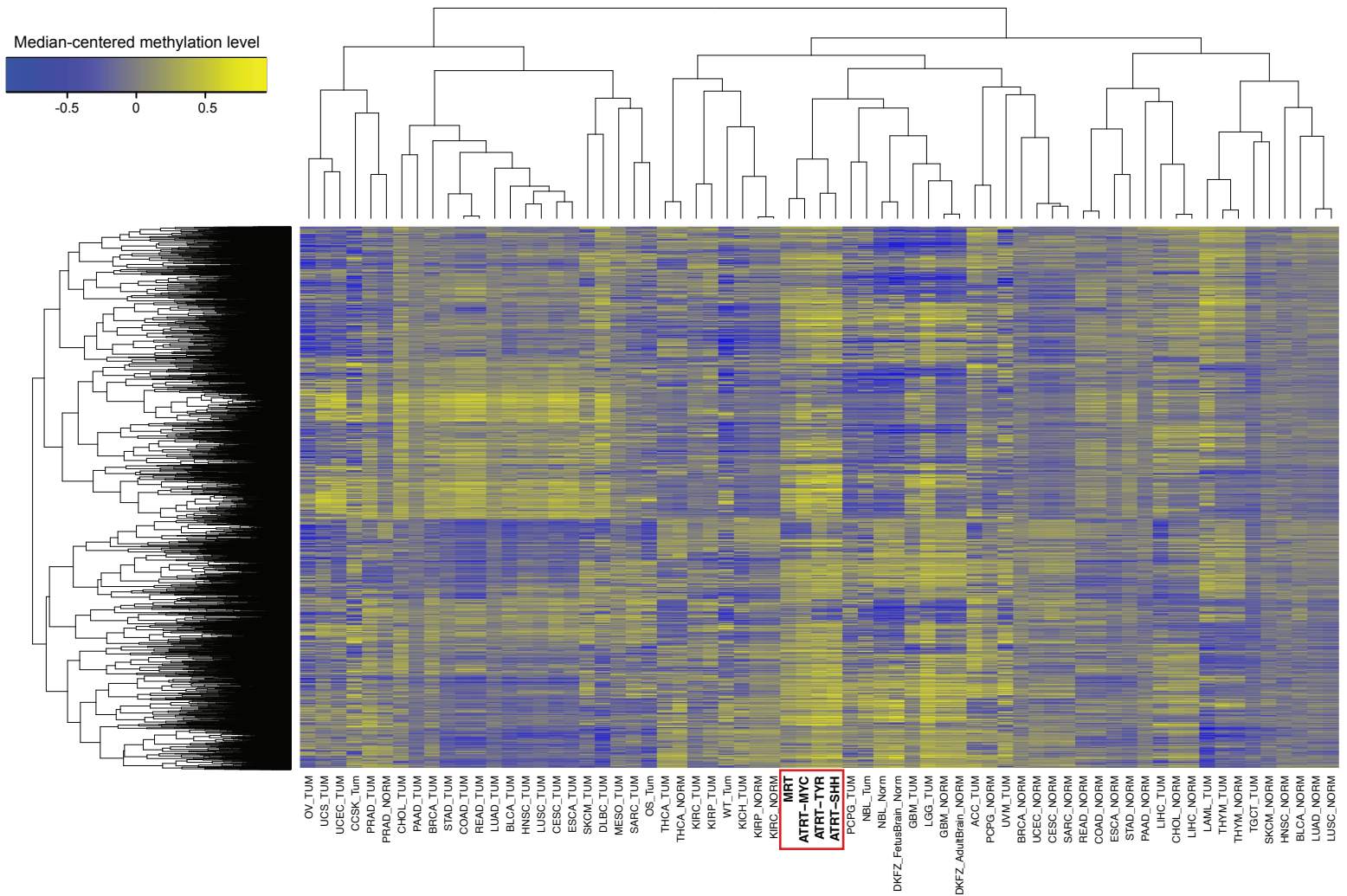


## Supplemental Information

### Identification and Analyses of Extra-Cranial and Cranial Rhabdoid Tumor Molecular Subgroups Reveal Tumors with Cytotoxic T Cell Infiltration

Hye-Jung E. Chun, Pascal D. Johann, Katy Milne, Marc Zapatka, Annette Buellesbach, Naveed Ishaque, Murat Iskar, Serap Erkek, Lisa Wei, Basile Tessier-Cloutier, Jake Lever, Emma Titmuss, James T. Topham, Reanne Bowlby, Eric Chuah, Karen L. Mungall, Yussanne Ma, Andrew J. Mungall, Richard A. Moore, Michael D. Taylor, Daniela S. Gerhard, Steven J.M. Jones, Andrey Korshunov, Manfred Gessler, Kornelius Kerl, Martin Hasselblatt, Michael C. Frühwald, Elizabeth J. Perlman, Brad H. Nelson, Stefan M. Pfister, Marco A. Marra, and Marcel Kool

# Figure S1

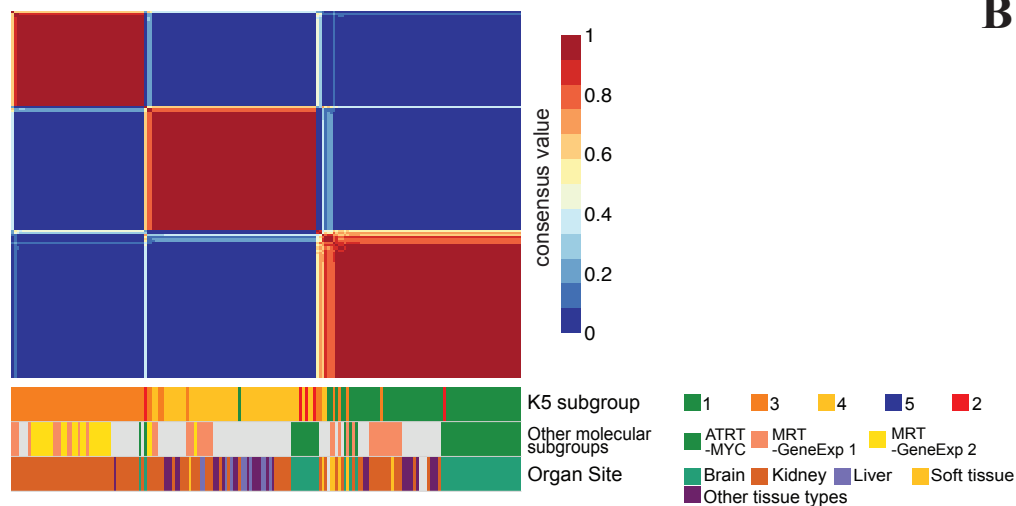


**Figure S1. Comparison of DNA methylation profiles from MRTs and ATRTs to TCGA cases revealed that RTs are distinct from adult tumor and differentiated tissue types, and show relative similarities to cancers originating from neural crest-derived cell types, brain cancers and normal brain tissues. Related to Figure 1.**

