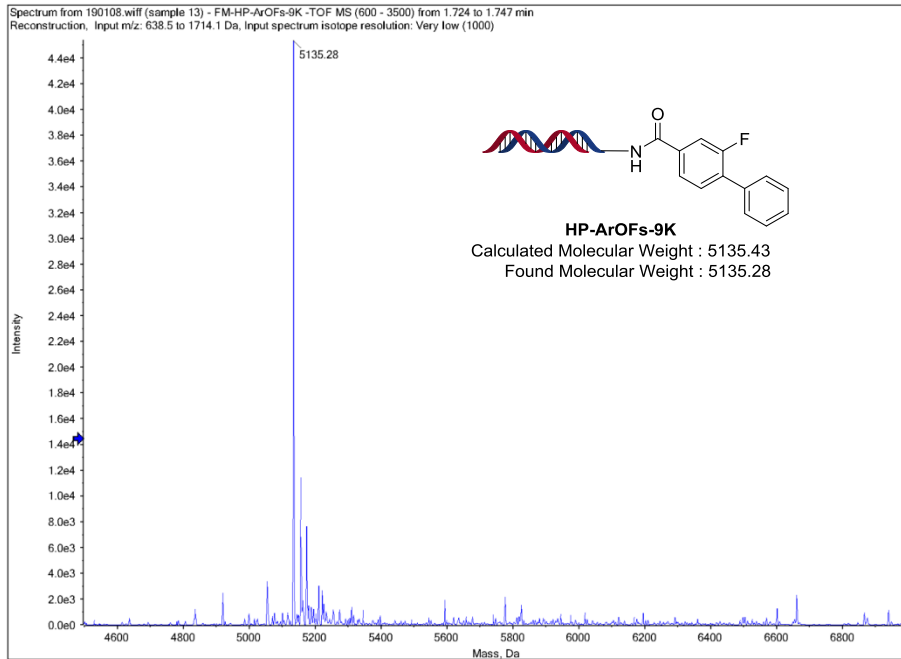
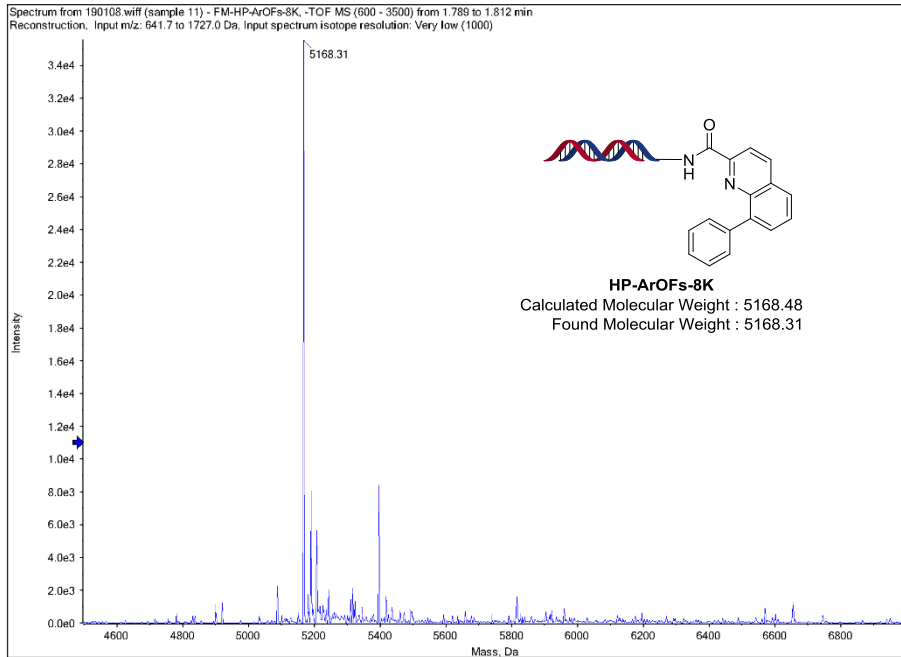




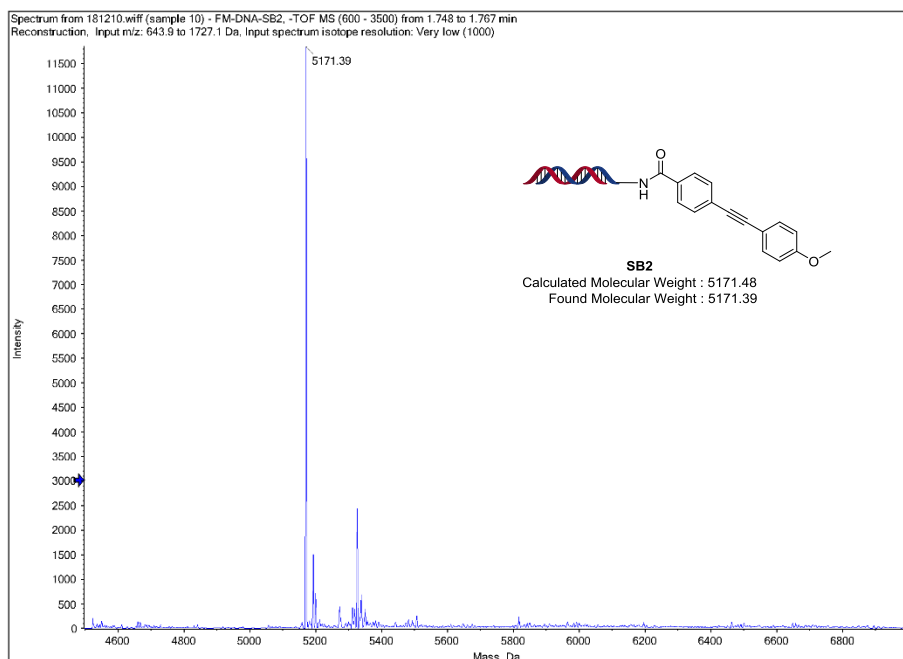
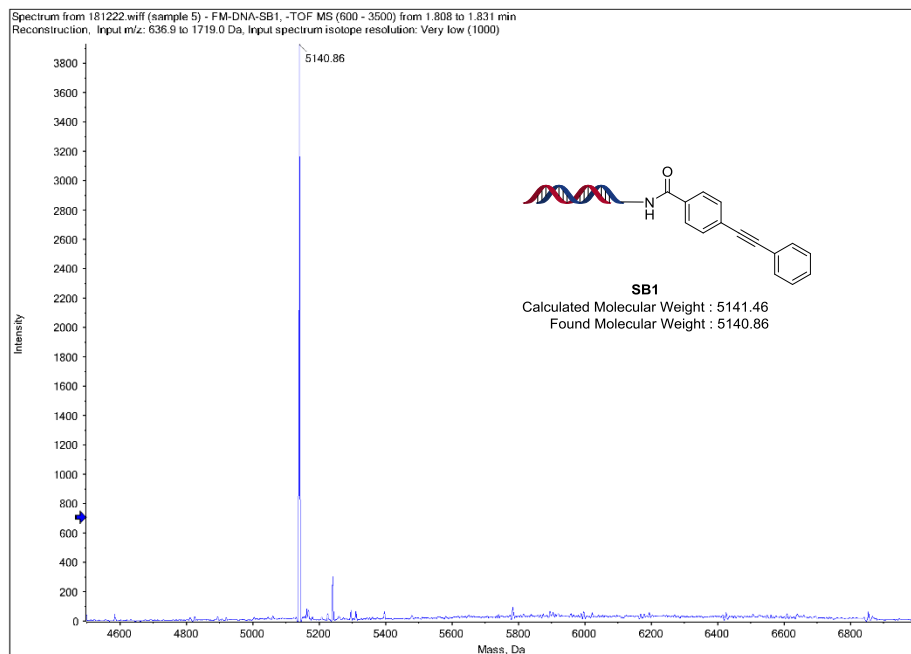


