Supplementary Information for

Photoperiodic and clock regulation of the vitamin A pathway in the brain mediates seasonal responsiveness in the monarch butterfly

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Datasets S1 to S2

Supplementary Information

Material and Methods

Animal husbandry

For photoperiodic experiments, wild-type and mutant monarchs maintained in 15 hours light: 9-hours dark at 25^oC were hand-paired. Fertilized females were placed in a cage to lay eggs on potted tropical milkweed plants (*Asclepias curassavica*). Eggs were transferred to petri dishes onto milkweed leaves and divided in two groups placed in Percival incubators respectively under 15-hours light: 9-hours dark (long photoperiod; LP) and under 10-hours light: 14-hours dark (short photoperiod; SP), both at constant temperature of 21°C and 70% humidity. Starting at the second instar, larvae were raised individually on semi-artificial diet until pupation. Post-eclosion, adult monarchs were housed in glassine envelopes and were manually fed a 25% honey solution daily until dissection.

For RNA-seq experiments, fall migratory monarch butterflies were captured on October 15, 2014 in College Station, Texas (latitude 30°37'N, longitude 96°20'W). After capture, migrants were housed indoors in glassine envelopes in a Percival incubator under fall-like conditions with light:dark (LD) cycle set to prevailing light conditions (11 h:13 h LD, 0730–1830 Central Standard Time), at a constant temperature of 21 $^{\circ}$ C with 70% humidity. They were manually fed a 25% honey solution every other day and dissected 8 days after capture. Monarchs used for the LP and SP data sets were raised in the same conditions as for the photoperiodic experiments, and were dissected 7 to 11 days posteclosion.

Methoprene/vehicle treatment

Monarch wild-type and *Cry2-/-* females were treated topically on their abdomens with 200µg of methoprene in 5µl of acetone or with 5µl of acetone alone on day 1 and day 3 after adult emergence, as previously described (1).

Compound eye painting and antennae removal

Painting of the compound eyes and removal of the antennae were performed on the

day of eclosion. For eye painting, the labial palps were first cut at the base to expose the entire compound eyes including the retina and the dorsal rim, which were then covered under a dissecting microscope with enamel based clear paint (Model master clear top coat; Testors no. 2736) or black paint (Glossy black; Testors no. 1147) using a fine paintbrush. Monarchs were harnessed to allow the paint to air-dry. The completeness of painting was verified the next day under a dissecting microscope, and touch ups were performed if needed. The light-permissive and light-blocking properties of the clear and black paints, respectively, had previously been characterized (2). Unpainted control monarchs were subjected to labial palps removal similar to painted monarchs. Antennae were removed by clipping them with scissors at the base of the flagellum just above the pedicel as described in (2).

RNA-sequencing experiments

Brains of monarchs raised indoors in LP and SP and of wild-caught migrants were dissected in 0.5X RNA later (Invitrogen) to prevent RNA degradation, the retinal pigmented photoreceptor layer was removed, and the brains were stored at -80° C until use. For each seasonal phenotype/photoperiodic condition, three pooled brains were collected in two replicates at Zeitgeber time (ZT)1, ZT4, ZT7, ZT10, ZT13, ZT16, ZT19, and ZT22. For each sample, total RNA was extracted using an RNeasy Mini kit (Qiagen). For samples from wild-caught migrants, polyA+ RNA was isolated from 2 µg of total RNA with NEBNext Poly(A) mRNA Magnetic Isolation Module (New England Biolabs), and multiplexed libraries were prepared using the NEBNext® Ultra™ II Directional RNA Library Prep Kit for Illumina and NEBNext Multiplex Oligos (New England Biolabs) and amplified with 12 PCR cycles, following the manufacturer's recommendations. RNA-seq datasets from brains of summer-like monarchs and clockdeficient mutants used in this study were already available (3). Butterflies were raised in 15:9 LD at 25°C, brains were collected at 3-hours intervals for summer-like monarchs starting at ZT1 and at 6-hours intervals for clock-deficient mutants starting at ZT4, and libraries were prepared as for wild-caught migrants. For samples from monarchs raised in LP and SP, multiplexed libraries were prepared by the Texas A&M AgriLife Genomics and Bioinformatics Facility using polyA+ RNA isolated from 1 µg of total RNA, and

multiplexed libraries were prepared using the TruSeq Stranded mRNA Library Prep Kit for Illumina, following the manufacturer's recommendations. Libraries quality and size distribution was verified on a Bioanalyzer, libraries were quantified by real-time quantitative PCR, and 16 multiplexed libraries were mixed in equimolar ratios and sequenced on a Hi-seq 2500 (Illumina) using 50bp single end reads.