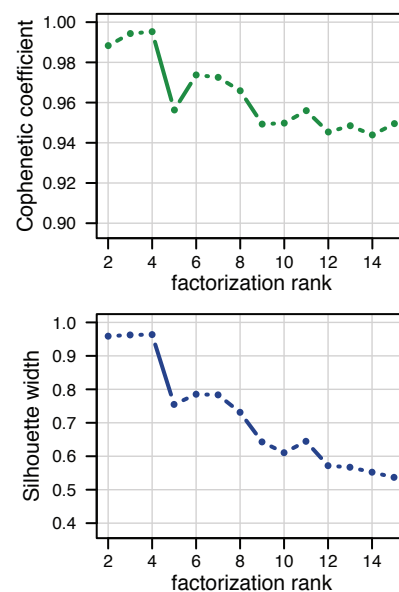
Unsupervised hierarchical clustering of 301 MRT and ATRT cases, 9,757 TCGA cases and 475 TARGET cases representing 33 different tumor types and 23 normal tissue types was performed using DNA methylation array data. Median methylation levels were used to represent each tumor and normal tissue type. The top 8,000 most variable median values were used for the clustering analysis. A suffix, “\_TUM”, indicates an adult tumor case, whereas “\_Tum” denotes a pediatric tumor case. “\_NORM” indicates a matched adult normal case, whereas “\_Norm” indicates a pediatric normal case. Disease abbreviations are listed in the STAR Methods.

# Figure S2

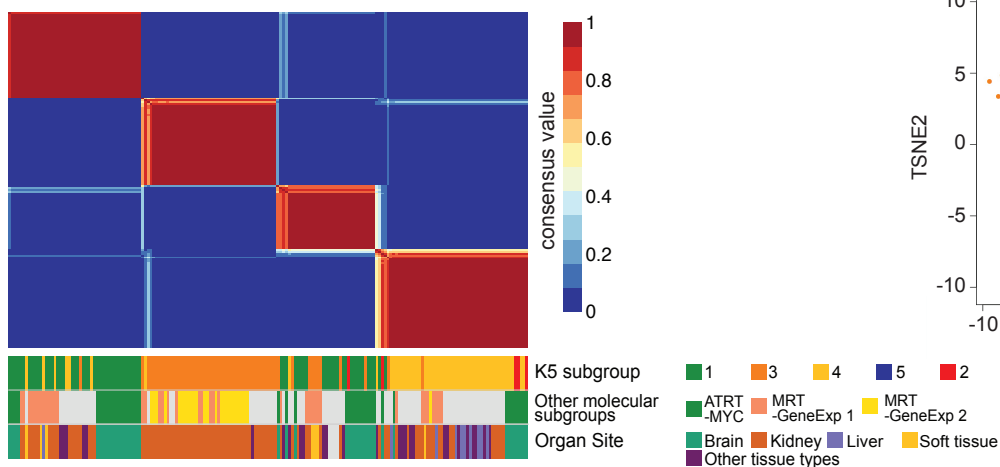
## A



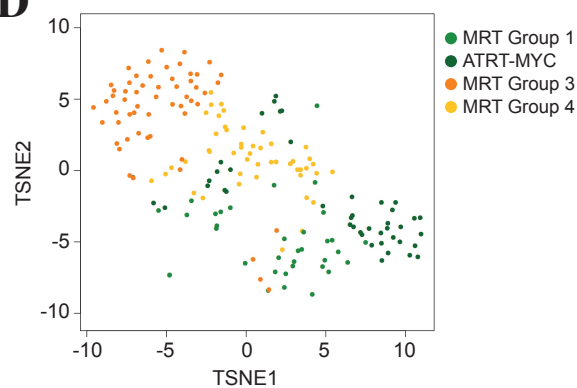
## B



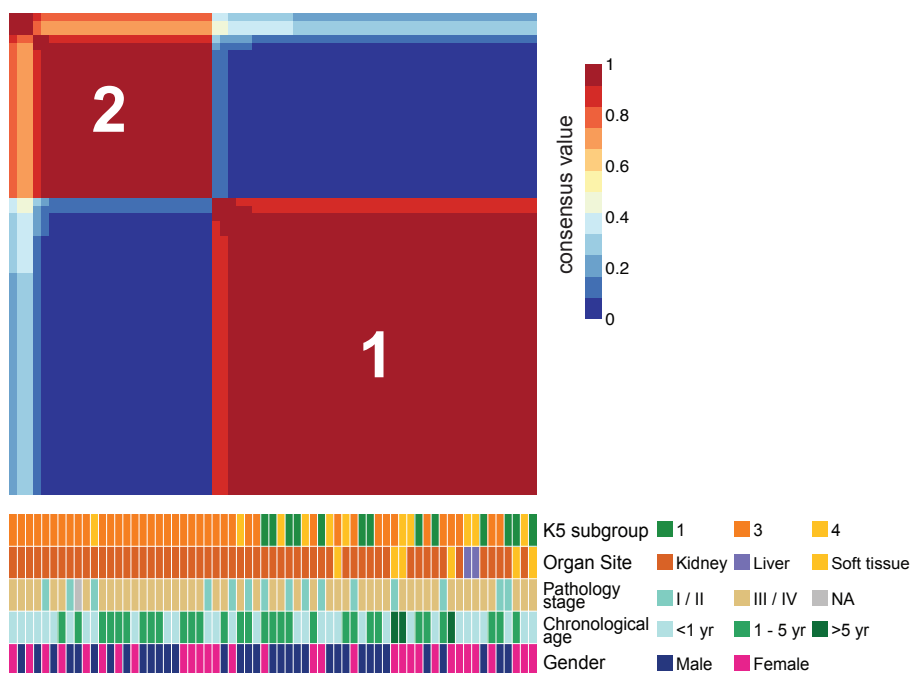
## C



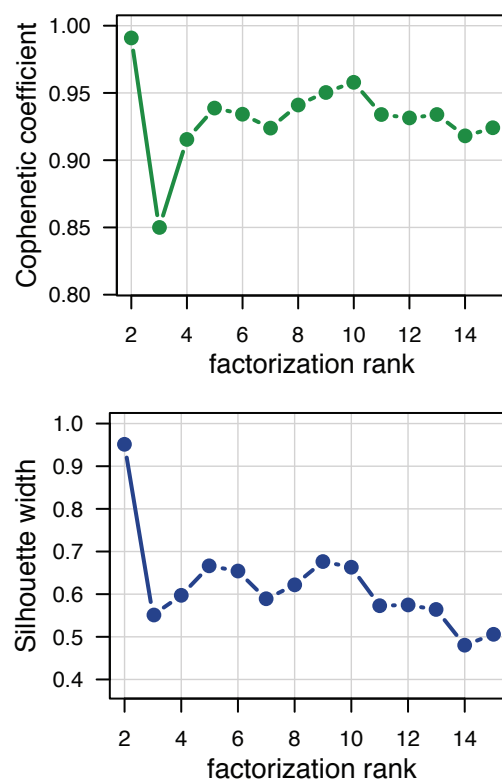
## D



## E



## F



**Figure S2. Clustering of MRT and ATRT-MYC cases reveals subgroups consistent with the five RT subgroups. Related to Figure 2.**

