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### Supporting Information

## A Scaffold-Diversity Synthesis of Biologically Intriguing Cyclic Sulfonamides

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#### Supporting Information

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#### 1. General Information

All commercially available compounds were used as provided without further purification unless otherwise noted. Solvents were purchased from Fisher, VWR and Acros in laboratory, reagent and anhydrous grade, as labelled by corresponding companies. If no additional information regarding treatment is given, the solvents were used directly from the container. Reagents were purchased from Alfa Aesar, Sigma Aldrich and VWR respectively. If it is stated, that a reaction was carried out under an Argon atmosphere, standard Schlenk techniques were used. Column chromatography was performed using silica gel (Acros, particle size 0.035-0.070 mm).

<sup>1</sup>H, <sup>13</sup>C and <sup>19</sup>F-NMR were recorded on a Bruker DRX400 (400 MHz), Bruker DRX500 (500 MHz), INOVA500 (500 MHz) or Bruker Biospin AVANCE HDX-III (700 MHz) using Chloroform*d* (CDCl<sub>3</sub>), CD<sub>2</sub>Cl<sub>2</sub>, (CD<sub>3</sub>)<sub>2</sub>CO, or (CD<sub>3</sub>)<sub>2</sub>SO as solvent at room temperature. <sup>1</sup>H and <sup>13</sup>C-NMR spectra were calibrated to the solvent signals of Chloroform-*d* (7.26 and 77.16 ppm), CD<sub>2</sub>Cl<sub>2</sub> (5.32 and 53.84 ppm), CD<sub>3</sub>CN (1.94 and 1.32/118.26 ppm) or (CD<sub>3</sub>)<sub>2</sub>SO (2.50 and 39.52 ppm).<sup>[1]</sup>The abbreviations *s*, *d*, *t*, *q* and *m* stand for singlet, doublet, triplet, quartet and multiplet in that order. High resolution mass spectra (HRMS) were recorded on a *LTQ Orbitrap* mass spectrometer coupled to an *Accela HPLC*-System (HPLC column: Hypersyl GOLD, 50 mm x 1 mm, particle size 1.9 µm; Ionization method: electron spray ionization). Preparative HPLC was performed using a 1260 Infinity II system by Agilent Technologies and Nucleodur C18 Gravity VP10/125 5 µm or Nucleodur C18 gravity VP21/125 5 µm columns by Macherey-Nagel.

#### 2. Experimental section - Synthesis

2.1. Branching Pathway

#### 2.1.1 Synthesis of Ketimines (1), Allenes (3) and Sulfonium Salts

CO<sub>2</sub>Et-1,<sup>[1]</sup> H-1,<sup>[2]</sup> Me-1<sup>[3]</sup>, 2-Pyr-1<sup>[4]</sup> were synthesized according to procedures previously reported in literature. Spectral data were in accordance with reported ones.



Allenes **3-OEt**, **3-OtBu** and **3-OBn** were synthesized according to reported literature procedures;<sup>[5][6]</sup> with recorded spectral data matching previously reported ones.

SMe<sub>2</sub> Organo Halide 
$$R^2$$
  $R^2 = CH_2CO_2Et$   
Acetone  $/$   $R^2 = Bn$ 

Sulfonium salts (carbethoxy)methyl dimethylsulfonium bromide and benzyl dimethylsulfonium bromide were synthesized and handled according to reported literature procedures; with recorded spectral data matching previously reported ones.<sup>[7][8]</sup>

#### 2.1.2 Synthesis of Aziridines 2a-2h



#### General Reaction Scheme

Aziridines were synthesized according to general scheme depicted above. Observed diastereoselectivity (*syn* in most examples) or lack of it in cases of **2c** and **2h** can be described and established by the procedure shown in Scheme S1, referring to general considerations of sulfur ylide mediated aziridination.<sup>[9]</sup>



Scheme S1: Model for diastereoselectivity of aziridination reaction.

#### (+)-7b-methyl-1-phenyl-1,7b-dihydroazirino[1,2-b]benzo[d]isothiazole 3,3-dioxide (2a)

Benzyl dimethylsulfonium bromide (1.4 eq., 100.0 mg, 0.43 mmol), Potassium carbonate (2.0 eq., 32.8 mg, 0.24 mmol ) and 3-methylbenzo[d]isothiazole 1,1-dioxide **1-Me** (1.0 eq., 21.0 mg, 0.12 mmol) were combined in Acetonitrile (970 µl). The reaction was stirred for 10 h at ambient temperature. Then, an additional 2.2 eq (157 mg, 0.68 mmol) of sulfonium salt were added to the mixture and the reaction stirred for 30 additional hours at ambient temperature. The reaction was concentrated and objected to silica gel column chromatography (7% to 14% EA/CyH) to yield 22 mg (0.08 mmol, 68%) of *syn-2a* and 3 mg (0.01 mmol, 9%) of *anti-2a*.



<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.77 (dt, *J* = 7.6, 0.9 Hz, 1H), 7.69 (td, *J* = 7.6, 1.2 Hz, 1H), 7.62 – 7.56 (m, 2H), 7.45 – 7.34 (m, 5H), 3.65 (s, 1H), 1.60 (s, 3H); <sup>13</sup>**C NMR** (126 MHz, Chloroform-d)  $\delta$  141.2, 133.8, 133.7, 132.6, 130.3, 128.8, 128.7, 127.7, 124.3, 123.66, 61.6, 56.9, 13.1; **HMRS (ESI):** Calculated for C<sub>15</sub> H<sub>14</sub> O<sub>2</sub> N S = [M+H]<sup>+</sup>: 272.07398, found: 272.07328



<sup>1</sup>**H NMR** (600 MHz, Chloroform-*d*)  $\delta$  7.73 (dt, *J* = 7.8, 0.9 Hz, 1H), 7.68 (ddd, *J* = 7.8, 7.1, 1.2 Hz, 1H), 7.46 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 7.41 (dt, *J* = 7.7, 1.0 Hz, 1H), 7.15 – 7.12 (m, 1H), 7.10 – 7.06 (m, 2H), 6.93 (dq, *J* = 7.3, 1.2 Hz, 2H), 4.21 (s, 1H), 2.07 (s, 3H); <sup>13</sup>**C NMR** (151 MHz, Chloroform-d)  $\delta$  137.7, 137.4, 133.4, 131.0, 130.4, 128.5, 128.3, 127.9, 125.6, 122.6, 62.2, 55.1, 19.7; **HMRS (ESI):** Calculated for C<sub>15</sub> H<sub>14</sub> O<sub>2</sub> N S = [M+H]<sup>+</sup>: 272.07398, found: 272.07359



(<u>+</u>)-7b-methyl-1-carbethoxy-1,7b-dihydroazirino[1,2-b]benzo[d]isothiazole 3,3-dioxide (2b)



(Carbethoxymethyl)dimethylsulfonium bromide (1.2 eq., 99 mg, 0.43 mmol, Potassium carbonate (2.0 eq., 99.0 mg, 0.72 mmol) and 3-methylbenzo[d]isothiazole 1,1-dioxide **1-Me** (1.0 eq., 65.0 mg, 0.36 mmol) were combined in Acetonitrile (1.3 ml). The reaction was concentrated and objected to silica gel column chromatography (15% to 21% EA/CyH) to yield 81.0 mg (0.30 mmol, 84%) of product aziridine.

