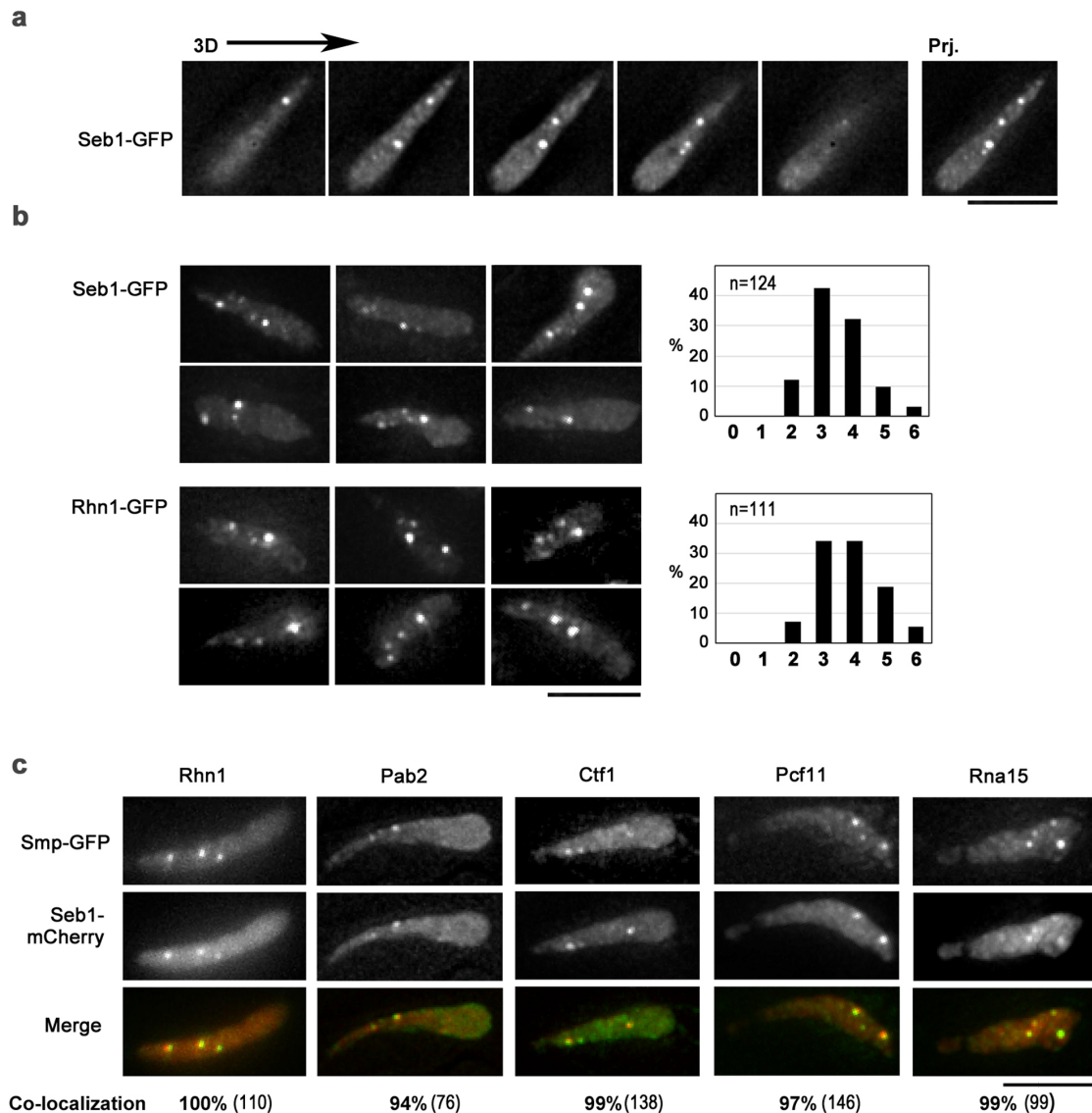


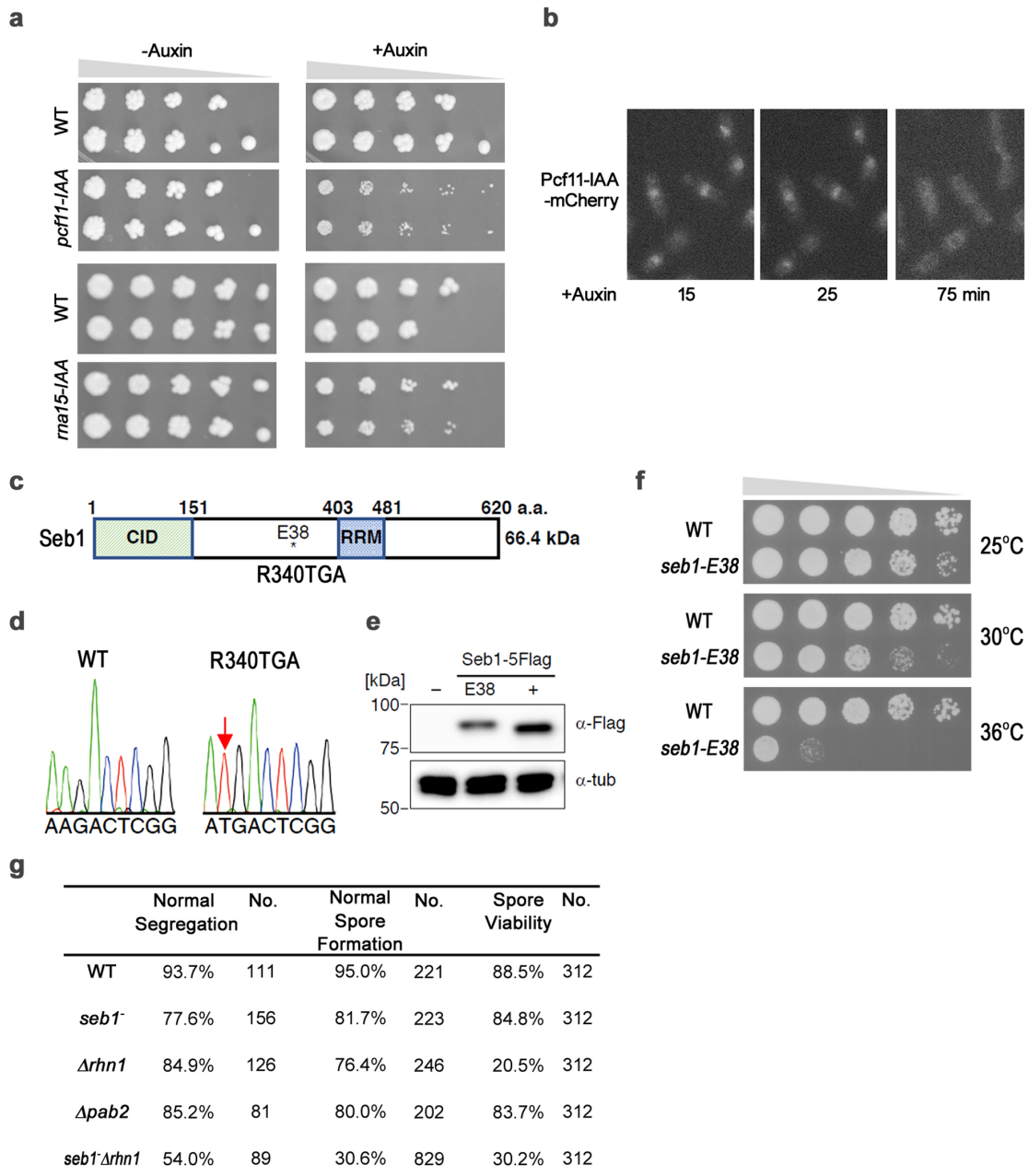
Chromosome-associated RNA-protein complexes promote pairing of homologous chromosomes during meiosis in *Schizosaccharomyces pombe*

