Role of Cellular Retention and Intracellular State in Controlling Gene Delivery Efficiency of Multiple Nonviral Carriers

Ujjwal Ranjan Dahiya <sup>1,2</sup> ,Sarita Mishra<sup>1,2</sup>, Sabyasachi Chattopadhyay<sup>1</sup>, Anupama Kumari<sup>1,2</sup>, Apurva Gangal<sup>1</sup>, Munia Ganguli <sup>1,2\*</sup>

<sup>1</sup>CSIR - Institute of Genomics and Integrative Biology, Mathura Road, New Delhi 110025, India.

<sup>2</sup>Academy of Scientific and Innovative Research (AcSIR), Anusandhan Bhawan, 2 Rafi Marg, New Delhi 110001, India.

\* Address for correspondence: Munia Ganguli, Lab-219, Discovery Genomics Building, CSIR-Institute of Genomics and Integrative Biology, Mathura Road, New Delhi- 110025. Tel: 011-29879225 Email id: <u>mganguli@igib.res.in</u>, <u>mganguli@igib.in</u>

## **Supplementary Information:**

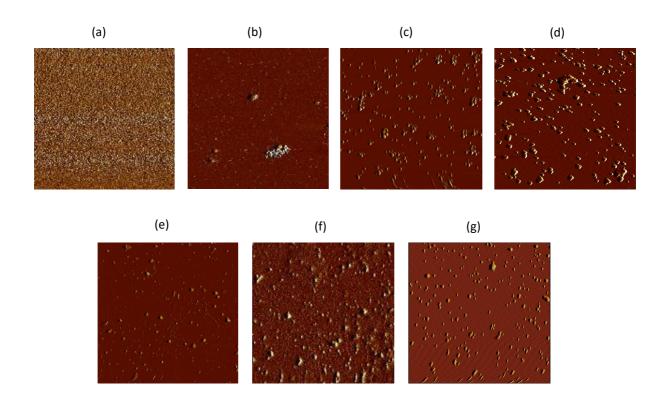


Figure S1: Characterization of shape and size of different nanocomplexes prepared at charge ratio Z (+/-) 5 using AFM (Atomic force microscopy): (a) control (bare mica) (b) M1 (c) M3 (d) M4 (e) Lipofectamine (f) M9 and (g) PEI nanocomplexes. Amplitude images of area  $10\mu m \times 10\mu m$  are shown in panels for different nanocomplexes.

| Primer         | Sequences            |  |  |
|----------------|----------------------|--|--|
| Forward primer | AGTCCCGTTGATTTTGGTGC |  |  |
| Reverse primer | CAGTACATCAATGGGCGTGG |  |  |

Table S1: Primer sequences designed for qPCR, against backbone of pMIR plasmid DNA.

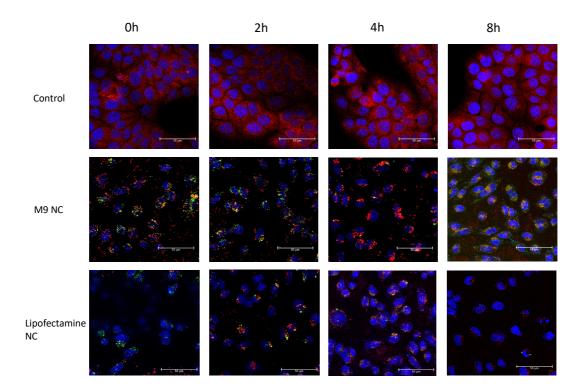
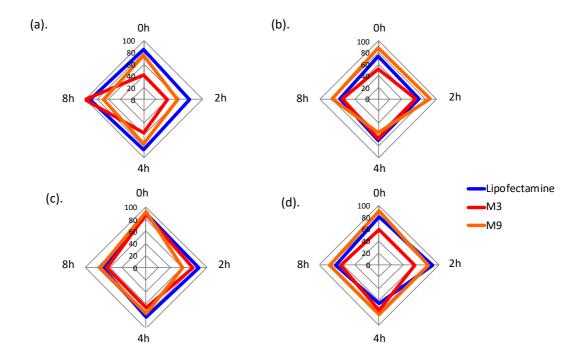
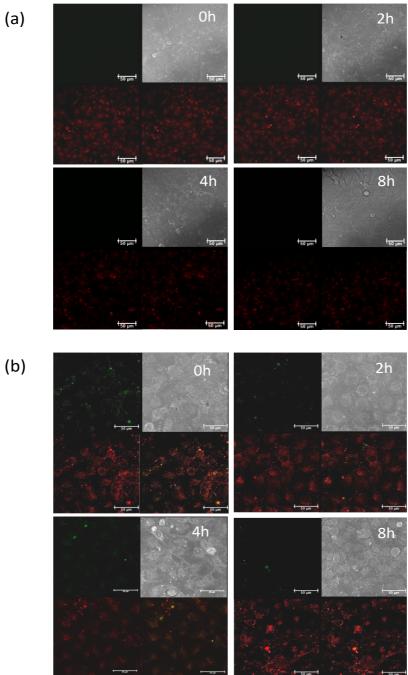


Figure S2: Confocal Imaging for tracking nanocomplex retention with time: B16-F10 cells were incubated with FITC labeled M9 and Lipofectamine nanocomplexes (NC) for 4h, followed by media change and fixing cells at 0, 2, 4 and 8h. Nucleus was stained with Hoechst and plasma membrane with Cell Mask orange plasma.