RNA-seq data processing, mapping and identification of cycling transcripts

The resulting sequencing files were checked for quality control and demultiplexed by the Texas A&M AgriLife Genomics and Bioinformatics Facility. Reads were mapped to the monarch genome (assembly v3; (4)) using TopHat2 (5) with parameters "--readrealign-edit-dist 2 -g 1 --b2-sensitive". On average across LP, SP, summer-like and migrant monarchs, 91.5% of the reads mapped to the monarch genome (Table S4). After mapping, expression levels of transcripts were quantified in each sample using Cufflinks (6, 7). To identify cycling transcripts, RAIN (8) and MetaCycle (9) were used with parameters "period=24, deltat=3, period.delta=3, nr.series=2, method='independent', peak.border=c(0.2, 0.8)" and "timepoints=seq(1, 46, by=3), adjustPhase="predictedPer", combinePvalue="fisher")", respectively. Only genes with three or more reads per kilo base per million mapped reads (RPKM) in at least one time point were considered as expressed and used for subsequent analysis. Rhythmically expressed genes were determined based on fold-change and *p*-values adjusted for multiple testing using Benjamini-Hochberg procedure to control for false discovery rate. Transcripts were considered rhythmically expressed when meeting the following criteria: (1) adjusted *p*values ≤ 0.05 for both RAIN and MetaCycle; and (2) fold-change (maximal/minimal experimental values within a time series) ≥ 1.3 . Of the 15,130 genes in the monarch genome, 68.3%, 66.8%, 66.6%, and 68.4% were respectively expressed in the brains of LP, SP, summer-like, and migrant monarchs, of which 2.70%, 2.66%, 2.80% and 3.04% were respectively determined as rhythmically expressed (Table S5).

Comparisons of rhythmic expression were performed between LP and SP monarchs, and between summer-like monarchs and wild-caught migrants. Genes were sorted based on their rhythmic expression in a photoperiod condition or a seasonal form and classified in three groups: (1) rhythmic in both LP and SP or summer-like and migrant monarchs,

(2) rhythmic only in LP or summer-like monarchs, and (3) rhythmic only in SP or migrant monarchs. Heatmaps depicting all categories were produced using heatmap.2 in gplots package for R. To identify the genes that may be responsible for photoperiodic responses, genes present in both comparisons were identified. Results of this overlap analysis are summarized in Fig. 2. The complete list of genes is provided in Table S1. Using BLAST (10), homologous proteins from *Drosophila* and mouse that best matched the protein sequences of the overlapped genes were used to annotate them. Enriched gene ontology of biological processes and KEGG pathway were determined using Metascape (metascape.org).

gRNA design and construction

The gRNA site for CRISPR/Cas9-mediated targeted mutagenesis of *ninaB1* was selected within exon 3 of the 14 exons-containing *ninaB1* using CHOPCHOP target site finder (http://chopchop.cbu.uib.no/index.php target site finder; (11, 12)). The gRNA expression vector was constructed by inserting annealed synthetic oligomers into the DR274 plasmid from Addgene (13) at the *BsaI* cleavage site. Oligomer sequences were as follow (F, forward primer; R, reverse primer): *F*, 5'- TAGGAGTGACAACTATACGACCCG-3' and *R*, 5'- AAACCGGGTCGTATAGTTGTCACT-3'.

Synthesis of Cas9 mRNA and sgRNA

In vitro transcription of *Streptococcus pyogenes* Cas9 mRNA was performed using the mMessage mMachine T3 transcription kit (Invitrogen) and pCS2-nCas9n expression plasmid from Addgene (14), as previously described (15). The resulting capped PolyA mRNAs were purified by acid-phenol-chloroform extraction and resuspended in RNasefree water following isopropanol precipitation. The sgRNA was *in vitro* transcribed using T7 RNA polymerase (Promega) from purified PCR products containing the T7 promoter, gRNA and gRNA scaffold amplified from the DR274 vectors using the following primers; F: 5'-ATTGAGCCTCAGGAAACAGC-3' and R: 5'-

AAAAGCACCGACTCGGTGCC-3'. The sgRNA was then treated with RQ1-DNase and purified by acid phenol-chloroform extraction and resuspended in RNase-free water after

isopropanol precipitation. Cas9 mRNAs and the sgRNA were quantitated by spectrophotometry (NanoDrop 1000) and diluted in RNase-free water to a final concentration of 0.25 μ g/ μ l for the sgRNA and 0.5 μ g/ μ l for Cas9 mRNAs.