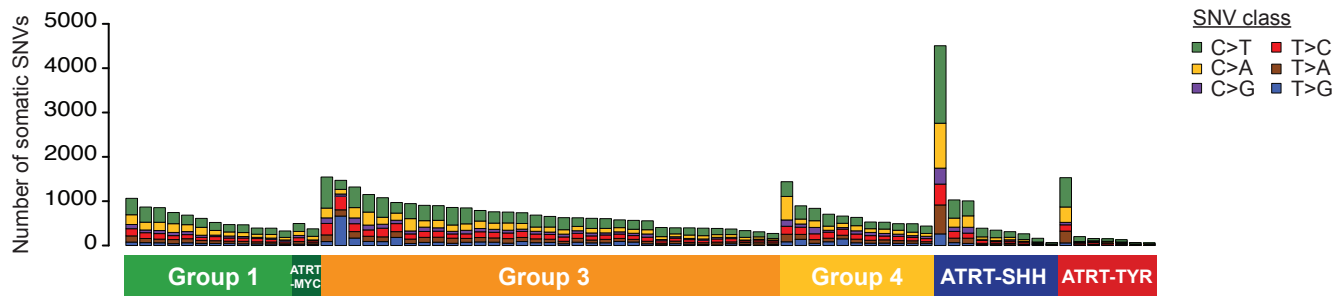
Unsupervised NMF analysis was performed using top 10,000 most variably methylated CpG sites from 140 MRT and 44 ATRT-MYC samples. The analysis revealed the most robust clustering solutions at  $k=3$  (**A**) and  $k=4$  (**C**). Cophenetic coefficients (top) and silhouette widths (bottom) indicating the robustness of NMF clustering from  $k=2$  to  $k=15$  are shown (**B**). Clinical information, and previously described ATRT subgroups and MRT gene-expression subgroups (in **Figure S2E**) are shown in colored tracks.

**(D)** t-SNE analysis was performed using the top 10,000 most variably methylated CpG sites from 140 MRT and 44 ATRT-MYC samples, and supported the existence of Groups 1, 3 and 4 also detected using NMF.

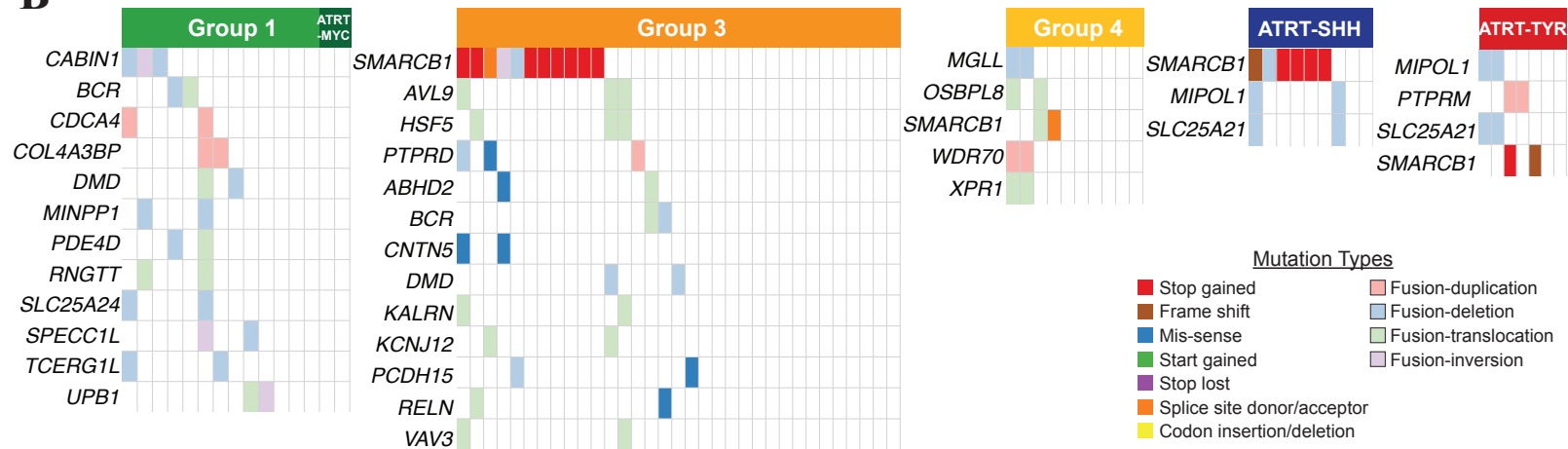
**(E)** NMF consensus matrix from clustering RNA-Seq data of 65 MRT cases (top). Clinical correlates are shown in the bottom panel. The gene expression subgroup 1 contains all extra-renal MRT cases, while gene expression subgroup 2 exclusively contains RTK cases. Metrics indicating the robustness of clustering solutions from  $k = 2$  to  $k = 15$  are shown in (**F**).

