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Supplemental Information

SNX27-Mediated Recycling

of Neuroligin-2 Regulates

Inhibitory Signaling

Els F. Halff, Blanka R. Szulc, Flavie Lesept, and Josef T. Kittler

Figure S1



Figure S1 (related to Figure 1): Internalized NL2 co-localizes and interacts with endosomal markers and recycling complexes. A. Confocal images of antibody feeding in COS-7 cells showing co-localization of

internalized ^{HA}NL2 (green) with Rab5^{dsRed} (top) or Rab11^{dsRed} (bottom, red). Scale bars, 15 μm (whole cells) and 5 μm (zooms). Arrowheads show examples of co-localization. **B.** Example fluorescent intensity line scans through clusters (corresponding to the lines shown in the merged zoomed images in panel A), showing overlap between internalized ^{HA}NL2 and Rab5^{dsRed} or Rab11^{dsRed}. **C.** Pearson's coefficient of co-localization between ^{HA}NL2 and SNX27^{GFP} in neurons (related to Fig.1A; n=4 cells from 2 independent experiments). **D.** Confocal image of antibody feeding in HeLa cells showing co-localization of internalized ^{HA}NL2 (red) with SNX27^{GFP}. Arrowheads show examples of co-localization. Scale bar, 10 μm. **E.** Pearson's coefficient of co-localization between ^{HA}NL2 and VPS35^{GFP} in neurons (related to Fig.1B; n=3 cells from 1 experiment). **F.** Confocal image of antibody feeding in HeLa cells showing co-localization of internalized ^{HA}NL2 (red) with VPS35^{GFP}. Arrowheads show examples of co-localization of internalized ^{HA}NL2 (red) with VPS35^{GFP}. Arrowheads show examples of co-localization of internalized ^{HA}NL2 (red) with VPS35^{GFP}. Arrowheads show examples of co-localization of internalized ^{HA}NL2 (red) with VPS35^{GFP}. Arrowheads show examples of co-localization of internalized ^{HA}NL2 (red) with VPS35^{GFP}. Arrowheads show examples of co-localization of internalized ^{HA}NL2 (red) with VPS35^{GFP}. Arrowheads show examples of co-localization. Scale bar, 10 μm. **G**. Full western blots related to the sections of western blots shown in Fig.1C-F. Red boxes indicate the part of the blot that was shown in Fig.1C-F. **H.** Coomassie gel of purified GST and GST-NL2^{CT}. Numbers on the left indicate molecular weight in kDa.

Values are mean <u>+</u> SEM; *p<0.05, **p<0.01; paired two-tailed *t*-test.

Figure S2



Figure S2 (related to Figure 2): Abolishing PDZ-ligand interaction between SNX27 and NL2 does not affect endosomal localization of SNX27 and NL2 mutants. A. Confocal images showing co-localization of WT and mutant SNX27^{GFP} with Early Endosome Antigen 1 (EEA1) in the soma of hippocampal neurons. Scale bar, 5 μm. **B.** Western blot showing co-immunoprecipitation from COS-7 cells co-expressing GFP-control with ^{HA}NL2WT (first lane) or SNX27^{GFP} with ^{HA}NL2 constructs (second and third lane). GFP and SNX27^{GFP} were pulled down using GFP-Trap beads (IP, immunoprecipitation). Numbers on the left indicate molecular weight in kDa. **C.** Pearson's coefficient of co-localization between SNX27^{GFP} and ^{HA}NL2WT or ^{HA}NL2ΔPDZL in HeLa cells (related to Fig. 2D; n=15 and 14 cells, respectively, from 2 independent experiments; unpaired two-tailed *t*-test). **D.** Confocal images of antibody feeding in hippocampal neurons showing co-localization of internalised ^{HA}NL2WT or ^{HA}NL2ΔPDZL (red) with SNX27^{GFP} (green) in the soma. Arrowheads show examples of colocalization. Scale bars, 25 μm (whole cells) and 5 μm (soma). **E.** Pearson's coefficient of co-localization between ^{HA}NL2WT or ^{HA}NL2ΔPDZL and SNX27^{GFP} in neurons (related to Fig.S2D; n=15 and 7 cells, respectively; twoway ANOVA with Bonferroni's correction).

Values are mean <u>+</u> SEM; n.s. non-significant; ****p<0.0001.

