

***New Phytologist* Supporting Information**

Article title: A LysM effector subverts chitin-triggered immunity to facilitate arbuscular mycorrhizal symbiosis

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The following Supporting Information is available for this article:

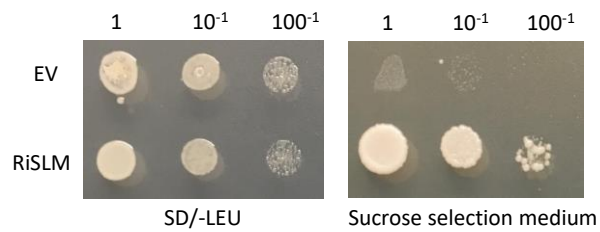


Fig. S1 Yeast signal sequence trap. pYST-02-RiSLM, representing a fusion of full length RiSLM (including its endogenous signal sequence) with an invertase, and the empty pYST-02 vector (EV) were transformed into *Saccharomyces cerevisiae* Y02321, and grown on SD/-Leu (containing glucose) and sucrose selection medium at different dilutions for 3 days at 30 °C.

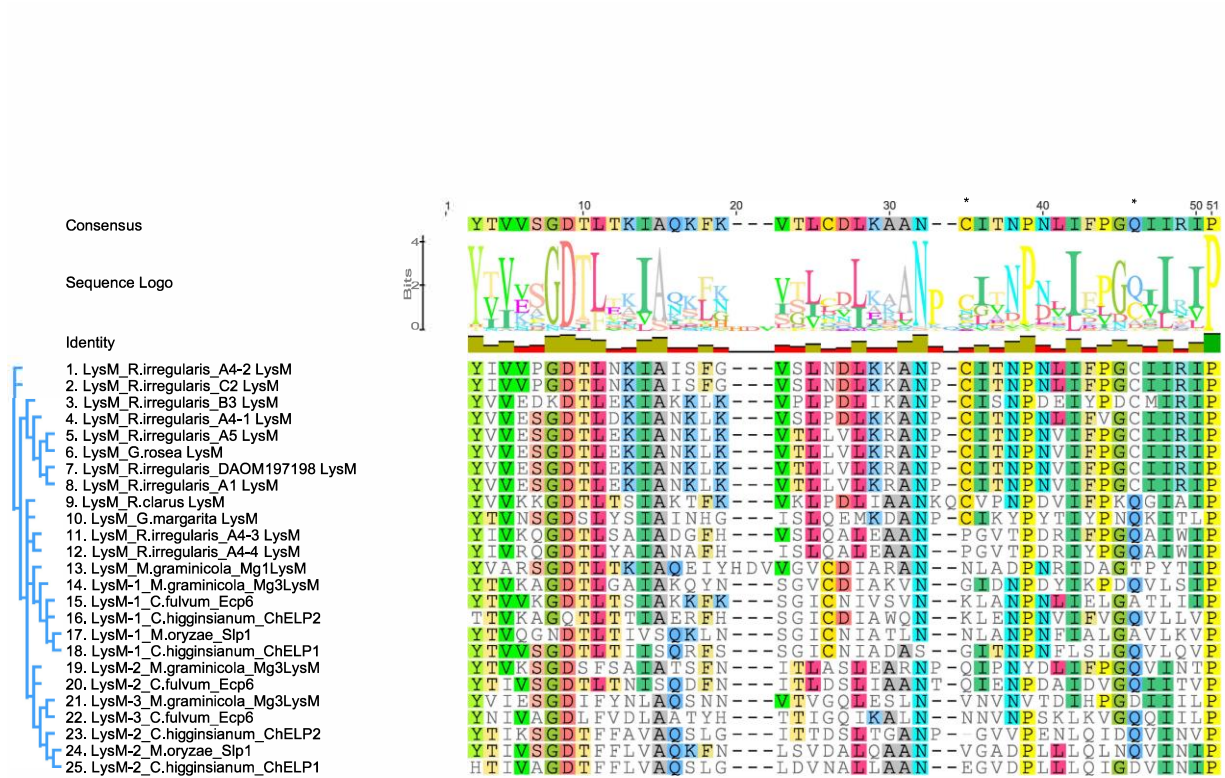


Fig. S2 Comparison of LysM domains from AM fungal or pathogenic LysM effectors.