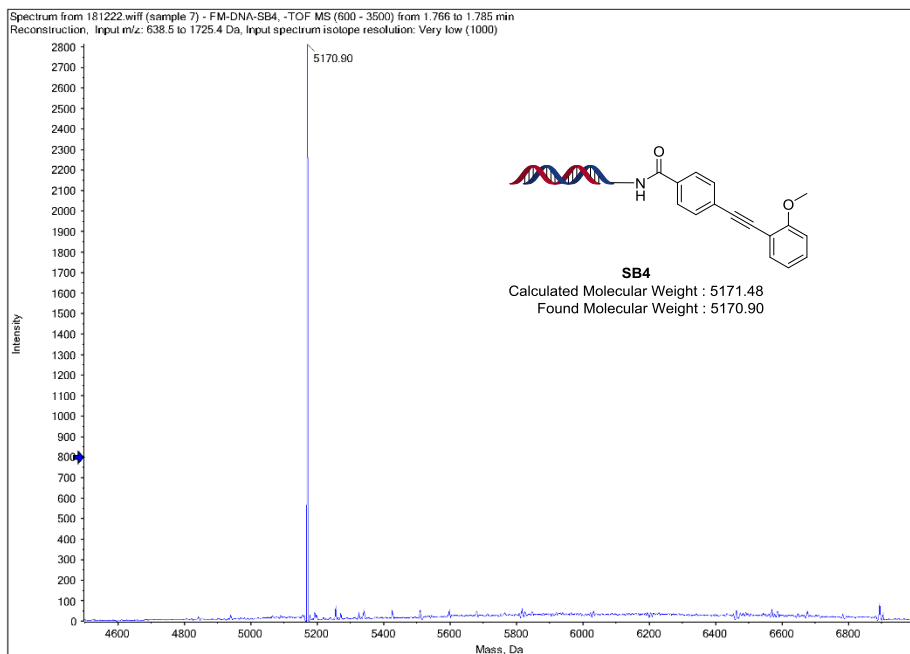
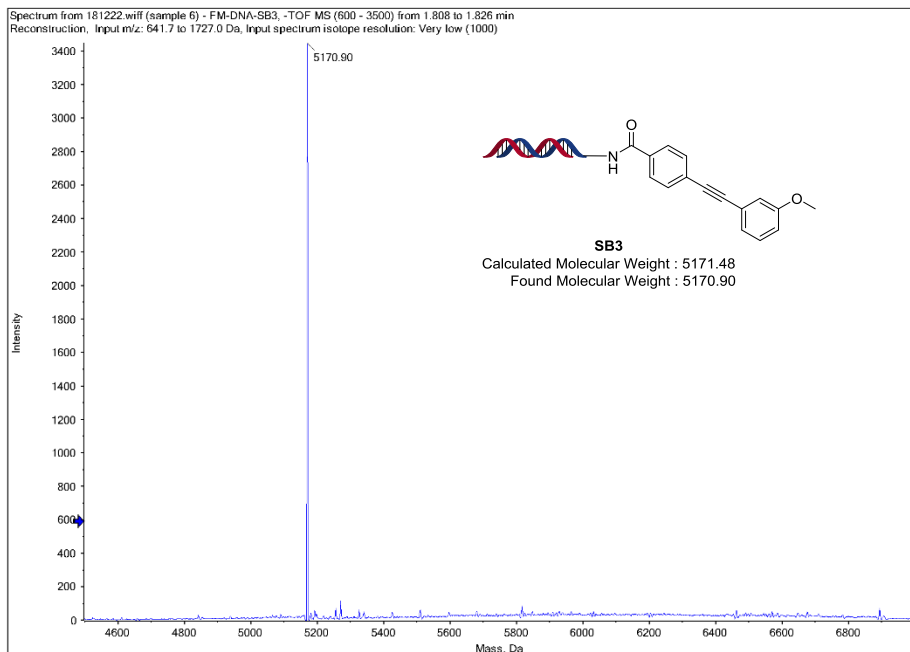


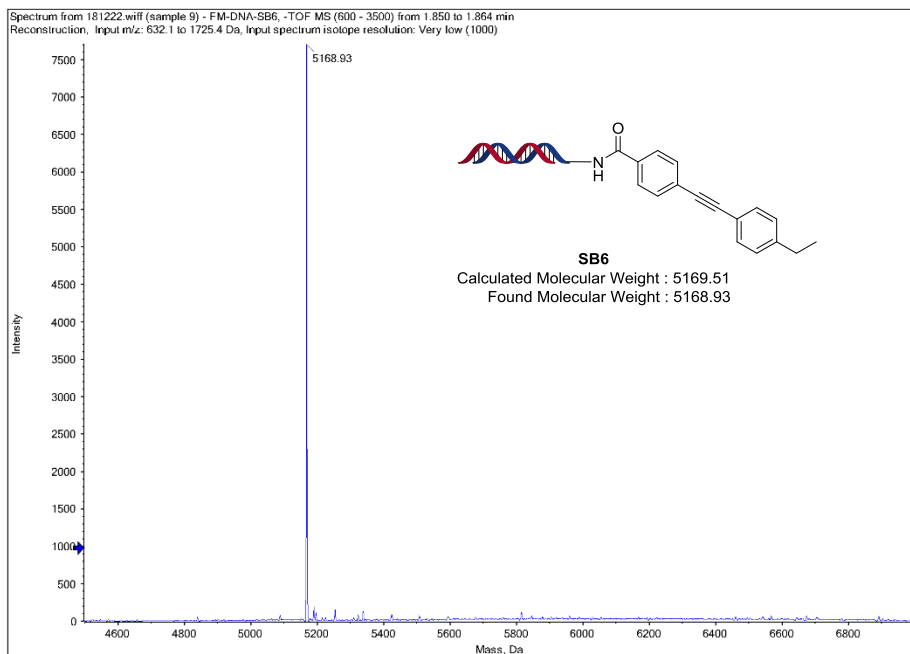
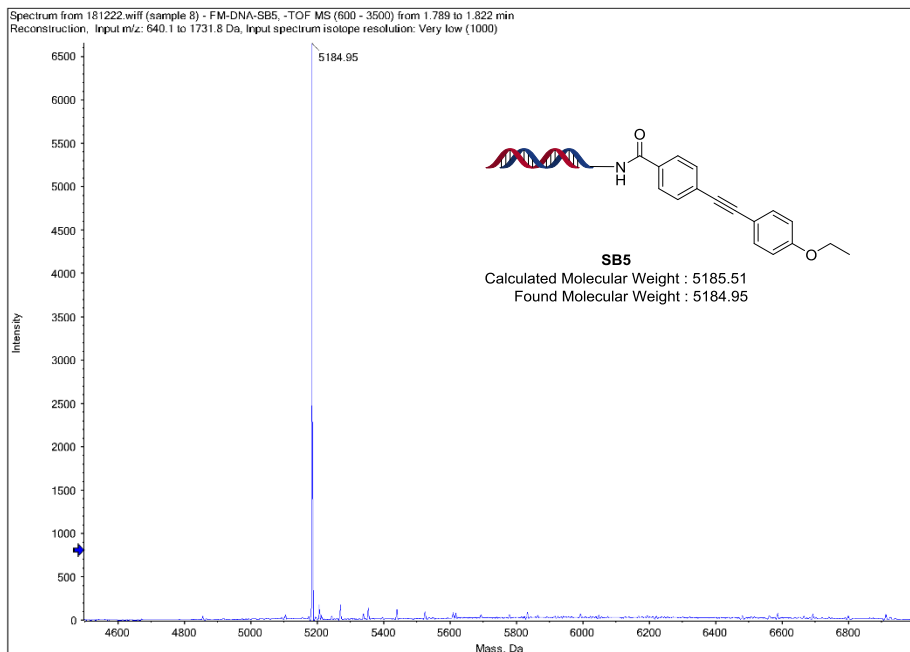


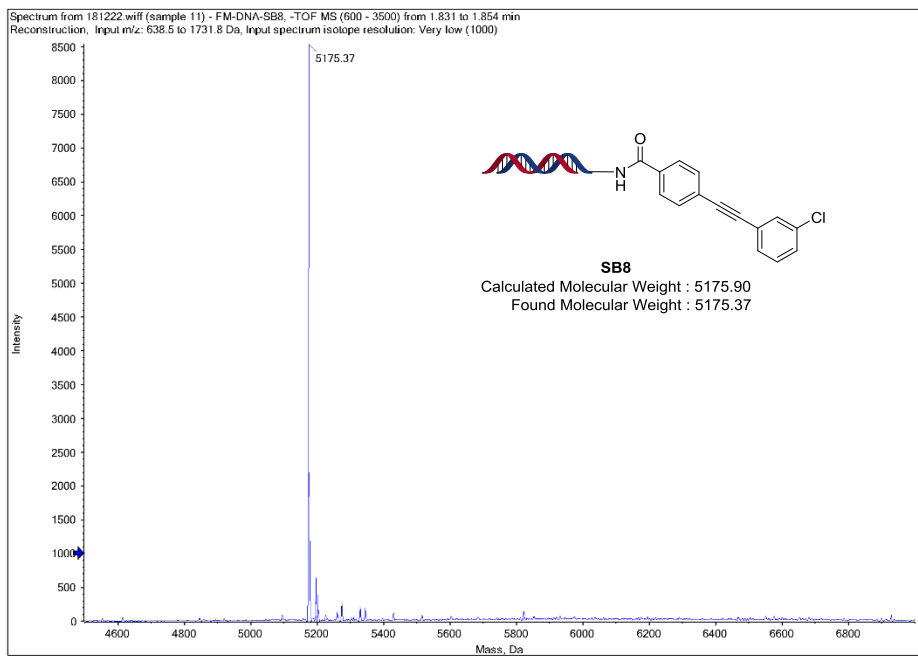
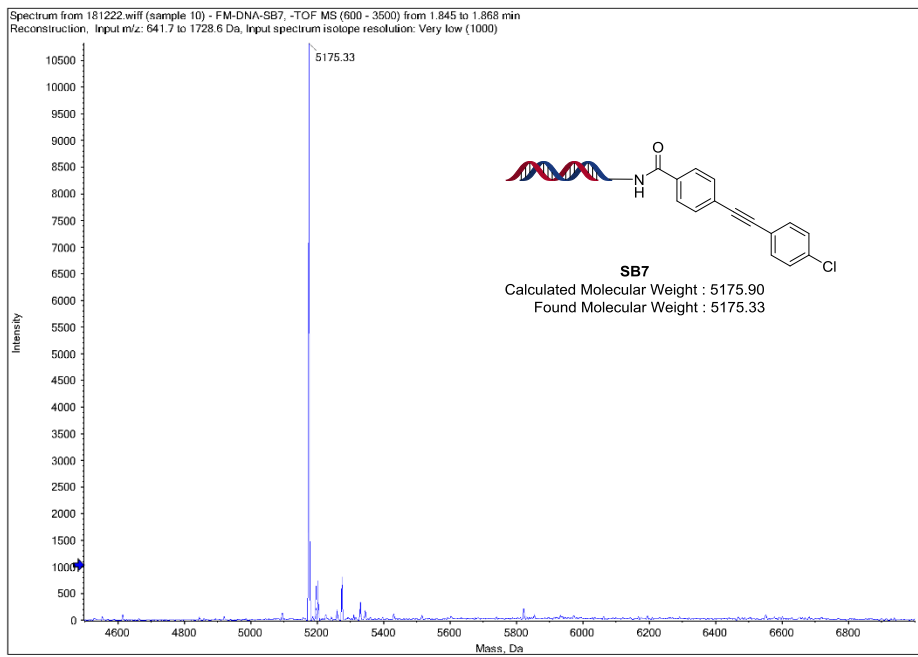


