

# Resonance Raman, EPR and MCD Spectroscopic Investigation of Diheme Cytochrome *c* Peroxidases from *Nitrosomonas europaea* and *Shewanella oneidensis*

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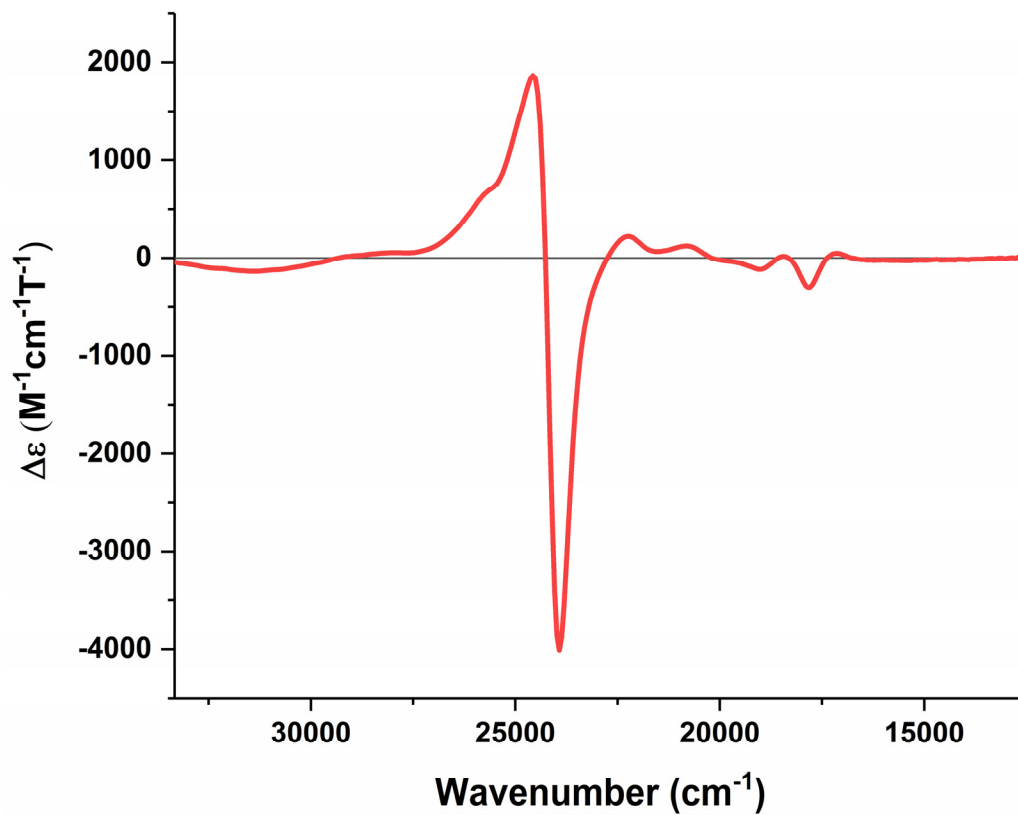
## Supporting Information

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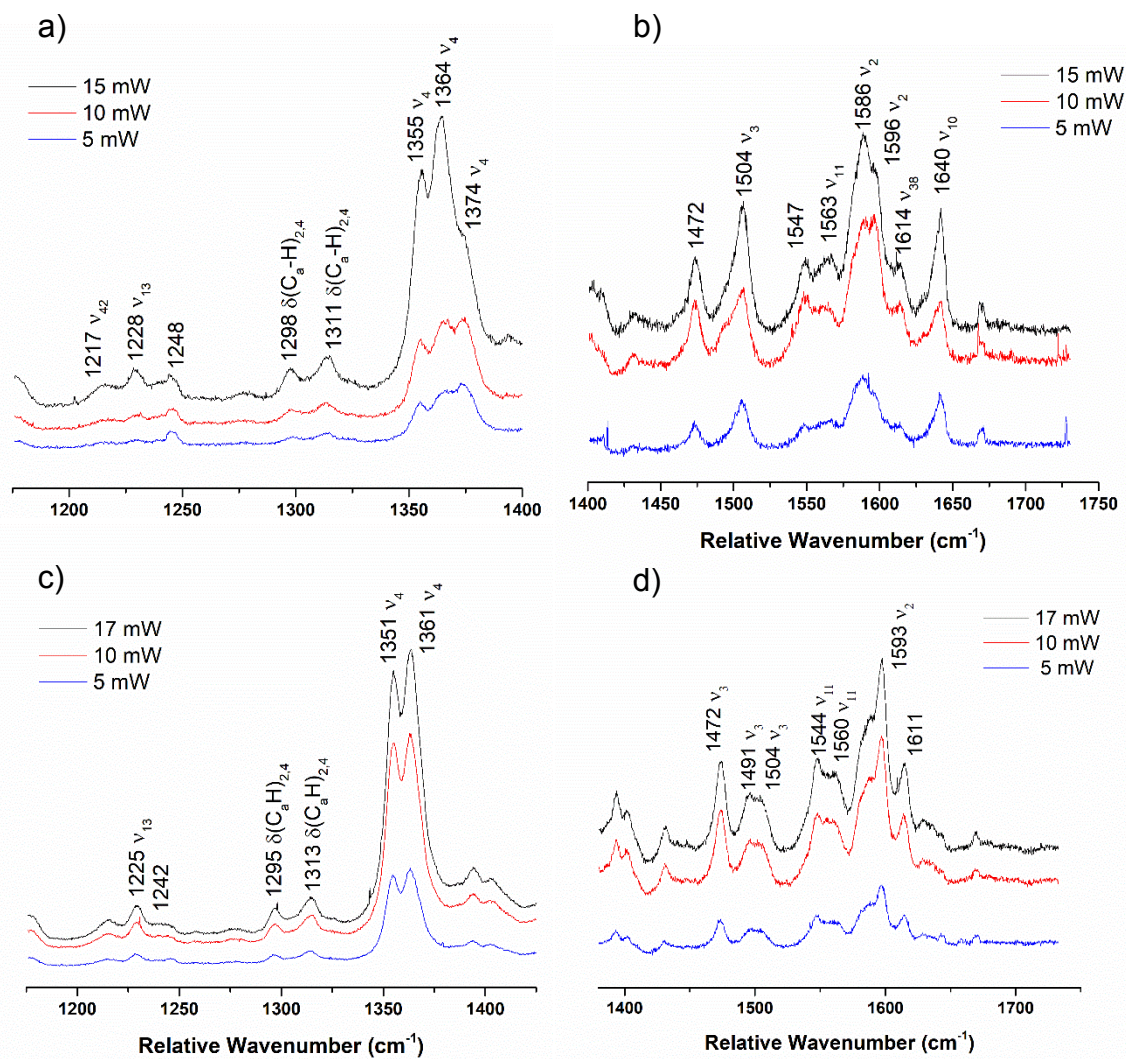
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**Table S1**