# Figure S3

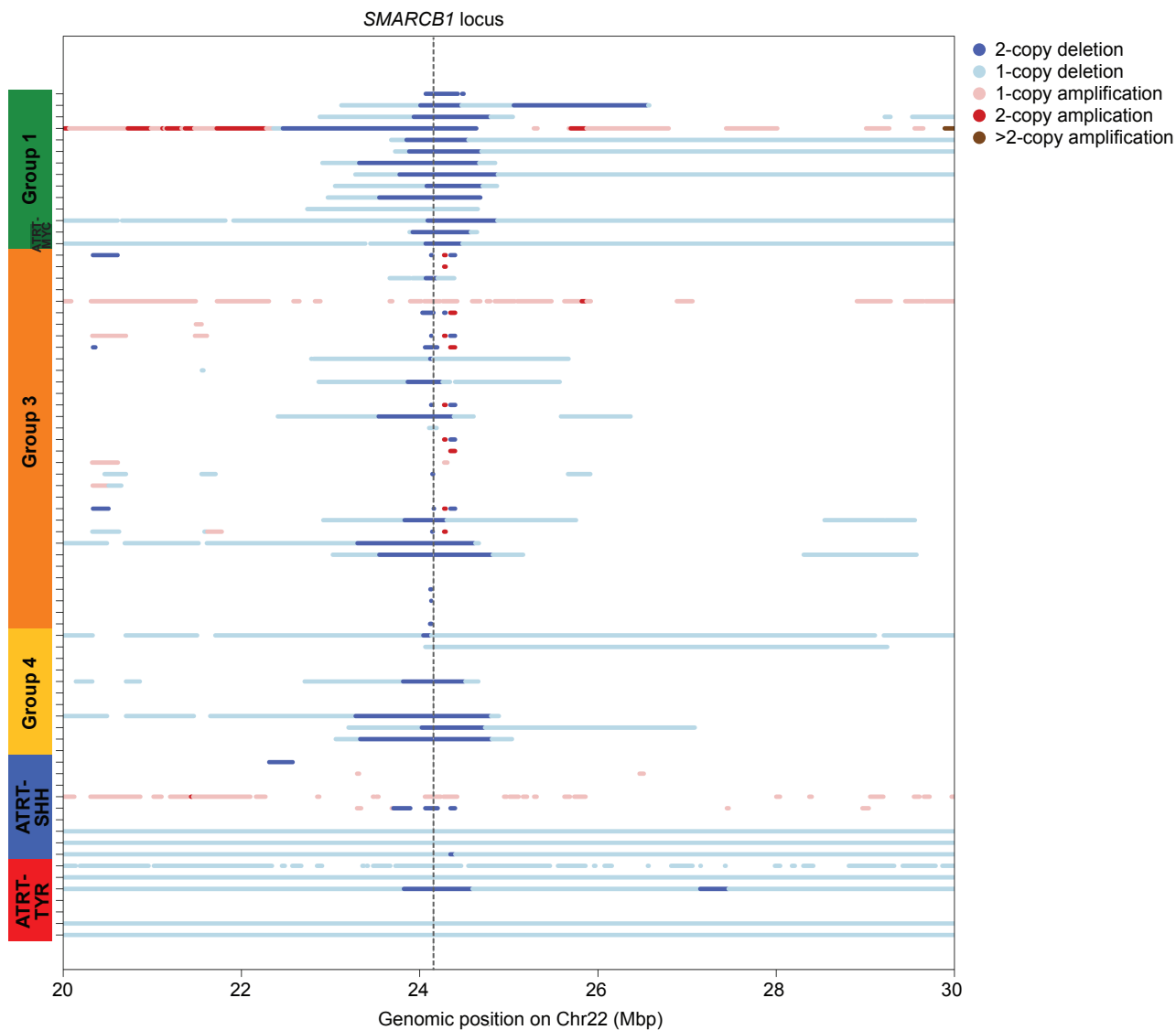
**A**



**B**



**C**



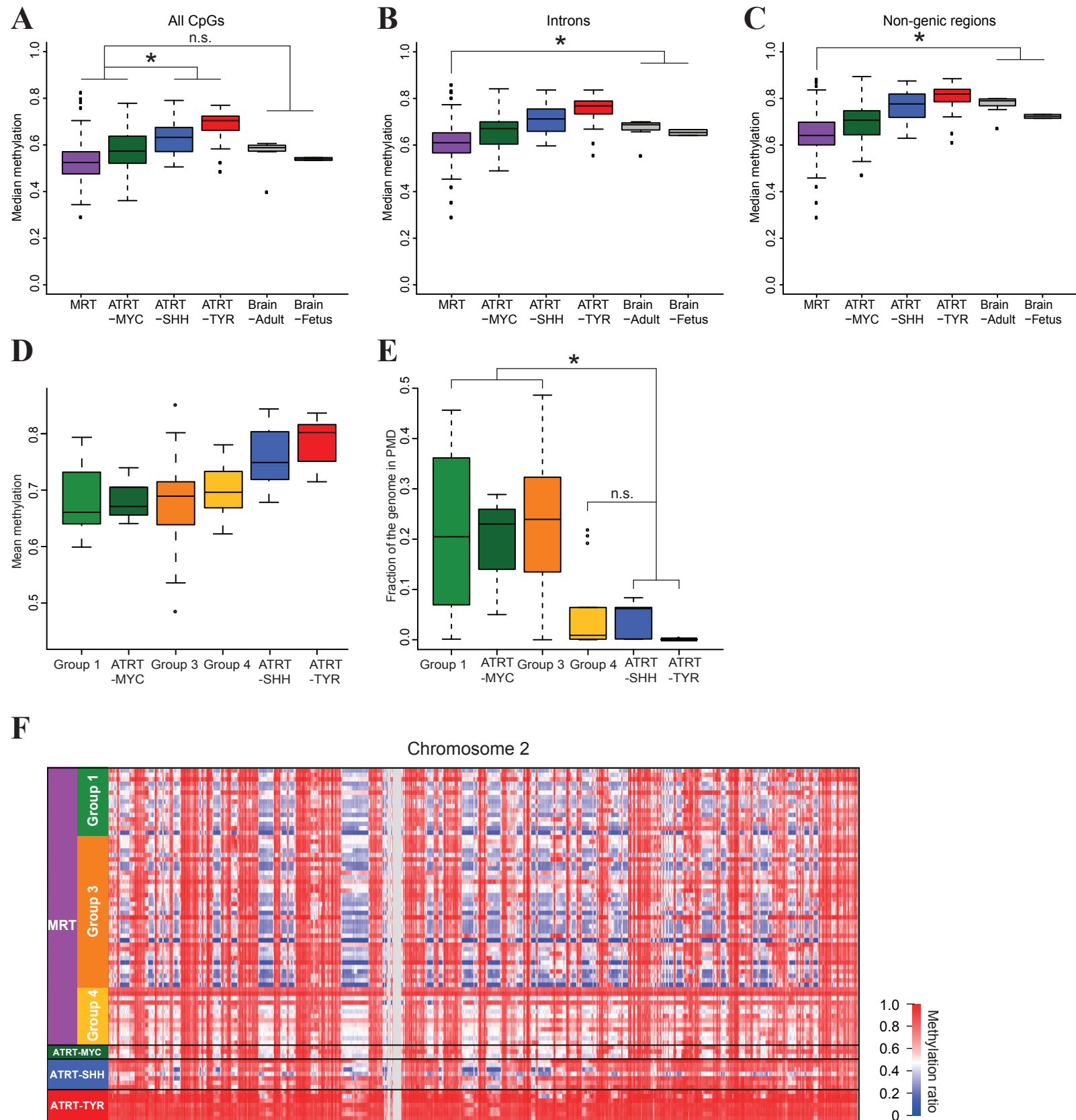
**Figure S3. Mutation profiles across RT subgroups. Related to Figure 2.**

**(A)** Barplot shows the number and classes of somatic single nucleotide variants (SNVs) found across RT subgroups.

**(B)** Oncoprints show recurrently mutated genes across RT subgroups. Different mutation types are indicated by colored boxes for MRT and ATRT cases.

