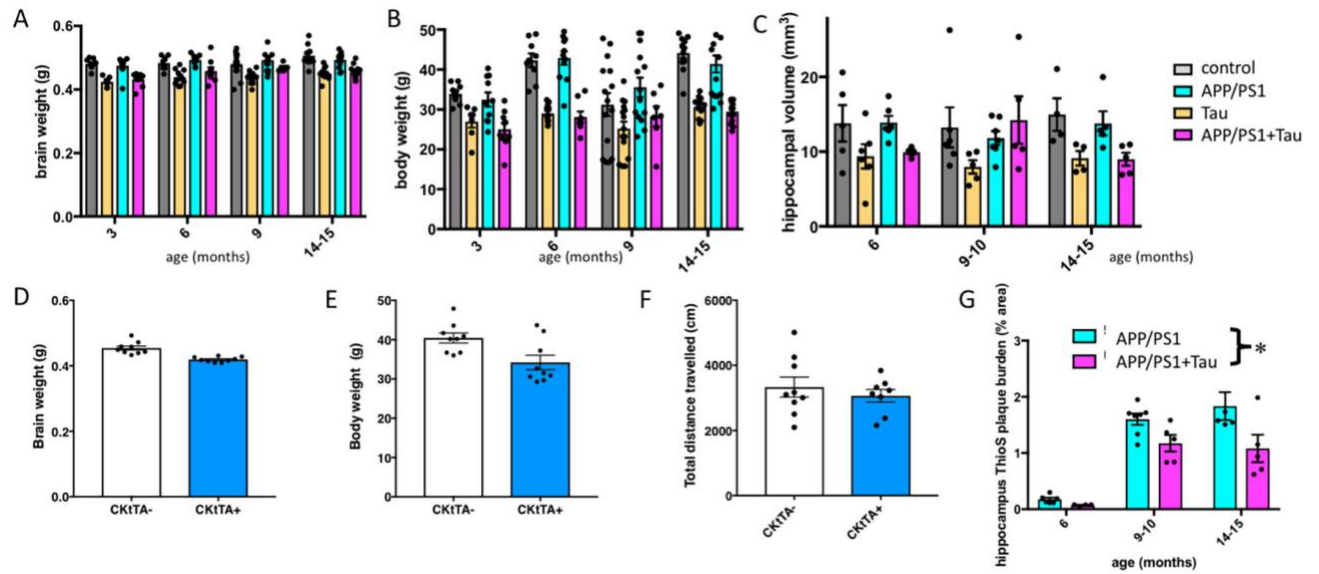


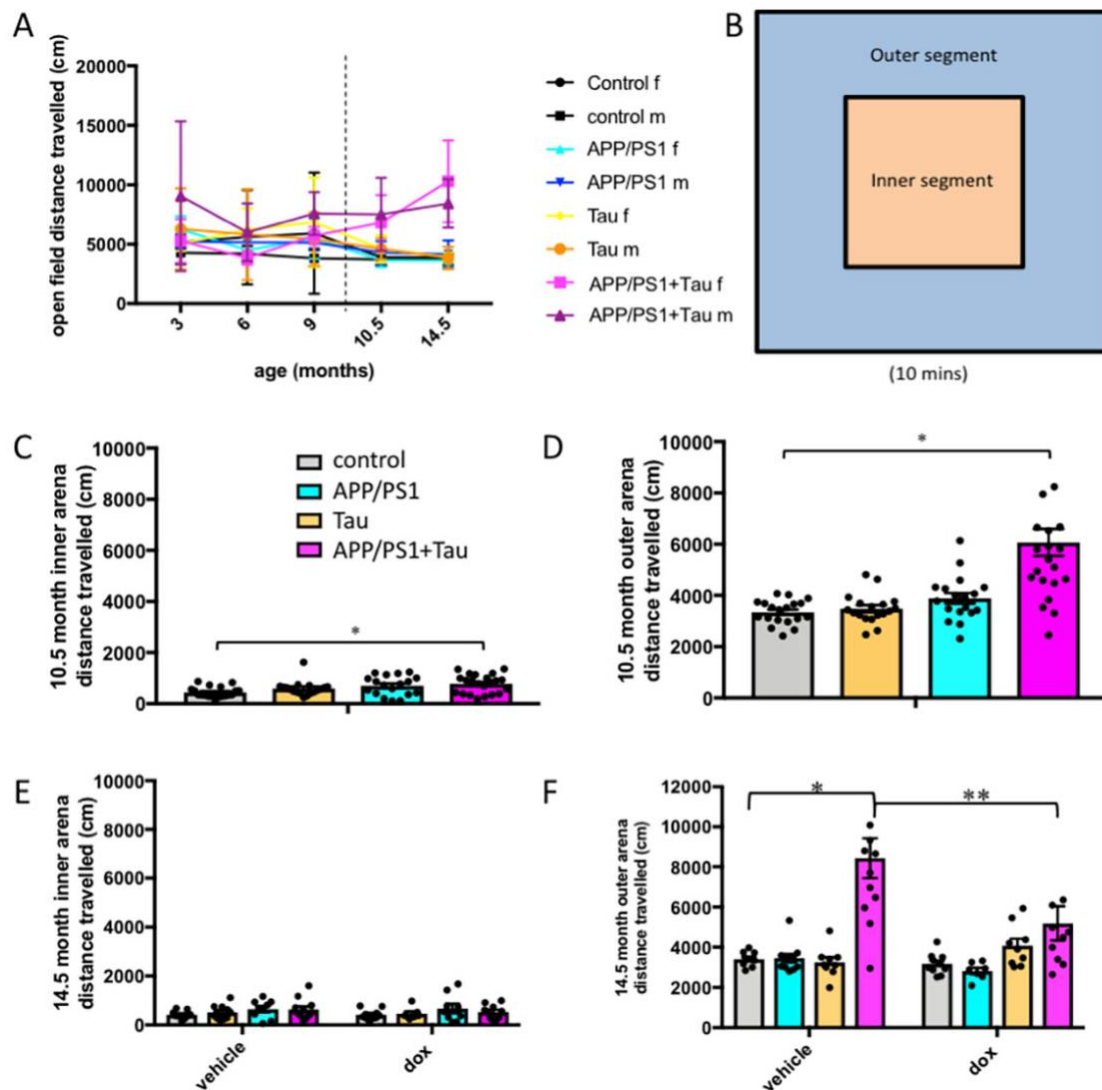
**Supplemental Information**

**Amyloid Beta and Tau Cooperate to Cause  
Reversible Behavioral and Transcriptional  
Deficits in a Model of Alzheimer's Disease**