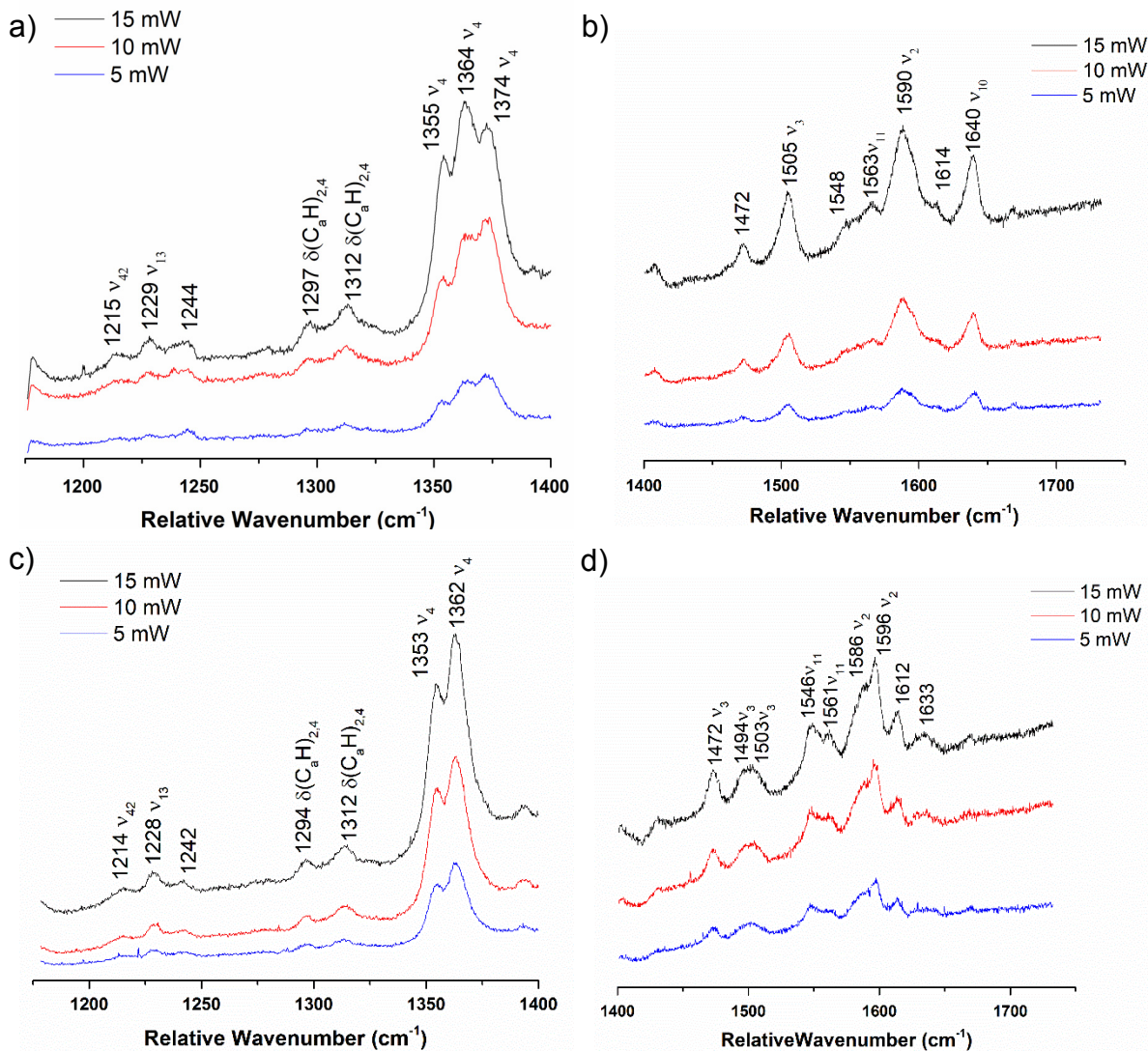
Bacterial CcPs that require reduction	Bacterial CcPs that are constitutively active
<i>Paracoccus denitrificans</i> <sup>6</sup>	<i>Nitrosomonas europaea</i> <sup>3</sup>
<i>Pseudomonas aeruginosa</i> <sup>7</sup>	<i>Methyloccocus casulatus</i> <sup>18</sup>
<i>Paracoccus pantrophus</i> <sup>8</sup>	
<i>Marinobacter hydrocarbonoclasticus</i> <sup>9</sup>	
<i>Rhodobacter capsulatus</i> <sup>10</sup>	
<i>Pseudomonas stutzeri</i> <sup>11</sup>	
<i>Geobacter sulfurreducens</i> <sup>12</sup>	
<i>Shewanella Oneidensis</i> <sup>13</sup>	