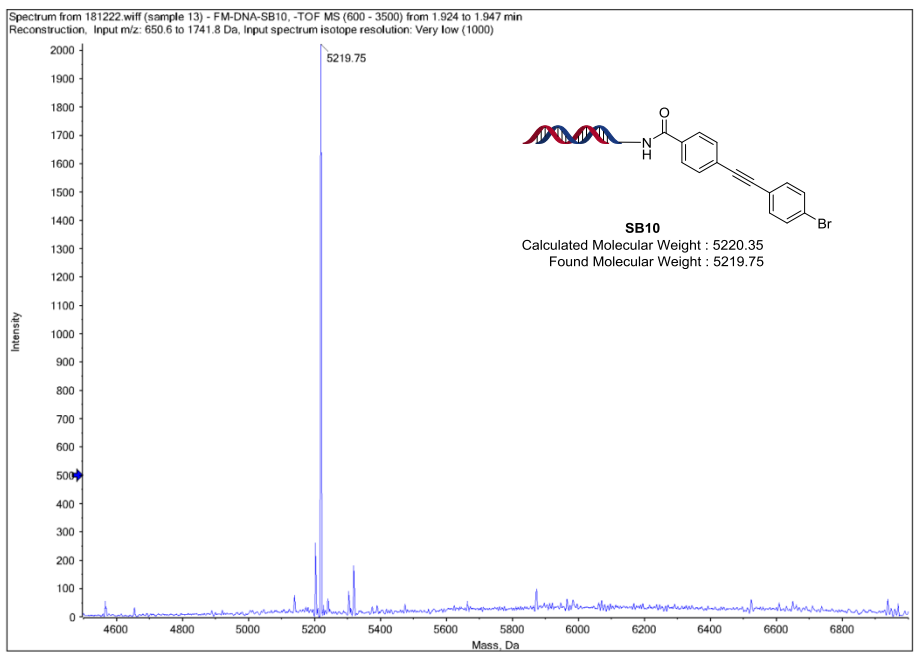
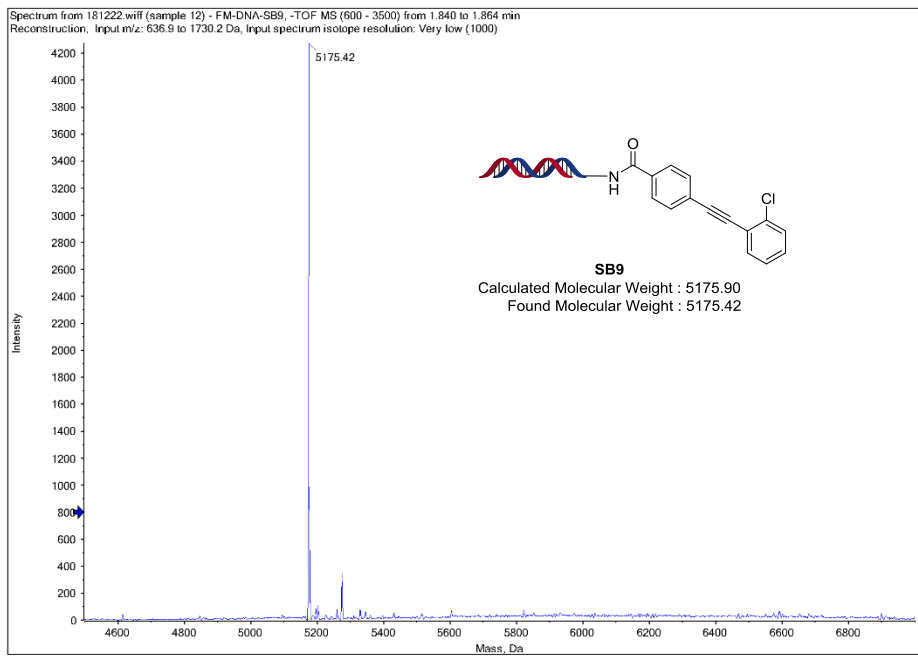
MS Spectra of On-DNA Sonogashira coupling products

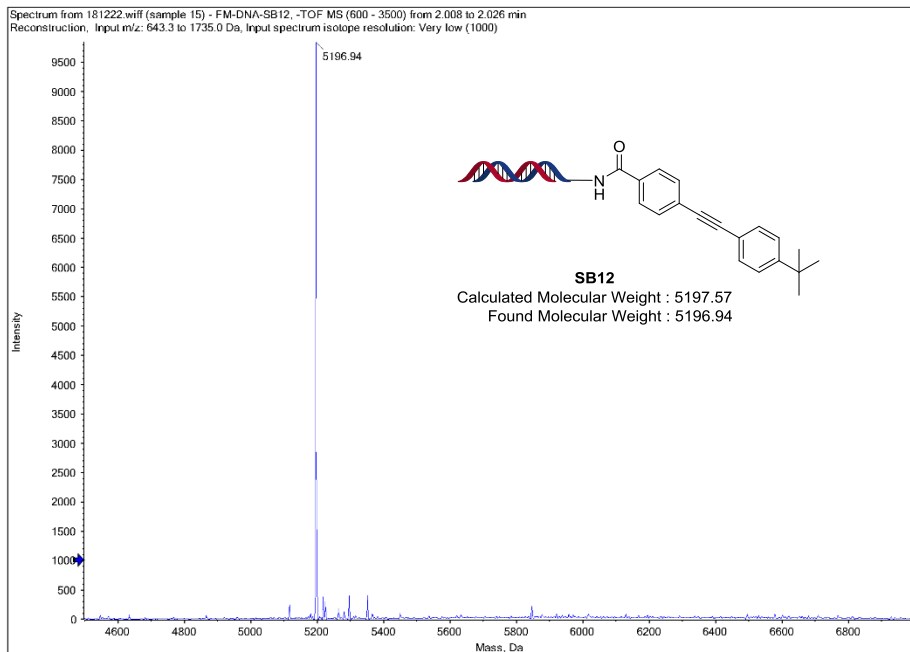
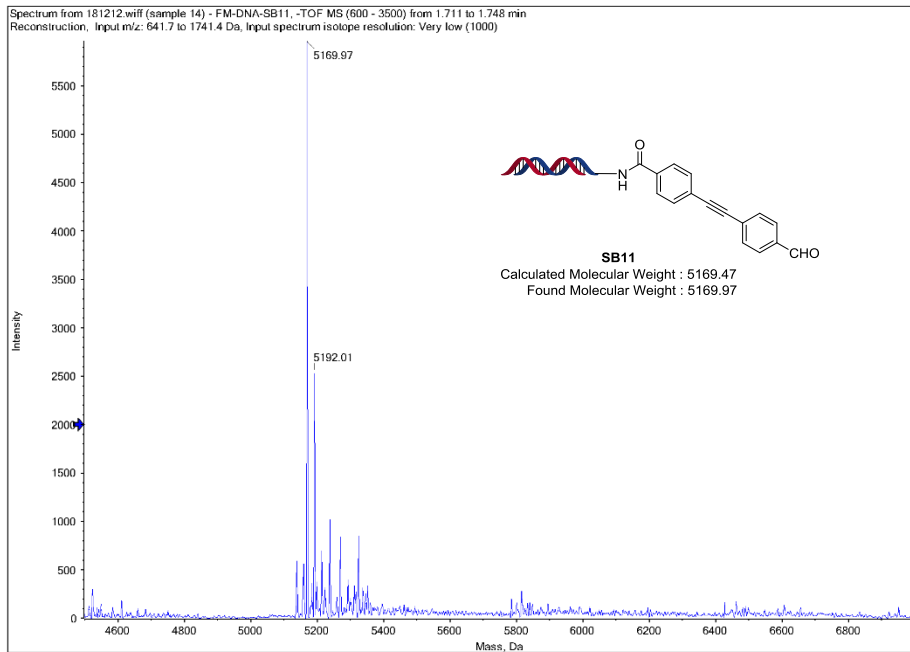