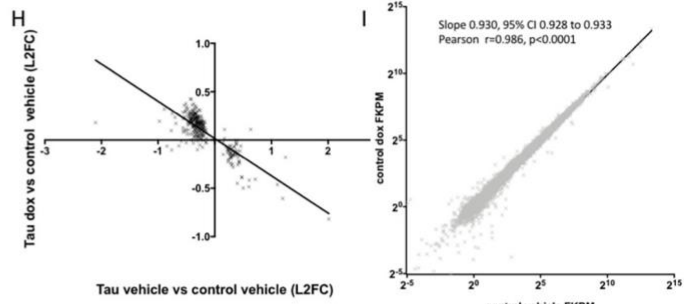
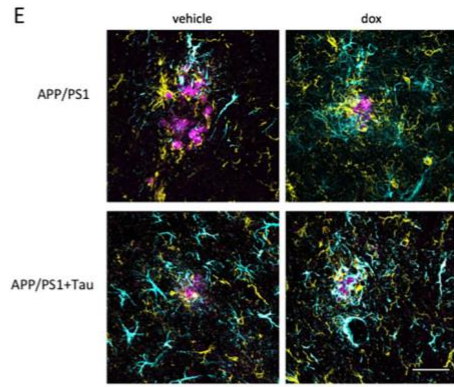
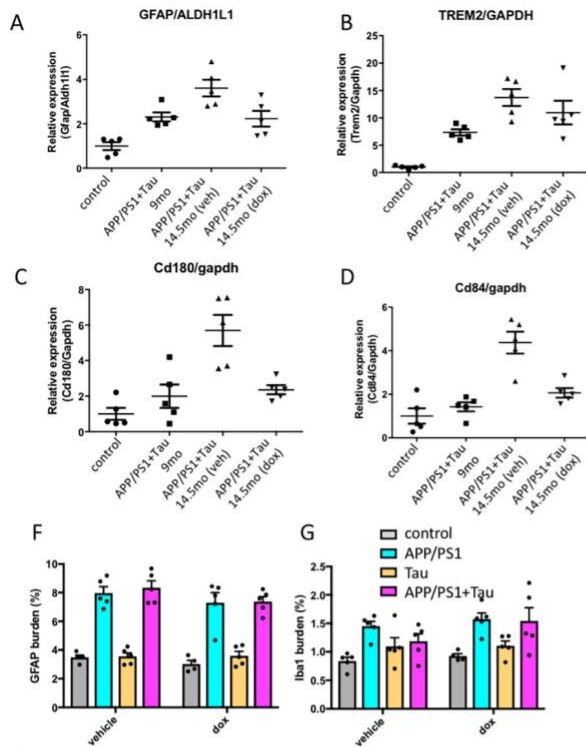
**Eleanor K. Pickett, Abigail G. Herrmann, Jamie McQueen, Kimberly Abt, Owen Dando, Jane Tulloch, Pooja Jain, Sophie Dunnett, Sadaf Sohrabi, Maria P. Fjeldstad, Will Calkin, Leo Murison, Rosemary J. Jackson, Makis Tzioras, Anna Stevenson, Marie d'Orange, Monique Hooley, Caitlin Davies, Marti Colom-Cadena, Alejandro Anton-Fernandez, Declan King, Iris Oren, Jamie Rose, Chris-Anne McKenzie, Elizabeth Allison, Colin Smith, Oliver Hardt, Christopher M. Henstridge, Giles E. Hardingham, and Tara L. Spires-Jones**



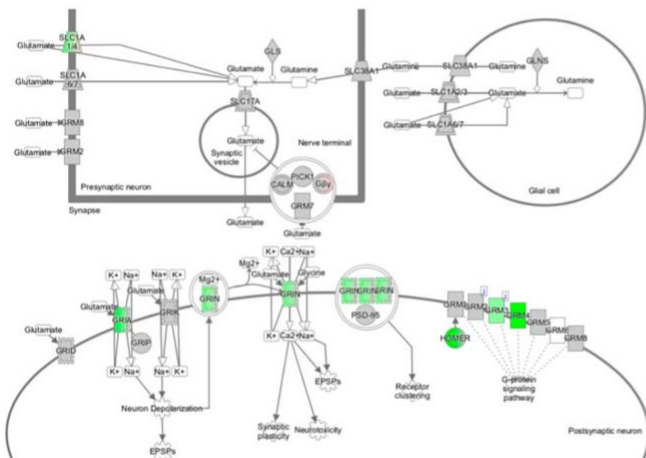
**Figure S1 related to Figure 1: No age-related loss of brain, weight, body weight, or hippocampal volume in APP/PS1+Tau mice.** There was a significant effect of genotype on brain weight (A,  $F[3,163]=40.28$ ,  $p<0.0001$ ), body weight B,  $F[3,153]=37.25$ ,  $p<0.001$ ), and hippocampal volume (C,  $F[3,50]=4.82$ ,  $p=0.005$ ) with both tau mice and APP/PS1+Tau mice exhibiting reductions compared to the other 2 genotypes. Despite reduced hippocampal volume in both APP/PS1+Tau and Tau mice, there was no age-related reduction in hippocampal volume (C, 2-way ANOVA effect of age  $F[3,50]=0.002$ ,  $p=0.997$ ). The reduction in brain and body weight is driven not by tau expression but by the CK1TA activator transgene that is needed to drive the tau transgene. 9 month old MAPTnullxCK1TA mice in the absence of the Tg21221 responder transgene have reduced brain (D, unpaired t-test  $t=5.144$   $df=16$ ,  $p<0.0001$ ) and body weights (E, unpaired t-test  $t=2.803$   $df=16$ ,  $p=0.012$ ). This loss of brain and body weight did not affect total distance travelled in the open field (F). Similar to the cortical data presented in Fig 1, hippocampal plaque burden was significantly smaller in APP/PS1+Tau mice compared to APP/PS1 mice (G, 2-way ANOVA effect of genotype  $F[1,26]=11.21$ ,  $p=0.002$ , effect of age  $F[2,26]=44.19$ ,  $p<0.0001$ ).



