

# Supplementary Information

## Functional Significance of U2AF1 S34F Mutations in Lung Adenocarcinomas

Mohammad S. Esfahani<sup>1,2,\*</sup>, Luke J. Lee<sup>1,\*</sup>, Young-Jun Jeon<sup>1,\*</sup>, Ryan A. Flynn<sup>3</sup>, Henning Stehr<sup>1,4</sup>, Angela B. Hui<sup>1</sup>, Noriko Ishisoko<sup>5</sup>, Eric Kildebeck<sup>6</sup>, Aaron M. Newman<sup>7,8</sup>, Scott V. Bratman<sup>7,§</sup>, Matthew H. Porteus<sup>6</sup>, Howard Y. Chang<sup>10</sup>, Ash A. Alizadeh<sup>1,2,10,#</sup>, Maximilian Diehn<sup>1,7,11#</sup>

<sup>1</sup>Stanford Cancer Institute, Stanford University, Stanford, USA

<sup>2</sup>Division of Oncology, Department of Medicine, Stanford University, Stanford, USA

<sup>3</sup>Department of Chemistry, Stanford University, Stanford, USA

<sup>4</sup>Department of Pathology, Stanford University, Stanford, CA

<sup>5</sup>Department of Bioengineering, Stanford University, Stanford, USA

<sup>6</sup>Department of Pediatrics, Stanford University, Stanford, USA

<sup>7</sup>Institute for Stem Cell Biology and Regenerative Medicine, Stanford University, Stanford, USA

<sup>8</sup>Department of Biomedical Data Science, Stanford University, Stanford, USA

<sup>9</sup>Howard Hughes Medical Institute, Stanford University, Stanford, CA, USA

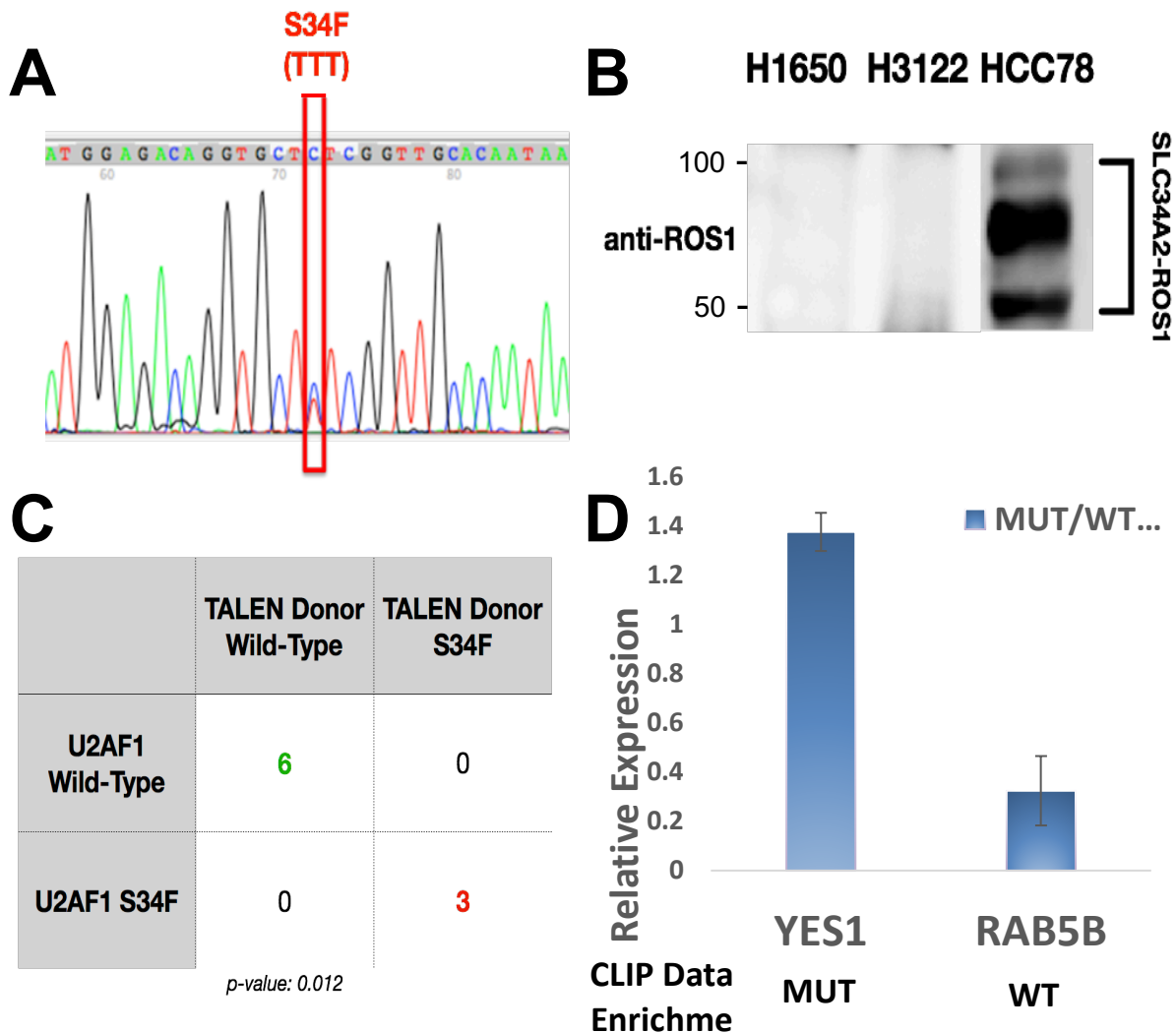
<sup>10</sup>Division of Hematology, Department of Medicine, Stanford University, Stanford, USA