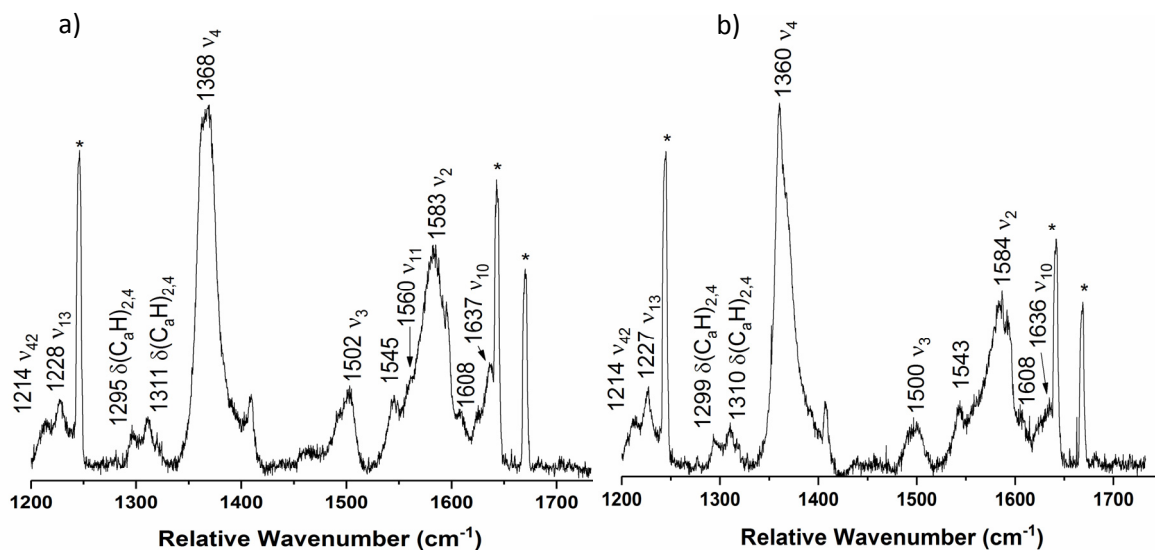
**Figure S1.** Near-UV/visible-region MCD spectrum of **low-spin ferric** Cytochrome  $b_5$ . The sample was 3.7  $\mu\text{M}$  with  $\sim 50\%$  glycerol in a TIP7 buffer at pH 7. This spectrum was recorded at 2 K.



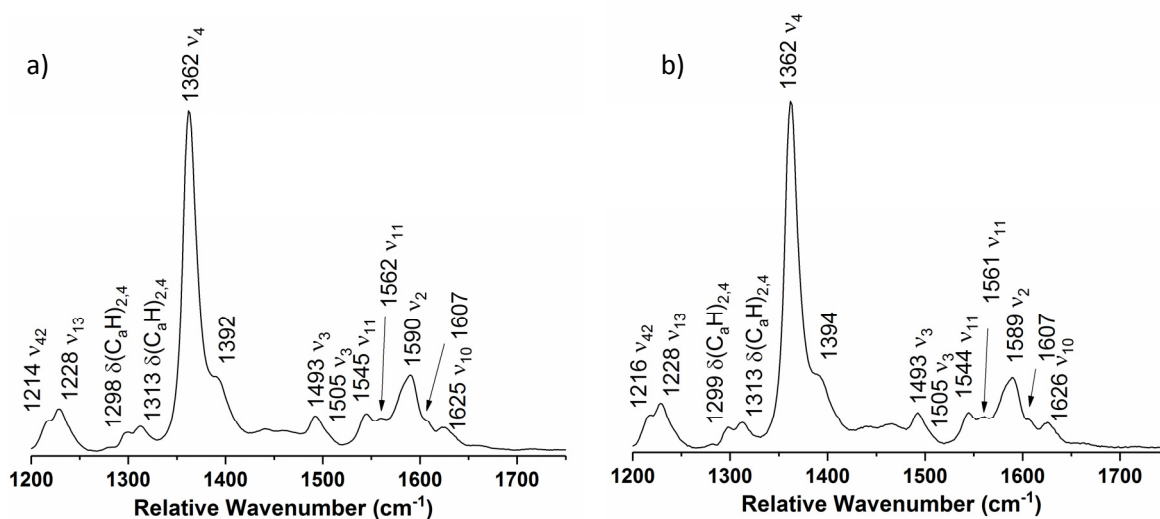
**Figure S2.** Power dependence resonance Raman spectra of **diferric NeCcP** from a) 1150 to 1400  $\text{cm}^{-1}$ , and b) 1400 to 1750  $\text{cm}^{-1}$  and the power dependence spectra of **semireduced NeCcP** from c) 1150 to 1400  $\text{cm}^{-1}$ , and d) 1400 to 1750  $\text{cm}^{-1}$ . The diferric sample was 420  $\mu\text{M}$  and the semireduced sample was 490  $\mu\text{M}$  in protein, both with 30% glycerol in pH 7 TIP7 buffer. The sample was excited at 413.1 nm using a Krypton ion gas laser at 77 K.



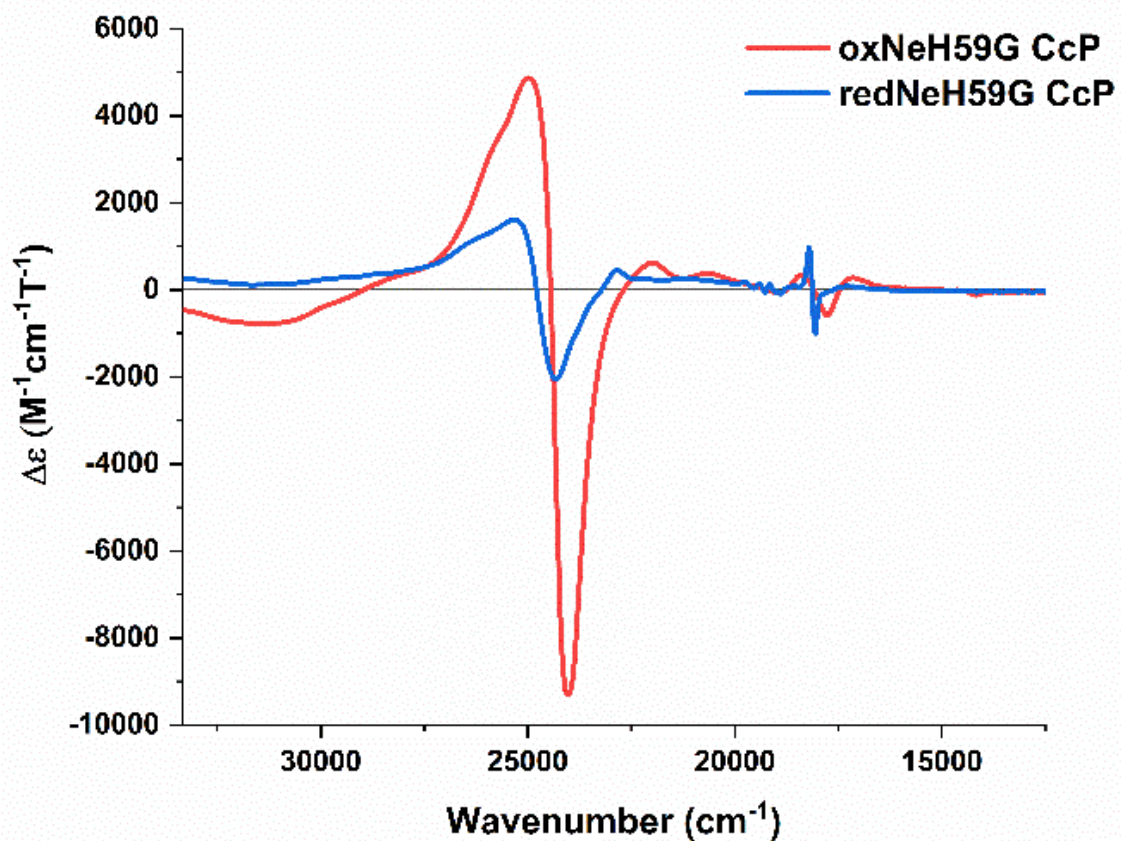
**Figure S3.** Power dependence resonance Raman spectra of **diferric *NeH59G CcP*** from a) 1150 to 1400 cm<sup>-1</sup>, and b) 1400 to 1750 cm<sup>-1</sup> and the power dependence spectra of **semireduced *NeH59G CcP*** from c) 1150 to 1400 cm<sup>-1</sup>, and d) 1400 to 1750 cm<sup>-1</sup>. The diferric sample was 419 μM and the semireduced sample was 460 μM in protein, both with 30% glycerol in pH 7 TIP7 buffer. The sample was excited at 413.1 nm using a Krypton ion gas laser at 77 K.