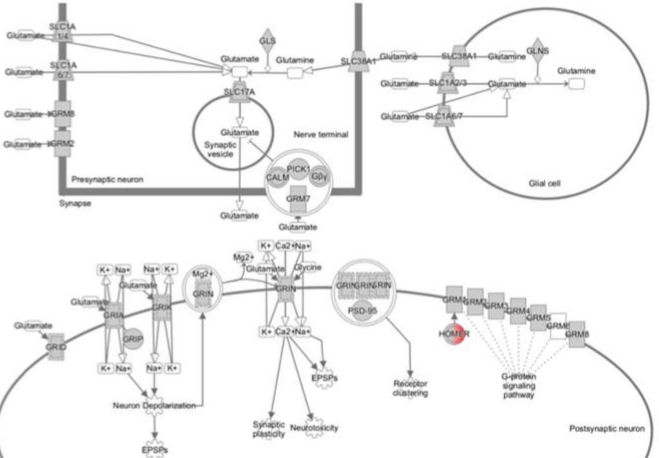
**Figure S2 related to Figure 2: Hyperactivity is not different between male and female mice and open field data indicate anxiety in APP/PS1+Tau mice.** The hyperactivity phenotype observed in APP/PS1+Tau mice is not different between male and female mice (A, one-Way ANOVA effect of sex  $p > 0.05$  at all age groups). To determine whether APP/PS1+Tau mice have an anxiety phenotype, open field data was analysed by the distance travelled in the inner segment versus outer segment of the arena (B). At 10.5 months, there was a significant difference between genotypes in the inner (C, ANOVA,  $F[3,69]=4.075$ ,  $p=0.010$ ) and outer (D, ANOVA  $F[3,69]=15.91$ ,  $p<0.0001$ ) portions of the arena. APP/PS1+Tau mice travelled significantly further in both the inner and outer arena compared to MAPTnull mice (\* Tukey's posthoc test  $p<0.01$ ). At 14.5 months, there were no significant differences between genotype and treatment in distance travelled in the inner arena (E). At 14.5 months of age, APP/PS1+Tau mice travel over 2 times farther in the outer portion of the arena (F) than other genotypes, a phenotype which recovers with dox treatment (2-way ANOVA genotype  $F[3,69]=19.548$ ,  $p<0.0001$ ; treatment  $F[1,69]=3.9990$ ,  $p=0.0497$ , interaction  $F[2,69]=4.770$ ,  $p=0.004$ ). \*, \*\* Tukey's multiple comparisons tests  $p=0.002$ ,  $p<0.0001$ ). Graphs depict mean  $\pm$  SEM. Individual points represent the mean value for each mouse. The dotted line in A indicates that a different cohort of mice was used at 3,6, 9 months of age and at 10.14 months of age.



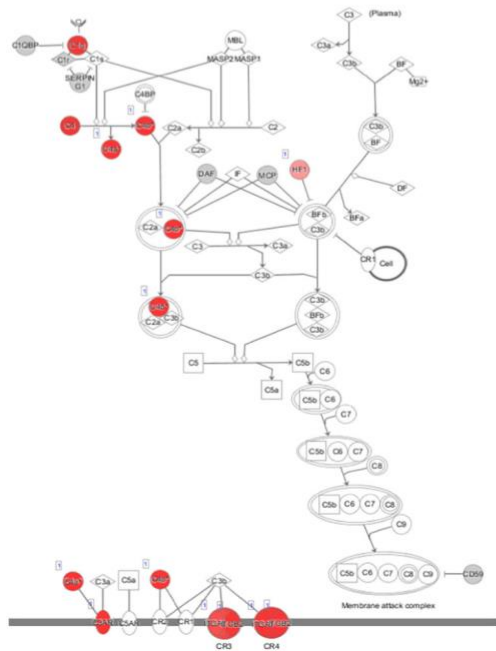
**J** APP/PS1+Tau vehicle vs control glutamate receptor signaling