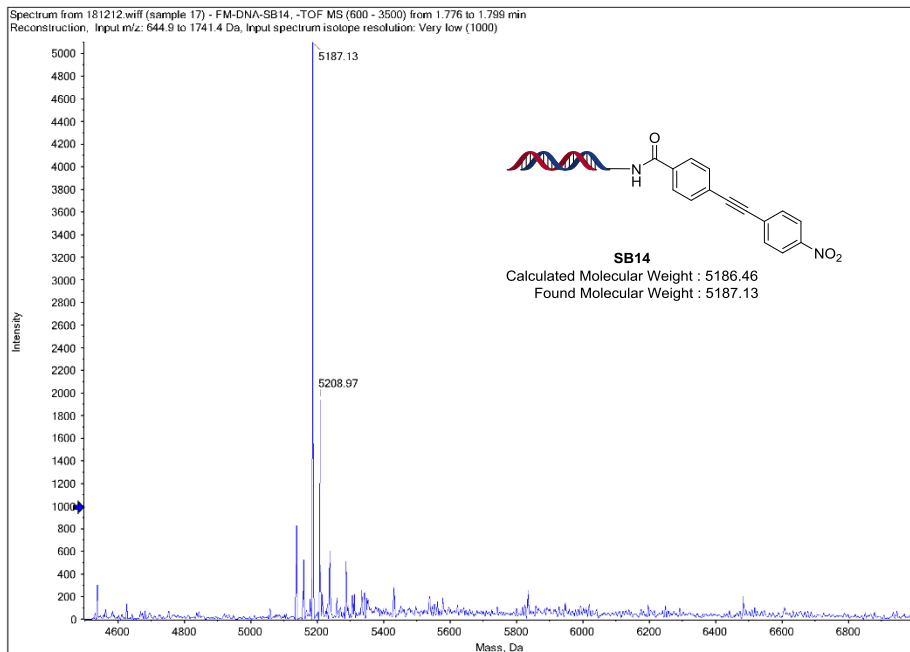
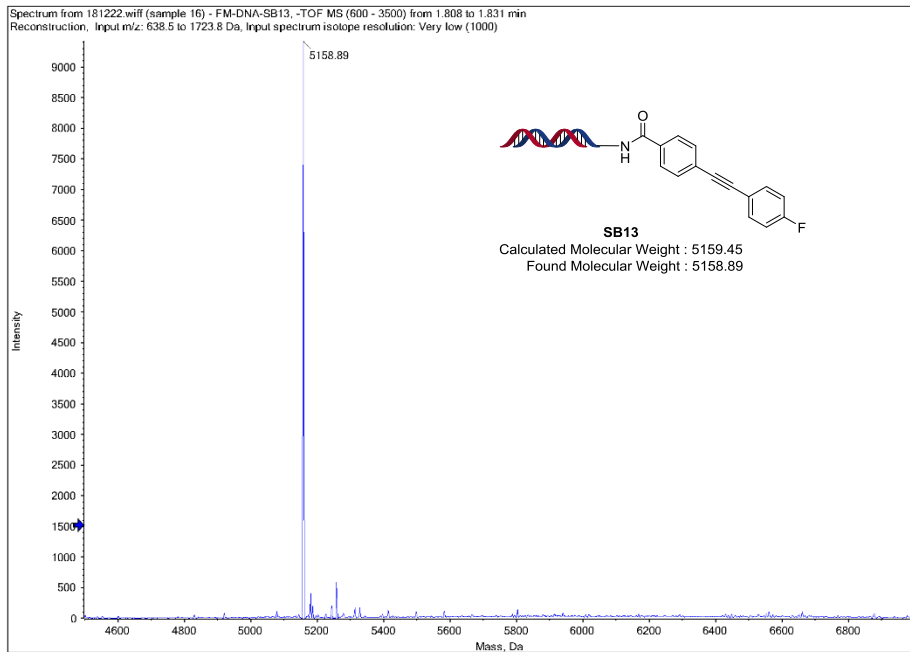


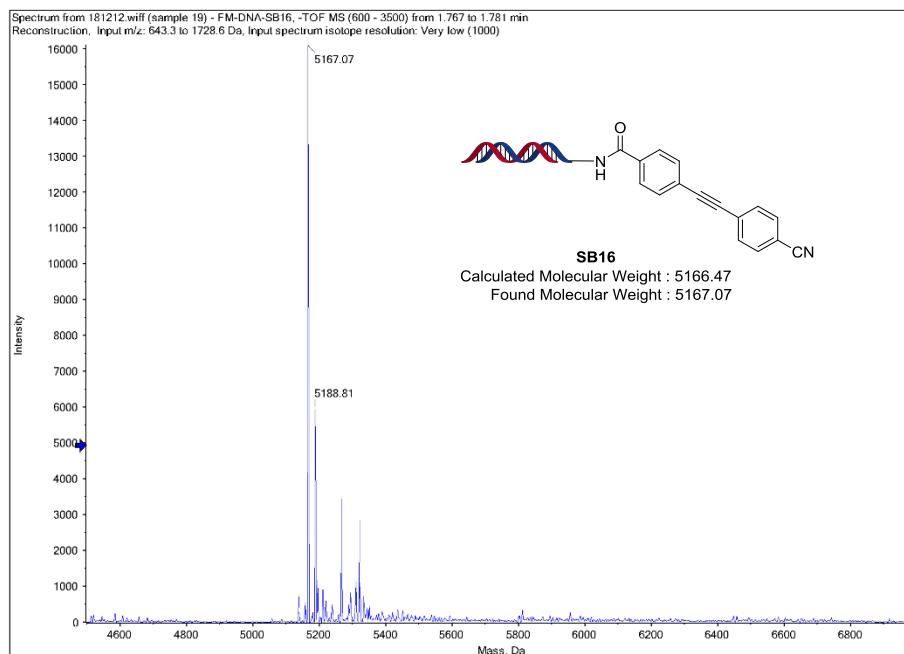
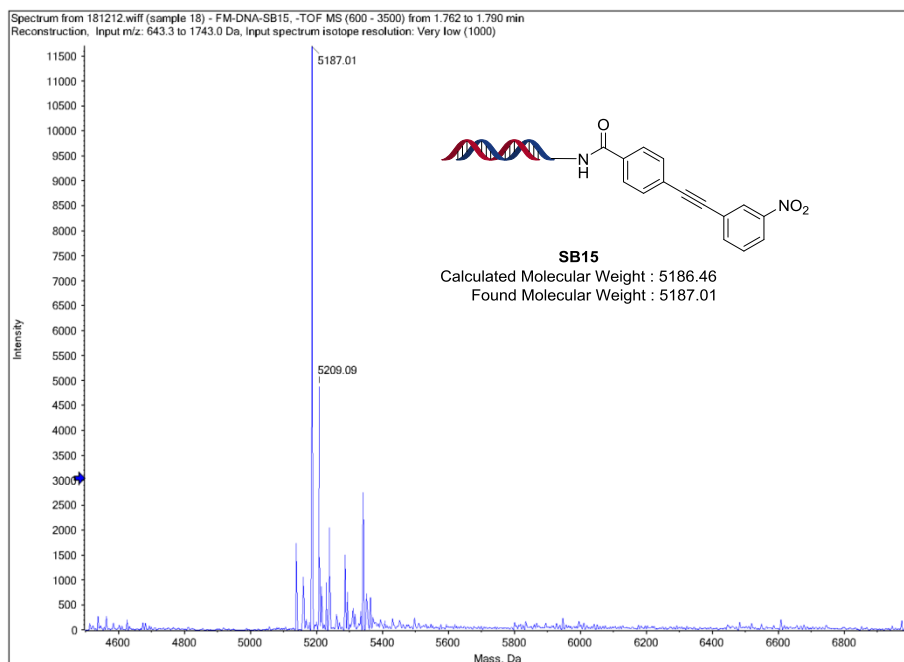