**Figure S4.** Room temperature resonance Raman spectrum of a) diferric *NeCcP* and b) diferric *NeH59G CcP*. The samples were  $\sim 420 \mu\text{M}$  in protein with 30% glycerol in pH 7 TIP7 buffer. The samples were rotated and excited at 413.1 nm using a Krypton ion gas laser. Plasma lines are indicated with an asterisk.

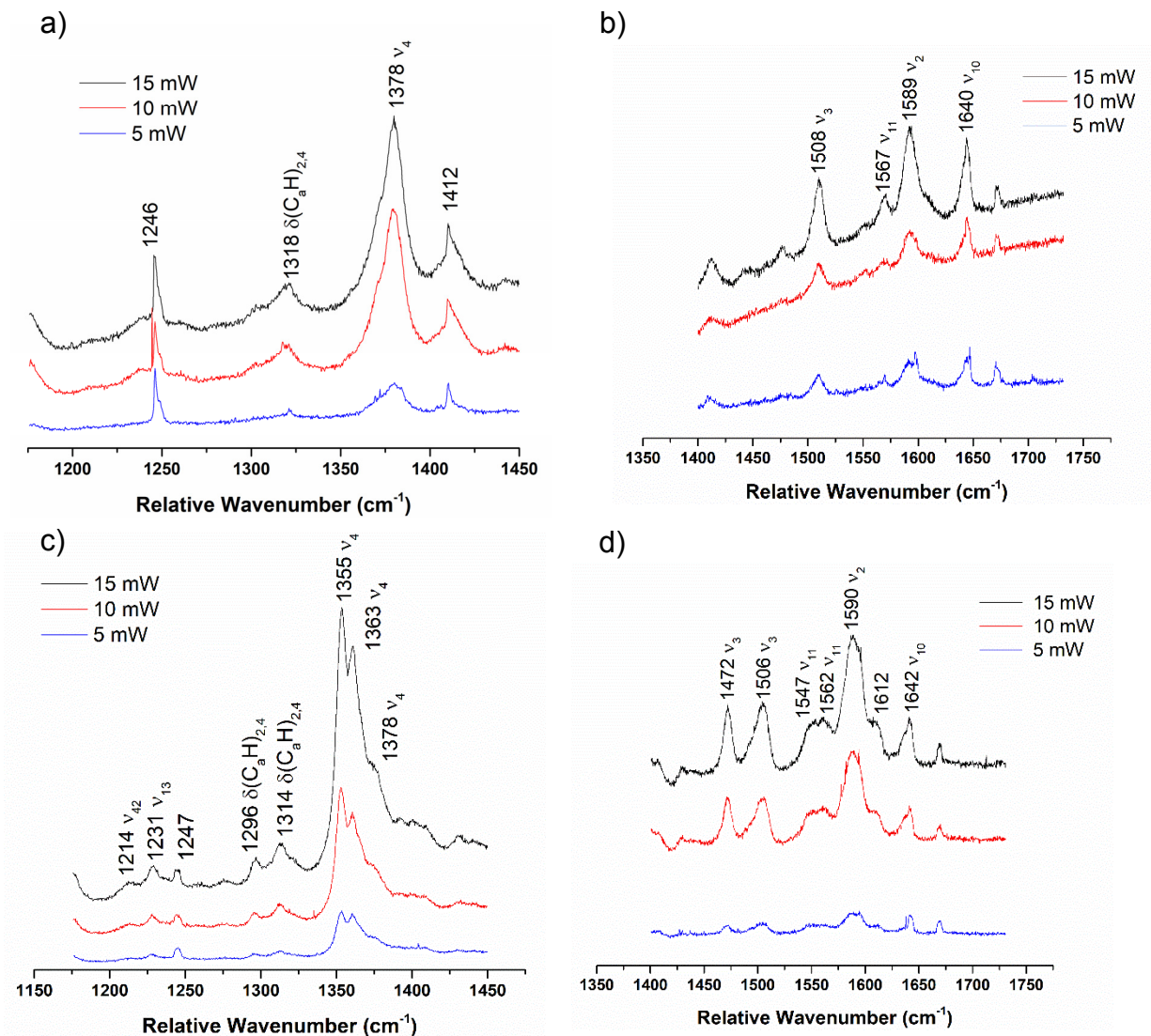


**Figure S5.** Room temperature resonance Raman spectrum of a) semireduced *NeCcP* and b) semireduced *NeH59G CcP*. The samples were  $\sim 200 \mu\text{M}$  in protein with 30% glycerol in pH 7 TIP7 buffer. The samples were excited at 406.7 nm using a tunable titanium:sapphire laser.