R<sub>*f*</sub> (30% EA/CyH) = 0.55; <sup>1</sup>**H NMR** (400 MHz, Chloroform-d) δ 7.74 – 7.64 (m, 2H), 7.61 – 7.54 (m, 2H), 4.28 (m, 2H), 3.15 (s, 1H), 1.91 (s, 3H), 1.32 (t, J = 7.1 Hz, 3H). <sup>13</sup>**C NMR** (101 MHz, Chloroform-d) δ 164.0, 139.2, 134.0, 133.2, 130.7, 124.9, 123.3, 62.4, 55.5, 53.9, 14.1, 13.3.

**HMRS (ESI):** Calculated for  $C_{12} H_{14} O_4 N S = [M+H]^+$ : 268.06381, found: 268.06385

(+)-1,7b-diphenyl-1,7b-dihydroazirino[1,2-b]benzo[d]isothiazole 3,3-dioxide(2c)



Benzyl dimethylsulfonium bromide (2.25 eq., 112.0 mg, 0.48 mmol was combined with Potassium carbonate (2.50., 74.0 mg, 0.53 mmol) in an oven-dried schlenk tube with a magnetic stirring bar and solved in 2.28 ml DMF. After 10 min. Ketimine (1.0 eq., 52.0 mg, 0.21 mmol mg) was added to the mixture. The reaction was stirred at ambient temperature overnight. After completion of conversion (TLC analysis), the reaction was quenched by adding saturated aqueous ammonium chloride solution (4 ml). 2 ml Ethyl Acetate were added and layers were separated. The aqueous layer was extracted twice more with 2 ml each. Combined organic layers were washed with water three times (10 ml each) and brine once. Concentration delivered crude material, which was objected to silica gel column chromatography (3:2 Ethyl acetate / Cyclohexane (v/v)) to deliver 67 mg (0.20 mmol, 94%) of aziridine.

#### Syn-2c



<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 7.66 – 7.61 (m, 2H), 7.61 – 7.56 (m, 1H), 7.52 – 7.41 (m, 6H), 7.20 – 7.09 (m, 5H), 4.65 (s, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 137.3, 136.7, 135.1, 133.2, 131.0, 130.4, 129.5, 129.1, 128.5, 128.5, 128.0, 127.9, 127.1, 122.2, 60.7, 60.2.

HMRS (ESI): Calculated for C<sub>20</sub> H<sub>16</sub> O<sub>2</sub> N S = [M+H]<sup>+</sup>: 334.08963, found: 334.09003





<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*) δ 7.83 – 7.80 (m, 1H), 7.59 – 7.56 (m, 2H), 7.36 – 7.33 (m, 1H), 7.31 – 7.27 (m, 5H), 7.23 – 7.16 (m, 3H), 7.12 (d, *J* = 7.7 Hz, 2H), 3.87 (s, 1H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 140.6, 133.6, 133.2, 132.6, 130.9, 130.3, 129.3, 129.0, 128.8, 128.7, 128.3, 127.8, 125.6, 123.5, 62.4, 62.3.

**HMRS (ESI):** Calculated for  $C_{20}$  H<sub>16</sub> O<sub>2</sub> N S = [M+H]<sup>+</sup>: 334.08963, found: 334.09043



(<u>+</u>)-Ethyl 7b-phenyl-1,7b-dihydroazirino[1,2-b]benzo[d]isothiazole-1-carboxylate 3,3dioxide (2d)



To (Carbethoxymethyl)dimethylsulfonium bromide (121 mg, 0.53 mmol, 1.35 eq.) was added sodium hydride (24 mg ,60 wt.% in paraffin oil, 1.55 eq.) in 3.8 ml *N*,*N*-DMF. The mixture was stirred for 5 min at ambient temperature and 3-(phenyl)- 1,2 benzothiazole 1,1-dioxide **1-Ph** (1.0 eq., 95 mg, 0.39 mmol) was added in one portion. The reaction was stirred for 10 h at ambient temperature. The reaction mixture was poured into 50 ml of saturated aqueous ammonium chloride solution and extracted with 10 ml Ethyl acetate (3 times). Combined organic layers were washed with water and brine (each 15 ml), dried over anhydrous sodium sulfate and concentrated. Objection to silica gel column chromatography (Ethyl Acetate/Cyclohexane 15-25%) furnished 32 mg (0.10 mmol, 25%) of desired aziridine.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.80 – 7.72 (m, 1H), 7.61 – 7.58 (m, 2H), 7.53 (dq, *J* = 5.1, 2.9 Hz, 2H), 7.46 – 7.40 (m, 4H), 4.10 – 3.96 (m, 2H), 3.50 (s, 1H), 0.96 (t, *J* = 7.2 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 163.4, 138.4, 133.8, 132.7, 130.8, 130., 129.5, 129.0, 128.1, 126.0, 123.3, 62.1, 58.7, 56.6, 13.7.

HMRS (ESI): Calculated for C<sub>17</sub> H<sub>16</sub> O<sub>4</sub> N S = [M+H]<sup>+</sup>: 330.07946, found: 330.07993

(+)-Diethyl azirino[1,2-b]benzo[d]isothiazole-1,7b(1H)-dicarboxylate 3,3-dioxide(2e)



(Carbethoxymethyl)dimethylsulfonium bromide (2.0 eq., 105 mg, 0.46 mmol) and Potassium carbonate (3.0 eq., 95.3 mg, 0.69 mmol) were combined in dry DMSO (2.3 ml) in a flame-dried 10 ml schlenk tubed equipped with a magnetic stirring bar under an Argon atmosphere. The mixture was stirred for 10 min and Ketimine **1-CO<sub>2</sub>Et** (1.0 eq., 55 mg, 230.0  $\mu$ mol) was added as a solid. The reaction was then monitored via TLC.