MAFFT was used to align all amino acid sequences. Neighbour-joining Tree was built using Geneious tree builder.

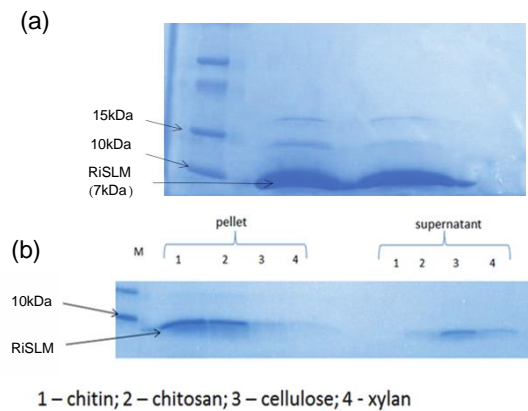


Fig. S3 Purified RiSLM protein binds chitin and chitosan. (a) Coomassie stained protein gel of RiSLM (size ~7kDa) purified from *Escherichia coli* ORIGAMI. (b) Coomassie stained protein gel of affinity precipitation assays showing that RiSLM binds to insoluble chitin and chitosan but not xylan and cellulose. For the latter two polymers the vast majority of the protein is retained in the supernatant.

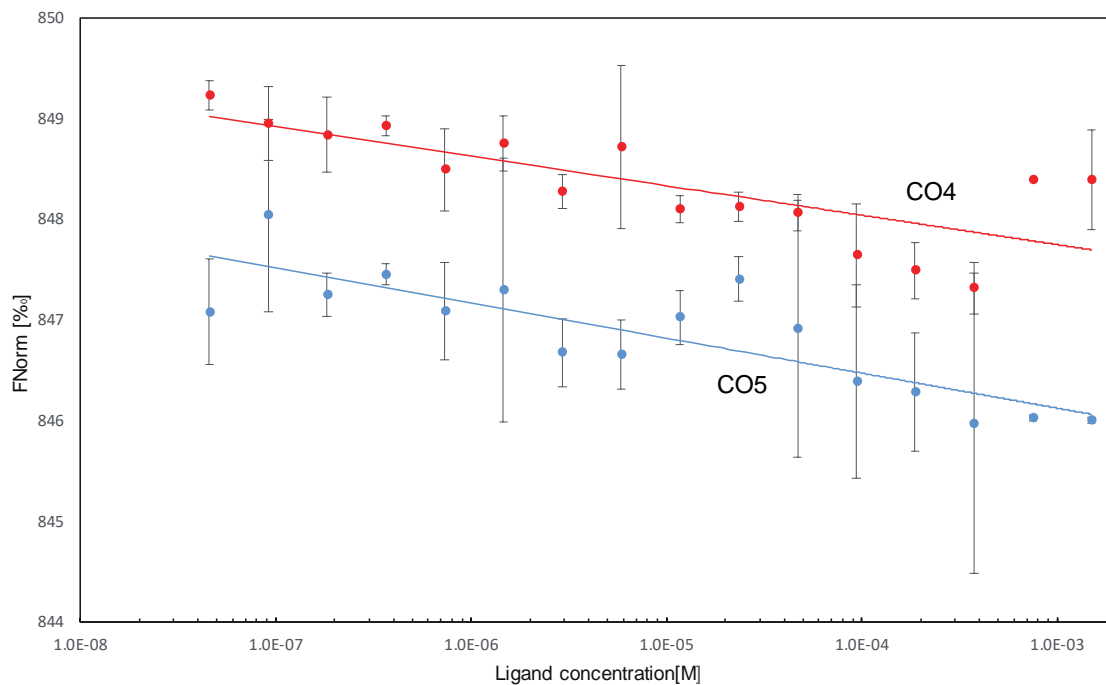


Fig. S4 RiSLM does not bind deacylated chitotetraose (CO4) and chitopentaose (CO5) as revealed by microscale thermophoresis. Binding experiments using microscale thermophoresis do not show any interaction with RiSLM in the range of concentrations tested (up to 1 mM). Error bars represent standard deviation from two independent measurements using two batches of independently purified proteins.