<sup>11</sup>Department of Radiation Oncology, Stanford University, Stanford, USA

## SI Guide

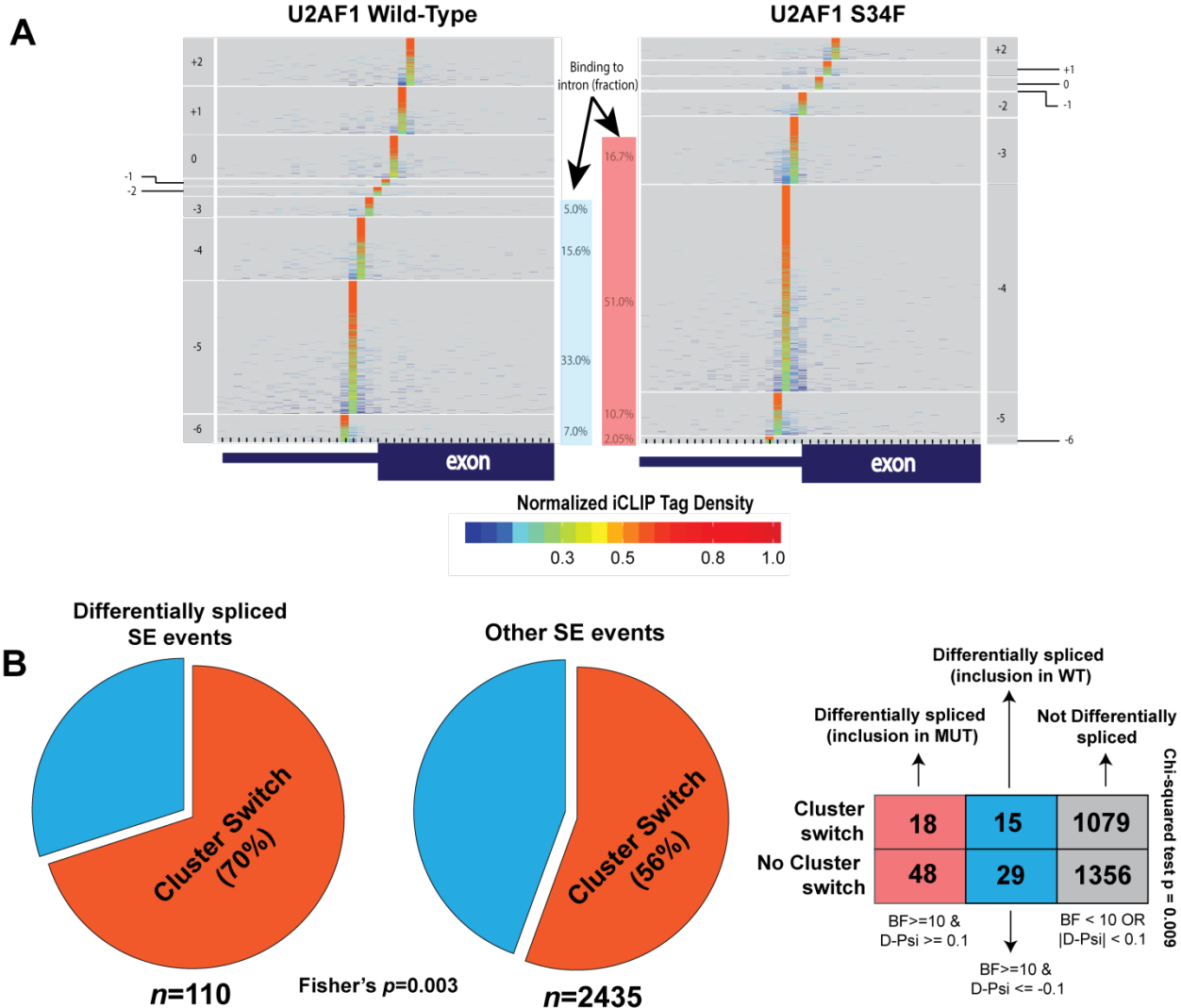
- |                        |  |
|------------------------|--|
| Supplementary Figure 1 | Confirmation of U2AF1 S34F mutation and SLC34A2-ROS1 translocation in HCC78 cells, TALEN homologous recombination results, and quality control for U2AF1 CLIP experiments. |
| Supplementary Figure 2 | Co-association of cross-linking cluster switching and significant alternative splicing.  |
| Supplementary Figure 3 | Sequence analysis of CLIP cross-linking peaks near splice junctions.   |
| Supplementary Figure 4 | Distribution of the frequency fold change of trinucleotides when measured by their cross-linking to mutant vs wild-type.   |
| Supplementary Figure 5 | Relative Expression of EMT Genes and relationship with cross-linking differences.  |
| Supplementary Figure 6 | Effect of wild-type and mutant U2AF1 overexpression on cell proliferation and effect of U2AF1 knockdown on the relative expression of SLC34A2-ROS1 isoforms.               |