After 3 h, starting material had been consumed and the reaction mixture was quenched with 2 ml saturated aqueous ammonium chloride solution and the resulting mixture was poured into 40 ml water in a separation funnel.

The mixture was extracted with Ethyl acetate (5 times with 9 ml each). Combined organic layers were washed with water and brine and dried over anhydrous sodium sulfate. Concentration delivered crude product. Objection to silica gel column chromatography (16 to 30% EA/CyH) furnished 28 mg aziridine (0.09 mmol, 37% yield).

 $R_f = 0.31 (30\% EA/CyH)$ ; <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.93 – 7.89 (m, 1H), 7.73 – 7.67 (m, 2H), 7.66 – 7.60 (m, 1H), 4.33 (qd, J = 7.1, 5.9 Hz, 2H), 4.24 (qd, J = 7.2, 0.6 Hz, 2H), 3.26 (s, 1H), 1.32 (t, J = 7.1 Hz, 3H), 1.28 (t, J = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 162.8, 162.7, 134.1, 132.9, 132.8, 131.6, 126.4, 123.4, 63.3, 62.8, 55.3, 53.8, 13.9, 13.9.

**HMRS (ESI):** Calculated for  $C_{14} H_{16} O_6 N S = [M+H]^+$ : 326.06928, found: 326.06938 Calculated for  $C_{14} H_{15} O_6 N Na S = [M+Na]^+$ : 348.05123, found: 348.05129

Ethyl 1-phenylazirino[1,2-b]benzo[d]isothiazole-7b(1H)-carboxylate 3,3-dioxide (2f)



To 3-carbethoxy 1,2 benzothiazole 1,1-dioxide **1-CO<sub>2</sub>Et** (1.0 eq., 24 mg, 0.10 mmol) in 0.8 ml Acetonitrile was added 2.1 eq. (benzyl)dimethylsulfonium bromide (49.1 mg, 0.21 mmol) and Potassium carbonate (2.05 eq., 28.4 mg, 0.21 mmol). The reaction was stirred for 14 h at ambient temperature.

The reaction mixture was poured into 10 ml of saturated aqueous ammonium chloride solution and extracted with 5 ml Ethyl acetate (3 times). Combined organic layers were washed with water and brine (each 15 ml), dried over anhydrous sodium sulfate and concentrated. Objection to silica gel column chromatography (11% to 29% EA/CyH) furnished 18 mg (0.05 mmol, 54%) aziridine.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  8.17 (dt, *J* = 7.9, 0.9 Hz, 1H), 7.78 (dt, *J* = 7.7, 0.9 Hz, 1H), 7.73 (td, *J* = 7.7, 1.2 Hz, 10H), 7.65 (td, *J* = 7.6, 1.1 Hz, 1H), 7.51 – 7.48 (m, 2H), 7.39 – 7.35 (m, 3H), 4.16 – 4.01 (m, 2H), 3.80 (s, 1H), 1.02 (t, *J* = 7.1 Hz, 3H).

 $^{13}\textbf{C}$  NMR (126 MHz, Chloroform-d)  $\delta$  163.4, 134.7, 133.8, 133.2, 131.0, 130.9, 129.3, 128.4, 127.6, 126.5, 123.4, 62.5, 61.7, 57.7, 13.8.

HMRS (ESI): Calculated for C<sub>17</sub> H<sub>16</sub> O<sub>4</sub> N S = [M+H]<sup>+</sup>: 330.07946, found: 330.07982

(<u>+</u>)-Ethyl 7b-(pyridin-2-yl)-1,7b-dihydroazirino[1,2-b]benzo[d]isothiazole-1-carboxylate 3,3-dioxide (2g)



(Carbethoxymethyl)dimethylsulfonium bromide (1.37 eq., 108 mg, 0.47 mmol), Potassium carbonate (2.0 eq., 95 mg ,0.69 mmol) and 3-methylbenzo[*d*]isothiazole 1,1-dioxide **1-Me** (1.0 eq., 84.0 mg, 0.34 mmol) were combined in Acetonitrile (1.275 ml). The reaction was concentrated and objected to silica gel column chromatography (18% to 36% EA/CyH) to yield 103 mg (0.31 mmol, 91%) of product aziridine.

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  8.63 (ddd, J = 4.9, 1.7, 1.0 Hz, 1H), 8.04 – 7.98 (m, 1H), 7.77 – 7.67 (m, 3H), 7.59 (pd, J = 7.4, 1.4 Hz, 1H), 7.31 (ddd, J = 7.2, 4.9, 1.6 Hz, 1H), 4.10 – 3.92 (m, 1H), 3.48 (s, 1H), 1.00 (t, J = 7.1 Hz, 2H).

<sup>13</sup>**C NMR** (101 MHz, Chloroform-d) δ 163.2, 151.7, 149.5, 137.8, 137.3, 134.0, 133.0, 131.1, 127.3, 124.2, 124.1, 123.3, 62.3, 56.8, 27.1, 14.0.

HMRS (ESI): Calculated for C<sub>16</sub> H<sub>15</sub> O<sub>4</sub> N<sub>2</sub> S = [M+H]<sup>+</sup>: 331.07470, found: 331.07463

### (<u>+</u>)-1-Phenyl-7b-(pyridin-2-yl)-1,7b-dihydroazirino[1,2-*b*]benzo[*d*]isothiazole 3,3-dioxide (2h)

Benzyldimethylsulfonium bromide (1.4 eq., 100.0 mg, 0.43 mmol), Potassium carbonate (2.0 eq., 32.8 mg, 0.24 mmol) and 2-Pyridinyl Ketimine **2-Pyr-1** (1.0 eq., 29.0 mg, 0.12 mmol) were combined in Acetonitrile (970  $\mu$ l). The reaction was stirred for 10 hours at ambient temperature. Then, additional 2.2 eq (157 mg, 0.68 mmol) of sulfonium salt were added to the mixture and the reaction stirred for 30 additional hours at ambient temperature. The reaction was stopped by concentration using a rotary evaporator and objected to silica gel column chromatography to yield 29 mg (0.09 mmol, 73%) of a 1:1 mixture of diastereomers.