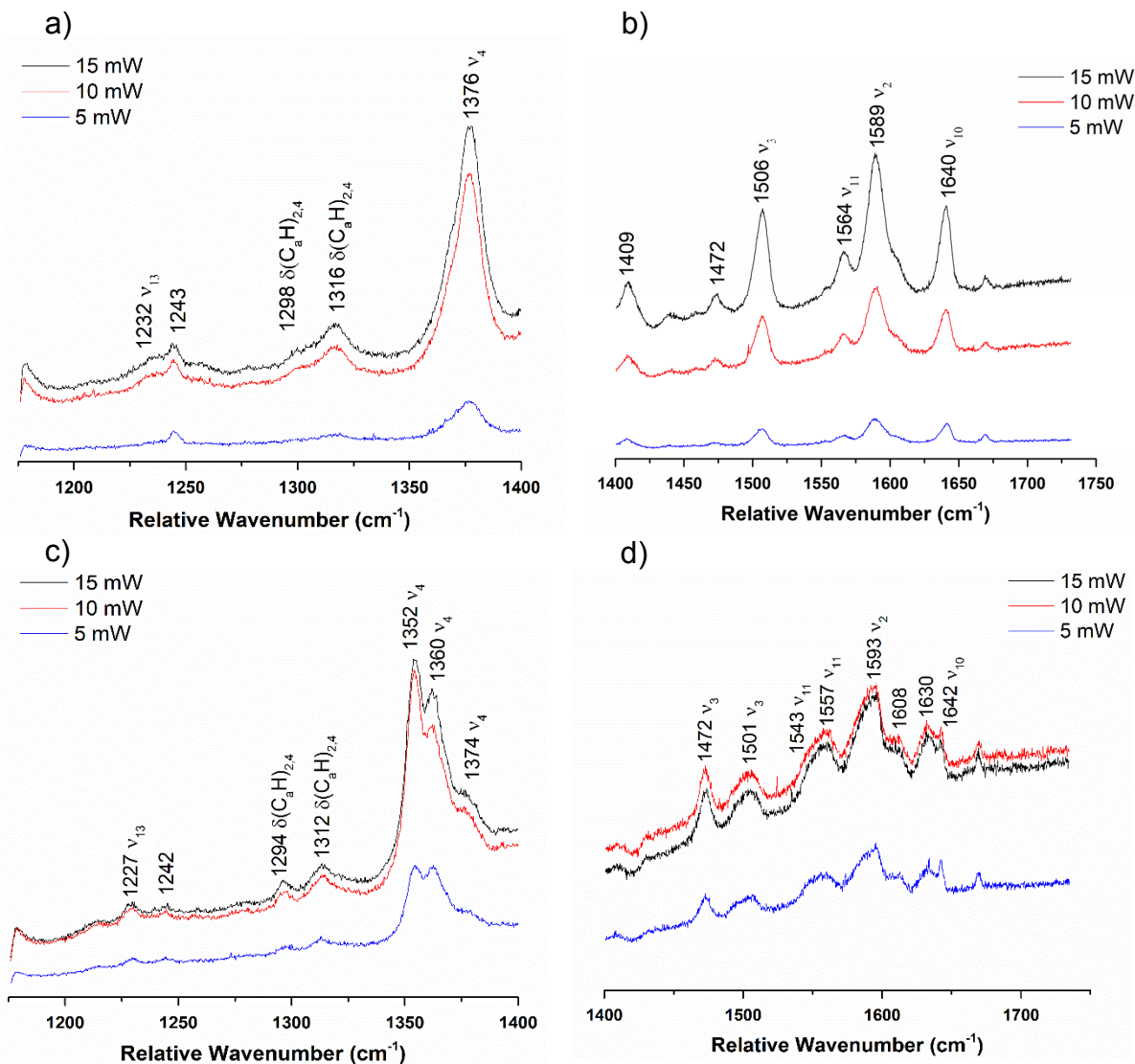


**Figure S6.** Near-UV/visible-region MCD spectra of the diferric and semi-reduced states of NeH59G CcP. Samples were 2-2.6  $\mu$ M with ~50% glycerol in a TIP7 buffer at pH 7. Spectra were recorded at 2 K.at pH 7. Spectra were recorded at 2 K.

