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Bromodomain inhibitor I-BET151 suppresses immune responses during fungal-immune interaction

Compound Number	Cytokine	Mean pIC50	SD	Range	Number of curves (n)	Mean Hill Slope	Mean asymptote max
GSK1210151A	ΤΝFα	6,5	0,43	5.9 - 7.8	68	1,3	105
GSK1210151A	IL-6	6,1	0,36	5.5 - 7.8	116	1,1	98

% Inhibition	[GSK1210151A] M				
	ΤΝFα	IL-6			
1,25	1,10E-08	1,50E-08			
2,5	1,89E-08	2,84E-08			
5	3,28E-08	5,46E-08			
10	5,83E-08	1,08E-07			
20	1,09E-07	2,25E-07			
30	1,65E-07	3,68E-07			
40	2,31E-07	5,49E-07			
50	3,16E-07	7,94E-07			
60	4,32E-07	1,15E-06			
70	6,07E-07	1,72E-06			
80	9,19E-07	2,80E-06			
90	1,71E-06	5,85E-06			
95	3,05E-06	1,15E-05			
97,5	5,30E-06	2,22E-05			

Figure S1. Target engagement plots for GSK1210151A (I-BET151).



Figure S2. Cytotoxicity assay for I-BET151 treatment. Detection of cell death by lactate dehydrogenase (LDH) assay in PBMCs treated with control medium, 1 μ M or 10 μ M I-BET151 and stimulated with RPMI or 10⁶ heat-killed *C. albicans* conidia during 48h (A) or 7 days (B) (mean ± SEM, n =3 (from 3 different individual donors)).



Figure S3. Morphological changes induced by trained immunity.

Morphology of cells after 24 h of training and 6 days of rest when the cells were trained with RPMI (negative control), 10 μ M I-BET151, 1 mg/ml β -glucan or 10 μ M I-BET151 + 1 mg/ml β -glucan. Pictures were taken at day 6, magnification x10.





(A) Cytotoxicity assay for the treatment of cells with three different concentrations of JQ1. Dead cells were used as a positive control. (B) IL-1 β , IL-6 and TNF α production by human monocytes treated or not with 0.05 μ M JQ1 and stimulated with 10⁶/ml heat-killed *C. albicans* conidia for 24 h. (C) IL-6 and TNF α production by human monocytes treated or not with 0.05 μ M JQ1 and stimulated with RPMI, 1 μ g/mL β -glucan or 10⁶/ml heat-killed *C. albicans* conidia, following a similar protocol displayed in Figures 3A and 3D. Cells were re-exposed to 10 ng/ml LPS after 6 days, and cytokines were measured 24 h later. Mean ± SEM, n=6 (from 6 different individual donors); pooled from 2 independent experiments with 3 individual donors each. *p<0.05, Wilcoxon signed-rank test.

1 Table 1. Primer pairs used for epigenetic study.

IL6.1	FW	TCGTGCATGACTTCAGCTTT 2
IL6.1	RV	GCGCTAAGAAGCAGAACCAC 3
IL6.2	FW	AGGGAGAGCCAGAACACAGA
IL6.2	RV	GAGTTTCCTCTGACTCCATCG 4
TNFA.1	FW	AGAGGACCAGCTAAGAGGGA
TNFA.1	RV	AGCTTGTCAGGGGATGTGG ⁵
TNFA.2	FW	GTGCTTGTTCCTCAGCCTCT
TNFA.2	RV	ATCACTCCAAAGTGCAGCAG
TNFA.3	FW	TGTCTGGCACACAGAAGACA 7
TNFA.3	RV	CCCTGAGGTGTCTGGTTTTC
TNFA.4	FW	AGCCAGCTGTTCCTCCTTTA 8
TNFA.4	RV	TTAGAGAGAGGTCCCTGGGG
TNFA.5	FW	TGATGGTAGGCAGAACTTGG
TNFA.5	RV	ACTAAGGCCTGTGCTGTTCC 10
TNFA.6	FW	CAGGCAGGTTCTCTTCCTCT
TNFA.6	RV	GCTTTCAGTGCTCATGGTGT 11

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14 Supplementary Methods

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16 LPS stimulated human blood assay

17 Blood was collected into 10 IU / mL sodium heparin and incubated for 30 min at 37°C in

18 a concentration response (10 μ M-0.51nM) of I-BET151, prior to the addition of 200 ng/mL

19 LPS. Supernatants were removed after 22h incubation 37°C and analysed for IL-6 and

20 TNFα using MesoScaleDescovery (MSD) technology. ICxx calculations were determined

using the formula ICxx = ((% inhibition / (100-% inhibition))^(1/Hill Slope))*IC50).

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23 Viability assays

24 Cell viability was assessed using CytoTox 96® Non-Radioactive Cytotoxicity Assay

- 25 (Promega). Released LDH in culture supernatants is measured with a 30-minute coupled
- enzymatic assay. The amount of color formed is proportional to the number of lysed cells.