#### *anti*-2h

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 8.53 – 8.50 (m, 1H), 8.15 (d, *J* = 7.7 Hz, 1H), 7.81 – 7.77 (m, 1H), 7.69 – 7.55 (m, 5H), 7.23 (dd, *J* = 6.8, 3.0 Hz, 2H), 7.16 (tt, *J* = 4.8, 2.7 Hz, 5H), 3.98 (s, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 151.5, 149.5, 138.7, 136.7, 133.5, 133.1, 131.9, 130.3, 128.5, 128.2, 127.4, 126.7, 123.8, 123.7, 123.2, 62.9, 61.3.

**HMRS (ESI):** Calculated for  $C_{19}$  H<sub>15</sub> O<sub>2</sub> N<sub>2</sub> S = [M+H]<sup>+</sup>: 335.08487 found: 335.08570 Calculated for  $C_{19}$  H<sub>15</sub> O<sub>2</sub> N<sub>2</sub> Na S = [M+Na]<sup>+</sup>: 357.06682 found: 357.06777



*syn*-2h



<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  8.74 (dt, *J* = 5.0, 1.3 Hz, 1H), 8.12 (d, *J* = 7.9 Hz, 1H), 7.79 (dd, *J* = 7.6, 1.8 Hz, 1H), 7.76 – 7.72 (m, 1H), 7.64 (td, *J* = 4.9, 2.6 Hz, 1H), 7.38 (ddd, *J* = 7.6, 4.9, 1.3 Hz, 1H), 7.20 – 7.07 (m, 5H), 4.70 (s, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 154.8, 149.8, 137.6, 137.1, 135.7, 133.3, 131.1, 130.5, 128.6, 128.1, 127.9, 124.1, 122.6, 122.3, 61.9, 59.4.

**HMRS (ESI):** Calculated for  $C_{19} H_{15} O_2 N_2 S = [M+H]^+$ : 335.08487, found: 335.08470



#### 2.1.3 Synthesis of Azetidines



Azetidines **4a** and **4b** were synthesized according to the procedure of Ye *et al.*<sup>[10]</sup> and results were in accordance with the reported yields and spectral data. For azetidines **4c-e**, same conditions with corresponding substrates and allenes were used.

(<u>+</u>)-tert-butyl (*E*)-2-(4,4-dioxido-8b-phenyl-1,8b-dihydro-2H-azeto[1,2-b]benzo[d]isothiazol-2-ylidene)acetate (4b)

Yield: 742 mg, 52% (3.7 mmol scale)

<sup>1</sup>**H NMR** (400 MHz, Chloroform-d)  $\delta$  7.80 (ddd, *J* = 7.8, 1.3, 0.7 Hz, 1H), 7.66 – 7.61 (m, 1H), 7.60 – 7.54 (m, 3H), 7.46 – 7.38 (m, 3H), 7.37 – 7.31 (m, 1H), 5.85 (t, *J* = 2.4 Hz, 1H), 4.01 (dd, *J* = 17.0, 2.4 Hz, 1H), 3.73 (dd, *J* = 17.0, 2.4 Hz, 1H), 1.42 (s, 9H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 165.7, 154.9, 143.3, 139.4, 136.5, 134.3, 130.2, 129.1, 128.8, 125.8, 125.0, 122.4, 106.8, 80.7, 77.1, 43.3, 28.4.

**HMRS (ESI):** Calculated for  $C_{21} H_{22} O_4 N S = [M+H]^+$ : = 384.12641, found: 384.12632 Calculated for  $C_{21} H_{22} O_4 N Na S = [M+Na]^+$ : = 406.10835, found: 406.10802 Calculated for  $C_{21} H_{22} O_4 N K S = [M+K]^+$ : = 422.08229, found: 422.08188

(<u>+</u>)-Ethyl (*E*)-2-(4,4-dioxido-8b-(pyridin-2-yl)-1,8b-dihydro-2H-azeto[1,2-b]benzo[d]isothiazol-2-ylidene)acetate (4c)<sup>[10]</sup>



<sup>1</sup>**H NMR** (400 MHz, Chloroform-d) δ 8.66 (ddd, *J* = 4.8, 1.7, 1.0 Hz, 1H), 7.87 (dt, *J* = 7.8, 0.9 Hz, 1H), 7.78 – 7.75 (m, 1H), 7.75 – 7.72 (m, 1H), 7.72 – 7.69 (m, 1H), 7.66 (ddd, *J* = 7.8, 4.7, 1.1 Hz, 1H), 7.60 – 7.55 (m, 1H), 7.26 (ddd, *J* = 7.2, 4.9, 1.5 Hz, 1H), 5.96 (t, *J* = 2.3 Hz,

1H), 4.09 (ddd, *J* = 17.7, 8.9, 2.9 Hz, 3H), 3.74 (dd, *J* = 17.4, 2.4 Hz, 1H), 1.23 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>C NMR (126 MHz, Chloroform-d) δ 166.0, 157.6, 156.1, 149.5, 141.6, 137.4, 136.2, 134.3, 130.3, 125.9, 123.3, 121.8, 120.3, 105.1, 76.0, 60.2, 42.9, 14.3.

**HMRS (ESI):** Calculated for  $C_{18} H_{17} O_4 N_2 S = [M+H]^+$ : = 357.09035, found: 357.09011

(<u>+</u>)-ethyl (*E*)-2-(8b-methyl-4,4-dioxido-1,8b-dihydro-2H-azeto[1,2-b]benzo[d]isothiazol-2-ylidene)acetate (4d)



Yield: 28% (0.12 mmol scale, 10 mg)

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 7.76 (d, *J* = 7.9 Hz, 1H), 7.72 – 7.67 (m, 1H), 7.58 (t, *J* = 7.6 Hz, 0H), 7.44 (d, *J* = 7.8 Hz, 1H), 5.87 (t, *J* = 2.3 Hz, 1H), 4.09 (q, *J* = 7.1 Hz, 2H), 3.53 (dd, *J* = 17.1, 2.3 Hz, 1H), 3.35 (dd, *J* = 17.1, 2.3 Hz, 1H), 1.90 (s, 3H), 1.22 (t, *J* = 7.1 Hz, 4H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 166.4, 156.0, 143.9, 137.2, 134.3, 130.1, 123.8, 122.3, 104.4, 73.6, 60.2, 42.6, 25.7, 14.4

HMRS (ESI): Calculated for C<sub>14</sub> H<sub>16</sub> O<sub>4</sub> N S = [M+H]<sup>+</sup>: = 294.07946, found: 294.07974