Egg microinjections

Eggs were collected and microinjected as previously described (16) with a mix of Cas9 mRNA at $0.5 \mu g/\mu l$, sgRNA at $0.25 \mu g/\mu l$, and blue food coloring for visual tracking of the injection. After injection, embryos were placed in an incubator at 25°C and 70% humidity. Developing embryos were transferred into individual small petri dishes containing milkweed leaves until larvae hatched. Larvae were fed milkweed leaves until the second larval instar before being transferred onto semi-artificial diet.

Analysis of CRISPR/Cas9-induced mutations and generation of a *ninaB1* **monarch loss-of-function line**

PCR fragments flanking the targeted region were amplified from genomic DNA extracted from larval sensors of potential founders at the fifth instar with the following primers: *ninaB1F*, 5'-GTTTCACTTGTACCGTGACTTC-3' and *ninaB1R,* 5'- GGATACTGTTTAGCCAGGTACC-3. PCR products were purified using 2 X modified Sera-Mag Magnetic Speed-beads (GE Healthcare) as previously described (16, 17), and resuspended in 10 µl of RNase-free water. Cas9-based cleavage assays of PCR products (250-350 ng) were performed using a recombinant Cas9 protein (750 ng) and the sgRNA (400-600 ng), as previously described (16). Digested products were purified using 2 X modified Sera-Mag Magnetic Speed-beads before being resolved with agarose gel electrophoresis and EtBr staining. Larvae presenting a high degree of targeting in somatic cells, estimated based on the relative abundance of uncleaved fragments, were selected and reared to adulthood. Surviving adults of opposite sexes presenting the highest level of somaticism were hand-paired in individual cages to establish a mutant line. Eggs were collected and the hatched larvae were screened for the presence of mutated alleles as described above. Uncleaved fragments corresponding to mutated alleles were gel purified and sequenced using one of the primers used for PCR amplification. A 7-base pair deletion causing a frameshift and the introduction of a premature stop codon was selected

to establish a mutant line.

Proboscis extension reflex assay

The proboscis extension reflex (PER) is a response in which stimulation of the chemoreceptors on the middle legs with a sugar solution (unconditioned stimulus, US) results in full extension of the proboscis (unconditioned response). Proboscis extension tests were conducted as previously described (2) by touching the middle leg with a cotton-tipped applicator soaked in 50% sucrose solution (wt/wt), with slight modifications. Individual butterflies were fed daily with 150μ of a 25% honey solution prior to the experiment. The day prior to stimulus conditioning, individuals were harnessed in 15ml polypropylene conical tubes as previously described (2) and starved for 24 hours. On the day of testing, individuals were checked for a positive proboscis extension reflex. Wild-type monarchs with eyes covered with either clear or black paint, and *ninaB1* -/- monarchs were conditioned to a colored stimulus (red flag; conditioned stimulus, CS) by presenting the stimulus for 5 sec (CS only), contacting the middle legs with sucrose solution for 15 sec $(CS + US)$, and removing the stimulus after 5 sec (US only). Individuals were then held for 5 min and this pairing procedure was performed again. US-CS pairing continued in this way for 13 to 14 trials a day for three consecutive days until the individual extended its proboscis upon the initial presentation of the CS, in which case a sucrose reward was given. Monarchs were considered to have a positive PER (conditioned response) if they fully extended their proboscis in response to the CS. On the second and third day of testing, butterflies were fed 50 µl of a 25% honey solution to prevent extreme starvation.

Real-time qPCR

To test for the presence of a functional circadian clock in *ninaB1* loss-of-function monarchs, brains of adult wild-type and *ninaB1* homozygous mutant monarchs entrained to seven 15:9 LD cycles were dissected under red light on the first day of transfer into DD at circadian time (CT) 0, CT4, CT8, CT12, CT16, and CT20. Dissections were performed in 0.5X RNA later (Invitrogen) to avoid RNA degradation, and brains free of eye photoreceptors were stored at -80°C until use. Total RNA was extracted using