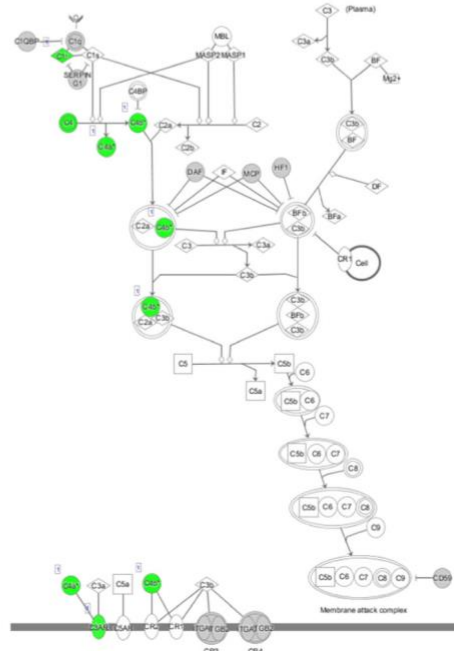
**K** APP/PS1+Tau dox vs control glutamate receptor signaling



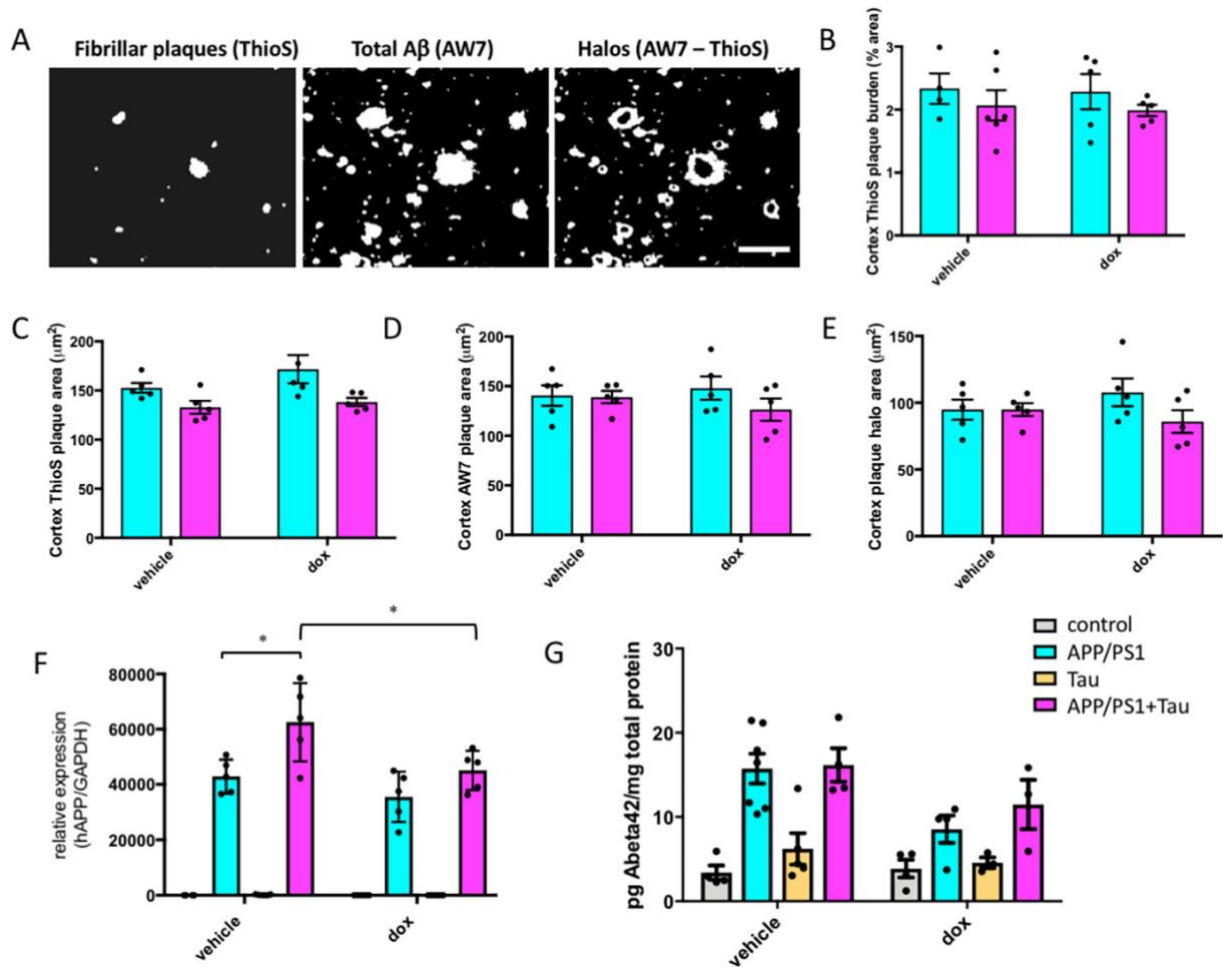
**L** APP/PS1+Tau vehicle vs control complement system