(<u>+</u>)-Ethyl (*E*)-2-(2-(benzyloxy)-2-oxoethylidene)-1,2-dihydro-8bH-azeto[1,2-b]benzo[d]isothiazole-8b-carboxylate 4,4-dioxide (4e)



Yield: 90% (0.34 mmol scale, 140 mg)

<sup>1</sup>**H NMR** (500 MHz, Chloroform-d)  $\delta$  7.80 (dt, *J* = 7.8, 0.9 Hz, 1H), 7.77 – 7.62 (m, 3H), 7.35 – 7.30 (m, 5H), 5.96 (t, *J* = 2.3 Hz, 1H), 5.09 (d, *J* = 2.0 Hz, 2H), 4.36 – 4.23 (m, 2H), 4.02 – 3.95 (m, 1H), 3.50 (dd, *J* = 17.3, 2.3 Hz, 1H), 1.31 (t, *J* = 7.2 Hz, 9H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 167.5, 165.8, 155.5, 137.2, 137.2, 135.8, 134.5, 131.2, 128.7, 128.4, 128.4, 125.3, 122.4, 104.9, 72.4, 66.3, 63.4, 40.9, 14.1.

**HMRS (ESI):** Calculated for  $C_{21}$  H<sub>20</sub> O<sub>6</sub> N S = [M+H]<sup>+</sup>: = 414.10058, found: 414.09967

Calculated for  $C_{21}$  H<sub>19</sub> O<sub>6</sub> N Na S = [M+Na]<sup>+</sup>: = 436.08253, found: 436.08153

Calculated for  $C_{21}$  H<sub>19</sub> O<sub>6</sub> N K S = [M+K]<sup>+</sup>: = 452.05647, found: 452.05550

(<u>+</u>)-Ethyl (*E*)-2-(2-(tert-butoxy)-2-oxoethylidene)-1,2-dihydro-8bH-azeto[1,2-b]benzo[d]isothiazole-8b-carboxylate 4,4-dioxide (4f)



Yield: 85% (70 µmol scale, 27 mg)

<sup>1</sup>**H NMR** (500 MHz, Chloroform-d) δ 7.81 (d, *J* = 7.8 Hz, 1H), 7.77 – 7.64 (m, 3H), 5.82 (t, *J* = 2.2 Hz, 1H), 4.37 – 4.23 (m, 2H), 3.96 (dd, *J* = 17.2, 2.2 Hz, 1H), 3.46 (dd, *J* = 17.2, 2.2 Hz, 1H), 1.41 (s, 9H), 1.31 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 167.6, 165.1, 153.6, 137.3, 137.2, 134.3, 131.1, 125.1, 122.3, 107.1, 80.8, 72.4, 63.2, 40.7, 28.2, 14.0.

HMRS (ESI): Calculated for  $C_{14}$  H<sub>13</sub> O<sub>6</sub> N S = [M-<sup>t</sup>Bu+H]<sup>+</sup>: = 324.05636, found: 324.05687 2.1.4 Synthesis of fused Pyrrolines by Phosphine Catalysis



Pyrroline 5a was synthesized according to the procedure reported by Chen et al.[10]

#### (<u>+</u>)-1-Benzyl 9b-ethyl benzo[d]pyrrolo[1,2-b]isothiazole-1,9b(3H)-dicarboxylate 5,5dioxide(5b)



Ketimine substrate **1-CO<sub>2</sub>Et** (1.0 eq., 90 mg, 0.38 mmol) and PPh<sub>3</sub> (0.35 eq., 34.5 mg, 0.13 mmol) were combined in dry Toluene (6 ml) in an oven-dried Schlenk tube under an Argon atmosphere. Benzyloxyallene (1.70 eq., 93.5 mg, 0.64 mmol) was added and the reaction was

stirred at same temperature and monitored by TLC. After completion of conversion (5 h), the reaction mixture was concentrated under reduced pressure.

Crude reaction mixtures was subjected to silica gel column chromatography (12% to 16% EA/CyH) to give [3+2] product in 82% yield (0.31 mmol, 128 mg).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 8.03 – 7.93 (m, 1H), 7.81 – 7.74 (m, 1H), 7.64 – 7.53 (m, 2H), 7.43 – 7.31 (m, 5H), 7.01 (t, *J* = 2.3 Hz, 1H), 5.29 (d, *J* = 12.2 Hz, 1H), 5.24 (d, *J* = 12.2 Hz, 1H), 4.87 (d, *J* = 18.2 Hz, 1H), 4.38 (d, *J* = 18.2 Hz, 1H), 4.21 (qd, *J* = 7.1, 4.2 Hz, 2H), 1.18 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 167.9, 161.8, 141.7, 136.5, 135.2, 135.1, 133.6, 133.4, 130.6, 128.8, 128.8, 128.5, 127.5, 121.4, 80.9, 67.3, 63.0, 54.8, 13.9. **HMRS (ESI):** Calculated for  $C_{21}$  H<sub>20</sub> O<sub>6</sub> N S = [M+H]<sup>+</sup>: = 414.10058, found: 414.10013 Calculated for  $C_{21}$  H<sub>19</sub> O<sub>6</sub> N Na S = [M+H]<sup>+</sup>: = 436.08253, found: 436.08184 Calculated for  $C_{21}$  H<sub>19</sub> O<sub>6</sub> N K S = [M+H]<sup>+</sup>: = 452.05593, found: 452.05647

#### 2.1.5 Diels-Alder reaction of Ketimines with Danishefsky's diene



#### General procedure:

To a solution of Ketimine **1** (1.0 eq) in Toluene or *N*,*N*-DMF in a 35 ml microwave tube equipped with a magnetic stirring bar, Danishefsky's diene was added, the vessel was flushed with Argon, closed and heated in a microwave at corresponding maximum temperature at 250 W for 45 min. After reaction, 8 ml of saturated aqueous NH<sub>4</sub>Cl solution was added and the mixture was extracted with Ethyl acetate (three times 5 ml). The combined organic layers were washed with brine, dried over anhydrous sodium sulfate and concentrated using a rotary evaporator. The crude material was objected to silica gel flash chromatography.

