β-catenin activation down-regulates cell-cell junction-related genes and induces epithelial-to-mesenchymal transition in colorectal cancers

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Supplementary Information

Supplementary Figure S1. qPCR analysis of Wnt/β-catenin signaling target genes, *CD44*, *CCND1*, *MMP7*, *BMP4*, *AXIN2*, and *CCNE2* in HCT116-P, HCT116-WT, and HCT116-MT cells. Error bars represent the standard deviation (SD) of the mean of two independent experiments.



Supplementary Figure S2. β -catenin activation promotes cell cycle upregulation and cell proliferation. **(a)** Cell proliferation assay performed on HCT116 cells containing a wild-type (WT) β -catenin allele (HCT116-WT, WT), a mutant β -catenin allele (HCT116-MT, MT), or both a wild-type and mutant allele (HCT116-P, Parent). Cells were seeded at a density of 3 × 10⁶ and counted manually 2 and 4 days after seeding. **(b)** Propidium iodide flow cytometric cell cycle analysis of all three β -catenin HCT116 cell lines.

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Major EMT factor ormalized to GAPDH

(normalized to GAPDH)				
Gene	F.C. (MT/WT)			
TWIST1	1.260202			
SNAI1	1.227645			
SNAI2	2.114922			
ZEB1	2.107306			

b

EMT hallmark Gene F.C. (MT/WT) LAMA3 -3.48248 LAMC2 -2.67062 SCG2 -2.35741 RGS4 -2.30539 ID2 2.27408 MATN2 -1.96483 CD44 -1.90887 EFEMP2 -1.73006 FUCA1 -1.72227 GADD45A -1.64045 PLAUR -1.6369 ACTA2 -1.60321 VEGFA -1.58174 PLOD3 -1.56025 DPYSL3 -1.53545 -1.52886 CYR61 TNFAIP3 1.509536 IL8 1.523821 RHOB 1.543268 FERMT2 1.546939 COL6A3 1.578306 TGM2 1.618568 SPP1 1.698225 DKK1 1.821872 CAP2 2.282853 VCAN 2.330703 MSX1 3.179599 VIM 7.576687

Supplementary Figure S3. Identification of EMT-related genes showing differential expression in HCT116-MT cells, compared to HCT116-WT cells in the microarray data (a) The expression of four main EMT factors (SNAI1, SNAI2, ZEB1, and TWIST1) was analyzed using microarray data by normalizing them to GAPDH expression. (b) 1,507 DEGs obtained from microarray data were compared to a set of 200 EMT-related genes obtained from KEGG database. Twenty-eight EMT-related genes were significantly changed in HCT116-MT cells, compared to HCT116-WT cells.



Supplementary Figure S4. HCT116-P, HCT116-MT, and HCT116-WT cells show distinct patterns of cell density-dependent β -catenin, Claudin-7, and E-cadherin mRNA expression. The mRNA expression levels of β -catenin, Claudin-7, and E-cadherin were measured by qRT-PCR in all three β -catenin HCT116 cell lines at low, moderate, and high cell density. Error bars represent the SD of the mean of three independent experiments. One-way analysis of variance (ANOVA) with a post-hoc test (Bonferroni) was performed to compare multiple means. **P* <0.01, ***P* <0.001, ****P* <0.0001. Statistical significance between low (L) and moderate (M) cell densities, and M and high (H) cell densities is shown. Statistical significance between L and M or M and H.



Supplementary Figure S5. Wnt3a treatment induced translocation of β -catenin and EMT transcription factors into the nucleus, as well as loss of E-cadherin and Claudin-7, accompanying transcriptional activation of downstream target genes of Wnt/ β -catenin signaling in HCT116-WT cells. (a) Immunofluorescence microscopy analysis of HCT116-WT cells with or without 2-hour Wnt3a treatment. (b) Western blotting analysis of β -catenin, SNAIL, SLUG, ZEB1, TWIST1, E-cadherin, and Claudin-7 using the nuclear fraction of HCT116-WT cells treated with Wnt3a for 2 and 4 hours. (c) qPCR analysis of *MMP7*, *CD44*, *AXIN2*, and *CCND1* in HCT116-WT cells treated with Wnt3a for 2 and 4 hours. Error bars represent the standard deviation (SD) of the mean of two independent experiments.