Figures S3: qPCR-based plasmid DNA release study: Amount of complexed plasmid DNA as percentage of total plasmid DNA at each time point is plotted in quadrilateral plots: (a) CHOK-1 (b) HaCaT (c) B16-F10 and (d) HEK-293 cells.



(a)

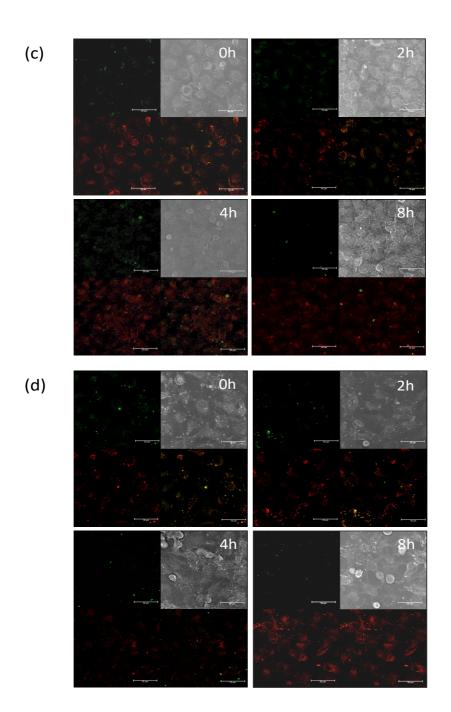


Figure S4: Lysosomal co-localization of nanocomplexes (NCs) with time: NCs formed using FITC-labeled DNA (green) were studied for co-localization with Lysosome (red labeled with Lysotracker Red) in B16-F10 cell line at 0, 2, 4 and 8h: (a) Control, (b) M3-NC, (c) M9-NC and (d) Lipofectamine NC.

| Nanocomplexes    | 0h          | 2h              | 4h          | 8h          |
|------------------|-------------|-----------------|-------------|-------------|
| M3 NC            | 0.51 ± 0.05 | 0.35 ± 0.05     | 0.30 ± 0.04 | 0.17 ± 0.04 |
| M9 NC            | 0.29 ± 0.04 | 0.31 ± 0.03     | 0.28 ± 0.06 | 0.22 ± 0.00 |
| Lipofectamine NC | 0.38 ± 0.08 | $0.40 \pm 0.04$ | 0.18 ± 0.02 | 0.34 ± 0.05 |

Table S2: Lysosomal co-localization of NCs with time was studied using confocal microscopy and respective Pearson's correlation coefficient estimated from three fields (~50 cells) is reported in Table (Experiments were performed in two independent sets. Data is represented in mean ± standard deviation).

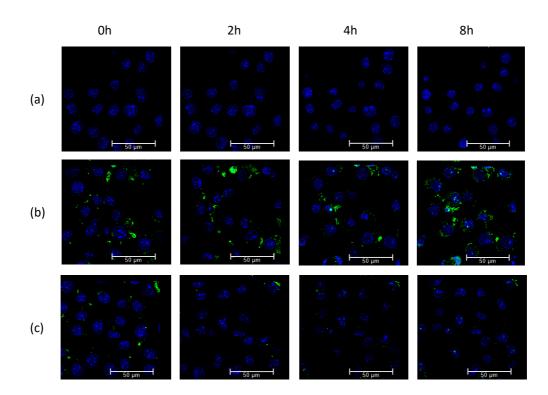


Figure S5: Nuclear co-localization of M9 and Lipofectamine nanocomplexes: Live cells confocal imaging was performed for studying nuclear localization of nanocomplexes for 8h. FITC-labeled nanocomplexes were incubated for 4h and imaging was done after media change: (a) control (b) M9 nanocomplexes (c) Lipofectamine nanocomplexes. B16-F10 Cells were imaged with same field in focus for 8h.

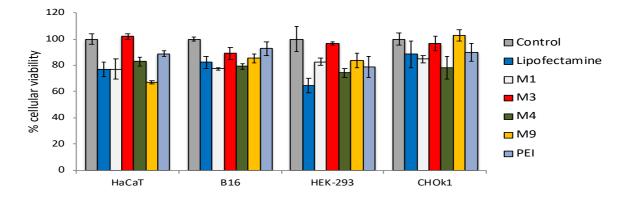


Figure S6: Cellular viability 24h after incubating the cells with the nanocomplexes. MTT assay was performed to assess the percentage viable cells with respect to control.