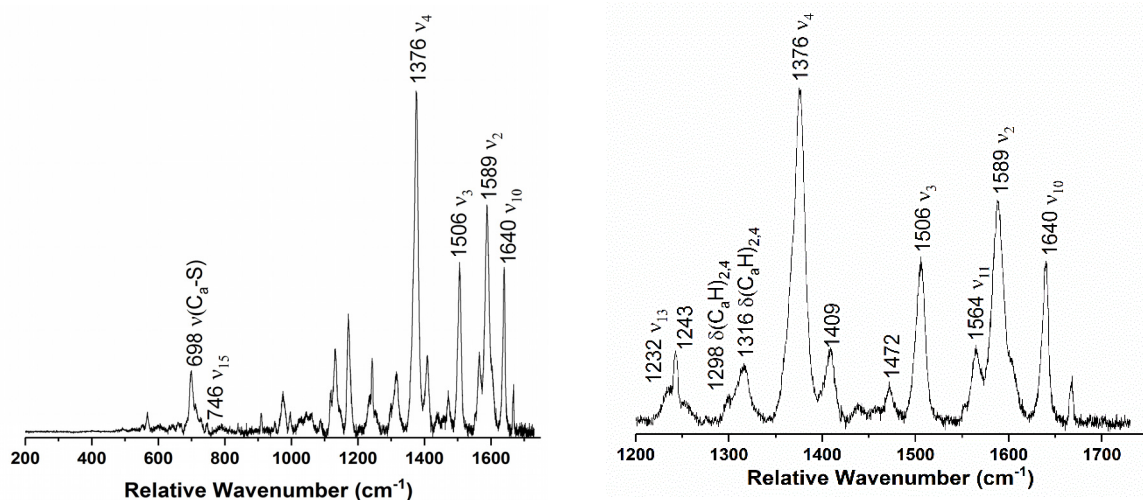




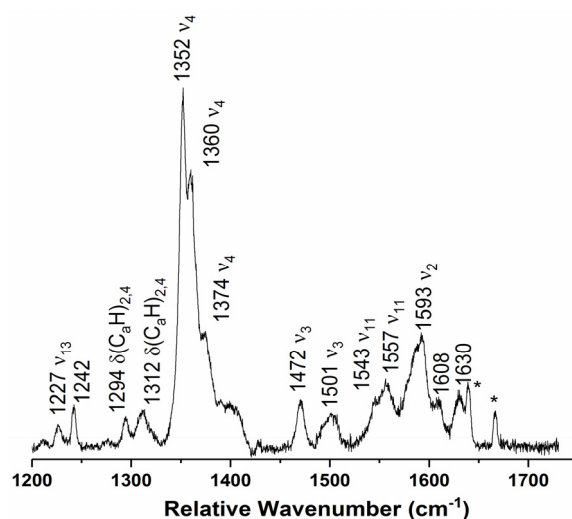
**Figure S7.** Power dependence resonance Raman spectra of **diferric SoCCP** from a) 1150 to 1400  $\text{cm}^{-1}$ , and b) 1400 to 1750  $\text{cm}^{-1}$  and the power dependence spectra of **semireduced SoCCP** from c) 1150 to 1400  $\text{cm}^{-1}$ , and d) 1400 to 1750  $\text{cm}^{-1}$ . The diferric sample was 330  $\mu\text{M}$  and the semi-reduced sample was 230  $\mu\text{M}$  in protein, both with 30% glycerol in pH 7 TIP7 buffer. The sample was excited at 413.1 nm using a Krypton ion gas laser at 77 K.



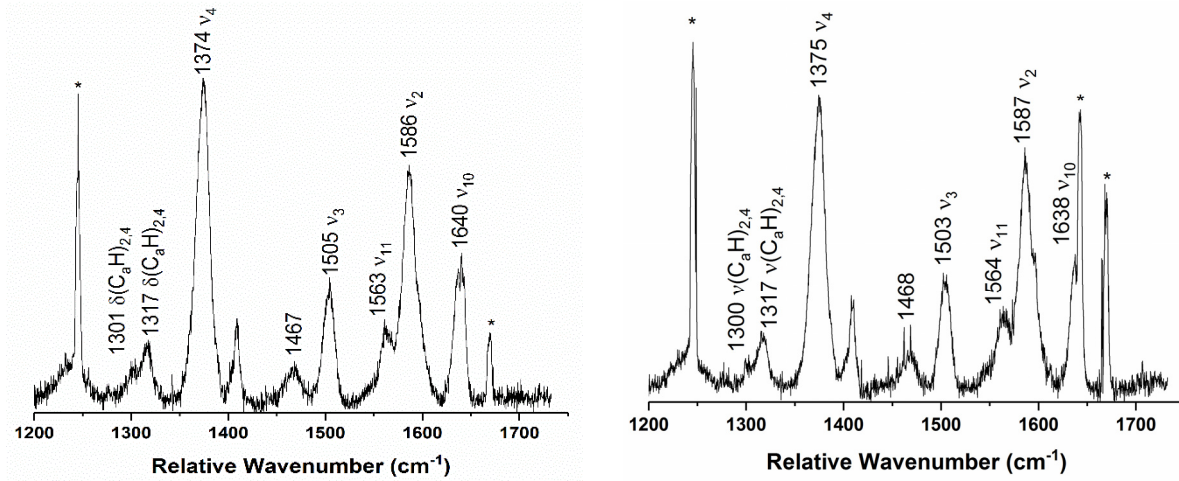
**Figure S8.** Power dependence resonance Raman spectra of **diferric SoH80G CcP** from a) 1150 to 1400  $\text{cm}^{-1}$ , and b) 1400 to 1750  $\text{cm}^{-1}$  and the power dependence spectra of **semireduced SoH80G CcP** from c) 1150 to 1400  $\text{cm}^{-1}$ , and d) 1400 to 1750  $\text{cm}^{-1}$ . The diferric sample was 280  $\mu\text{M}$  and the semi-reduced sample was 340  $\mu\text{M}$  in protein, both with 30% glycerol in pH 7 TIP7 buffer. The sample was excited at 413.1 nm using a Krypton ion gas laser at 77 K.



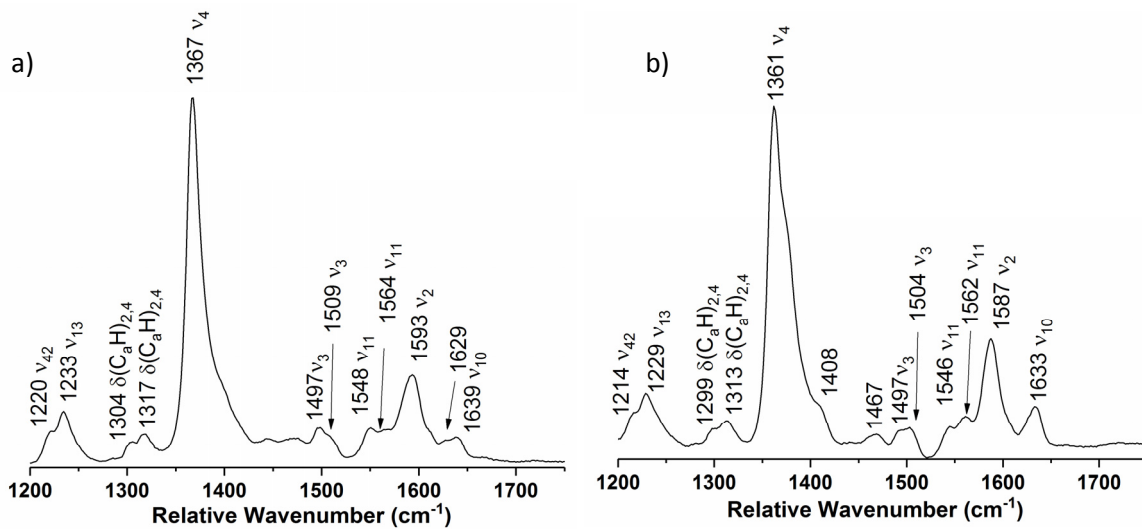
**Figure S9.** Resonance Raman spectrum of **diferric SoH80G CcP** from a) 200 to 1750 cm<sup>-1</sup>, and b) enlarged view of the 1200 to 1750 cm<sup>-1</sup> region. The sample was 280 μM in protein with 30% glycerol in pH 7 TIP7 buffer. The sample was excited at 413.1 nm using a Krypton ion gas laser at 77 K.