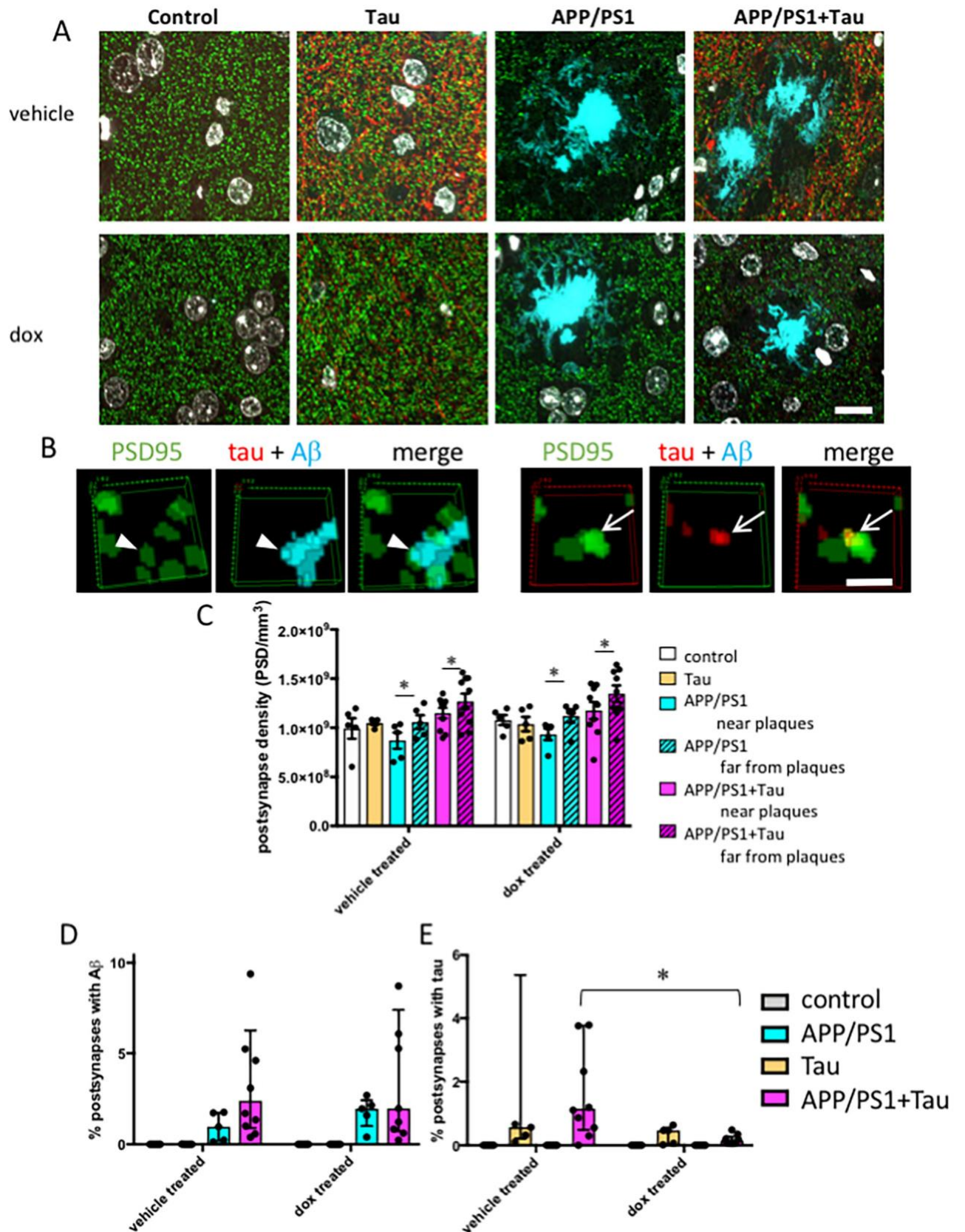
**M** APP/PS1+Tau dox vs control complement system