Ding *et al.*



Supplementary Figure 1. Chromosomal accumulation of Smp proteins

(a) Optical section images of Seb1-GFP in the horsetail nucleus were acquired at a 0.35 μm focus interval. Optical section images were projected to produce the projection image (Prj). (b) Examples of projection images for Seb1-GFP and Rhn1-GFP (left). The number distribution of Seb1-GFP and Rhn1-GFP dots (right). The “dots” were identified using the 2D polygon finder function of softWoRx software (GE Healthcare); the fluorescence intensity threshold of a “dot” was set as 1.5 times higher than the average intensity in the nucleus. “n” in the bar graphs represents the number of horsetail nuclei examined. Source data are provided as a Source Data 6. (c) Co-localization of GFP-fused Smp proteins with Seb1-mCherry in the horsetail nucleus. The number below indicates percentage of Smp dots colocalized with Seb1 dots; the number in parentheses indicates the number of cells examined. Scale bars represent 5 μm (a-c).



Supplementary Figure 2. Mutant characterization

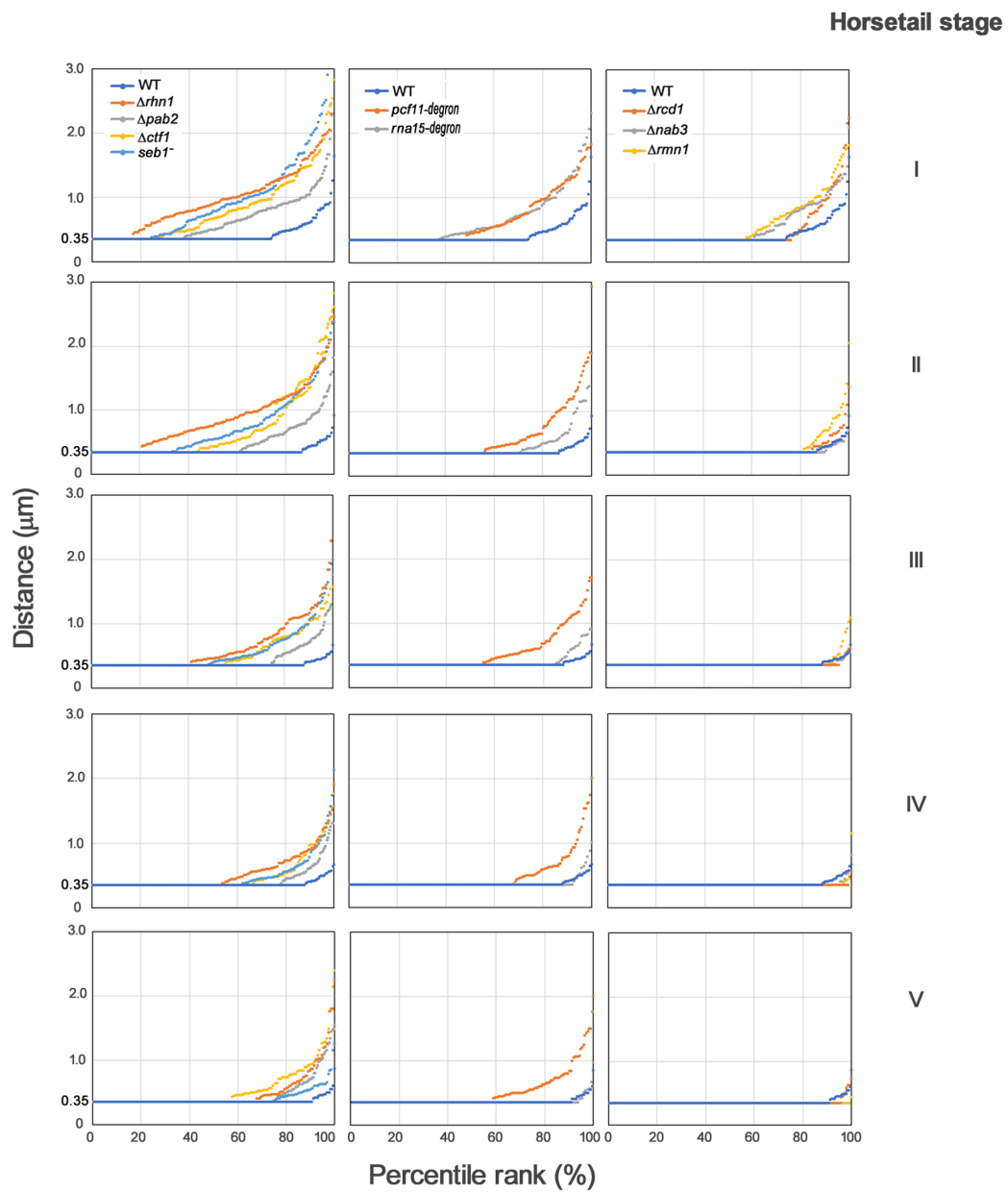
(a) Growth assay of Pcf11 and Rna15 degron strains (*pcf11-IAA*, *rna15-IAA*) in the culture medium with or without Auxin. The plates were incubated for 3 days at 30°C.

(b) An example of live cell imaging of Pcf11-IAA-mCherry degradation after adding Auxin and incubation at 26°C in EMM2-N medium. The fluorescent signals disappeared from the nuclei at 75 min after incubation in the medium containing Auxin.

(c) Diagram of Seb1 protein feature. CID: CTD-interacting domain. RRM: RNA recognition motif. *: E38 (R340TGA) mutation.