Supplementary Figure S6. Expression of nuclear β-catenin, Claudin-7, and E-cadherin in colorectal cancer (CRC) tissues. **(a)** Representative immunohistochemical analysis of CRC tissues with negative, low, or high nuclear β-catenin expression. Nuclear β-catenin expression was classified as negative (detected 0% of tumor cells), low (detected in 1–29% of tumor cells), or high (detected in ≥30% of tumor cells). **(b)** Proportion of the three nuclear β-catenin expression groups identified in our cohort of 101 CRC tumor samples. **(c)** Representative immunohistochemical analysis of CRC tissues with low or high expression of Claudin-7 and E cadherin. If the H-score (see Materials and Methods) of Claudin-7 or E cadherin was >200, the sample was classified as high expression (right, upper and lower), and if the H-score ≤200, the sample was classified as low expression (right, upper and lower). **(d)** The mean percentage of nuclear β-catenin-staining in CRC samples that displayed metastasis after operation and in those that did not.



Supplementary Figure S7. Kaplan–Meier analysis of 101 CRC patients according to nuclear β -catenin expression. Using 101 CRCs, cases with no nuclear β -catenin expression (n = 33), cases with low nuclear β -catenin expression (n = 53), and cases with high nuclear β -catenin expression (n = 15) were subjected to the overall survival (P = 0.009) and disease-free survival (P = 0.134) analysis.



Supplementary Figure S8. Localization and nuclear expression of β-catenin, and Wnt pathway activity in cell lines with Wnt pathway related mutations or not. **(a,b)** Immunofluorescence microscopy and western blotting analysis of β-catenin in two APC mutant CRC cell lines (DLD-1 and LoVo), CRC cell lines with no Wnt pathway related mutations (RKO and HCT8), a hepatocellular carcinoma (HCC) cell line with *AXIN1* mutation (Hep3B), a HCC cell line with *CTNNB1* mutation (HepG2), and a CRC cell line with *CTNNB1* mutation (LS174T). Nuclear fraction of cell lysates was used for western blotting analysis. Black arrow indicates mutant β-catenin. **(c)** WNT pathway activity was measured by TOP/FOP luciferase reporter assay. Error bars represent the standard deviation (SD) of the mean of two independent experiments.



Supplementary Figure S9. Solid-pseudopapillary neoplasm (SPN) tissues show specific and strong nuclear β -catenin expression, accompanied by complete loss of both E-cadherin and Claudin-7 expression. Expression of β -catenin, E-cadherin, and Claudin-7 was measured by immunohistochemistry in 15 SPN samples. Representative protein-specific and hematoxylin and eosin staining (H&E) results are shown.



Supplementary Figure S10. Model describing the relationship between β -catenin activation status and EMT progression in CRCs.

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Gene	siRNA sequence
SNAI1	5'- GAUGCAGUUCCGCUCCAUU d(UU)-3'
SNAI2	5'- GAGCAAUCAAGCAGAUCAU d(UU)-3'
ZEB1	5'- CGCUCUGUUGAAGGCUGGA d(UU)-3'
TWIST1	5'- GTGTATGTGCGCCAAAGTA d(UU)-3'