**Figure S3 related to Figure 3: RT-PCR validation of RNA-seq results** at 9 months and 14.5 months of age indicate that the upregulated genes GFAP (A), Trem2 (B), Cd180 (C), and Cd84 (D) increase between 9 and 14.5 months and that this is prevented by dox treatment. The percentage area occupied by GFAP labelled astrocytes (cyan, E) and Iba1 labelled microglia (magenta E) was higher in 14.5 month old mice in genotypes with plaques but did not change with tau transgene suppression (F, GFAP 2-way ANOVA effect of genotype  $F[3,31]=75.16$ ,  $p<0.001$ , treatment  $F[1,31]=3.22$ ,  $p=0.082$ ; G, Iba1 2-way ANOVA effect of genotype  $F[3,31]=9.05$ ,  $p=0.0002$ , treatment  $F[1,31]=2.48$ ,  $p=0.13$ ). Dox treatment significantly rescues transcriptional changes in Tau mice at 14.5 months (H) without affecting control mice (I). Dox treated control data correlate very closely with values from vehicle treated control animals (J). To visualise recovery of networks with dox treatment, the changes in APP/PS1+Tau vehicle treated mice compared to controls are shown for glutamate signalling (j) and complement system (l) and dox treated APP/PS1+Tau mice compared to controls are shown in k and m. Transcripts labelled in red are increased and those labelled in green are decreased. Scale bar represents 40  $\mu\text{m}$ .



**Figure S4 related to Figures 3 and 4: Tau suppression does not affect amyloid pathology.** Tau suppression did not change amyloid pathology in APP/PS1+Tau mice. In 14.5 month old mice, fibrillar plaques were measured with ThioS, total A $\beta$  with AW7 immunostaining, and oligomeric A $\beta$  halos were measured by subtracting the fibrillar cores from total Ab staining (a). None of the amyloid plaque measurements was changed by doxycycline treatment (b-e). APP mRNA levels (f) were increased by 30% in APP/PS1+Tau mice, an effect which was ameliorated by dox treatment (2-way ANOVA genotype  $F[3,31]=153.5$ ,  $p<0.0001$ , treatment  $F[1,31]=7.912$ ,  $p=0.0084$ , interaction  $F[3,31]=3.468$ ,  $p=0.0279$ , \* post-hoc Tukey's test  $p<0.01$ ). Soluble A $\beta$ 42 ELISA on brain homogenates (g) showed a significant effect of genotype since mice without the human APP/PS1 transgene have low levels of A $\beta$  (2-way ANOVA effect of genotype  $F[3,26]=13.14$ ,  $p<0.0001$ ). There was a significant effect of dox treatment when all genotypes were considered (2-way ANOVA effect of treatment  $F[1,26]=6.11$ ,  $p=0.02$ ), however post-hoc analyses reveal that A $\beta$ 42 levels were not changed in APP/PS1+Tau mice compared to APP/PS1 mice an neither APP/PS1+Tau nor APP/PS1 A $\beta$ 42 levels were significantly reduced by dox treatment (Tukey's multiple comparisons tests  $p>0.05$ ). Scale bar represents 30  $\mu\text{m}$ .