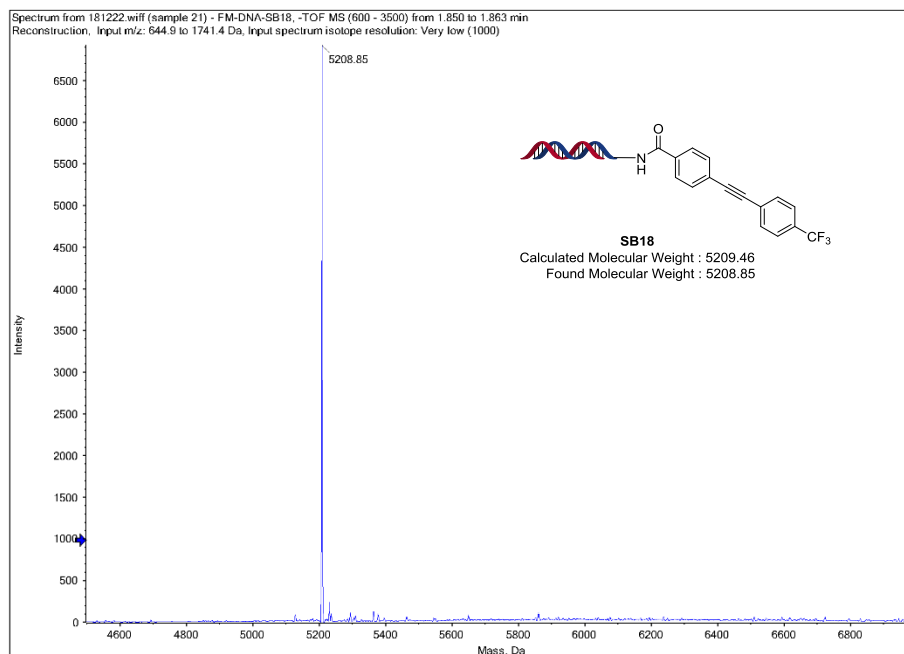
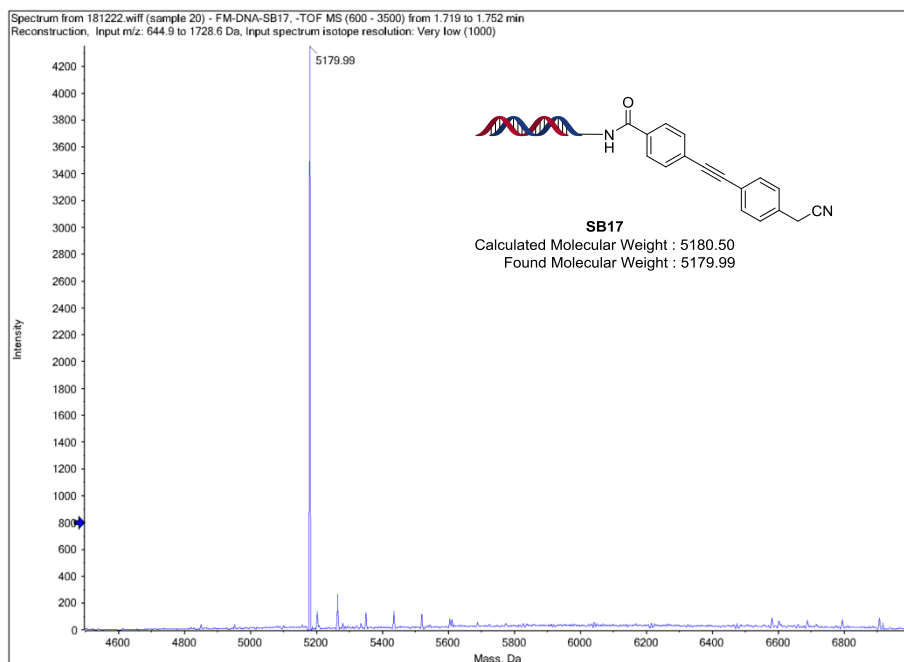


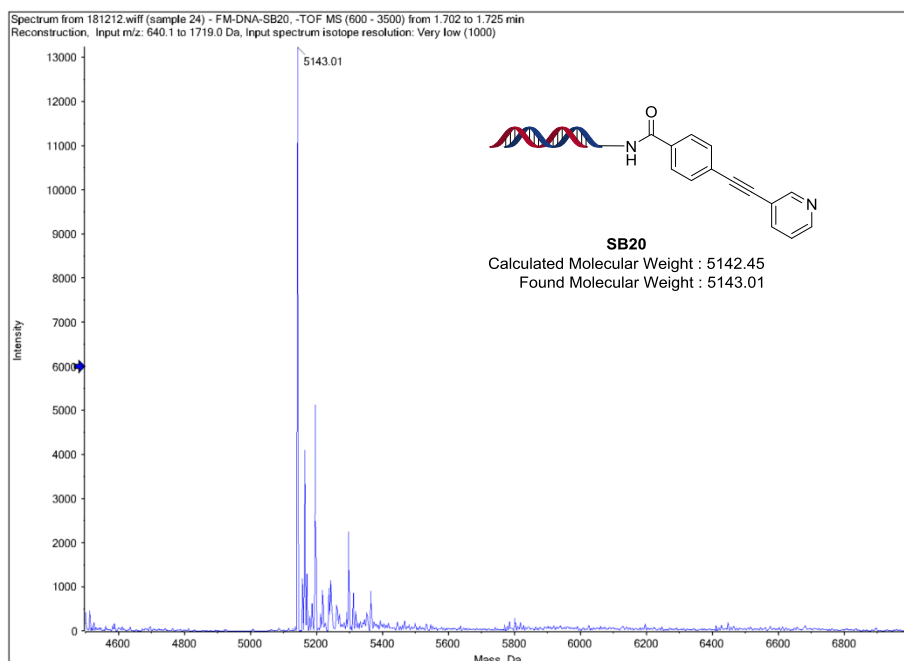
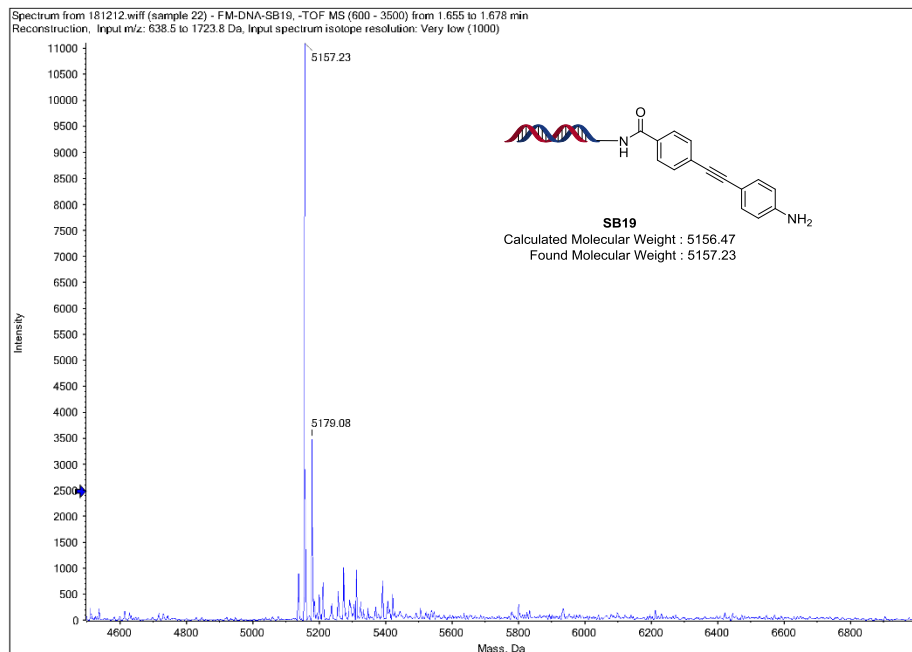


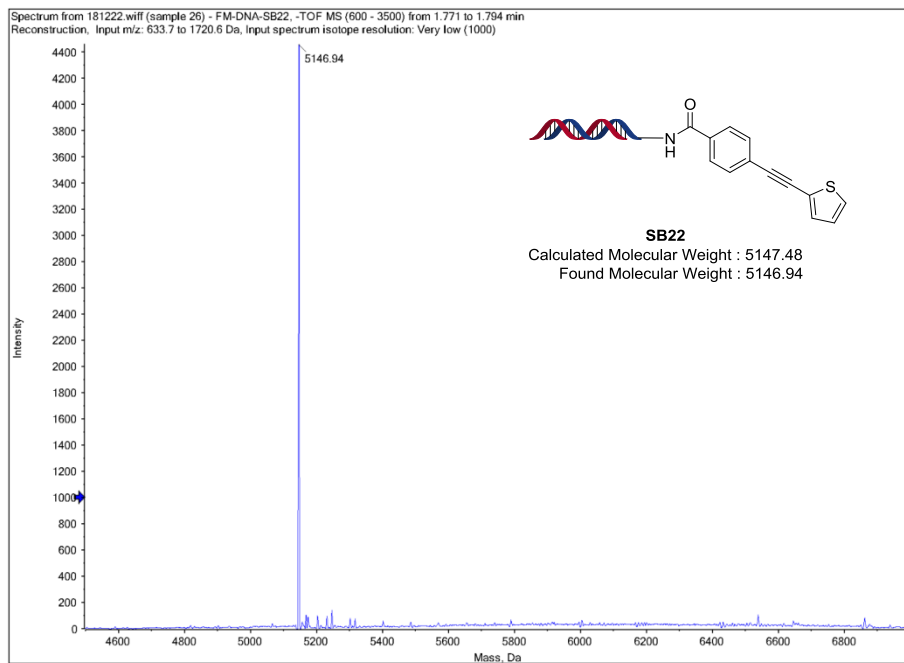
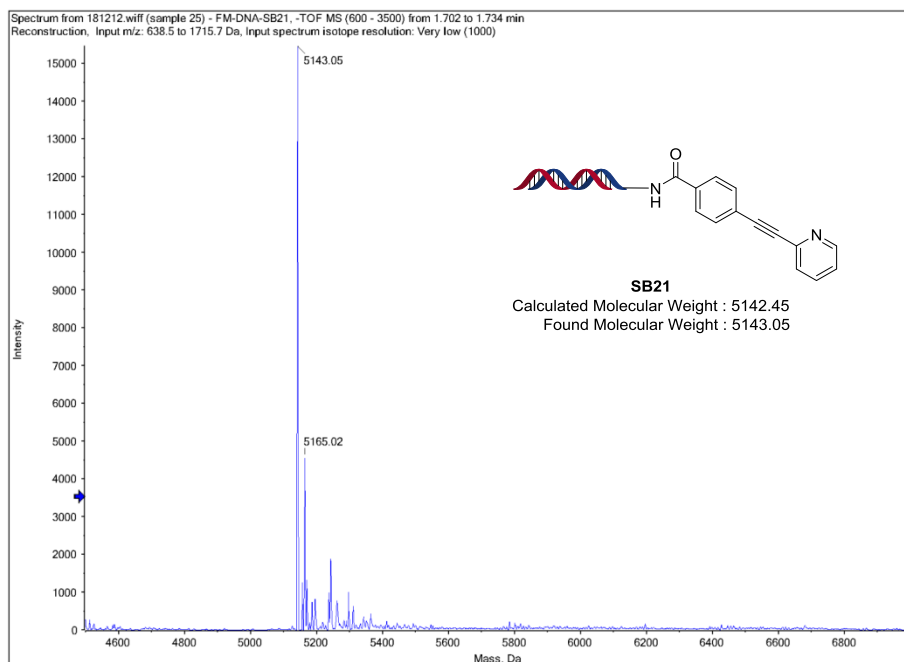