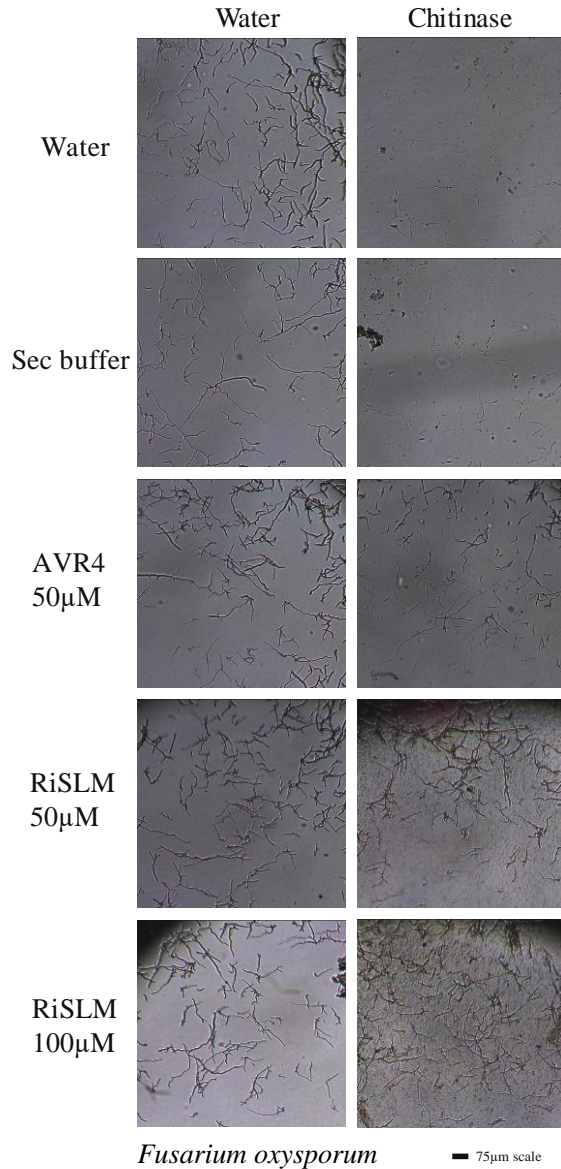


Fig. S5 RiSLM protects hyphae from the pathogenic fungus *Fusarium oxysporum f.sp lycopersici* against plant chitinases. Germinated hyphae are treated with 50 μM AVR4, 50 μM RiSLM or 100 μM RiSLM and incubated at room temperature for 2 hours, and subsequently treated with a tomato chitinase preparation or water for 4 hours. Water or buffer used for size-exclusion chromatography (SEC) were used as negative controls. The Avr4 effector from *Cladosporium fulvum* was used as positive control.