RNeasy Mini kit (Qiagen), treated with RQ1 Dnase (Promega), and random hexamers (Promega) were used to prime reverse transcription with Superscript II Reverse Transcriptase (Thermo Scientific), all according to the manufacturers' instructions. Quantifications of gene expression were performed on a QuantStudio™ 6 Flex Real-Time PCR System (Thermo Scientific) using iTaq Universal SYBR Green Supermix (Bio-Rad). PCR reactions were assembled by combining two master mixes: the first mix contained 5 µl of iTaq Universal SYBR® Green Supermix (Bio-Rad) and forward and reverse primers (5 µmol each) per reaction and the second mix contained approximately 7 ng of cDNA template and the water needed to bring each reaction to a final volume of 10 µl. The monarch *per* and control *rp49* primers were as follows (F, forward primer; R, reverse primer): *perF*, 5'-AGTGAAGCGTCCCTCAAAACA-3'; *perR*, 5'- TGGCGACGAGCATCTGTGT-3'; *rp49F*, 5'-TGCGCAGGCGTTTTAAGG-3'; *rp49R*, 5'-TTGTTTGATCCGTAACCAATGC-3'. The near 100% efficiency of each primer set was validated by determining the slope of Ct versus dilution plot on a dilution series. Individual reactions were used to quantify each RNA level in a given cDNA sample, and the average Ct from duplicated reactions within the same run was used for quantification. The data were normalized to *rp49* as an internal control, and normalized to the mean of one sample within a set for statistics.

Supplementary Figures

Fig. S1

Fig. S1. Application of a juvenile hormone analogue restores high levels of oocyte production in monarch *Cry2 -/-***.** Number of mature oocytes produced 10 days post adult emergence in wild-type or *Cry2 -/-* female monarchs treated with either an acetone vehicle or with methoprene, a juvenile hormone analogue (JHA). Monarch were raised in LP at 25°C. Legends as in Figure 1. Interaction genotype x treatment, Two-way ANOVA, Tukey's pairwise comparisons, $p < 0.05$.

Fig. S2. Temporal mRNA expression profiles of *period***,** *timeless***,** *vrille***, and** *clockwork orange* **in brains of monarchs in different photoperiods.** A) mRNA expression levels in brains of monarchs raised in LP and SP (*top*) and of summer-like and migrant monarchs (*bottom*). For each condition, two biological replicates are plotted. White bars: light; black bars: dark. B) Relative mRNA expression levels from pooled biological replicates of LP and summer-like monarchs (solid line), and SP and migrants (dashed line). Data represent the mean \pm s.e.m. Yellow bars: long day; blue bars: short day; black bars: dark. Interactions seasonal morph x photoperiod, Two-way ANOVAs: *period*, *p* < 0.0001; *timeless*, non significant; *vrille*, *p* < 0.005; *clockwork orange*, *p* < 0.0001. Tukey's pairwise comparisons, $*, p < 0.05;$ ****, $p < 0.0001$.

Fig. S3

Fig. S3. The reproductive state exhibited by photoperiod-impaired *ninaB1* **loss-offunction could be caused by redundant function of ninaB1 and ninaB2.** (A, B)

Number of mature oocytes produced 10 days post adult emergence in *ninaB1* homozygous loss-of-function and wild-type sibling females, all raised in LP and SP at 21° C after either one backcross (A) or three backcrosses (B). Legends as in Figure 4B. Interaction genotype x photoperiod, Two-way ANOVA, Tukey's pairwise comparisons, *p* < 0.05. (C) Comparisons of *Drosophila* ninaB (Dm; AAF54978) and monarch (Dp) ninaB1 (DPOGS212590), ninaB2 (DPOGS212591), and ninaB3 (DPOGS212592) sequences. *Top*, phylogenetic relationship. *Bottom*, sequences alignment. Identical and similar residues found in at least three out of four sequences are shaded in black and gray,

respectively. The glutamic acid residue highlighted in red, which is conserved between Dm-ninaB, Dp-ninaB1 and Dp-ninaB2 but absent in Dp-ninaB3, corresponds to the residue providing the enzymatic activity of Dm-ninaB (18). (D) Visualization of RNAseq signal in brains of wild-type monarchs over 24 hours in LP on scaffold DPSCF300245 that contains the three copies of *ninaB* present in the monarch genome. Of the two detectable expressed copies, only *ninaB1* displays rhythmic expression. Two biological replicates are plotted consecutively from top to bottom. White bar: light; dark bar: night. (E) mRNA expression levels of *ninaB1* and *ninaB2* in the brain of wild-type monarchs over the course of the day in LP. For each gene, two biological replicates are plotted. White bar: light; black bar.