| Supplementary Figure 7 | Relative expression of SLC34A2-ROS1 isoforms in HCC78 RNA-Seq data from Fei et al.   |
| Supplementary Figure 8 | Western blot of the ROS1 short and long isoforms with Histone H3 used as the loading control.  |

38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65  
66  
67  
68  
69  
70  
71  
72  
73  
74  
75  
76  
77  
78  
79  
80  
81  
82  
83  
84  
85  
86  
87  
88



**Supplementary Figure 1. Confirmation of U2AF1 S34F mutation and SLC34A2-ROS1 translocation in HCC78 cells, TALEN homologous recombination results, and quality control for U2AF1 CLIP experiments.**

- (A) Sanger sequencing of exon 2 of U2AF1 in HCC78 cells. Box indicates presence of heterozygous S34F mutation (TTT).
- (B) Western blot with anti-ROS1 antibody detecting the presence of the SLC34A2-ROS1 fusion in HCC78 cells. Negative controls cell lines H1650 and H3122 are shown.
- (C) Summary of homologous recombination events involving U2AF1 wild-type and S34F donor constructs in HCC78. TALEN, Transcription activator-like effector nuclease.
- (D) RNA immunoprecipitation quantitative PCR (RIP-qPCR) validation of CLIP binding for wild-type and mutant U2AF1.