**(C)** Copy number alteration events at the *SMARCB1* locus (Chr22: 20,000,000 – 30,000,000 bp) across RT subgroups. Somatic copy number alterations were identified using WGS data from 74 tumor-normal pairs.

# Figure S4



**Figure S4. Analyses of DNA methylation array and WGBS data consistently reveal that MRT and ATRT-MYC cases show relative hypomethylation compared to ATRT-SHH and -TYR cases. Related to Figure 3.**

DNA methylation levels were profiled for 301 RT cases and 10 normal brain cases using Illumina 450K and 850K arrays. Boxplots show distributions of median methylation levels of all CpG sites targeted by the arrays **(A)**, median methylation levels in introns of protein-coding genes **(B)**, and in non-genic regions **(C)**; \* Wilcoxon p-value < 0.05; n.s. = not significant).

**(D)** Boxplot shows mean global DNA methylation levels based on WGBS data across five RT subgroups.

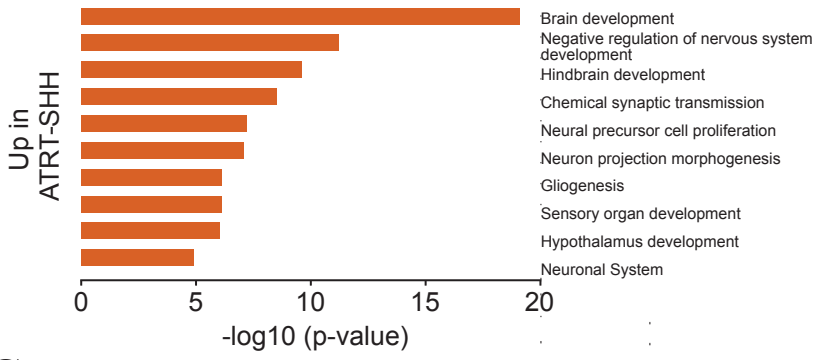
**(E)** Boxplot shows the fraction of a genome covered by PMDs across five RT subgroups (\* Wilcoxon p-value < 0.05; n.s. = not significant).

**(F)** Heatmap shows CpG methylation levels across chromosome 2 in RT subgroups using WGBS data (red = 100%; blue = 0% methylation).