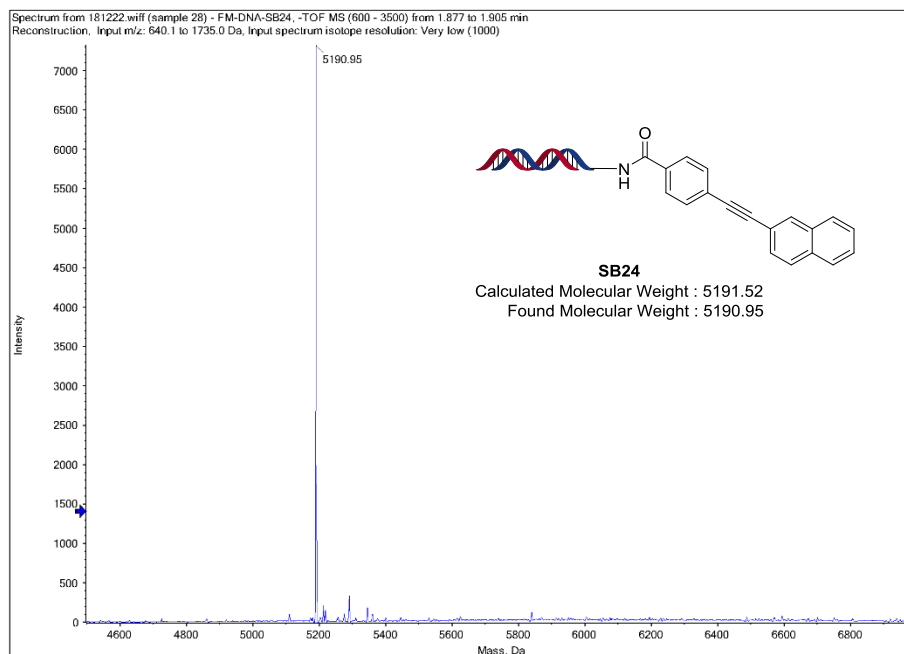
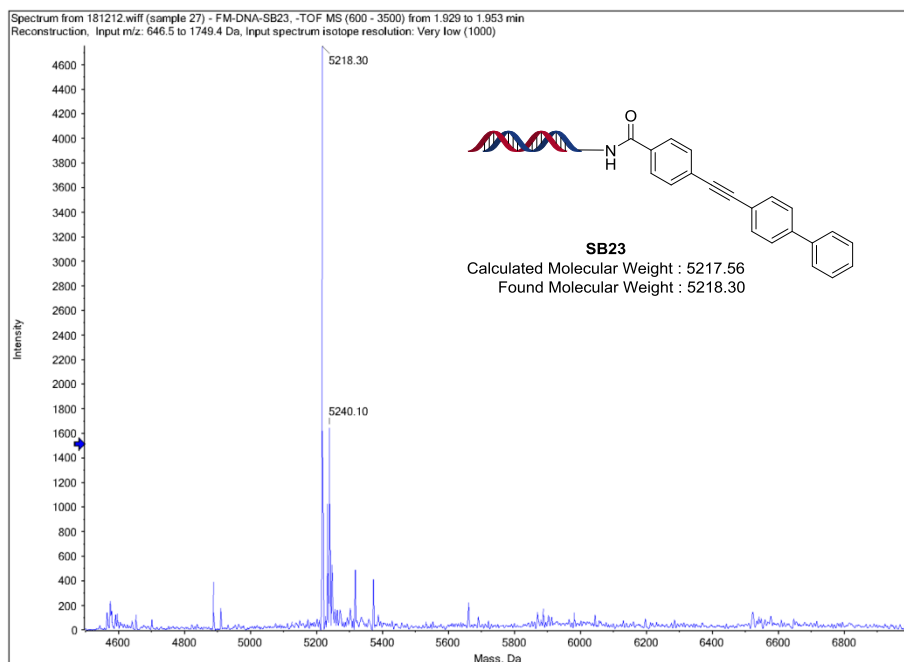


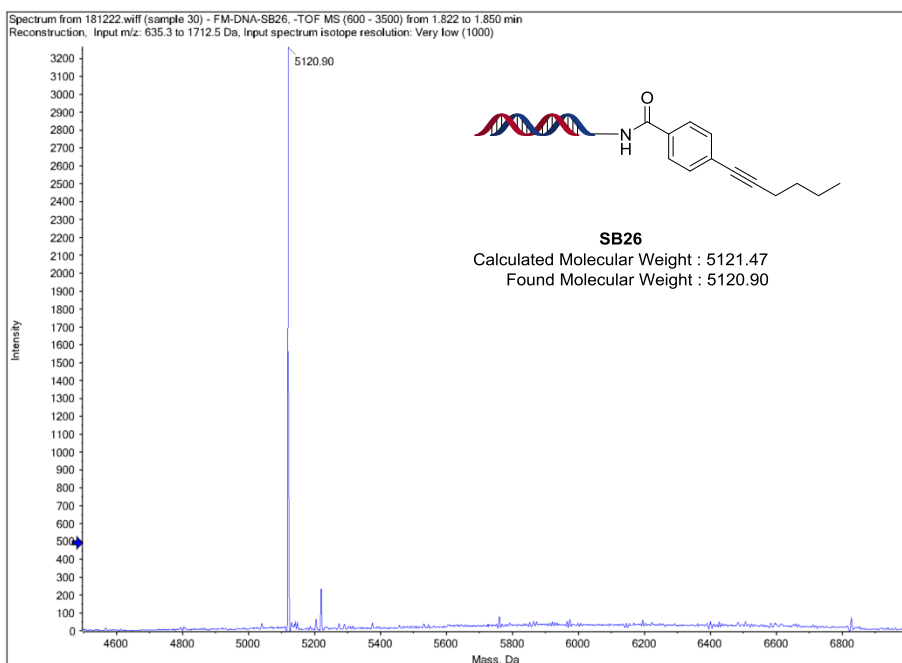
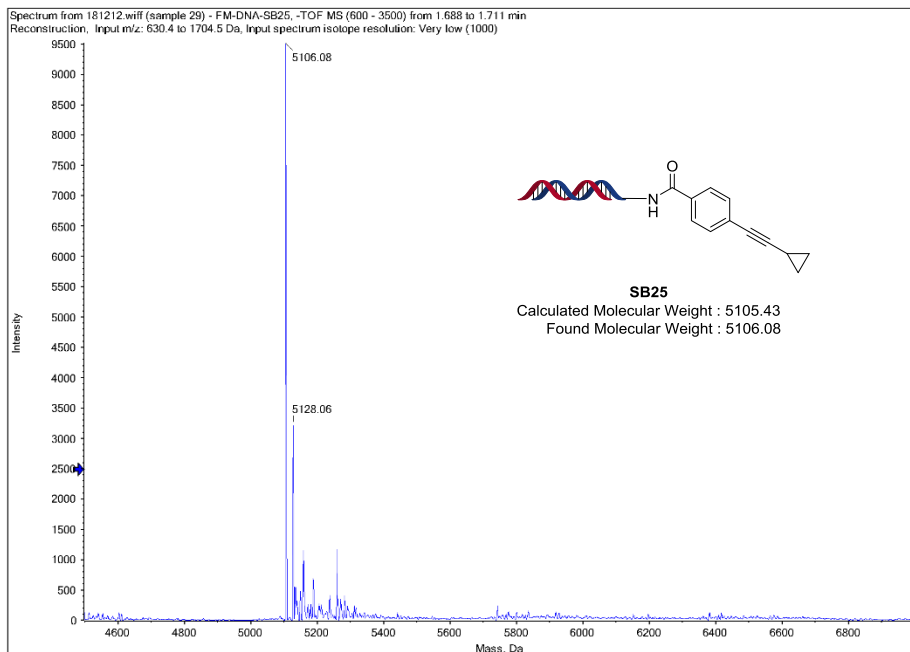