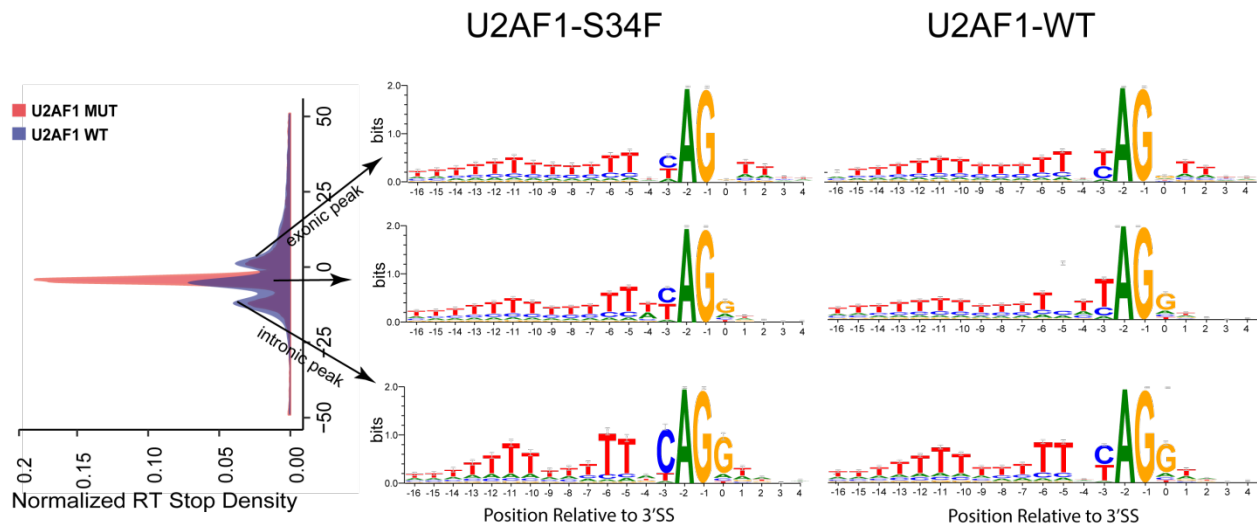


89  
90  
91  
92  
93  
94  
95  
96  
97  
98  
99  
100  
101  
102  
103  
104  
105  
106  
107  
108

**Supplementary Figure 2. Co-association of cross-linking cluster switching and significant alternative splicing.**

(A) Heatmaps corresponding to wild-type and S34F mutant, showing the extent of binding to the neighboring positions next to the 3'SSs. Major binding occurs at position -5 and -4 for wild-type and mutant, respectively. Each row represents one coding 3' splice site.

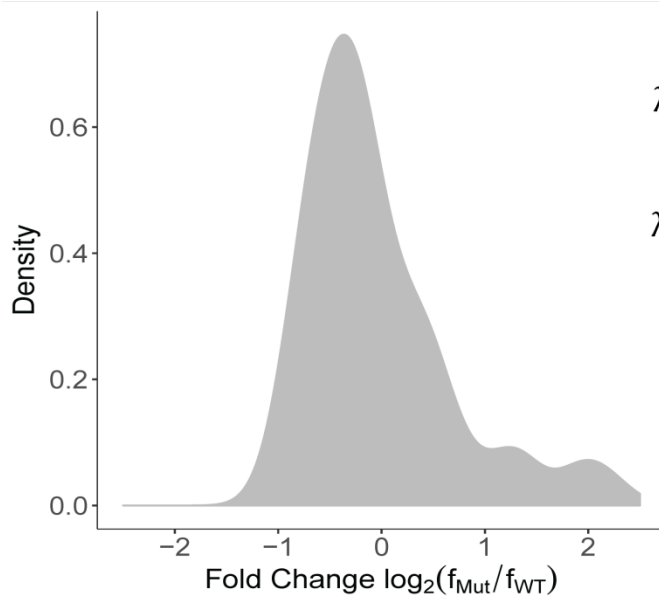
(B) Differentially spliced events are defined as having  $\Delta\Psi \geq 10\%$  and Bayes factor of at least 10. Cluster switch is defined as splice sites which appear in different clusters in wild-type versus mutant (excluding cluster 10). Differentially spliced skipped exon (SE) events between wild-type and mutant conditions were more likely to display cluster switching ( $P=0.003$  by Fisher's exact test).