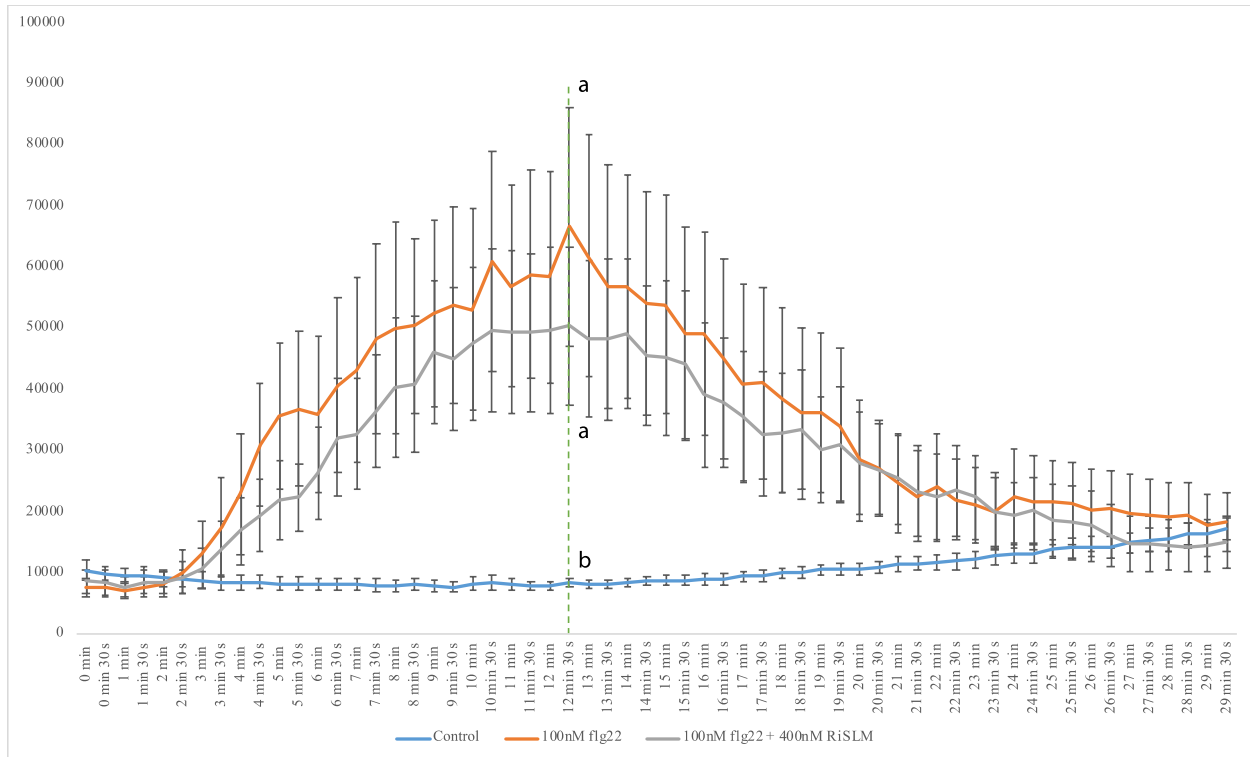


Fig. S6 RiSLM does not suppress flg22-induced reactive oxygen species (ROS) production.

Medicago truncatula A17 root pieces were treated with 100nM flg22 with or without 400nM RiSLM . Error bars represent standard error from 6 (control and 100nM flg22 + 400nM RiSLM treated) or 5 (100nM flg22 treated) biological replicates. Different letters indicate significant difference (LSD $p < 0.05$) between different treatments at the indicate time point (green dotted line).