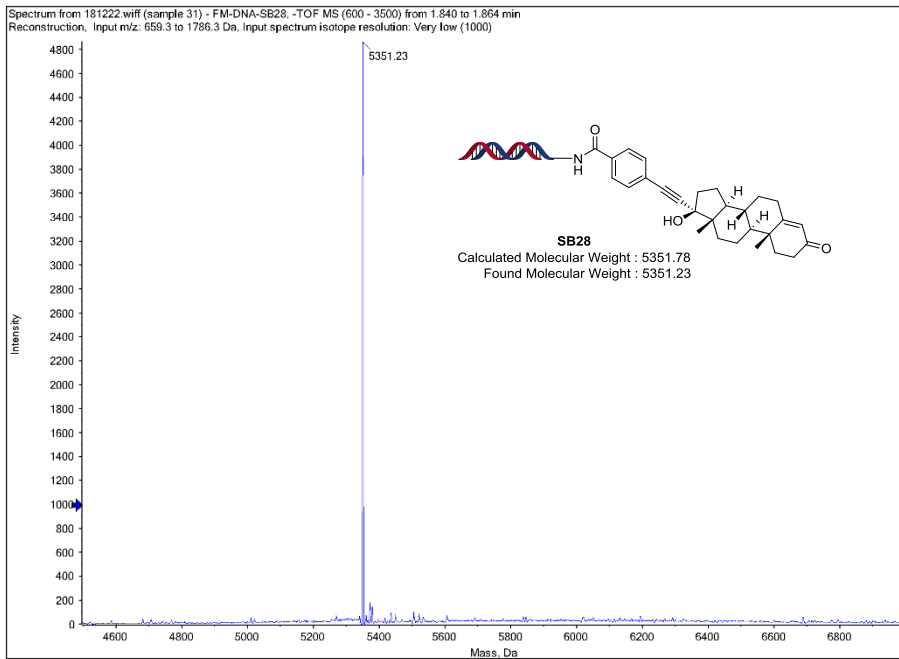
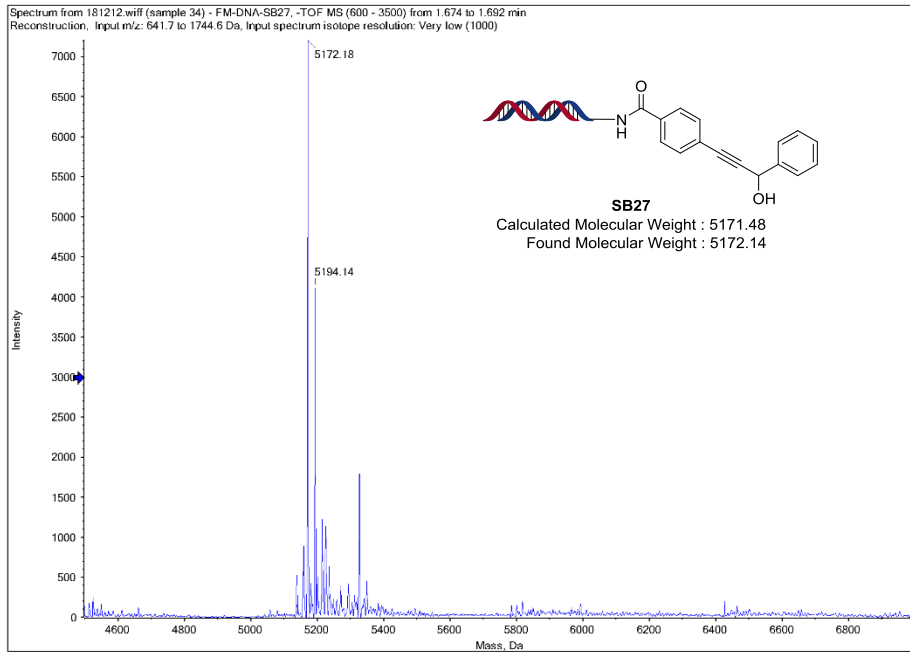


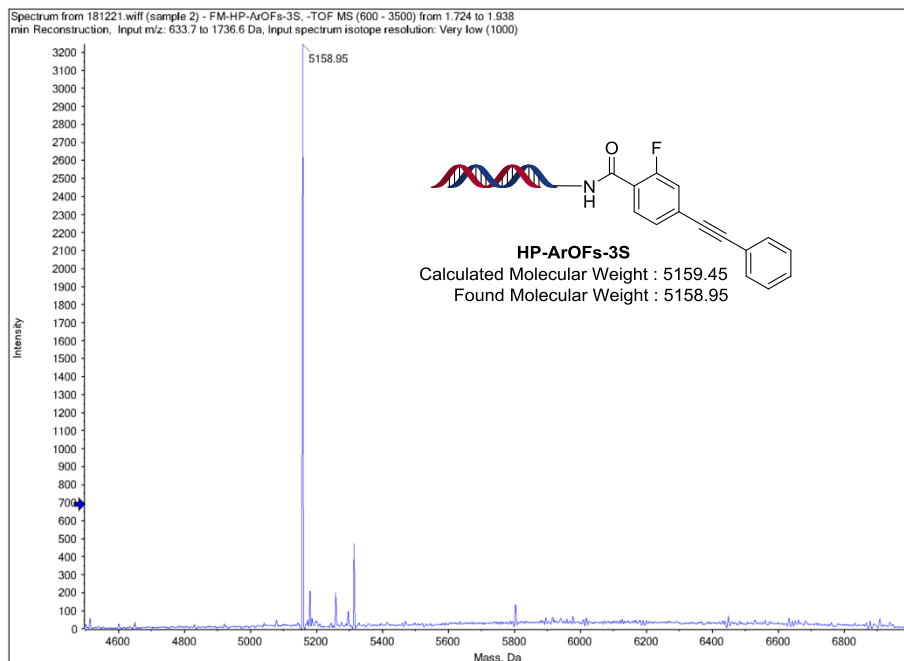
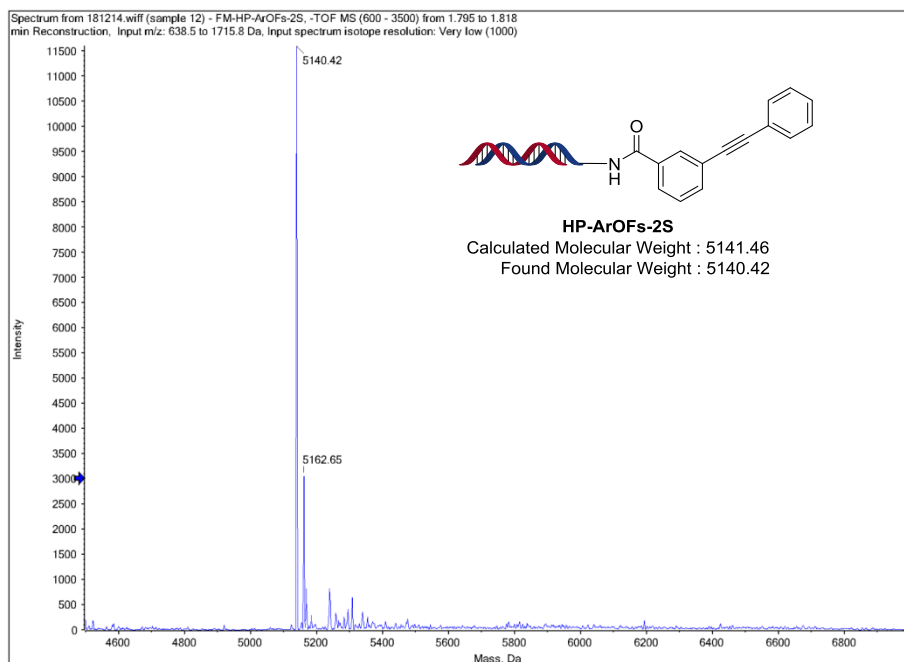


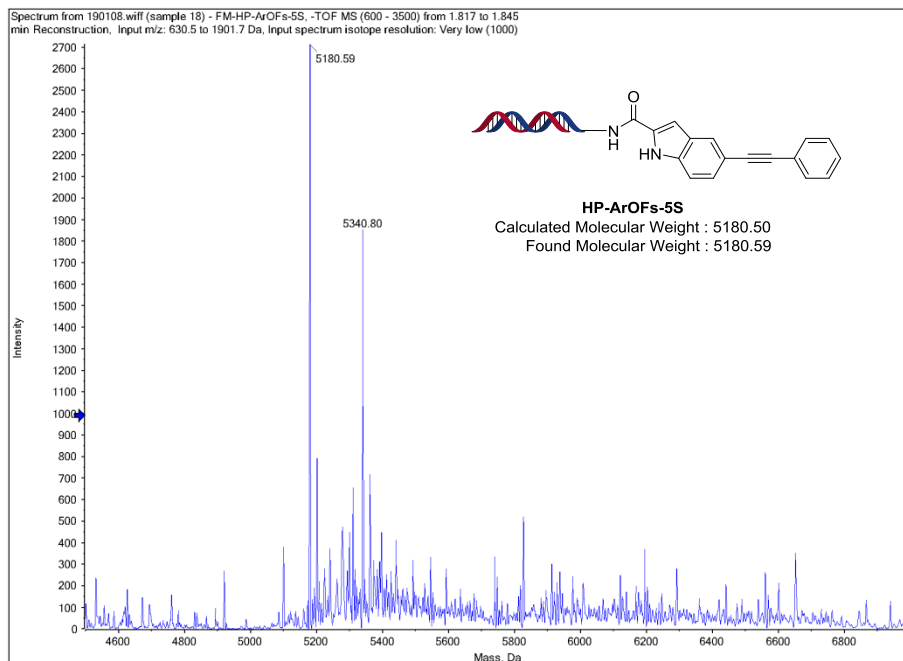
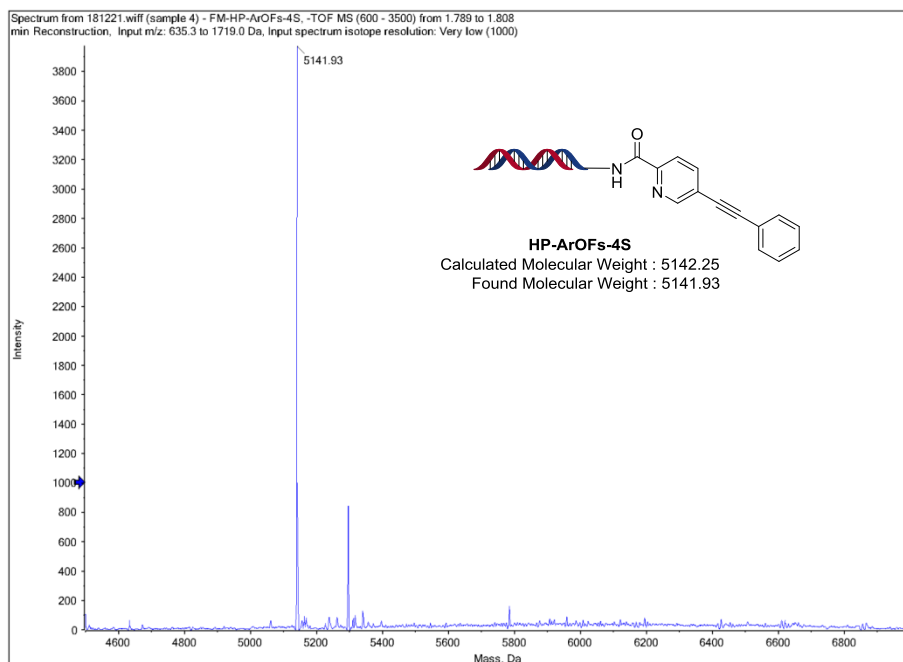