Supplementary Table S1. siRNA sequences against the four EMT factors

Pathway analysis of down-regulated genes							
Category	Term	P-Value	Benjamini				
KEGG_PATHWAY	Antigen processing and presentation	5.00E-09	7.80E-07				
KEGG_PATHWAY	Cell adhesion molecules (CAMs)	1.00E-05	7.90E-04				
KEGG_PATHWAY	Type I diabetes mellitus	1.70E-05	8.80E-04				
KEGG_PATHWAY	Allograft rejection	3.00E-05	1.20E-03				
KEGG_PATHWAY	Graft-versus-host disease	5.90E-05	1.80E-03				
KEGG_PATHWAY	Lysosome	9.80E-05	2.50E-03				
KEGG_PATHWAY	Viral myocarditis	4.10E-04	9.10E-03				
KEGG_PATHWAY	Autoimmune thyroid disease	5.20E-04	1.00E-02				
KEGG_PATHWAY	Systemic lupus erythematosus	6.60E-04	1.10E-02				
KEGG_PATHWAY	Other glycan degradation	5.50E-03	8.20E-02				
KEGG_PATHWAY	Small cell lung cancer	4.30E-02	4.60E-01				
KEGG_PATHWAY	ECM-receptor interaction	4.30E-02	4.60E-01				
KEGG_PATHWAY	Asthma	4.50E-02	4.50E-01				
KEGG_PATHWAY	Tight junction	4.90E-02	4.50E-01				
Pathway analysis of up-reg	gulated genes						
Category	Term	P-Value	Benjamini				
KEGG_PATHWAY	Ribosome	9.20E-06	1.30E-03				
KEGG_PATHWAY	Spliceosome	1.50E-03	1.10E-01				
KEGG_PATHWAY	Cell cycle	4.50E-03	1.90E-01				
KEGG_PATHWAY	Purine metabolism	7.60E-03	2.40E-01				
KEGG_PATHWAY	DNA replication	8.10E-03	2.10E-01				
KEGG_PATHWAY	Limonene and pinene degradation	1.20E-02	2.50E-01				
KEGG_PATHWAY	Pancreatic cancer	1.30E-02	2.40E-01				
KEGG_PATHWAY	Pyrimidine metabolism	1.90E-02	2.90E-01				
KEGG_PATHWAY	Proteasome	2.40E-02	3.30E-01				
KEGG PATHWAY	Fatty acid elongation in mitochondria	3.00E-02	3.60E-01				

Supplementary Table S2. Pathway analysis of up- or down-regulated genes (|fold change|>1.5) in HCT116-MT cells compared to HCT116-WT cells

		β-catenin nuclear accumulation							
	-	Absent		Low		High		_ Total	p-value
		n = 33, 32.6%		n = 53, 52.5%		n = 15, 14.9%			
Age		60.18±11.15		57.64±11.13		61.07±11.52		101	0.449
Sav	Male	16	48.5%	27	50.9%	8	53.3%	51	0.948
Sex	Female	17	51.5%	26	49.1%	7	46.7%	50	
Levelier	Right side	17	51.5%	14	26.4%	5	33.3%	36	0.060
Location	Left side	16	48.5%	39	73.6%	10	66.7%	65	
MSI	MSS and MSI low	27	81.8%	52	98.1%	14	93.3%	93	0.013
	MSI high	6	18.2%	1	1.9%	1	6.7%	8	
Pre-operative CEA level	≤5ng/ml	27	81.8%	35	66.0%	11	73.3%	73	0.281
	>5ng/ml	6	18.2%	18	34.0%	4	26.7%	28	
Gross type	Exophytic	28	84.8%	38	71.7%	11	73.3%	77	0.364
	Non exophytic	5	15.2%	15	28.3%	4	26.7%	24	
0.	<5cm	12	36.4%	33	62.3%	5	33.3%	50	0.026
Size	≥5cm	21	63.6%	20	37.7%	10	66.7%	51	
Pathologic diagnosis	Adenocarcinoma	31	93.9%	51	96.2%	13	86.7%	95	0.288
	Mucinous adenocarcinoma	1	3.0%	1	1.9%	2	13.3%	4	
	Medullary carcinoma	1	3.0%	1	1.9%	0	0.0%	2	
Metastasis after operation	Absent	31	93.9%	46	86.8%	11	73.3%	88	0.142
	Present	2	6.1%	7	13.2%	4	26.7%	13	
E-cadherin	High expression	25	75.8%	39	73.6%	7	46.7%	71	0.093
	Low expression	8	24.2%	14	26.4%	8	53.3%	30	
Claudin7	High expression	20	60.6%	34	64.2%	7	46.7%	61	0.474
	Low expression	13	39.4%	19	35.8%	8	53.3%	40	

Supplementary Ta	able S3. The cl	inicopathologic f	features of 101	CRCs according	to nuclear	ß-catenin exp	ression status

Supplementary Information (Blots)

Fig. 1C

















Supplementary Fig. 8