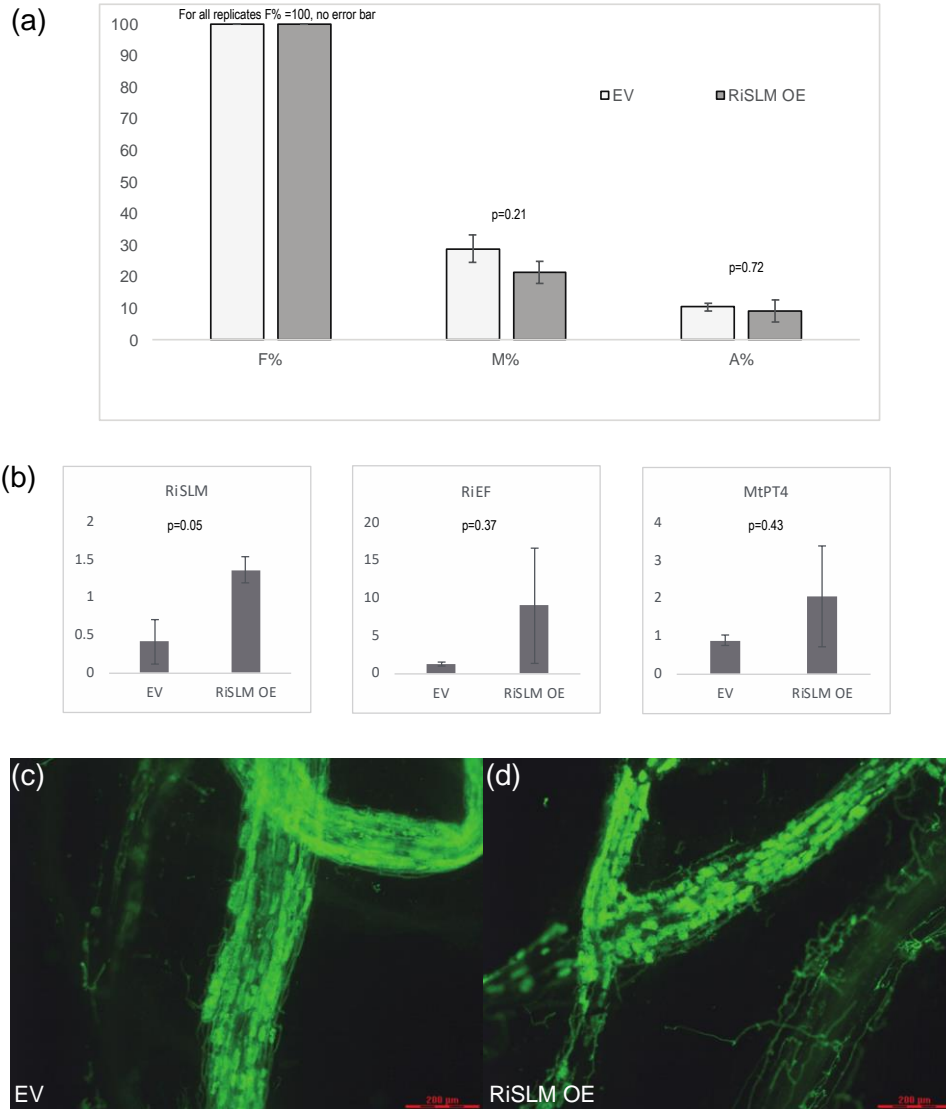


Fig. S7 Figure S7. *RiSLM* overexpression does not enhance mycorrhization in *Medicago*

***truncatula*.** (a) Frequency (F%), mycorrhization intensity in the root (M%), or arbuscule

abundance in the root (A%) are not affected by overexpressing of *RiSLM*. (b) qPCR analysis of

control and *RiSLM* overexpressing roots showing *RiSLM* expression level relative to

Rhizophagus irregularis elongation factor *RiEF* and *MtPT4* expression levels relative to

Medicago elongation factor *MtEF*. Error bar represents standard error from 3 replicates. (c, d)

WGA-alex488 staining of mycorrhization in *RiSLM* overexpressed roots (d) or control roots (c).

Scale bar = 200 μm.

Table S1 Primers used in this research.

Primer	Sequence	Note
RiSLM_SP-F	CTGAATTCATGAGACTCAATCAATCTTTGCTC	for cloning into pYST-02
RiSLM_SP-R	AAGCGGCCGCAAACATCTAGTAGCATTAGGTA TCC	
MtEPI-qPCR-F	GGTTGTGTGTGTAACAGGTGCTTCA	qPCR primers for three defence marker genes
MtEPI-qPCR-R	AGGGTCACGAACAGTGGCTCTA	
MtPAL-qPCR-F	GCAAGGGCCGGTGTGAAGC	
MtPAL-qPCR-R	GGCAGCACCAAAGCCAGTGGT	
MtTHA-qPCR-F	GGCGCAATCCCACCAGCAAC	
MtTHA-qPCR-R	ACCACTCCCACCTTGTGGCG	
MtVapyrin-qPCR-F	TCATCCTCCACAACAACAAGGT	qPCR primers for three symbiotic marker genes
MtVapyrin-qPCR-R	TCAAGCACTTCTCTTATGTCATCCATTG	
MtPUB1-qPCR-F	GTAGCAGCAAAAGGCACAAG	
MtPUB1-qPCR-R	ATATCGGTTCGAACCAGTTGC	
MtTUBB1-qPCR-F	GAACAGTTCTCTGCCATGTTTCAGAA	
MtTUBB1-qPCR-R	TCTTCATAAACTGGGGCATCCTGGT	
MtEF-qPCR-F	GATTGCCACACCTCTCACAT	qPCR primer for Medicago EF
MtEF-qPCR-R	TCAGCGAAGGTCTCAACCAC	
RiEF-qPCR-F	AACCCCTTCGTCTTCCACTT	qPCR primer for Rizophagus EF
RiEF-qPCR-R	ATTGTTTCGTGGTGCATTCA	
RiSLM-qPCR-F	AACGTAATCTTTCCCGTTG	qPCR primer for RiSLM
RiSLM-qPCR-R	TGGTGTATTTCTCTCACGGCTA	
RiSLM-HIGS-F	AAAAAAGCAGGCTTCATGAGACTCAATCAATCTTTG CTC	primers to make RiSLM HIGS construct
RiSLM-HIGS-R	CAAGAAAGCTGGGTCGATTTGTAATACAAGGATTTG CTCT	
RiSLM_F_EcoR1	GAATTCGCAATTAAGACATGTACTCC	primers used to clone RiSLM into pET-SUMO
RiSLM_R_Hind3	AAGCTTATTATTA AAAACATCTAGTAGCATTAG GTATCC	
F-RiSLM	AGAAGACTCAGGTGCAATTAAGACATGTACTC CG	Primers for LO golden gate cloning (<i>RiSLM</i> overexpression)
R-RiSLM	GGAAGACAGCGAAAAACATCTAGTAGCATTAG G	