 $a₂₄$ a 26 \bullet $\ddot{\cdot}$ \cdot Number of mature oocytes **b** 29 100 ೲ $\ddot{\cdot}$ °。 ÷. \bullet 25 b $\ddot{}$ $\ddot{\bullet}$ $\circ \circ \circ$ 50 \circ° \bullet k $-2.2 - 2.22$ \circ $\begin{bmatrix} 0 \\ 0 \\ 0 \\ 0 \end{bmatrix}$ $\mathbf 0$ LP SP LP SP Black painted eyes
Antenna-less **Black painted eyes**

Fig. S4. The antennae are not necessary for photoperiodic responses. Number of mature oocytes produced 10 days post adult emergence in wild-type female monarchs with eyes painted black and with or without antennae, all raised in LP and SP at 21°C. Legends as in Figure 4C. Interaction genotype x photoperiod, Two-way ANOVA, Tukey's pairwise comparisons, $p < 0.05$.

Fig. S4

Table S1. List of genes with similar or differential temporal expression patterns in the brain of monarchs raised in LP and SP. R-R: Genes rhythmic in both conditions; R-AR: Genes rhythmic in LP monarchs and arrhythmic in SP monarchs; AR-R: Genes arrhythmic in LP monarchs and rhythmic in SP monarchs. The annotation is based on *Drosophila* (no shading) or mouse (gray shading) orthologues. NA, monarch genes without orthologues in *Drosophila* or the mouse.

Table S2. List of genes with similar or differential temporal expression patterns in the brain of summer-like monarchs and wild-caught migrants. R-R: Genes rhythmic in both conditions; R-AR: Genes rhythmic in summer-like monarchs and arrhythmic in migrants; AR-R: Genes arrhythmic in summer-like monarchs and rhythmic in migrants. The annotation is based on *Drosophila* (no shading) or mouse (gray shading) orthologues. NA, monarch genes without orthologues in *Drosophila* or the mouse.

Table S3. List of genes with similar temporal expression patterns in the brains of monarchs raised in LP and in summer-like monarchs and/or in the brains of monarchs raised in SP and in wild-caught migrants. R-R: Genes rhythmic in all four conditions; R-AR: Genes rhythmic in LP and summer-like monarchs, and arrhythmic in SP monarchs and migrants ; AR-R: Genes arrhythmic in LP and summer-like monarchs, and rhythmic in SP monarchs and migrants. The annotation is based on *Drosophila* (no shading) or mouse (colored shading) orthologues. NA, monarch genes without orthologues in *Drosophila* or the mouse.

Samples	Number of reads	Mapped reads	Mapping percentage
LP ZT01 R1	13,522,929	12,569,145	92.9
LP ZT01 R2	13,678,471	12,641,190	92.4
LP ZT04 R1	15,821,730	14,679,157	92.8
LP ZT04 R2	16,370,285	15,082,230	92.1
LP ZT07 R1	18,287,614	16,960,023	92.7
LP ZT07 R2	18,547,268	17,043,095	91.9
LP ZT10 R1	13,373,316	12,350,949	92.4
LP ZT10 R2	16,462,573	15,232,242	92.5
LP ZT13 R1	15,378,742	14,253,836	92.7
LP ZT13 R2	18,124,188	16,640,088	91.8
LP ZT16 R1	16, 141, 542	14,943,063	92.6
LP ZT16 R2	17,448,980	15,981,062	91.6
LP ZT19 R1	14,581,672	13,501,921	92.6
LP ZT19 R2	15,306,522	14,013,930	91.6
LP ZT22 R1	15,207,486	14,104,449	92.7
LP ZT22 R2	16,168,931	14,936,831	92.4
SP ZT01 R1	10,939,912	10,068,478	92
SP ZT01 R2	11,026,339	10,288,670	93.3
SP ZT04 R1	11,957,374	11,060,333	92.5
SP ZT04 R2	11,984,004	11,214,567	93.6
SP ZT07 R1	14,661,160	13,598,461	92.8
SP ZT07 R2	11,503,579	10,754,899	93.5
SP ZT10 R1	11,591,287	10,850,911	93.6
SP ZT10 R2	11,747,485	10,928,317	93
SP ZT13 R1	13,794,223	12,895,039	93.5
SP ZT13 R2	11,163,015	10,402,241	93.2
SP ZT16 R1	12,703,894	11,832,060	93.1
SP ZT16 R2	12,174,241	11,372,549	93.4
SP ZT19 R1	11,931,961	11,165,608	93.6
SP ZT19 R2	11,276,364	10,498,869	93.1
SP ZT22 R1	9,356,277	8,741,841	93.4
SP ZT22 R2	9,315,476	8,698,696	93.4
Summer ZT01 R1	8,329,736	7,474,898	89.7
Summer ZT01 R2	11,307,582	9,768,700	86.4

Table S4. RNA-seq mapping summary for LP, SP, summer-like (summer), and migrant monarchs. R: replicate.