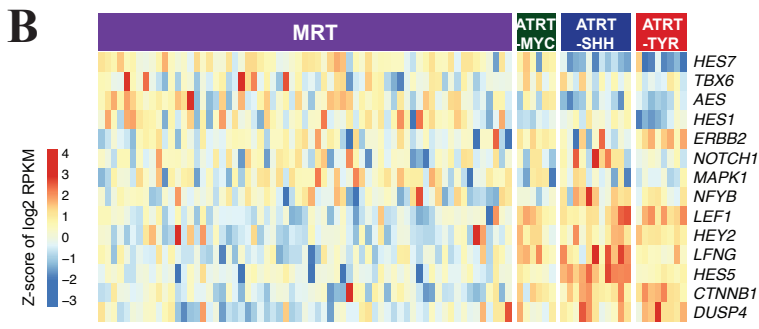


# Figure S5

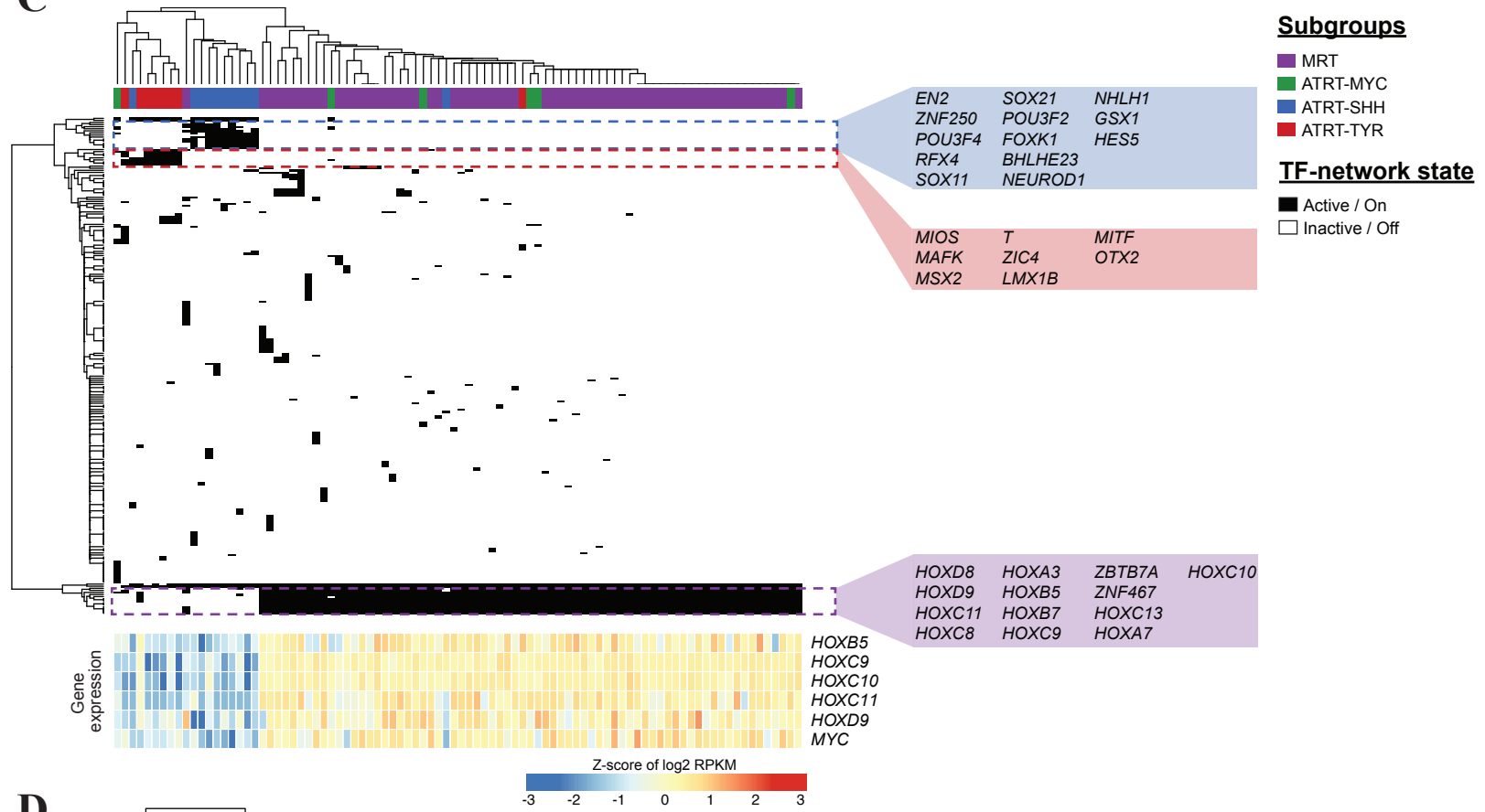
## A



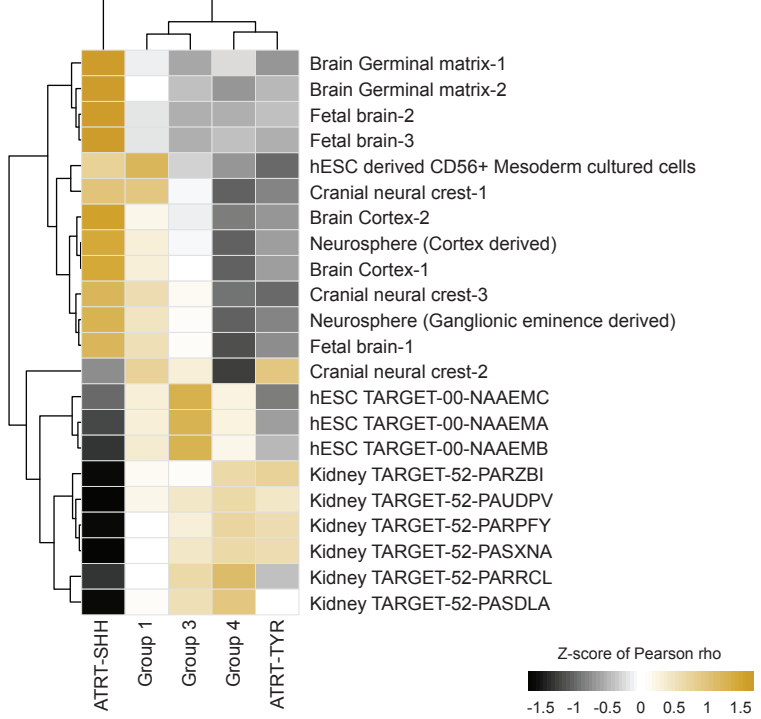
## B



## C



## D



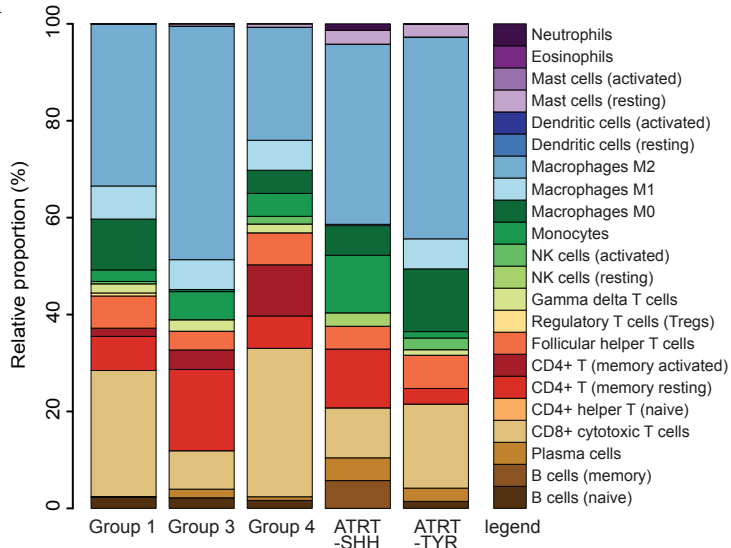
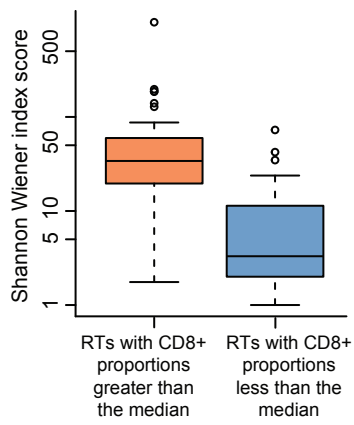
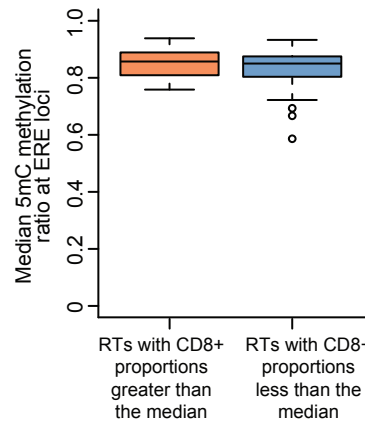
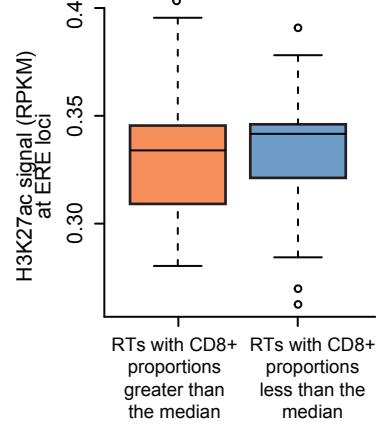
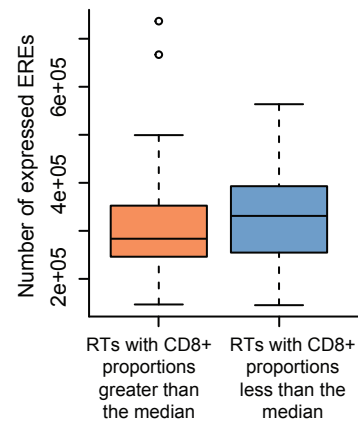
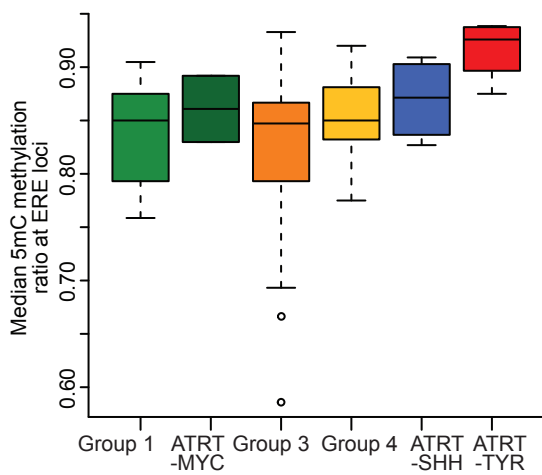
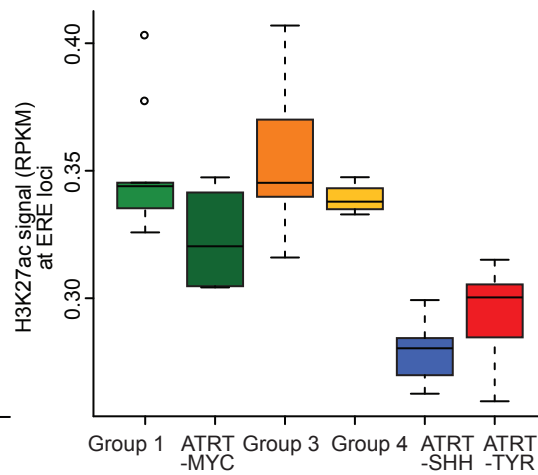
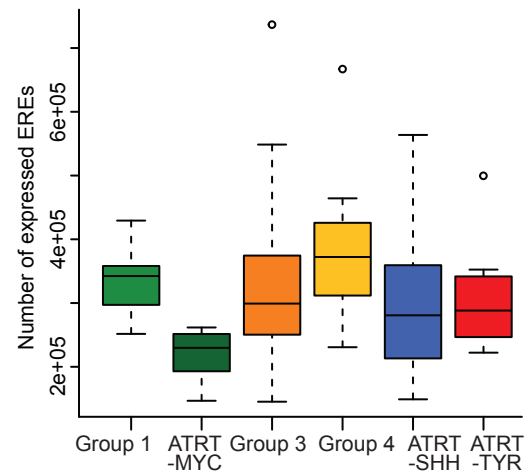
**Figure S5. Analyses of TF-regulatory networks and transcriptome comparisons to different progenitor cell types. Related to Figure 5.**