Table S2 Coding sequences of LysM effectors from different AM fungal species or isolates.

RiSLM [organism= <i>Rhizophagus irregularis</i>] isolate DAOM197198 LysM effector RiSLM completely CDS	ATGAGACTCAATCAATCTTTGCTCCTCTTAACCGTCCTTTTCG CCCTTATTGCTGTTGCTTCATGTGCAATTAAGACATGTA GGTTTATGTTGTTGAGAGTGGTGATACCCTTGAAAAATCGC GAATAAACTTAAGGTGACCTTGCTAGTTTTGAAAAGAGCAA TCCTTGATTACAAATCCGAACGTAATCTTTCCCGGTTGTATT ATCCGGATACCTAATGCTACTAGATGTTTTTAA
RiSLM_A1 [organism= <i>Rhizophagus irregularis</i>] isolate A1 LysM effector completely CDS	ATGAGACTCAATCAATCTTTGCTCCTCTTAACCGTCCTTTTCG CCCTTATTGCTGTTGCTTCATGTGCAATTAAGACATGTA GGTTTATGTTGTTGAGAGTGGTGATACCCTTGAAAAATCGC GAATAAACTTAAGGTGACCTTGCTAGTTTTGAAAAGAGCAA TCCTTGATTACAAATCCGAACGTAATCTTTCCCGGTTGTATT ATCCGGATACCTAATGCTACTAGATGTTTTTAA
RiSLM_A5 [organism= <i>Rhizophagus irregularis</i>] isolate A5 LysM effector completely CDS	ATGAGACTCAATCAATCTTTGCTCCTCTTAACCGTCCTTTTCG CCCTTATTGCTGTTGCTTCATGTGCAATTAAGACATGTA GGTTTATGTTGTTGAGAGTGGTGATACCCTTGAAAAATCGC GAATAAACTTAAGGTGACCTTGCTAGTTTTGAAAAGAGCAA TCCTTGATTACAAATCCGAACGTAATCTTTCCCGGTTGTATT ATCCGGATACCTAATGCTACTAGATGTTTTTAA
RiSLM-1_A4 [organism= <i>Rhizophagus irregularis</i>] isolate A4 LysM effector-1 completely CDS	ATGAGACTCAATCAATCTTTGCTCCTCTTAACCATCCTTTTCG CCCTTATTGCTGTTGCTTCATGTGCAGTTAAGACATGTA GGTTTATGTTGTTGAGAGTGGTGATACCCTTGATAAAATCGC GAATAAACTTAAGGTGTCCTTGCCAGATTTGAAAAAAGCAA TCCTTGATTACAAATCCGAACCTAATCTTTGTCGGTTGTATT ATCCGGATACCTAATGCTACTGGATGTTTTTAA
RiSLM-2_A4 [organism= <i>Rhizophagus irregularis</i>] isolate A4 LysM effector-2 completely CDS	ATGAGACTCAATAAATCTTTGCTCCTCTTAACCATCCTTTTCG CCCTTATTGCTGTTGCTTCATCTCAGCGTCTGACGACATGTA TCGGGTTTATATTGTTGTGCCTGGTGATACCCTTAATAAAATC GCGATAAGTTTTGGTGTGTCCTTGAATGACTTGAAAAAAGCA AATCCTTGATTACAAATCCGAACCTAATCTTTCCCGGTTGCA TTATCCGGATACCTAATCGTACTCAATGTTTTTAA
RiSLM-3_A4 [organism= <i>Rhizophagus irregularis</i>] isolate A4 LysM effector-3 completely CDS	ATGAGACTCAGCCTATTTTTTCTCCCCTTAACCATCATTTTCG CCCTTATTGCTGCTGTTGCTTCGGCTGGTTGGTATATAGTTAA ACAGGGTGATACCCTTTCTGCTATCGCGGATGGGTTTCATGTA TCCTTGCAAGCTTTGGAAGCCGCTAATCCGGGAGTTACTCCG GACAGAATCTTTCTGGACAAGCTATCTGGATACCCTAA
RiSLM-4_A4 [organism= <i>Rhizophagus irregularis</i>] isolate A4 LysM effector-4 completely CDS	ATGAGACTCAGCCTATTTTTTCTCCCCTTAACCATCCTTTTCG CCTTATTGCTTCTGTTGCTTCGGCTGGTTGGTATATAGTTAGA CAGGGTGATACCCTTTATGCTATCGCGAATGCGTTTCATATAT CCTTGCAAGCTTTGGAAGCCGCTAATCCGGGAGTTACTCCGG ACAGAATCTATCCTGGACAAGCTATCTGGATACCCTAA
RiSLM_B3	ATGAGACTCAATCTATCTTTGCTCCCCTTGACCATCCTTTTCG