Dataset S1. RPKM values of genes with similar or differential temporal expression patterns in the brain of monarchs raised in LP and SP. Legends as in Table S1.

Dataset S2. RPKM values of genes with similar or differential tenmporal expression patterns in the brain of summer-like monarchs and wild-caught migrants. Legends as in Table S2.

References

- 1. Zhu H, Gegear RJ, Casselman A, Kanginakudru S, & Reppert SM (2009) Defining behavioral and molecular differences between summer and migratory monarch butterflies. *BMC Biol* 7:14.
- 2. Merlin C, Gegear RJ, & Reppert SM (2009) Antennal circadian clocks coordinate sun compass orientation in migratory monarch butterflies. *Science* 325(5948):1700-1704.
- 3. Lugena AB, Zhang Y, Menet JS, & Merlin C (2019) Genome-wide discovery of the daily transcriptome, DNA regulatory elements and transcription factor occupancy in the monarch butterfly brain. *PLoS Genetics* 15(7):e1008265.
- 4. Zhan S & Reppert SM (2013) MonarchBase: the monarch butterfly genome database. *Nucleic Acids Res* 41(Database issue):D758-763.
- 5. Kim D*, et al.* (2013) TopHat2: accurate alignment of transcriptomes in the presence of insertions, deletions and gene fusions. *Genome Biol* 14(4).
- 6. Trapnell C*, et al.* (2012) Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. *Nat Protoc* 7(3):562-578.
- 7. Trapnell C*, et al.* (2010) Transcript assembly and quantification by RNA-Seq reveals unannotated transcripts and isoform switching during cell differentiation. *Nat Biotechnol* 28(5):511-U174.
- 8. Thaben PF & Westermark PO (2014) Detecting Rhythms in Time Series with RAIN. *Journal of Biological Rhythms* 29(6):391-400.
- 9. Wu G, Anafi RC, Hughes ME, Kornacker K, & Hogenesch JB (2016) MetaCycle: an integrated R package to evaluate periodicity in large scale data. *Bioinformatics* 32(21):3351-3353.
- 10. Altschul SF, Gish W, Miller W, Myers EW, & Lipman DJ (1990) Basic Local Alignment Search Tool. *Journal of Molecular Biology* 215(3):403-410.
- 11. Labun K, Montague TG, Gagnon JA, Thyme SB, & Valen E (2016) CHOPCHOP v2: a web tool for the next generation of CRISPR genome engineering. *Nucleic Acids Research* 44(W1):W272-W276.
- 12. Montague TG, Cruz JM, Gagnon JA, Church GM, & Valen E (2014) CHOPCHOP: a CRISPR/Cas9 and TALEN web tool for genome editing. *Nucleic Acids Research* 42(W1):W401-W407.
- 13. Hwang WY*, et al.* (2013) Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nat Biotechnol* 31(3):227-229.
- 14. Jao LE, Wente SR, & Chen WB (2013) Efficient multiplex biallelic zebrafish genome editing using a CRISPR nuclease system. *P Natl Acad Sci USA* 110(34):13904-13909.
- 15. Zhang Y, Markert MJ, Groves SC, Hardin PE, & Merlin C (2017) Vertebrate-like CRYPTOCHROME 2 from monarch regulates circadian transcription via independent repression of CLOCK and BMAL1 activity. *Proc Natl Acad Sci U S A* 114(36):E7516-E7525.
- 16. Markert MJ*, et al.* (2016) Genomic Access to Monarch Migration Using TALEN and CRISPR/Cas9-Mediated Targeted Mutagenesis. *G3-Genes Genom Genet* 6(4):905-915.
- 17. Rohland N & Reich D (2012) Cost-effective, high-throughput DNA sequencing libraries for multiplexed target capture. *Genome Res* 22(5):939-946.
- 18. von Lintig J, Dreher A, Kiefer C, Wernet MF, & Vogt K (2001) Analysis of the blind Drosophila mutant ninaB identifies the gene encoding the key enzyme for vitamin A formation invivo. *Proc Natl Acad Sci U S A* 98(3):1130-1135.