#### (+)-10,10a-dihydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5-dioxide (6a)



#### **Conditions:**

Ketimine (1.0 eq., 148 mg, 0.89 mmol) in 3 ml *Toluene* (to give 0.30 M solution) Danishefsky's diene (1.4 eq., 240 µl, 1.24 mmol)

Conditions: 140 °C (250 W) for 30 min

Silica gel flash chromatography (EA/ CyH 18%-26%) to furnish 200 mg of Sulfonamide (96%, 0.85 mmol).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.89 (dt, *J* = 7.9, 0.9 Hz, 1H), 7.76 (td, *J* = 7.6, 1.1 Hz, 1H), 7.67 (d, *J* = 7.8 Hz, 1H), 7.64 (d, J = 7.9 Hz, 1H), 7.47 (d, *J* = 7.8 Hz, 1H), 5.58 (dd, *J* = 8.0, 1.1 Hz, 1H), 5.35 (dd, *J* = 15.2, 4.3 Hz, 1H), 3.06 (ddd, *J* = 16.0, 4.3, 1.1 Hz, 1H), 2.72 (dd, *J* = 16.0, 15.2 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 190.8, 138.1, 134.8, 134.4, 134.2, 130.6, 123.7, 122.1, 108.4, 57.3, 41.1.

**HMRS (ESI)**: Calculated for  $C_{11} H_{10} O_3 N S = [M+H]^+$ : = 236.03759 found: 236.03744

#### (<u>+</u>)-10a-phenyl-10,10a-dihydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5dioxide (6b)



#### **Conditions:**

Ketimine (1.0 eq., 400 mg, 1.64 mmol) in 3.5 ml Toluene (to give a 0.45 M solution),

Danishefsky's diene (1.25 eq., 398  $\mu l,$  2.06 mmol),

Conditions: 180 °C (250 W) for 45 min,

Silica gel flash chromatography (EA/ CyH 20%-34%) to furnish 315 mg of sulfonamide (62%, 1.01 mmol).

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.91 – 7.88 (m, 1H), 7.66 (td, *J* = 7.6, 1.4 Hz, 1H), 7.61 (td, *J* = 7.6, 1.2 Hz, 1H), 7.56 (d, *J* = 7.8 Hz, 1H), 7.52 – 7.48 (m, 3H), 7.39 – 7.28 (m, 7H), 5.53 (dd, *J* = 7.8, 1.1 Hz, 2H), 3.66 (dd, *J* = 16.1, 1.1 Hz, 1H), 3.05 (d, *J* = 16.1 Hz, 1H).

<sup>13</sup>**C NMR** (101 MHz, Chloroform-d) δ 190.4, 140.5, 137.9, 136.8, 134.8, 132.2, 130.4, 129.5, 129.3, 126.3, 124.2, 122.1, 109.9, 69.3, 46.4.

**HMRS (ESI): Calculated for**  $C_{17} H_{14} O_3 N S = [M+H]^+$ : = 312.06889, found: 312.06872

(<u>+</u>)-10a-(pyridin-2-yl)-10,10a-dihydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5dioxide (6c)



#### **Conditions:**

Ketimine (1.0 eq., 240 mg, 0.98 mmol) in 4 ml Toluene:N,N DMF 7:1 (v/v, as Ketimine is not soluble in neat toluene, to give 0.25 M solution)

Danishefsky's diene (1.5 eq., 285 µl, 1.47 mmol)

Conditions: 140 °C (250 W) for 45 min

Silica gel flash chromatography (EA/ CyH 20%-32%) to furnish 238 mg of Sulfonamide (78%, 0.76 mmol).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  8.62 (d, *J* = 4.8 Hz, 1H), 7.85 (d, *J* = 7.8 Hz, 1H), 7.75 (d, *J* = 7.9 Hz, 1H), 7.70 - 7.58 (m, 4H), 7.53 (d, *J* = 8.1 Hz, 1H), 5.52 (dd, *J* = 7.9, 1.0 Hz, 1H), 4.23 (dd, *J* = 15.7, 1.1 Hz, 1H), 2.93 (d, *J* = 15.7 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 190.6, 156.6, 150.0, 139.0, 137.7, 136.5, 134.7, 132.1, 130.5, 125.0, 123.6, 121.7, 119.8, 109.7, 69.8, 46.1.

**HMRS (ESI)**: Calculated for  $C_{17} H_{16} O_3 N_2 S = [M+H]^+$ : = 313.06414 found: 316.06401

(<u>+</u>)-10a-ethyl-10,10a-dihydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5-dioxide--carbon dioxide (6d)



#### **Conditions:**

Ketimine **1-CO<sub>2</sub>Et** (1.0 eq., 248 mg, 1.00 mmol) in 3.4 ml Toluene (0.29 M), Danishefsky's diene (1.4 eq., 271  $\mu$ l, 1.40 mmol), Conditions: 140 °C (250 W) for 45 min, Silica gel flash chromatography (EA/ CyH 20%-34%) to furnish 290 mg of desired sulfonamide (94%, 0.94 mmol)

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.89 (d, *J* = 7.6 Hz, 1H), 7.77 (t, *J* = 7.6 Hz, 1H), 7.74 – 7.62 (m, 3H), 5.58 (d, *J* = 8.0 Hz, 1H), 4.28 – 4.12 (m, 2H), 3.61 (d, *J* = 16.1 Hz, 1H), 2.83 (d, *J* = 16.1 Hz, 1H), 1.21 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 194.8, 169.8, 164.2, 137.1, 135.7, 133.7, 130.8, 124.3, 122.0, 104.9, 65.6, 63.5, 58.0, 14.1.

**HMRS (ESI)**: Calculated for  $C_{14} H_{14} O_5 N S = [M+H]^+$ : = 308.05872, found: 308.05897

### (<u>+</u>)-10a-methyl-10,10a-dihydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5-dioxide (6e)



#### **Conditions:**

Ketimine (1.0 eq., 200.0 mg, 1.10 mmol) in 5 ml *Toluene* (to give 0.22 M solution) Danishefsky's diene (1.25 eq., 267 µl, 1.38 mmol)

Conditions: 140 °C (250 W) for 30 min

Silica gel flash chromatography (EA/ CyH 22%-34%) to furnish 19 mg of Sulfonamide (7% yield, 0.08 mmol)

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*)  $\delta$  7.87 (d, *J* = 7.8 Hz, 1H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.65 (t, *J* = 7.6 Hz, 1H), 7.57 (d, *J* = 7.9 Hz, 1H), 7.43 (d, *J* = 7.8 Hz, 1H), 5.58 (d, *J* = 7.9 Hz, 1H), 2.88 (d, *J* = 15.7 Hz, 1H), 2.81 (d, *J* = 15.7 Hz, 1H), 1.71 (s, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 190.7, 141.3, 136.7, 134.6, 133.2, 130.4, 130.3, 122.8, 122.1, 106.9, 64.1, 47.3, 24.3.