[organism= <i>Rhizophagus irregularis</i>] isolate B3 LysM effector completely CDS	CCCTTATTGCTGTTGCTTCATGTGCAACTATGAAATGTGCTCC GGTTTATGTTGTTGAGGATAAAGATACTCTTGAAAAATCGC GAAAAACTTAAGGTGCCCTTGCCAGATTTGATAAAAGCTAA TCCTTGTATTTCAAATCCGGACGAAATCTATCCTGATTGCATG ATCCGGATACCTAAACAACTAAATGCACTAAGGTTTGTATT TTATCTTTATTGTTTATATTTATCGATTACTACTAA
RiSLM_C2 [organism= <i>Rhizophagus irregularis</i>] isolate C2 LysM effector completely CDS	ATGAGACTCAATCAATCTTTGCTCCTCTTAACCATCCTTTTCG CCCTTATTGCTGTTGCATCATCTCAGCGTCTGACGACATGTAT TCAGGTTTATATTGTTGTGCCTGGTGATACCCTTAATAAAATC GCGATAAGTTTTGGTGTGTCCTTGAATGACTTGAAAAAGCA AATCCTTGTATTACAAATCCGAACCTAATCTTTCCCGGTTGCA TTATCCGGATACCTAATCGTACTAAATGTTTTTAA
RcSLM [organism= <i>Rhizophagus clarus</i>] LysM effector completely CDS	ATGAAACACAATCTATTTTTGCTCTCCTTAACCATCCTTTTCG CGCTTAGTGCTTTTACTGATGTTACTTCGGCTGCTGCGAAAGG GTTTAGATATGTTGTTAAAAAGGGTGATACCCTTACCAGCAT CGCGAAAACTTTTAAAGTAAAATTGCCTGATTTGATCGCGGC TAATAAGCAGTGTGTTCCAAATCCGGACGTAATCTTTCCCAA ACAAGGTATCGCGATACCTCAGTACTGTCCCGTCTGTCCGTA G
GrSLM [organism= <i>Gigaspora rosea</i>] LysM effector completely CDS	ATGAGACTCTGTTTATTTTTGCTTCCCTTAACCATCATTTTCGC CTTTATTGCTGCTGTTGCTTCAATCAATTGTTGCTATACAGTT AACTCAGGCGATTCCCTTTATTCCATCGCAATTAATCATGGCA TAAGCTTGCAAGAAATGAAAGATGCTAATCCATGCATTAAT ACCCGTACACTATTTATCCAATCAAAAAATCACGCTCCCTA ACCAACCATACTGTTGA
GmSLM [organism= <i>Gigaspora margarita</i>] LysM effector completely CDS	ATGAGACTCAATCAATATTTGCTCCTCTTAACCGTCCTTTTCG CCCTTATTGCTGTTGCTTCATGTGCAATTAAGACATGTACTCC GGTTTATGTTGTTGAGAGTGGTGATACCCTTGAAAAATCGC GAATAAACTTAAGGTGACCTTGCTAGTTTTGAAAAGAGCAAA TCCTTGTATTACAAATCCGAACGTAATCTTTCCCGGTTGTATT ATCCGGATACCTAATGCTACTAGATGTTTTTAA