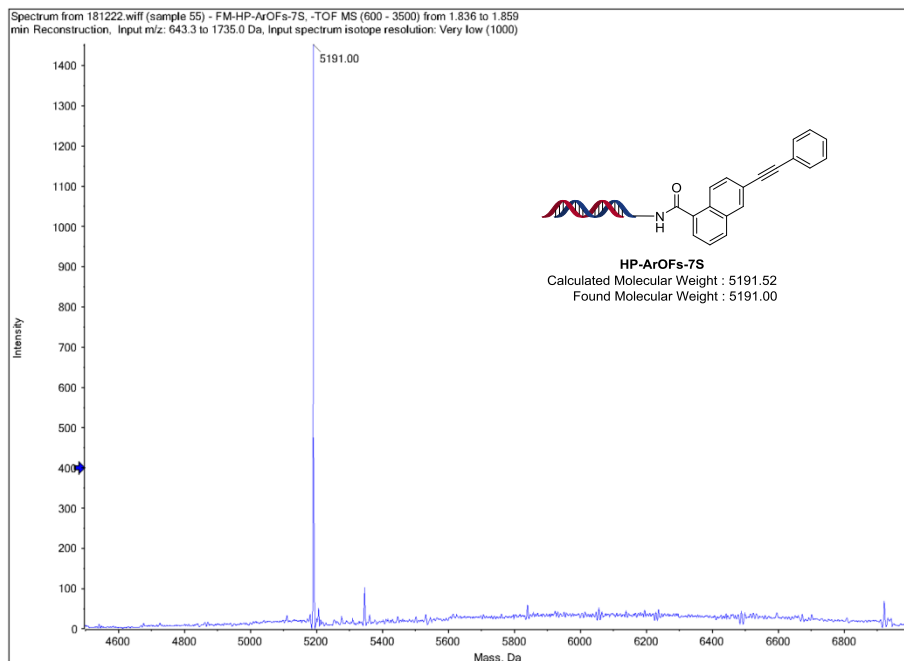
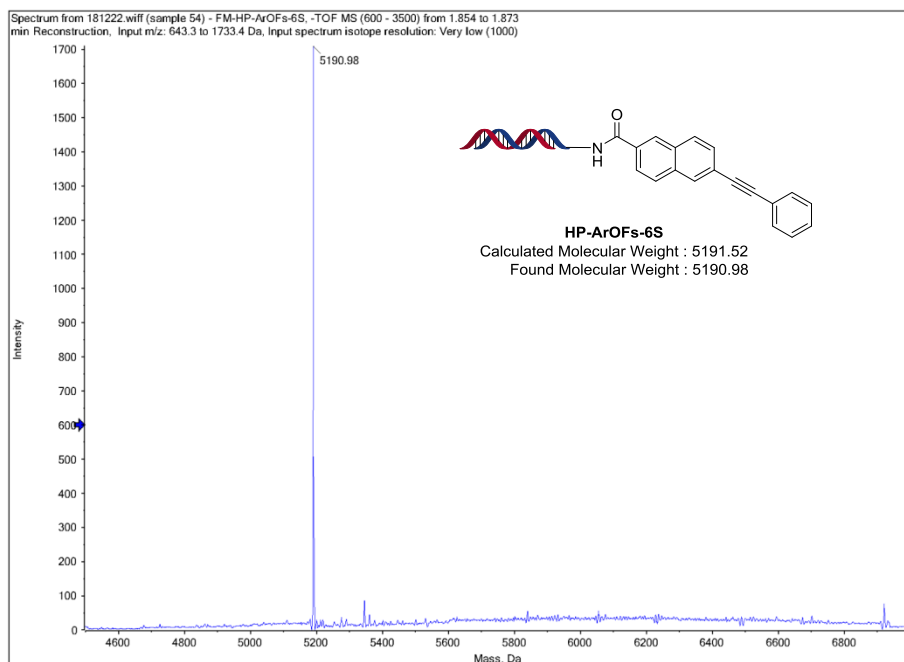


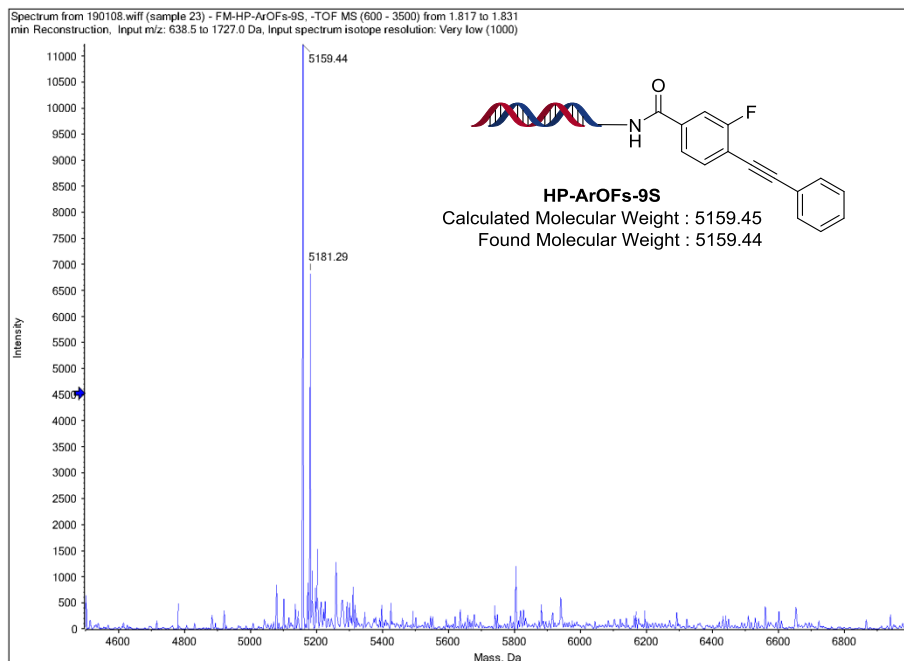
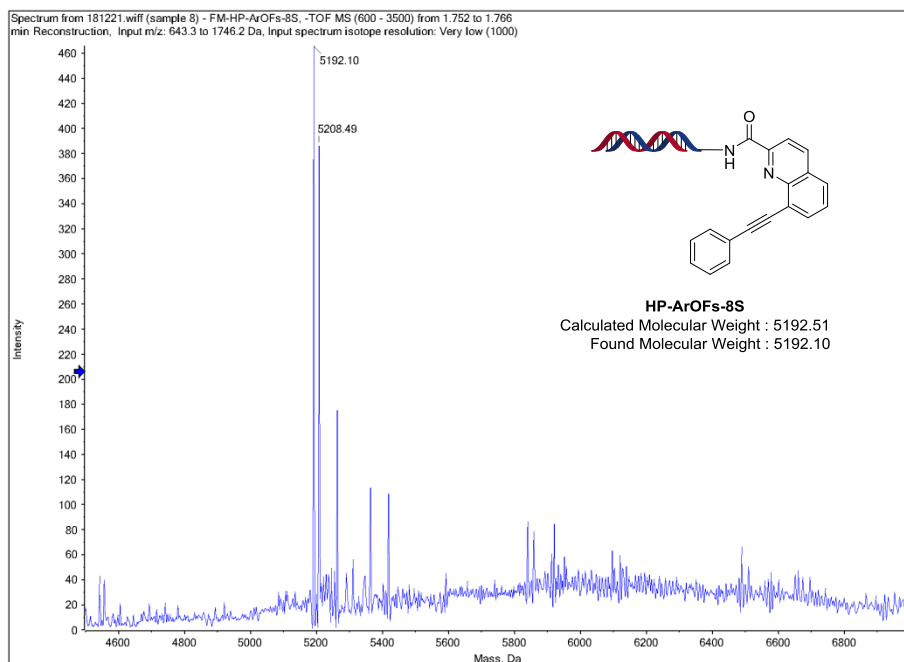












MS Spectra of On-DNA Buchwald amination products