**Figure S5 related to Figure 4: Tau suppression reduces post-synaptic accumulation of tau.** To investigate post-synapse loss and synaptic proteins, array tomography ribbons from 14.5 month old mice were stained for postsynaptic terminals (green), human tau (red), and amyloid beta (AW7, cyan).

Maximum intensity projections of 10 serial 70 nm sections are shown in a. Three-dimensional reconstructions of 5 consecutive serial sections from processed image stacks of a APP/PS1+Tau mouse (b) demonstrate post-synaptic terminals positive for tau (arrows) or A $\beta$  (arrowheads). Quantification reveals significant post-synapse loss near plaques in APP/PS1 and APP/PS1+Tau mice which is not rescued by lowering tau levels with doxycycline (dox) treatment (c). The percentage of post-synapses positive for A $\beta$  is not different between APP/PS1 mice and APP/PS1+Tau mice, nor is it affected by dox treatment (d). The percentage of post-synapses containing tau is significantly lowered by dox treatment in APP/PS1+Tau mice (e, \* Mann-Whitney U test p=0.004). Data represent mean + SEM (c) and median + interquartile range (d-e). Scale bars represent 10  $\mu$ m in a, 1  $\mu$ m in b.

**Table S5 related to STAR methods - oligonucleotides**

<b>Genotyping primers</b>
PSEN1dE9 forward primer: GGCTACCATTAAGTCAGTCAGCTTT
PSEN1dE9 reverse primer: CCCACAGTCTCGGTATCTTCTG
APPSwe forward primer: CCGACATGACTCAGGATATGAAGTT
APPSwe reverse primer: CCGACATGACTCAGGATATGAAGTT
CkTTA forward primer: TGCCAACAAGGTTTTTCACTAGAGA
CkTTA reverse primer: CTCTTGATCTTCCAATACGCAACCTA
MAPT forward primer: CTGCTCCAAGACCAAGAAGGA
MAPT reverse primer: TGTGTATGTCCACCCCACTGA
<b>RNASeq validation QPCR primers</b>
<i>Gapdh</i> forward primer: GGGTGTGAACCACGAGAAAT
<i>Gapdh</i> reverse primer: CCTTCCACAATGCCAAAGTT
<i>MAPT</i> forward primer: CCAATCACTGCCTATACCC
<i>MAPT</i> reverse primer: CCACGAGAATGCGAAGGA
Human mutant <i>APP</i> forward primer: CCGACATGACTCAGGATATGAAGTT
Human mutant <i>APP</i> reverse primer: CCTTTGTTTGAAACCCACATCTTCTG
<i>Trem2</i> forward primer: CTGGAACCGTCACCATCACTC
<i>Trem2</i> reverse primer: CGAAACTCGATGACTCCTCGG
<i>Gfap</i> forward primer: GCAAAAGCACCAAAGAAGGGGA
<i>Gfap</i> reverse primer: ACATGGTTCAGTCCCTTAGAGG
<i>Aldh1l1</i> forward primer: CATCCAGACCTTCCGATACTTC
<i>Aldh1l1</i> reverse primer: ACAATACCACAGACCCCAAC
<i>Cd180</i> forward primer: CCAAAGCCAACATCGGTTAGACAC
<i>Cd180</i> reverse primer: CAGAGACCCTCAAACACGGCAGG
<i>Cd84</i> forward primer: GCTGAAGTTACCATAACCCAGG
<i>Cd84</i> reverse primer: CAAAAGTAAATCCAAGGCCCG