**HMRS (ESI)**: Calculated for  $C_{12}H_{12}O_3 N S = [M+H]^+$ : = 250.05324 found: 250.05298

#### 2.1.6 [3+2] Cycloaddition of Ketimines with Azomethine ylides



#### General procedure for synthesis of fused imidazolidines 8a-e

*N*-(Methoxymethyl)-*N*-(trimethylsilylmethyl)benzylamine (82.0  $\mu$ l, 0.30 mmol) and corresponding imine (0.10 mmol) were combined in CH<sub>2</sub>Cl<sub>2</sub> (0.7 mL) in an oven-dried Schlenk tube and a mixture of TFA and CH<sub>2</sub>Cl<sub>2</sub> (10% v/v in 36  $\mu$ l DCM, 0.5 eq.) was added *via* syringe. The resultant mixture was stirred at 0° C (ice-water bath) for 1 h. The crude reaction mixture was purified by flash chromatography on silica gel (EtOAc/Petroleum Ether) to afford corresponding tricyclic sulfonamides.

#### (+)-2-benzyl-1,2,3,9b-tetrahydrobenzo[d]imidazo[1,5-b]isothiazole 5,5-dioxide (8a)



Yield: 37%, 11 mg, 0.04 mmol

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.78 (d, *J* = 7.6 Hz, 1H), 7.63 (td, *J* = 7.6, 1.1 Hz, 1H), 7.57 – 7.52 (m, 1H), 7.33 (dd, *J* = 7.7, 1.0 Hz, 1H), 7.30 – 7.22 (m, 4H), 7.18 (d, *J* = 7.2 Hz, 1H), 5.07 (dd, *J* = 7.0, 2.8 Hz, 1H), 4.74 (d, *J* = 8.6 Hz, 1H), 3.87 (d, *J* = 8.6 Hz, 1H), 3.60 (d, *J* = 13.2 Hz, 1H), 3.52 (d, *J* = 13.2 Hz, 1H), 3.22 (dd, *J* = 10.5, 2.8 Hz, 1H), 3.17 (dd, *J* = 10.5, 7.0 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 139.9, 137.3, 136.2, 133.4, 129.6, 128.5, 128.5, 127.5, 123.8, 121.6, 71.0, 63.2, 58.0, 57.2.

**HMRS (ESI)**: Calculated for  $C_{16} H_{17} O_2 N_2 S = [M+H]^+$ : = 301.10053, found: 301.10053

(<u>+</u>)-2-benzyl-9b-phenyl-1,2,3,9b-tetrahydrobenzo[d]imidazo[1,5-b]isothiazole 5,5dioxide (8b)



Yield: 83%, 73 mg, 0.19 mmol

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 7.74 (d, *J* = 7.7 Hz 1H), 7.66 (d, *J* = 7.9 Hz, 2H), 7.56 (t, *J* = 8.0 Hz, 1H), 7.49 (d, *J* = 7.4 Hz, 1H), 7.45 (d, *J* = 8.5, 1H), 7.36 (t, *J* = 7.3 Hz, 2H), 7.32 – 7.22 (m, 4H), 7.14 (d, *J* = 6.8 Hz, 2H), 4.98 (d, *J* = 9.0 Hz, 1H), 3.92 (d, *J* = 8.9 Hz, 1H), 3.84 (d, *J* = 10.1 Hz, 1H), 3.59 (d, *J* = 13.2 Hz, 2H), 3.50 (d, *J* = 13.2 Hz, 2H), 3.30 (d, *J* = 10.2 Hz, 2H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 144.0, 142.7, 137.7, 135.4, 133.8, 129.7, 129.0, 128.7, 128.6, 128.2, 127.7, 125.9, 124.3, 121.3, 77.4, 71.6, 66.4, 56.7

**HMRS (ESI): Calculated for**  $C_{22}$   $H_{21}$   $O_2$   $N_2$   $S = [M+H]^+$ : = 377.13183, found: 377.13178

(<u>+</u>)-2-benzyl-9b-(pyridin-2-yl)-1,2,3,9b-tetrahydrobenzo[d]imidazo[1,5-b]isothiazole 5,5dioxide (8c)



Yield: 67%, 28 mg, 74 µmol

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  8.57 (ddd, J = 4.9, 1.8, 0.9 Hz, 1H), 7.95 (dt, J = 8.0, 1.1 Hz, 1H), 7.85 (dt, J = 7.9, 0.9 Hz, 1H), 7.72 (dt, J = 7.7, 0.9 Hz, 1H), 7.65 (td, J = 7.8, 1.8 Hz, 1H), 7.57 (td, J = 7.6, 1.2 Hz, 1H), 7.50 (td, J = 7.5, 1.1 Hz, 1H), 7.28 – 7.21 (m, 3H), 7.18 (dd, J = 7.5, 4.8 Hz, 1H), 7.14 – 7.09 (m, 2H), 4.96 (dd, J = 9.0, 1.3 Hz, 1H), 3.97 (d, J = 9.0 Hz, 1H), 3.88 (dd, J = 11.0, 1.3 Hz, 1H), 3.58 (d, J = 13.3 Hz, 1H), 3.46 (d, J = 13.3 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 161.0, 149.5, 142.8, 137.5, 137.4, 134.7, 133.6, 129.7, 128.7, 128.5, 127.5, 125.3, 122.8, 121.1, 120.2, 77.4, 72.0, 65.7, 57.1.

**HMRS (ESI)**: Calculated for  $C_{21} H_{20} O_2 N_3 S = [M+H]^+$ : = 378.12707, found: 378.12685

#### (<u>+</u>)-2-benzyl-9b-methyl-1,2,3,9b-tetrahydrobenzo[d]imidazo[1,5-b]isothiazole 5,5dioxide (8d)



Yield: 48%, 18 mg, 0.05 mmol

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.74 – 7.70 (m, 1H), 7.63 – 7.51 (m, 3H), 7.24 – 7.16 (m, 3H), 7.08 – 7.04 (m, 2H), 4.76 (dd, *J* = 9.1, 0.5 Hz, 1H), 4.18 (tq, *J* = 7.1, 3.6 Hz, 2H), 4.01 (d, *J* = 9.1 Hz, 1H), 3.52 (d, *J* = 13.1 Hz, 1H), 3.41 (d, *J* = 0.7 Hz, 2H), 3.39 (s, 1H), 1.20 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 169.7, 138.2, 137.1, 135.7, 133.7, 130.6, 128.6, 128.6, 127.7, 124.7, 121.7, 75.2, 72.1, 63.0, 62.7, 57.2, 14.0.