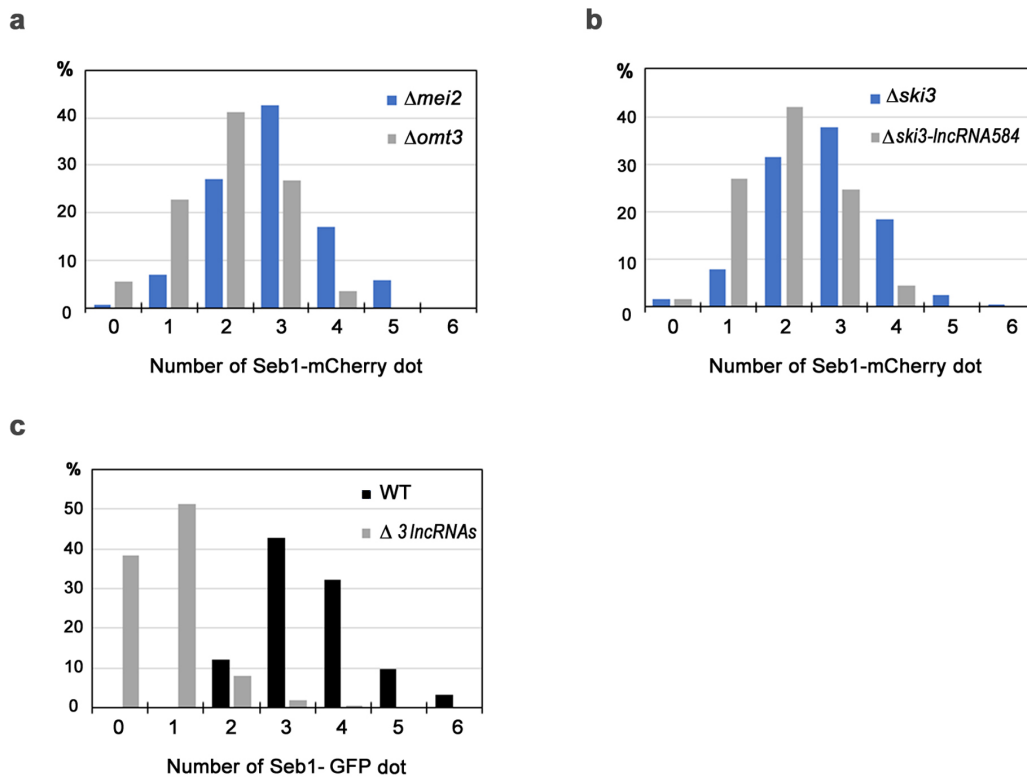
(d) Sequencing of *seb1-E38* mutation. The *E38* allele bears a nonsense mutation converting R340 to a termination codon (TGA). **(e)** Immunoblot analysis of the Seb1-

E38 mutant. α -Tubulin was used as a loading control. (-) No tag strain (FY648), (E38) Seb1-E38-5Flag (EON125), (+) Seb1-5Flag (EON06). Primary antibody: (α -Flag) anti-Flag M2 antibody (1/1000 dilution; A8592, Merck), (α -tub) anti- α -tubulin (1/1000 dilution; T5168, Merck). Secondary antibody: Peroxidase-conjugated AffiPure anti-mouse IgG (1/1000 dilution; 515-035-072, Jackson). The Seb1-E38 mutant strain expresses a reduced amount of the full-length Seb1 protein. Source data are provided as a Source Data file S2. **(f)** Seb1-E38 growth assay. Five-fold-diluted cultures of the indicated strains were plated on YEA medium plates and incubated for 2–3 days at the indicated temperature. **(g)** Meiotic chromosome segregation (after meiotic division II, using *cen2-lacO/lacI-GFP* as marker), spore formation and spore viability in wild type cells and Smp mutants.



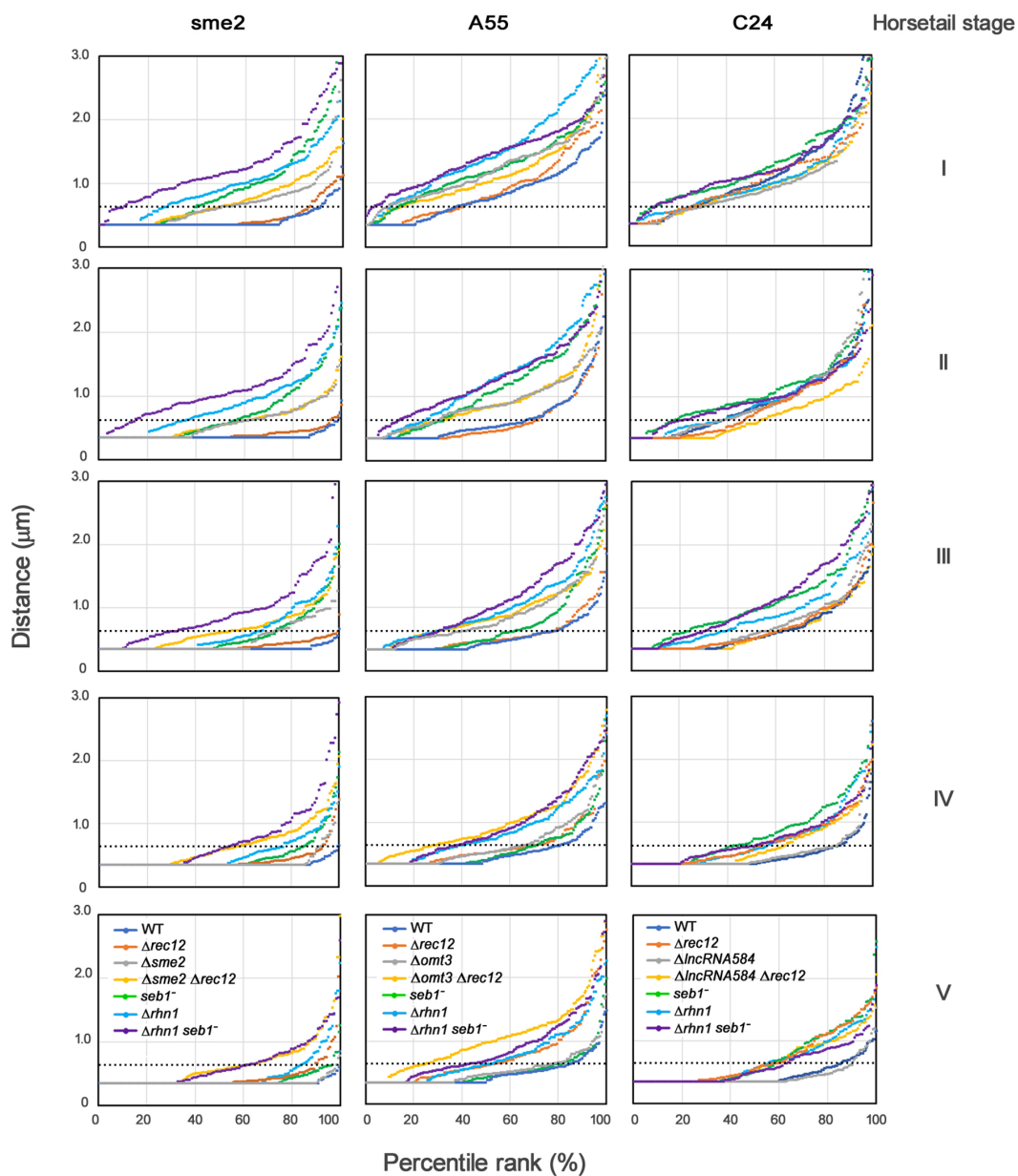
Supplementary Figure 3. Percentile plot of the distance

