A multigenotype therapeutic human papillomavirus vaccine elicits potent T cell responses to conserved regions of early proteins

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Gating strategy for ICS. Single cell suspensions of PBMCs, splenocytes or cervicovaginal lymphocytes were prepared and stimulated for 6.5 hours with 5GHPV3 peptide pools E1, E2, E4/5, E6 and E7 (10μ g/ml), negative control (0.1% DMSO) and positive controls (SEB, 10μ g/ml). Following surface and intracellular staining cells were acquired using a CyAn flow cytometer. Lymphocytes (A) were gated on FS lin and SS lin, singlets (B) were gated on FS lin and FS area, live cells (C) were gated on FS lin and aqua live/dead, CD3+ cells (D) were gated on FS lin and CD3 and CD4 cells (E) were gated on CD3 and CD8.



Induction of T ce II responses to 5GHPV3 administered in different vaccination regimens, as measured by IFN-γ Elispot. C57BL/6 mice (6/group) were primed intramuscularly with DNA-5GHPV3 (100µg), MVA-5GHPV3 (1x106 pfu) or ChAdOx1-5GHPV3 (1x108 IU) and then boosted intramuscularly with a heterologous or homologous vaccine two weeks later. Tail vein bleeds were performed two weeks post-prime and one and two weeks post-boost. PBMCs were stimulated with peptides spanning the entire immunogen sequence, pooled according to protein source. Data show responses to each peptide pool. Horizontal lines show mean with standard deviation.

Supplementary Figure 3



ICS was performed on PBMC from C57BL/6 mice (6/group) vaccinated with the CM regimen at one and six weeks post-boost following stimulation with 5GHPV3 E6 and E7 peptide pools. Data shown are CD8+ T cells (A) and CD8-T cells (B).

Supplementary Figure 4



CD1 mice (10/group) were primed intramuscularly with either DNA-5GHPV3 (100 μ g) or ChAdOx1-5GHPV3 (1x10⁸ IU) and then boosted two weeks later with MVA-5GHPV3 (1x106 pfu). PBMC responses were determined by IFN- γ Elispot assays at two weeks post-prime (top) and two (middle) and three (bottom) weeks post-boost after stimulation with peptides spanning the entire immunogen sequence, pooled according to protein source. Responses to each peptide pool are shown.

Supplementary Table 1

Number of full length protein sequences collected from NCBI protein database of high quality sequencing

| Protein | Genotype | | | | |
|---------|----------|----|----|-----|-----|
| | 16 | 18 | 31 | 52 | 58 |
| E1 | 126 | 49 | 24 | 27 | 53 |
| E2 | 195 | 56 | 26 | 32 | 54 |
| E4 | 197 | 48 | 24 | 24 | 60 |
| E5 | 1205 | 78 | 90 | 218 | 185 |
| E6 | 566 | 70 | 85 | 193 | 199 |
| E7 | 126 | 49 | 24 | 27 | 53 |

Supplementary Table 2

| Reagent pane | l for intracellular | cytokine staining |
|---------------------|---------------------|-------------------|
|---------------------|---------------------|-------------------|

| Specificity | Clone | Fluorochrome | Supplier |
|-------------|----------|--------------------|---------------|
| CD3 | 17A2 | Alexa Fluor 750 | eBioscience |
| CD8 | 53-6.7 | PerCP Cy5.5 | BioLegend |
| CD4 | RM4-5 | PECF594 | BD Bioscience |
| CD29 | ΗΜ β1-1 | PE (phycoerythrin) | BD Bioscience |
| CD49 | R1-2 | FITC | BioLegend |
| CD44 | IM7 | BB515 | BD Bioscience |
| CD62L | MEL-14 | PE (phycoerythrin) | BD Bioscience |
| CD127 | SB/199 | BV421 | BD Bioscience |
| IFN-γ | XMG1.2 | APC | BD Bioscience |
| IL-2 | JES6-5H4 | PE | BD Bioscience |
| ΤΝFα | MP6-XT22 | FITC | BioLegend |
| CD107 | 1D4B | BV421 | BD Bioscience |

Supplementary Table 3

Baseline characteristics of study participants

| | Cohort 1 | Cohort 2 |
|--|------------|------------------------------|
| n | 106 | 34 |
| Age, mean (range) | 21 (16-24) | 32 (25-47) |
| Vaccinated (at least 1 dose, bi/quadrivalent) | 93 (88%) | 7 (21%) |
| Baseline HR HPV DNA positive | 27ª (25%) | 31 ^{<i>b</i>} (91%) |
| Age at sexual debut, mean (range) | 17 (14-24) | 17 (14-29) |

a All hrHPV were Other (non-16, non-18) types

b hrHPV types were: Other (n = 23), Other + HPV16 (n = 3), Other + HPV18 (n = 1), HPV16 only (n = 2), HPV18 only (n = 2)