110  
 111  
 112  
 113  
 114  
 115  
 116  
 117  
 118

**Supplementary Figure 3. Sequence analysis of CLIP cross-linking peaks near splice junctions.**

Weblogo plots of the three cross-linking peaks near splice junctions observed in Fig. 3B. The three rows correspond to intronic (left-side peak), “main” (closer to 3’ splice site from the intronic side) and exonic (right-side peak) peaks, respectively. The left and right columns correspond to mutant and wild-type, respectively. In all peaks, CAG is enriched in mutant compared to wild-type.



$$\lambda_{AAG} = \text{avg} \left[ \log_2 \frac{f_{Mut,i}^{AAG}}{f_{WT,i}^{AAG}} \right]_{i \in \{-1,0,+1\}} = 0.3216$$

$$\lambda_{TAG} = \text{avg} \left[ \log_2 \frac{f_{Mut,i}^{TAG}}{f_{WT,i}^{TAG}} \right]_{i \in \{-1,0,+1\}} = -0.833$$

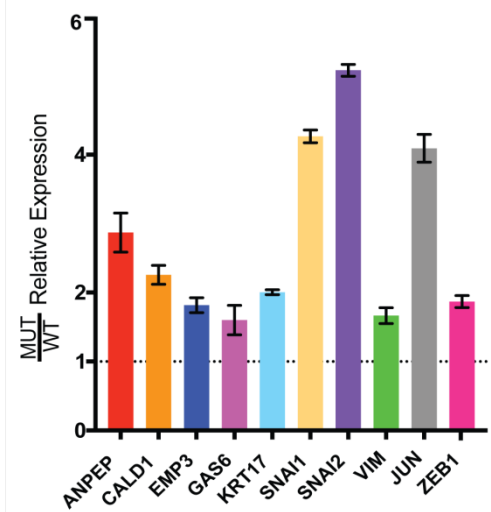
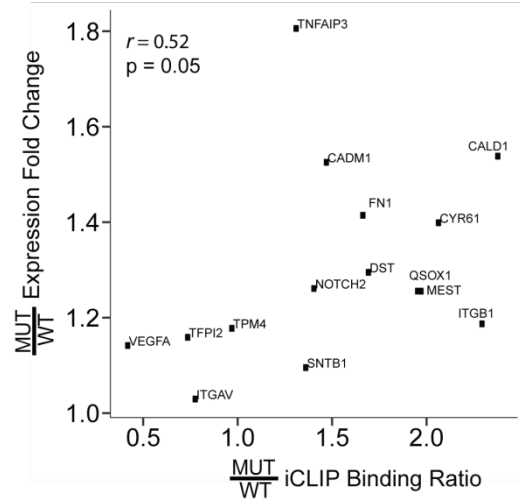
$$\lambda_{CAG} = \text{avg} \left[ \log_2 \frac{f_{Mut,i}^{CAG}}{f_{WT,i}^{CAG}} \right]_{i \in \{-1,0,+1\}} = 0.418$$

$$\kappa_{AAG:TAG} := \lambda_{AAG} - \lambda_{TAG} = 1.15$$

119  
120  
121  
122  
123  
124  
125  
126  
127  
128  
129  
130  
131  
132  
133  
134  
135  
136  
137

**Supplementary Figure 4. Distribution of the frequency fold change of trinucleotides when measured by their cross-linking to mutant vs wild-type.**

The distribution of the scores, denoted by  $\lambda$  and defined as the average of mutant to wild-type fold change over a window of  $-/+1$ bp around the TSS ( $i$  denotes the relative distance from the RT site), corresponding to 64 possible trinucleotides. The three trinucleotides highlighted are: CAG (enriched in mutant), TAG (enriched in wild-type) and AAG (slightly in favor of mutant). Relative preference of AAG over TAG in mutant is 2.2-fold larger.

**A****B**138  
139

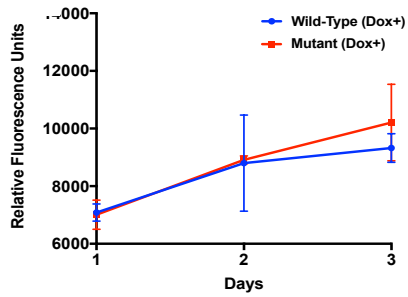
140 **Supplementary Figure 5. Relative Expression of EMT Genes and relationship with cross-**  
 141 **linking differences.**

142 (A) Relative expression of EMT genes taking ratios of mutant to wild-type U2AF1  
 143 overexpression in HCC78. Experiments were performed in triplicate.