Percentile rank plots of the distance between homologous *sme2* loci in every horsetail stage shown in Fig. 1c. The threshold of pairing is 0.35 μm as labeled on the Y axis. Source data are provided as a Source Data file 1b.



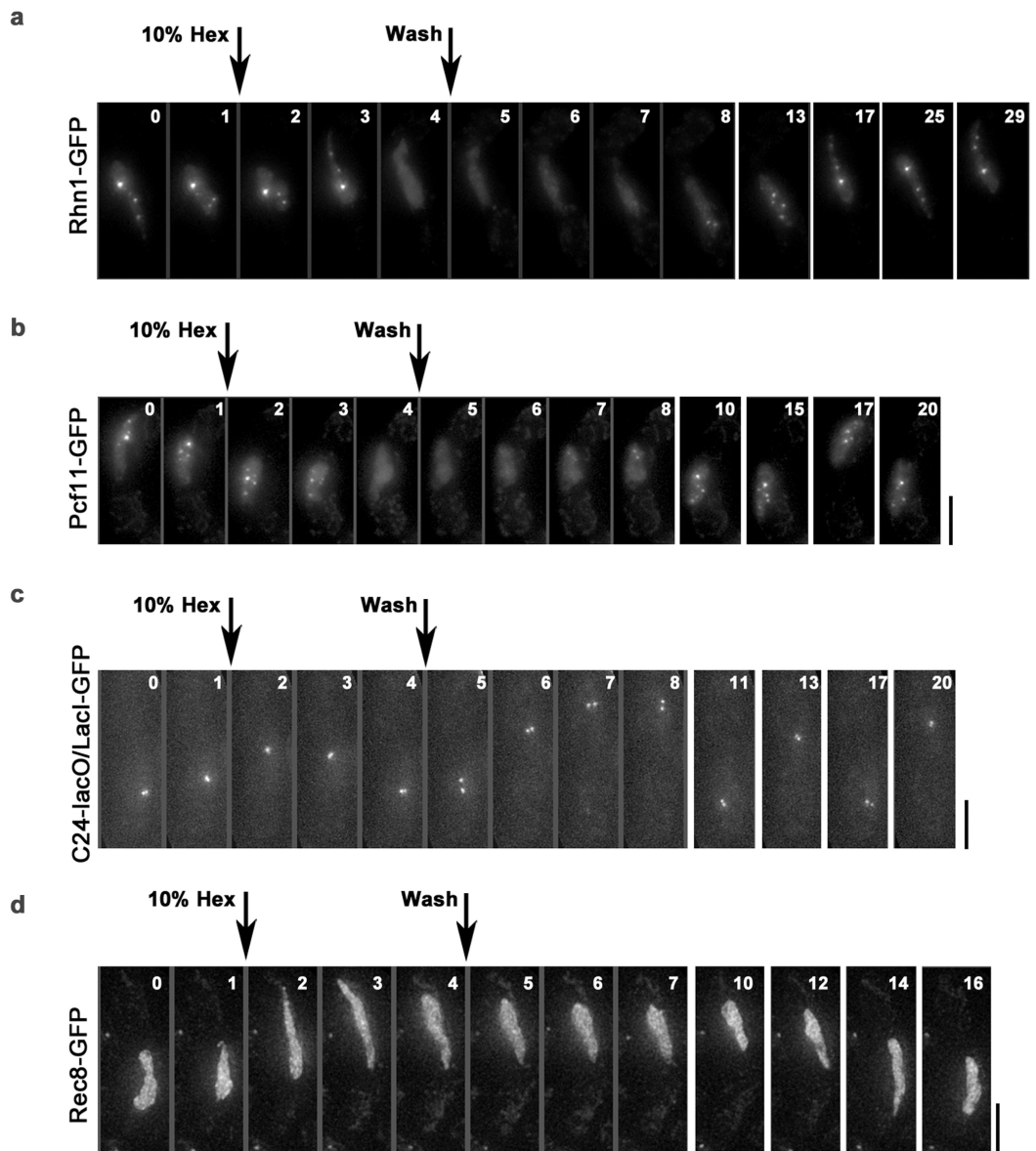
Supplementary Figure 4. Quantification of the Smp protein dots

(a) Number of Seb1-mCherry dots in the horsetail nuclei of $\Delta mei2$ and $\Delta omt3$ cells. (b) Number of Seb1-mCherry dots in the horsetail nuclei of $\Delta ski3$ and $\Delta ski3-lncRNA584$ cells. (c) Number of Seb1-GFP dots in the horsetail nuclei of wild type and in a strain lacking all the three lncRNAs, *sme2* RNA, *omt3* RNA, and lncRNA584. Source data are provided as a Source Data file S4.



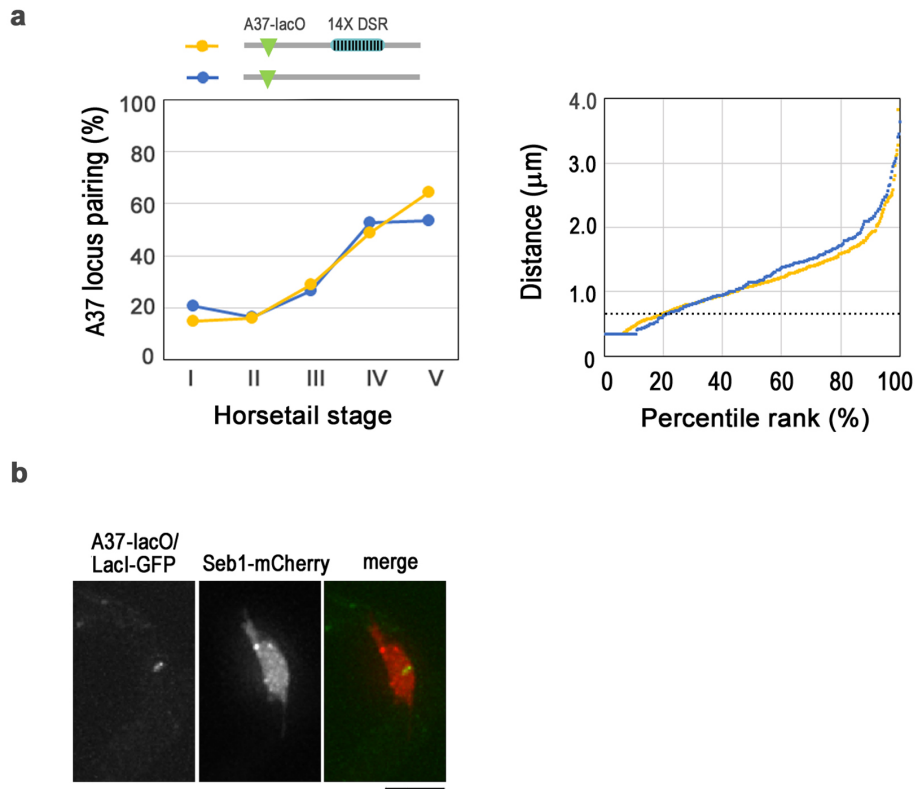
Supplementary Figure 5. Percentile plot of the distance