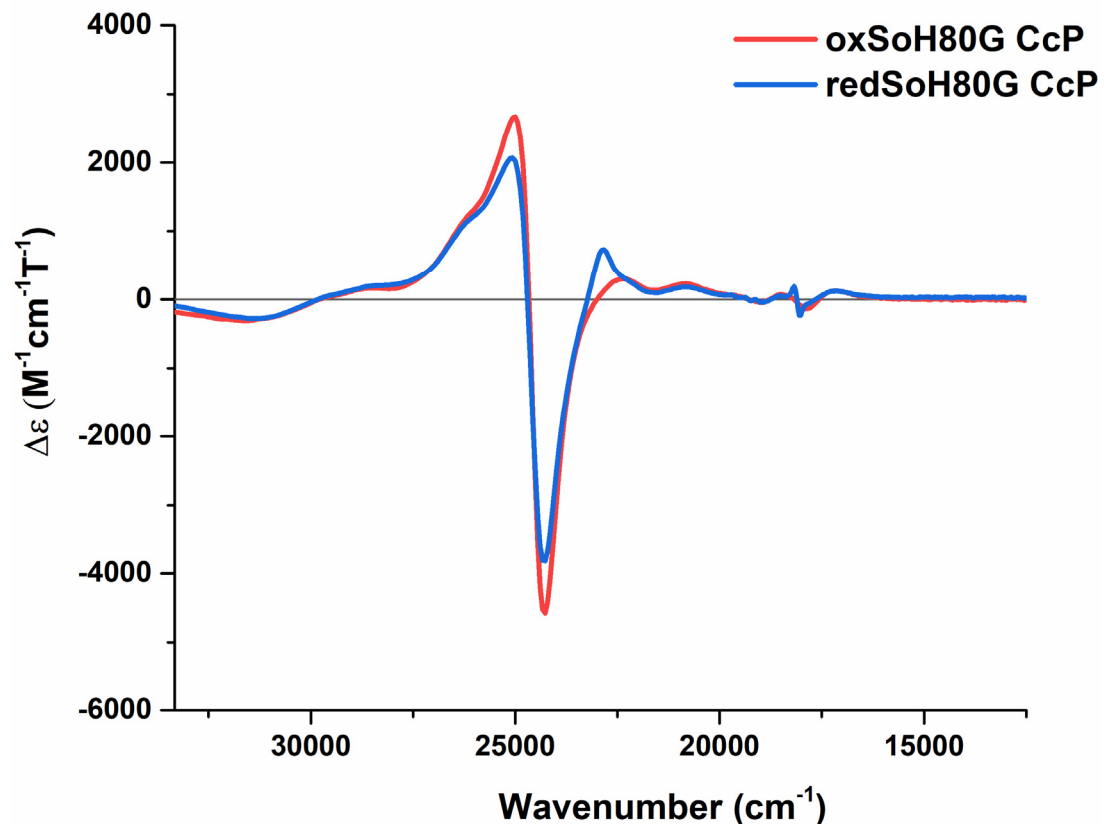
**Figure S10.** Resonance Raman spectrum of **semi-reduced SoH80G CcP** from 1200 to 1750 cm<sup>-1</sup> region. The sample was 280 μM in protein with 30% glycerol in pH 7 TIP7 buffer. The sample was excited at 413.1 nm using a Krypton ion gas laser at 77 K.



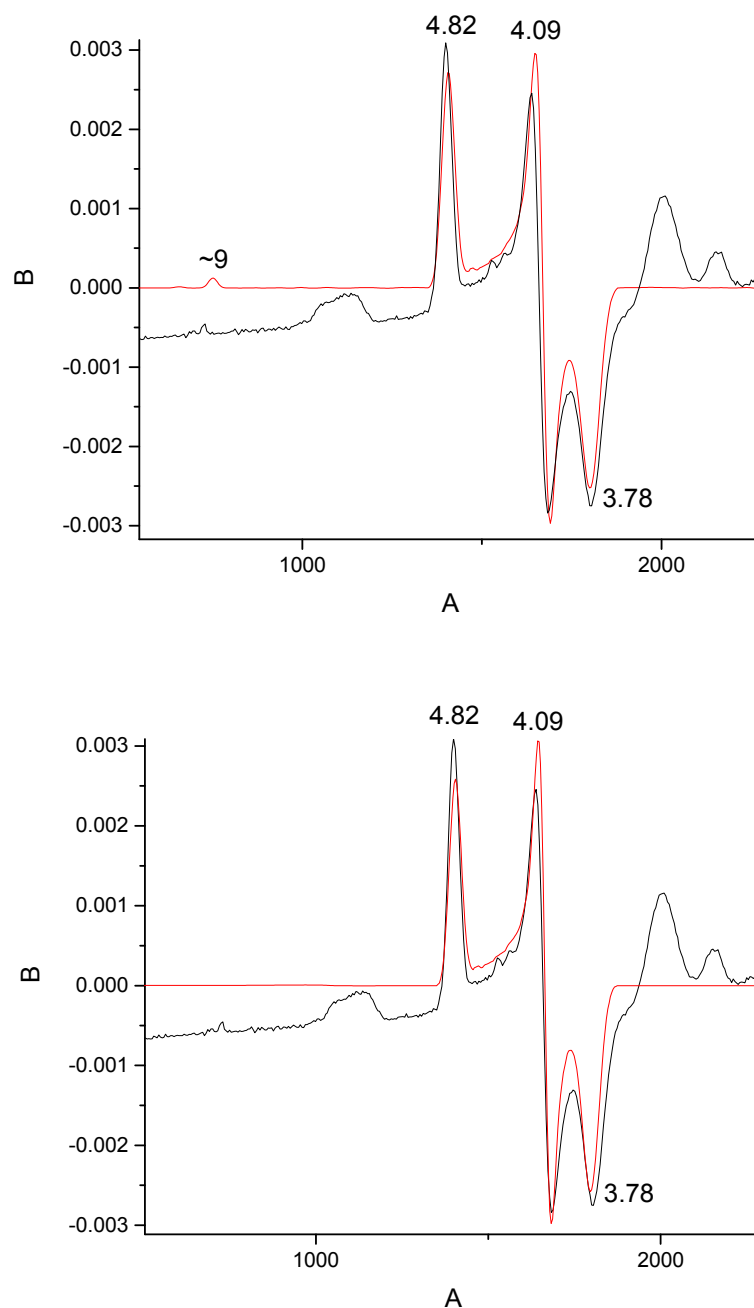
**Figure S11.** Room temperature resonance Raman spectrum of a) *diferric SoCcP* and b) *diferric SoH80G CcP*. The samples were 280-330  $\mu\text{M}$  in protein with 30% glycerol in pH 7 TIP7 buffer. The samples were rotated and excited at 413.1 nm using a Krypton ion gas laser. Plasma lines are indicated with an asterisk.