**(A)** Bar plot shows the significance of a pathway enrichment test ( $-\log_{10}$  of p-values) for genes that were over-expressed in ATRT-SHH compared to ATRT-TYR.

**(B)** Heatmap shows expression levels of genes that interact with *HES7* in MRT and ATRT. Relative over-expression of *HES7* and under-expression of its negative downstream target genes including *LEFI*, *DUSP4*, and *CTNNB1* are shown in MRT and ATRT-MYC compared to ATRT-SHH and -TYR.

**(C)** Unsupervised clustering of enrichment of TF-regulatory networks i.e. groups of TF and genes with TF motifs that show co-expression patterns across MRTs and ATRTs (top panel). ATRT subgroups and MRT cases are indicated by different colors. Networks unique to the ATRT-MYC and MRT group, ATRT-SHH, and ATRT-TYR are indicated in dashed boxes. TFs for these networks are listed in larger fonts. Expression levels of TFs whose regulatory networks contained *MYC* as a putative direct target gene are shown in a heatmap (bottom panel).

**(D)** Unsupervised hierarchical clustering that compared transcriptome profiles of RTs against various progenitor cell types including human embryonic stem cell (hESC) lines from the TARGET consortium, mesodermal progenitors (i.e. hESC-derived CD56+ mesoderm cultured cells), cranial neural crest cell cultures, neural progenitors (i.e. neurospheres derived from ganglionic eminence and cortex, germinal matrix), fetal brains, adult brains (brain cortex), and pediatric kidneys from the TARGET consortium. Pearson correlation coefficients were transformed to Z scores to show the deviation from the mean for each cell types.

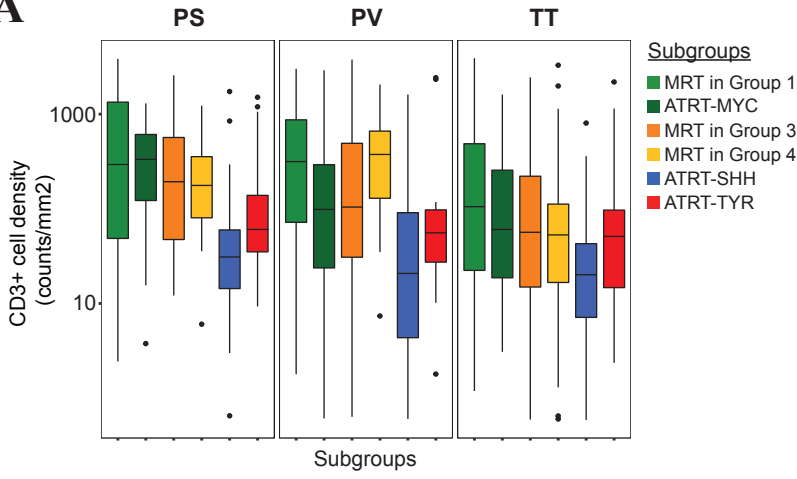
**Figure S6****A****B****C****D****E****F****G****H**

**Figure S6. Analyses of tumor purity levels, filtered pathway enrichment, CIBERSORT-based immune cell type deconvolution, and epigenetic profiles at ERE loci. Related to Figure 6.**

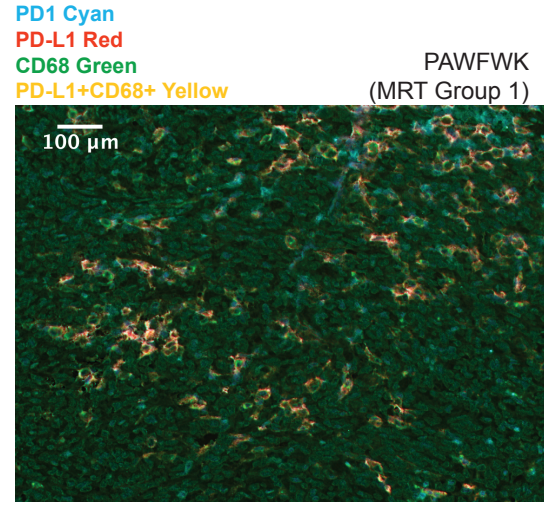
(A) Stacked bar plot shows relative proportions of 22 immune cell types predicted using CIBERSORT. Boxplots show distributions of Shannon Wiener index scores representing TCR diversity (B; higher Shannon Wiener index scores indicate greater TCR diversity); median DNA methylation levels at known ERE loci (C); H3K27ac signal density (D); and numbers of EREs that had normalized read coverage greater than the median (representing the level of transcript abundance in RPM; E) in RTs with CD8+ cytotoxic T cell proportions greater or less than the median of the cohort. Boxplots show distributions of median DNA methylation levels at known ERE loci (F); H3K27ac signal density (G); and numbers of EREs that were expressed at the level greater than the median (H) across RT subgroups.