Percentile rank plots of the distance between homologous *sme2*, A55 and C24 loci in every horsetail stage shown in Fig. 4. The dotted line at 0.67 μm on the Y axis indicates the threshold of pairing. Source data are provided as a Source Data file 4.



Supplementary Figure 6. Liquid-phase separation of Smp proteins and its role in pairing of homologous loci

(a) Selected projected time-lapse images of Rhn1-GFP in living cells at the horsetail stage. (b) Selected projected time-lapse images of Pcf11-GFP in living cells at the horsetail stage. (c) Selected projected time-lapse images of C24-lacO/LacI-GFP in living cells at the horsetail stage. (d) Selected projected time-lapse images of Rec8-GFP in living cells at the horsetail stage. Three-dimensional images were taken every minute. Microfluidic yeast plates (CellASIC) were used for cell culture on the microscope stage. 1,6-Hexanediol (10%) was added or removed, as indicated by the arrows. The solution in the cell culture chamber was changed in < 1 min. The scale bars represent 5 μ m.



Supplementary Figure 7. Effect of DSR repeats on homologous pairing.

(a) Pairing frequency at the A37 locus in wild type (blue) and in the 14×DSR motif (TTAAAC) repeats inserted strain (yellow) (left panel). A DNA fragment containing 4×U1A and 14×DSR sequences²⁸ was inserted in the genome at about 60 kb away from A37 *lacO* insertion (Supplemental Table 1). Right panel shows percentile rank plots of the distance between the A37 loci in the first 3 substages. The dotted line at 0.67 μm on the Y axis indicates the threshold of pairing. Source data are provided as a Source Data file S7. **(b)** Double staining of A37-*lacO*/LacI-GFP with Seb1-mCherry. No Seb1-mCherry dot was formed at the A37 locus.

Supplementary Table 1. Insertion positions of *lacO* arrays in *lacO*/LacI-GFP library

Chromosome I		Chromosome II		Chromosome III	
Name	Genomic position	Name	Genomic position	Name	Genomic position
A01	50946	B01	110029	C01	42728
A02	94498	B02	159887	C02	113974
A03	217912	B03	277440	C03	214549
A04	272653	B04	343050	C04	319994
A05	325930	B05	449000	C05	397988
A06	425597	B06	539260	C06	505789
A07	463596	B07	636874	C07	558957
A08	553980	B08	739653	C08	666416
A09	639405	B09	828428	C09	756290
A10	667382	B10	934372	C10	853882
A11	765526	B11	1006688	C11	971822
A12	842624	B12	1028562	C12	1064740
A13	912607	B13	1136375	C13	1140428
A14	981422	B14	1224061	C14	1244477
A15	1027014	B15	1277831	C15	1316291
A16	1089958	B16	1363490	C16	1448223
A17	1210928	B17	1463491	C17	1524066
A18	1276130	B18	1554538	C18	1632149
A19	1329091	B19	1648000	C19	1727794
A20	1381967	B20	1727771	C20	1822398
A21	1485261	B21	1823968	C21	1928730
A22	1572568	B22	1915147	C22	2023785
A23	1664658	B23	1962938	C23	2107032
A24	1760902	B24	2074884	C24	2209985
A25	1866360	B25	2159853	C25	2335879
A26	1921920	B26	2254416	C26	2418000
A27	2052316	B27	2366555		
A28	2110800	B28	2470824		
A29	2215220	B29	2553418		
A30	2304577	B30	2647289		
A31	2385377	B31	2750390		
A32	2438381	B32	2847916		
A33	2527713	B33	2938954		
A34	2600539	B34	3040956		
A35	2669250	B35	3140566		
A36	2786827	B36	3237304		
A37	2882384	B37	3337527		
A38	2937494	B38	3516819		
A39	2996013	B39	3530977		
A40	3059909	B40	3615774		
A41	3184877	B41	3659380		
A42	3281070	B42	3762502		
A43	3404852	B43	3862500		
A44	3497734	B44	3925880		
A45	3559368	B45	3987400		
A46	3675200	B46	4110630		
A47	3739076	B47	4172323		
A48	3830016	B48	4255523		
A49	3920465	B49	4344090		
A50	4004001	B50	4403495		
A51	4106221				
A52	4196586				
A53	4299146				
A54	4450168				
A55	4510983				
A56	4577519				
A57	4648678				
A58	4701979				
A59	4782589				
A60	4852463				
A61	4964028				
A62	5024382				
A63	5096313				
A64	5177335				
A65	5256188				
A66	5393083				
A67	5479451				