Figure S3



Figure S3 (related to Figure 3): Overexpression of SNX27^{GFP}WT but not SNX27^{GFP}H112A affects miniature IPSCs in hippocampal neurons. A. Confocal images of hippocampal neurons overexpressing SNX27^{GFP}WT (left) or SNX27^{GFP}H112A (right), showing its distribution throughout the neuron. Scale bar, 25 μ m. B. Quantification of overlapping presynaptic GAD65 and postsynaptic γ 2 clusters in hippocampal neurons overexpressing SNX27^{GFP}H112A, or mock transfected (Related to Fig.3A-H; n=23,26,24 cells; Kruskal-Wallis test with Dunn's correction). C. Total charge transfer calculated for mIPSC of hippocampal

neurons that were mock transfected (Control), or overexpressing SNX27^{GFP}WT (WT) (related to Fig.3I-K; n=13 and 22 cells, respectively; Mann-Whitney test). **D.** Kinetics data related to whole-cell patch clamp recordings depicted in Fig. 3I-K. Rise time (Left) and decay time (Right) show no significant difference (n=13 and 22 cells, respectively; unpaired two-tailed *t*-tests). **E.** Representative traces of mIPSC patch-clamp recordings from hippocampal cultures, mock transfected (Control), or overexpressing SNX27^{GFP}H112A (H112A). **F-G.** Pooled data (left) and cumulative probability plot (right) of mIPSCs amplitude (**F**) and frequency (**G**) of hippocampal neurons that were mock transfected (Control), or overexpressing SNX27^{GFP}H112A (H112A) (n=11 cells each; unpaired two-tailed *t*-tests). **H.** Total charge transfer calculated for mIPSC of hippocampal neurons (related to panel E-G; unpaired one-tailed *t*-test) and decay time (Right; unpaired two-tailed *t*-test). Values are mean \pm SEM; *p<0.05, **p<0.01.

Figure S4



Figure S4 (related to Figure 4): The effects of SNX27 knockdown can be rescued by SNX27^{GFP}WT but not SNX27^{GFP}H112A overexpression. A. Confocal images of hippocampal neurons transfected with control or SNX27-specific shRNAi (co-expressing dsRed), combined with murine SNX27^{GFP} (green) to assess the efficiency of knockdown. Scale bar, 25 µm. B. Quantification of relative SNX27^{GFP} intensity in hippocampal neurons, transfected as in panel A, shows a 84% decrease in GFP intensity with SNX27-specific shRNAi compared to control shRNAi (n=18 and 22 cells, respectively; unpaired two-tailed t-test). C. Ouantification of overlapping presynaptic GAD65 and postsynaptic γ^2 clusters in hippocampal neurons transfected with control or SNX27specific shRNAi (related to Fig.4A-G; n=22 and 23 cells, respectively; unpaired one-tailed t-test). D. Total mIPSC charge transfer calculated for mIPSC of hippocampal neurons transfected with control or SNX27-specific shRNAi (related to Fig.4H-J; n=23 and 20 cells, respectively; unpaired two-tailed *t*-test). E. Kinetics data related to wholecell patch clamp recordings depicted in Fig. 4H-J: rise time (left) and decay time (right) (n=23 and 20 cells, respectively; unpaired two-tailed t-tests). F-H. Confocal images of 30 µm dendritic sections of hippocampal neurons transfected with SNX27 shRNAi only (KD, F), or combined with RNAi-resistant SNX27^{GFP}WT (KD+WT, G) or SNX27^{GFP}H112A (KD+H112A, H). Neurons were stained for synaptic NL2 (NL2^{EXT}), GAD65, gephyrin, and the GABAAR y2 subunit. Arrowheads show synaptic clusters. Scale bar, 4 µm. I-J. Quantification of cluster number (left) and area (right) in hippocampal neurons transfected as in panel F-H. Staining is analysed for $\gamma 2$ (I; n=23,22,24 cells; one-way ANOVA with Bonferroni's correction) and gephyrin (J; n=22,24,19 cells; cluster number, Kruskal-Wallis test; cluster area, one-way ANOVA). See Fig.4K-L for quantification of NL2EXT and GAD65. K. Quantification of overlapping presynaptic GAD65 and postsynaptic $\gamma 2$ clusters in hippocampal neurons transfected as in panel F-H (n=23,22,28 cells; one-way ANOVA). L. Kinetics of mIPSCs related to whole-cell patch clamp recordings depicted in Fig. 4M-P: rise time (left) and decay time (right) (n=16,15,13 cells; both one-way ANOVA). M. Schematic representation of the proposed molecular model, summarizing the data from Fig.3 and Fig.4, and explaining how SNX27-mediated recycling of NL2 modulates inhibitory signaling. Increased SNX27 activity and NL2 recycling enhances stabilization of the postsynaptic scaffold via recruitment of gephyrin and GABAARs, resulting in increased mIPSC amplitude. SNX27 knockdown and impaired NL2 recycling destabilizes the presynaptic terminal, leading to reduced mIPSC frequency. Values are mean <u>+</u> SEM; *=p<0.05, **=p<0.01, ***=p<0.001, ****=p<0.0001.