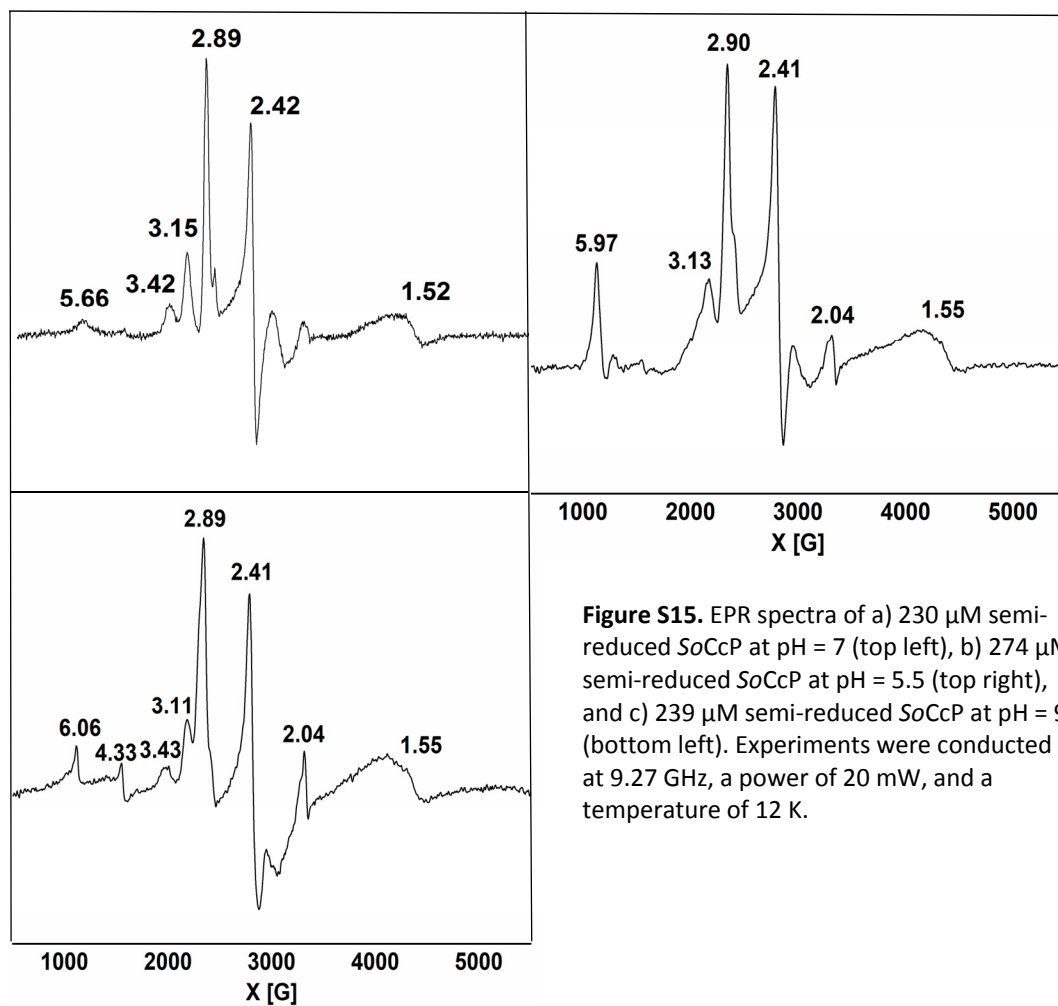
**Figure S12.** Room temperature resonance Raman spectrum of a) *semireduced SoCcP* and b) *semireduced SoH80G CcP*. The samples were  $\sim 100 \mu\text{M}$  in protein with 30% glycerol in pH 7 TIP7 buffer. The samples were excited at 406.7 nm using a tunable titanium:sapphire laser.



**Figure S13.** Near-UV/visible-region MCD spectra of the diferric and semi-reduced states of SoH80G CcP. Not all of the H-heme was reduced in the semi-reduced sample, some H-heme remained ferric. The samples were 1.2-1.4  $\mu M$  with  $\sim 50\%$  glycerol in a TIP7 buffer at pH 7. Spectra were recorded at 2K.



**Figure S14.** Fit of the highly rhombic high-spin ferric signal in the EPR spectrum of diferric NeH59G CcP (see Figure 3C). Top: fit using  $g_x = 1.94$ ,  $g_y = 1.90$ ,  $g_z = 2.11$ ,  $D = +1 \text{ cm}^{-1}$ ,  $E/D = 0.28$ ,  $T = 12 \text{ K}$ . Bottom: fit with the same parameters, except that  $D = -2.5 \text{ cm}^{-1}$ . Only the fit with a small, positive  $D$  value is able to reproduce the small signal around  $g \sim 9$ . In addition, using a larger, positive  $D$  increases the intensity of this feature, which is not in agreement with experiment. It can therefore be concluded that the highly rhombic species responsible for this signal must have a  $D$  of about  $+1 \text{ cm}^{-1}$ .



**Figure S15.** EPR spectra of a) 230  $\mu\text{M}$  semi-reduced SoCcP at pH = 7 (top left), b) 274  $\mu\text{M}$  semi-reduced SoCcP at pH = 5.5 (top right), and c) 239  $\mu\text{M}$  semi-reduced SoCcP at pH = 9 (bottom left). Experiments were conducted at 9.27 GHz, a power of 20 mW, and a temperature of 12 K.