Supplementary Table 2. Strains used in this study

Figure 1a	
YY982	<i>h90 ade6-216 leu1 mei2⁺-mCherry::HygR leu1⁺::nmt1p-rcd1⁺-YFP-FLAG-His6</i>
YY630	<i>h90 ade6-216 leu1 ura4 lys1 mei2⁺-mCherry::HygR pTB46(seb1⁺-GFP)</i>
YY660	<i>h90 ade6-216 leu1 mei2⁺-mCherry::HygR leu1⁺::nmt1p-nab3⁺-YFP-FLAG-His6</i>
YW372-1C	<i>h90 ade6-216 leu1 lys1 ura4-D18 mei2⁺-mCherry::HygR rnm1⁺-GFP::KanR</i>
YW371-1A	<i>h90 ade6-216 leu1 lys1 ura4-D18 mei2⁺-mCherry::HygR pab2⁺-GFP::KanR</i>
YY983	<i>h90 ade6-216 leu1 mei2⁺-mCherry::HygR leu1⁺::nmt1p-rna15⁺-YFP-FLAG-His6</i>
YY984	<i>h90 ade6-216 leu1 mei2⁺-mCherry::HygR leu1⁺::nmt1p-rhn1⁺-YFP-FLAG-His6</i>
YY990	<i>h90 ade6-216 leu1 mei2⁺-mCherry::HygR leu1⁺::nmt1p-pcf11⁺-YFP-FLAG-His6</i>
YW370-1A	<i>h90 ade6-216 leu1 lys1 ura4-D18 mei2⁺-mCherry::HygR pla1⁺-GFP::KanR</i>
YY991	<i>h90 ade6-216 leu1 mei2⁺-mCherry::HygR leu1⁺::nmt1p-ctf1⁺-YFP-FLAG-His6</i>
Figure 1b	
YY548-13C	<i>ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW002	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrhn1:: NatR</i>
YW012	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δpab2:: NatR</i>
YW373-4	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δctf1:: HygR</i>
YW079-5B	<i>h90 ade6-216 leu1 ura4-DS/E sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP seb1-5FLAG- HygR seb1-E38</i>
YW061	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP lys1⁺:: Padh15-skp1-NLS-AtTIR1-NLS pcf11⁺-IAA17-mCherry:: HygR</i>
YW162-3	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP lys1⁺:: Padh15-skp1-NLS-AtTIR1-NLS rna15⁺-IAA17-mCherry:: HygR</i>
YW001	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrnm1:: NatR</i>
YW005	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δnab3:: NatR</i>
YW008	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrcd1:: NatR</i>
Figure 1c	
YY906	<i>h90 ade6-216 leu1 ura4-D18 lys1 U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry</i>
YW006	<i>h90 ade6-216 leu1 ura4-D18 lys1 U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry Δpab2:: NatR</i>
YW004	<i>h90 ade6-216 leu1 ura4-D18 lys1 U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry Δrhn1::NatR</i>
YW283-16B	<i>h90 ade6-216 leu1 ura4-DS/E U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry seb1-5FLAG-HygR seb1-E38</i>
YW375-5B	<i>h90 ade6-216 leu1 ura4-D18 lys1 U1Atag-sme2⁺:: HygR aur1::Pnmt1-U1Ap- mCherry Δctf1::HygR</i>
YW007	<i>h90 ade6-2106 leu1 ura4-D18 lys1 U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry Δnab3:: NatR</i>
YW009	<i>h90 ade6-2106 leu1 ura4-D18 lys1 U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry Arcd1:: NatR</i>
YW010	<i>h90 ade6-2106 leu1 ura4-D18 lys1 U1Atag-sme2⁺::HygR aur1::Pnmt1-U1Ap- mCherry Δrnm11::NatR</i>
Figure 2b-d	

YW298	<i>h90 ade6-210 leu1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP seb1⁺-mCherry:: HygR</i>
YW260	<i>h90 ade6-216 leu1 ura4-D18 C24[::ura4⁺ KanR -lacOp] his7⁺::LacI-GFP seb1⁺-mCherry:: HygR</i>
YW309-13C	<i>h90 ade6-210 leu1 ura4-D18 seb1⁺-mCherry::HygR pab2⁺-GFP::KanR sme2proxy[::ura4⁺- KanR-lacOp] A55[::ura4⁺- KanR-lacOp] C24[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>

Figure 2e

YW145	<i>h-/h- ade6-216/ade6-210 lys1/lys1⁺::mat-Pc pat1::NatR/pat1::NatR pat1-as(L95A)::HygR/pat1-as(L95A)::HygR rhn1⁺-GFP::KanR/rhn1⁺-GFP::KanR</i>
YW146	<i>h-/h- ade6-216/ade6-210 ura4/ura4⁺ lys1/lys1⁺::mat-Pc pat1::NatR/pat1::NatR pat1-as(L95A)::HygR/pat1-as(L95A)::HygR pab2⁺-GFP::KanR/pab2⁺-GFP::KanR</i>
YW147	<i>h-/h- ade6-216/ade6-210 lys1/lys1⁺::mat-Pc pat1::NatR/pat1::NatR pat1-as(L95A)::HygR/pat1-as(L95A)::HygR seb1⁺-GFP::KanR/seb1⁺-GFP::KanR</i>

Figure 2f-g

YW344-14B	<i>h90 ade6-216 leu1 ura4 Δmei2::HygR A55[::ura4⁺-KanR-lacOp] his7⁺::lacI-GFP aur1-r::mei2⁺ seb1⁺-mCherry::HygR</i>
YW346-4	<i>h90 ade6-210 leu1 ura4 seb1⁺-mCherry::HygR A55[::ura4⁺- KanR -lacOp] his7⁺::lacI-GFP Δomt3::NatR</i>
YW345-19B	<i>h90 ade6-210 leu1 ura4 lys1 C24[::ura4⁺-lacOP- KanR] his7⁺::lacI-GFP seb1⁺-mCherry::HygR Δski3::HygR</i>
YW349	<i>h90 ade6-210 leu1 ura4 lys1 C24[::ura4⁺-lacOP- KanR] his7⁺::lacI-GFP seb1⁺-mCherry::HygR Δski3&SPNCRNA.584::NatR</i>

Figure 3

YY548-13C	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW289-5B	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>
YW592-2	<i>h90 ade6-216 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW002	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrhn1:: NatR</i>
YW079-8B	<i>h90 ade6-216 leu1 ura4-DS/E seb1-5FLAG-HygR seb1-E38 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW012	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δpab2:: NatR</i>
YW299-4C	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δrhn1:: HygR</i>
YW302-4D	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP seb1-5FLAG-HygR seb1-E38</i>
YW414-18	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δpab2:: HygR</i>
YW199-24	<i>h90 ade6-216 leu1 lys1 ura4-D18 C24[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δrhn1:: HygR</i>
YW288-10C	<i>h90 ade6-216 leu1 lys1 ura4-D18 C24[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP seb1-5FLAG-HygR seb1-E38</i>
YW417-38	<i>h90 ade6-216 leu1 lys1 ura4-D18 C24[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δpab2:: HygR</i>

Figure 4

YY548-13C	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW417	<i>h90 ade6-210 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrec12:: KanR</i>
YY656-7B	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δsme2:: HygR</i>