# Figure S7

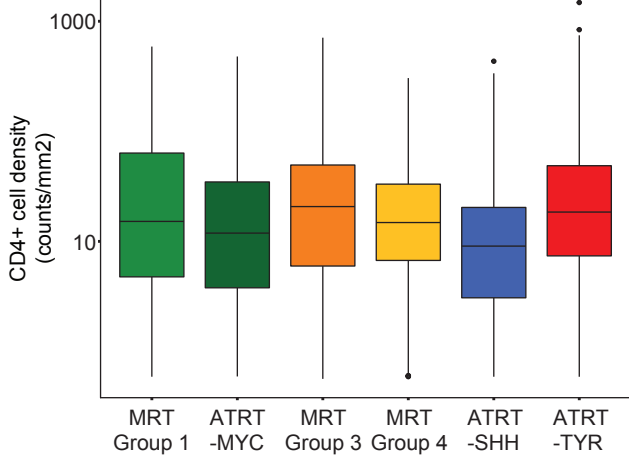
## A



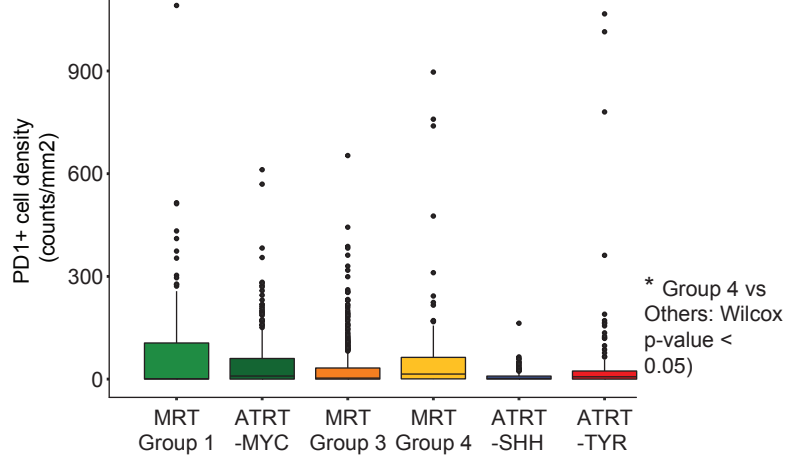
## B



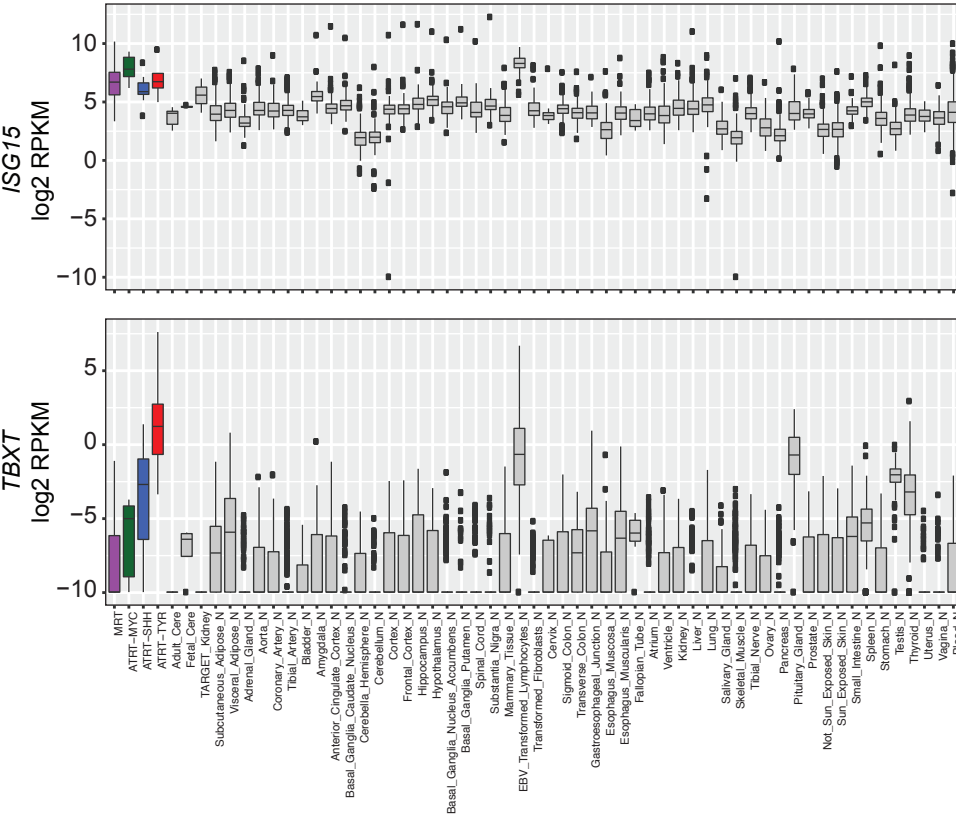
## C



## D



## E



**Figure S7. IHC validation experiment results. Related to Figure 7.**

**(A)** Boxplots show distributions of CD3+ leukocyte density (y axis, log10 scale) in tumor-enriched (TT), peri-stromal (PS), and peri-vascular (PV) tissues. MRT cases in Groups 1, 3, 4 and ATRT-MYC cases showed significantly higher CD3+ cell densities compared to ATRT-SHH and -TYR in all regions (Wilcoxon p-values = 1.041e-11, 2.932e-11, and 1.026e-09, respectively). CD3+ T cell density is the sum of CD3+CD8+ and CD3+CD8- cell counts per mm<sup>2</sup>.

**(B)** Example of a case with high PD-L1+ CD68+ immune cell infiltration, revealed by multiplex IHC staining of PD-L1 (red) and CD68 (green). Overlapping colors from PD-L1+CD68+ cells are shown in yellow. A scale bar: 100µm.

**(C)** Boxplot shows distributions of CD4+ helper T cell densities across MRTs in Groups 1, 3, 4 and ATRT-MYC, -SHH, and -TYR (y axis, log10 scale).

**(D)** Boxplot shows distributions of PD1-expressing leukocyte densities, with Group 4 exhibiting the highest median level among the RT subgroups (\* Wilcoxon p-value < 0.05).

**(E)** Boxplot distributions of expression levels of *ISG15* (top), and *TBXT* (bottom) across MRT, ATRT-MYC, -SHH, -TYR, adult (n = 5; labeled “Adult\_Cere”) and fetal cerebellum samples (n = 4; labeled “Fetal\_Cere”), normal kidney samples (n = 6; labeled “TARGET\_Kidney”), and samples from 52 normal tissue types from GTEx (n = 2,500; with a suffix, “\_N”).