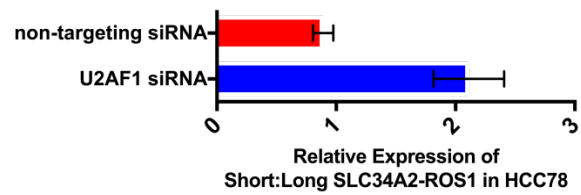
144 (B) Scatterplot of the EMT genes from the GSEA analysis that were present in both RNA-seq  
 145 and iCLIP data sets. The Y-axis depicts the magnitude of differential expression between  
 146 mutant and wild-type U2AF1 overexpression (in fold change); the X-axis shows the mutant to  
 147 wild-type crosslinking ratio (in fold change; Spearman correlation of 0.52, P=0.05).

148  
149

**A**



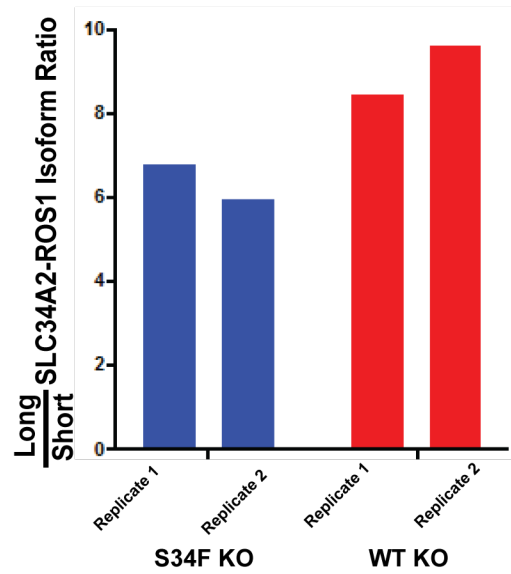
**B**



150  
151  
152  
153  
154  
155  
156  
157  
158  
159  
160

**Supplementary Figure 6. Effect of wild-type and mutant U2AF1 overexpression on cell proliferation and effect of U2AF1 knockdown on the relative expression of SLC34A2-ROS1 isoforms.**

- (A) Cell growth assay after doxycycline-induced overexpression of wild-type and mutant U2AF1 measured by relative fluorescence units over 72 hours.
- (B) siRNA knockdown of total U2AF1 using siRNA targeting the 3'UTR. Relative expression of short and long SLC34A2-ROS1 fusions were quantified by qPCR at exons 34 and 32, respectively. The error bars in both panels are used to show mean and standard deviations.



161

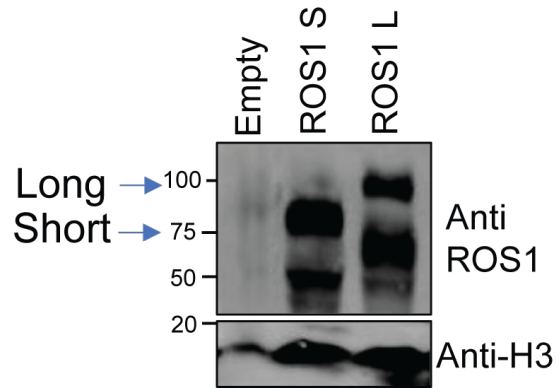
162

163 **Supplementary Figure 7. Relative expression of SLC34A2-ROS1 isoforms in HCC78 RNA-**  
164 **Seq data from Fei et al.**

165 Summary of long and short ROS1 fusion expression using data from Fei et al. “S34F KO”  
166 indicates cells with CRISPR-Cas9 knock-out using mutation-specific guide RNA; “WT KO”  
167 indicates use of control guide RNA against GFP.

168





169

170 **Supplementary Figure 8. Western blot of the ROS1 short and long isoforms with Histone**  
171 **H3 used as the loading control.** The lowest molecular weight species seen below the long /  
172 short bands is a presumed degradation product of both ROS1 isoforms.