YY665-16C	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δsme2:: HygR rec12-152::LEU2</i>
YW002	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrhn1:: NatR</i>
YW079-5B	<i>h90 ade6-216 leu1 ura4-DS/E sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP seb1-5FLAG- HygR seb1-E38</i>
YW287-15D	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrhn1:: NatR seb1-5FLAG- HygR seb1-E38</i>
YW289-5B	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>
YW286-1	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP rec12-152::LEU2</i>
YW346-4	<i>h90 ade6-216 leu1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δomt3:: NatR</i>
YW429	<i>h90 ade6-210 leu1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δomt3:: HygR rec12-152::LEU2</i>
YW299-4C	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δrhn1:: HygR</i>
YW302-4D	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP seb1-5FLAG- HygR seb1-ts125</i>
YW334-30B	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δrhn1:: HygR seb1-5FLAG- HygR seb1-ts125</i>
YW592-2	<i>h90 ade6-216 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YY792-2	<i>h90 ade6-210 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP rec12-152::LEU2</i>
YW351-4	<i>h90 ade6-210 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP ΔlncRNA584:: NatR</i>
YW430-8	<i>h90 ade6-210 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP rec12-152::LEU2</i>
YW350-2A	<i>h90 ade6-210 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP Δrhn1:: HygR</i>
YW288-2C	<i>h90 ade6-216 leu1 lys1 ura4-DS/E C24[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP seb1-5FLAG- HygR seb1-E38</i>
YW291-4B	<i>h90 ade6-216 leu1 lys1 ura4-D18 C24[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δrhn1:: HygR seb1-5FLAG- HygR seb1-E38</i>

Figure 5

YW096-6B	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR</i>
YY906-1	<i>h90 ade6-210 leu1 lys1 ura4-D18 U1Atag-sme2::HygR aur1::Pnmt1-U1Ap-mCherry</i>
YW289-5B	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>
YW286-1	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP rec12-152::LEU2</i>
YY548-13C	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW417	<i>h90 ade6-210 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP ΔRec12:: KanR</i>
YW592-2	<i>h90 ade6-216 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>
YW430-8	<i>h90 ade6-210 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP rec12-152::LEU2</i>

Figure 6

YW289-5B	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>
YW346-4	<i>h90 ade6-216 leu1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δomt3:: NatR</i>
YW445-2	<i>h90 ade6-210 leu1 ura4-D18 lys1 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δsme2:: HygR Δomt3:: sme2⁺-NatR</i>

YW441-1B	<i>hm ade6-210 leu1 ura4-D18 lys1 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δsme2:: HygR</i>
YW446-2D	<i>hp ade6-210 leu1 ura4-D18 lys1 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP Δsme2:: HygR Δomt3:: sme2⁺-NatR</i>
Supplementary Figure 1	
YY831-7B	<i>h90 ade6-210 leu1 lys1 seb1⁺-GFP::KanR</i>
YW055-10	<i>h90 ade6-216 leu1 lys1 rhn1⁺-GFP::KanR</i>
YW097-4B	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR pab2⁺-GFP::KanR</i>
YW364-1	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR rna15⁺-GFP::KanR</i>
YW096-5C	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR rhn1⁺-GFP::KanR</i>
YW365-2	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR pcf11⁺-GFP::KanR</i>
YW369-1	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR ctf1⁺-GFP::KanR</i>
Supplementary Figure 2	
YW061	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP lys1⁺:: Padh15-skp1-NLS-AtTIR1-NLS pcf11⁺-IAA17-mCherry:: HygR</i>
YW162-3	<i>h90 ade6-216 leu1 ura4-D18 sme2proxy[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP lys1⁺:: Padh15-skp1-NLS-AtTIR1-NLS rna15⁺-IAA17-mCherry:: HygR</i>
EON125	<i>hp ade6-210 otrR(SphI)::ura4⁺ leu1-32 ura4-DS/E seb1-5Flag-hphMX6 seb1-E38</i>
L968	<i>h90</i>
Supplementary Figure 4	
YW448-9A	<i>h90 ade6-210 leu1 lys1 seb1⁺-GFP::KanR Δsme2::ura4⁺ Δomt3:: KanR ΔlncRNA584:: HygR</i>
Supplementary Figure 6	
PY183	<i>h90 ade6-216 leu1-32 rec8⁺::GFP-KanR</i>
Supplementary Figure 7	
YY699-C6	<i>h90 ade6-216 leu1 lys1 ura4-D18 A37[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>
YW388-2	<i>h90 ade6-216 leu1 lys1 ura4-D18 A37[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP C02::4XU1A-14DSR-NatR</i>
YW404-22	<i>h90 ade6-216 leu1 lys1 ura4-D18 A37[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP C02::4XU1A-14DSR-NatR seb1⁺-mCherry::HygR</i>
Video 1	
YW096-6B	<i>h90 ade6-216 leu1 lys1 ura4-D18 seb1⁺-mCherry::HygR</i>
Video 2	
YW055-10	<i>h90 ade6-216 leu1 lys1 ura4-D18 rhn1⁺-GFP::KanR</i>
Video 3	
YW052-1	<i>h90 ade6-216 leu1 lys1 ura4-D18 rna15⁺-GFP::KanR</i>
Video 4	
YY906-1	<i>h90 ade6-210 leu1 lys1 ura4-D18 U1Atag-sme2::HygR aur1::Pnmt1-U1Ap-mCherry</i>
Video 5	
YW289-5B	<i>h90 ade6-216 leu1 lys1 ura4-D18 A55[::ura4⁺- KanR-lacOp] his7⁺::lacI-GFP</i>
Video 6	
YW592-2	<i>h90 ade6-216 leu1 ura4-D18 lys1 C24[::ura4⁺- KanR-lacOp] his7⁺::LacI-GFP</i>

All strains are constructed in this study, except for PY183 (Watanabe & Nurse, 1999).

Supplementary Table 3. List of primers for gene disruption

mei2 deletion

Forward1: GCATTGAGCCGTCATTGATC

Reverse1: cggcggggacgaggcaagctTTACGAAAGAGATCCTGTATTG

Forward2: gataactaacgccgccatccagtTTATGTGTTAAAACGGCAACAA

Reverse2: CTAACAACCATCTAACCCTTCC

omt3 deletion

Forward: CGCCGCATTTCGATTTTAAATGTATCACGTATAAATCGGTAAATATGTGTGTACCACGGT
TGTAGAAGTGCTCGAAGACAAagcttgccctcgtccccgccg

Reverse: CCTAATTTTTTAAAAGTTTATCGACTTGGCTTCAACACATAGTGCTTCTATAAGTAAACA
AACATGTTTTTAAAACAAGATAactggatggcggcggttagt

ski3 deletion

Forward1: CACTCCTTGGGTTCTCTATC

Reverse1: cggcggggacgaggcaagctGACAATGGCAGTTTATTATG

Forward2: gataactaacgccgccatccaGAACTTGTATTTATTATCAA

Reverse2: GAATCTGACAGTTCATTAGC

ski3-lncRNA584 deletion

Forward1: CACTCCTTGGGTTCTCTATC

Reverse1: cggcggggacgaggcaagctGACAATGGCAGTTTATTATG

Forward2: gataactaacgccgccatccaGAACTTGTATTTATTATCAA

Reverse2: AACCTTTTACTTGTATTATAAGCGCCAATAATTGGCTGTTTGAATTTCAACCCACTGC
TAAATCGATGACGAATTACGCTGactggatggcggcggttagt

rhn1 deletion

Forward: TTTATTACTTATGCTTATGATTTTCGTTTGCATGCTAACAGCAGTGTGTCGTAAGAAA
CTCACCATCAGCATAAAACAAGagcttgccctcgtccccgccg

Reverse: TGACTAAGACTGCCACATTGATTGACAGTTTAAAAAAGAAAAATAATGAAAAC
TAAGCATTACATTTTCATGGATTactggatggcggcggttagta

pab2 deletion

Forward1: TATAAGCAGGCAGTCCAAATCCT

Reverse1: ttaattaacccggggatccgTTTTTCAAATTTTCTGCTAC

Forward2: gataactaacgccgccatccaGTAATCGCTTTGATGACTT

Reverse2: GAATTACATTATGTCAGCATAT

ctf1 deletion

Forward1: GTACTTACCATGAATTGTCCC

Reverse1: cggcggggacgaggcaagctGCTGGTAGTCGCGTCTTTTCG

Forward2: gataactaacgccgccatccaGTCTAATTTTAGTAATATCA

Reverse2: GTTGATGTTACCACCTAC

rdl deletion

Forward1: ACGCATAGCACGAGGACTGAAATT

Reverse1: ttaattaacccggggatccgTTTTTCTAGAATGCGGGAAA

Forward2: gataactaacgccgccatccaCTGCCCTTCGTTTCTATCTG

Reverse2: ATCAGAGTAAGTTCTGTCCT

nab3 deletion

Forward1: CTTTACACATTTGGTTGAGTGT

Reverse1: ttaattaacccggggatccgTTATTGATTGACTTGAAAAG

Forward2: gataactaacgccgccatccaCAAGTGTTTTAGGTAGCAC

Reverse2: CAGTAAAAATCAAGTATCGAAAC

rmn1 deletion

Forward1: TCTCATAATCGTCAGAGTTTGA

Reverse1: ttaattaacccggggatccgTGTGATGGAGCAAAGTGCT

Forward2: gataactaacgccgccatccaCTGCATTGTATGCGGCTGTC

Reverse2: CATGAGTTTTCTCTCCTAGCCG

Supplementary Table 4. List of smFISH probes

sme2_1	ataccactaagtctgttta	ncRNA130_1	gttattaatgctcctctt	ncRNA584_1	cgltctatcctaagagtgga
sme2_2	cggcagaagattgaccaaca	ncRNA130_2	gaattcccaacctctttta	ncRNA584_2	tgctgacactctacgatcat
sme2_3	gcatattcgccttacaata	ncRNA130_3	cgcttttactctttttgt	ncRNA584_3	tatagtgccacgaatcgttc
sme2_4	accaactaaagcgatcttc	ncRNA130_4	atctttatcatgcataca	ncRNA584_4	tctgcacttttgagctttt
sme2_5	gaccatttcaaaatgttgca	ncRNA130_5	tcaaaaaccatcatcagcct	ncRNA584_5	goggcataaatattgtccac
sme2_6	taccgaatccagcttttga	ncRNA130_6	agcaacatcgctaatgatca	ncRNA584_6	cagattggcaaatatccggat
sme2_7	cagagcttagaagacaaggt	ncRNA130_7	ttcactgactctgcctttac	ncRNA584_7	caatccaaccactcactca
sme2_8	taactggaccccatcaagaa	ncRNA130_8	actcagcaaaagcttaactct	ncRNA584_8	cttacctagagccattttt
sme2_9	taaaccaacttgggggttg	ncRNA130_9	tacgaacgcattgtcccaa	ncRNA584_9	tgcttaccgtgtacgaatc
sme2_10	tctaagctactattcatcca	ncRNA130_10	acaattcgtcacgtacatca	ncRNA584_10	gatgctctgtaggaactgt
sme2_11	agtagattccatcagtcata	ncRNA130_11	tcaggcctcattttttta	ncRNA584_11	actcatgaggcactagttc
sme2_12	tgcaaccaaaaagtgtacca	ncRNA130_12	accatagagatagaagcggt	ncRNA584_12	tcagctctatggttgcaaa
sme2_13	catgtaagtgtcttcaagg	ncRNA130_13	gttcttctcctcaaatctca	ncRNA584_13	cttcacgaccagacgattg
sme2_14	ttcagctatccgcaaatgt	ncRNA130_14	acaagattcctcttaggttt	ncRNA584_14	gcatttgctactactgtgta
sme2_15	agtcgttttattcttttct	ncRNA130_15	gtcatgtgatttttcatcc	ncRNA584_15	accgttatgtctcaaagtct
sme2_16	gtttcaacaatagttcaggt	ncRNA130_16	gtttcgctccaagatgat	ncRNA584_16	gcttctagttgacttact
sme2_17	tctgttcaggaatacgttt	ncRNA130_17	aagctctcaactcctttc	ncRNA584_17	ctgtatgctctgcgattgaa
sme2_18	gtttcgcatacaactttca	ncRNA130_18	aattcctttgctgactgaca	ncRNA584_18	gtcttggtgcaatagctatt
sme2_19	gcgtttaacaaactgcggg	ncRNA130_19	aaggtttcttcacgtcact	ncRNA584_19	caagccgaaggaacaatt
sme2_20	tggttcagcacgtttcaa	ncRNA130_20	ttctcgagaagcatgaagca	ncRNA584_20	agtatccaaggtgtatggg
sme2_21	ttggttgtagggtttaacg	ncRNA130_21	ataaaaacccaagaccgct	ncRNA584_21	tgagcgattcaaatgccgac
sme2_22	tgaattagctgtctgtggt	ncRNA130_22	ccaaatcaaatgggacct	ncRNA584_22	ttaaaccgactacccttgga
sme2_23	ccaatgaacataaccattgt	ncRNA130_23	ttaggttctcactgcagcaac	ncRNA584_23	gatgcaaaaagccttactcgg
sme2_24	aagagggagcacagcacaac	ncRNA130_24	ggtgagacttagcttacgag	ncRNA584_24	cttcctactagaacacctt
sme2_25	atgtgtgctattctctct	ncRNA130_25	aacctcctgtttgcataaga	ncRNA584_25	tctggggaaaatccaaccg
sme2_26	gaaggcatgttgaattgga	ncRNA130_26	acatggatcaatcatccatt	ncRNA584_26	catgcagtaaacaccgagga
sme2_27	attaatagctgtccttgcg			ncRNA584_27	ttgatctattcggaaggca
sme2_28	ccttaaagtggttttaagc			ncRNA584_28	ttctatctctattgaccag