**HMRS (ESI)**: Calculated for  $C_{19} H_{21} O_4 N_2 S = [M+H]^+$ : = 373.12165, found: 373.12118 **Calculated for**  $C_{19} H_{20} O_4 N_2 Na S = [M+H]^+$ : = 395.10360, found: 395.10346

(<u>+</u>)-2-benzyl-9b-methyl-1,2,3,9b-tetrahydrobenzo[d]imidazo[1,5-b]isothiazole 5,5dioxide (8e)



Yield: 73%, 23 mg, 0.07 mmol

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.75 (dt, *J* = 7.8, 0.9 Hz, 1H), 7.62 (td, *J* = 7.5, 1.2 Hz, 1H), 7.52 (td, *J* = 7.6, 1.0 Hz, 1H), 7.31 (dt, *J* = 7.8, 0.9 Hz, 1H), 7.27 – 7.21 (m, 3H), 7.11 (dd, *J* = 7.9, 1.7 Hz, 2H), 4.75 (dd, *J* = 8.6, 1.1 Hz, 1H), 3.83 (d, *J* = 8.6 Hz, 1H), 3.57 – 3.44 (m, 2H), 3.38 (dd, *J* = 10.0, 1.2 Hz, 1H), 2.78 (d, *J* = 10.0 Hz, 1H), 1.71 (s, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 145.0, 137.3, 136.0, 133.6, 129.3, 128.5, 128.5, 127.5, 122.9, 121.4, 72.1, 71.1. 65.3, 56.7, 27.3.

**HMRS (ESI)**: Calculated for  $C_{17} H_{19} O_2 N_2 S = [M+H]^+$ : = 315.11618, found: 315.11605

#### 2.1.7 Reaction of Pyruvate derived α-silyl imines with Ketimines



#### (<u>+</u>)-diethyl-1-methyl-2,3-dihydrobenzo[d]imidazo[1,5-b]isothiazole-1,9b(1H)dicarboxylate 5,5-dioxide (10)

#### <u>Condition A (affording ~ 1:1 d.r.)</u>

To a solution of ketimine **1-CO<sub>2</sub>Et** (19 mg, 80 µmol, 1.0 equiv.) in 500 µl dry Et<sub>2</sub>O was added silylimine (365.470 µl, 90 mg per ml in diethyl ether solution, 158.83 µmol, 2.0 equiv.) followed by the addition of AgOAc (30mol%, 4.0 mg, 23.82 µmol) and PPh<sub>3</sub> (30mol%, 6.3 mg, 23.82 µmol). The reaction was stirred at 21° C and monitored *via* TLC for completion. The reaction mixture was then quenched by the addition of brine and transferred to a separation funnel. EA was added and layers were separated. The aqueous layer was extracted twice more with EA. Combined organic layers were washed with water and brine once more each and dried over Na<sub>2</sub>SO<sub>4</sub>. The solvent was removed *in vacuo* and the residue was objected to silica gel column chromatography (25% to 27% to 29% to 32% to 39% EA/CyH, 1 CV each) to elute two products *syn*-10 and *anti*-10 in 38% (11 mg, 0.03 mmol) and 31% (9 mg, 0.02 mmol) yield, respectively.

#### **Optimized Condition B** (affording 100:15 d.r.)

AgNO<sub>3</sub> (20mol%, 9.9 mg, 58.5 µmol) and tri(4-trifluoromethylphenyl)phosphine (20mol% 27 mg, 58.5 µmol, ) were combined in 1.84 ml dry MeCN at 0 °C. Ketimine **1-CO<sub>2</sub>Et** (70 mg, 293 µmol, 1.0 equiv.) and silylimine (2.0 eq., 585.16 µl, 585.16 µmol, 1 mM in diethyl ether) were subsequently added. The reaction was stirred at 0° C for 2 h and monitored *via* TLC for completion. HCl (20 µl) was added to the reaction mixture and stirred for 20 more minutes. The reaction mixture was then quenched by the addition of 50 µl 1 M aq. HCl. The mixture was concentrated and redissolved in Chloroform/Ethanol (9:1) and passed through a short pad of silica gel. The crude material was then analyzed by proton NMR to show a diastereomeric ratio of 100:15 in favor of **syn-10**. Objection to silica gel column chromatography (using gradient 25% to 27% to 29% to 32% to 39% EA/CyH) then furnished 57 mg (154.7 µmol, 53% combined yield) of fused imidazoline.



#### s*yn*-10

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.96 (d, J= 7.8 Hz, 1H), 7.74 (d, J = 7.8 Hz, 1H), 7.69 (td, J = 7.6 Hz, 1H), 7.64 (td, J = 7.6 Hz, 1H), 4.75 (d, J = 8.5 Hz, 1H), 4.59 (d, J = 8.5 Hz, 1H), 4.33 (qd, J = 7.2, 3.2 Hz, 2H), 4.18 (tq, J = 7.1, 3.6 Hz, 2H), 1.40 (t, J = 7.1 Hz, 3H), 1.23 (d, J = 7.1 Hz, 3H), 0.95 (s, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 171.3, 136.6, 133.3, 133.3, 131.1, 127.4, 121.4, 80.4, 72.0, 63.2, 62.8, 62.7, 20.2, 14.2, 14.0.

**HMRS (ESI)**: Calculated for  $C_{16} H_{21} O_6 N_2 S = [M+H]^+$ : = 369.11148, found: 369.11103 Calculated for  $C_{16} H_{20} O_6 N_2 Na S = [M+H]^+$ : = 391.09343, found: 391.09237 Calculated for  $C_{16} H_{20} O_6 N_2 K S = [M+H]^+$ : = 407.06737, found: 407.06662

Xray Deposition number at the Cambridge Crystallographic Data Centre: CCDC 1910465





anti-10

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.79 (d, *J* = 7.9 Hz, 1H), 7.69 (d, *J* = 7.3 Hz, 1H), 7.63 – 7.56 (m, 2H), 5.14 (d, *J* = 9.8 Hz, 1H), 4.43 (d, *J* = 9.8 Hz, 1H), 4.30 (qd, *J* = 7.2, 1.6 Hz, 2H), 3.83 – 3.74 (m, 1H), 3.72 – 3.63 (m, 1H), 1.63 (s, 3H), 1.35 (t, *J* = 7.2 Hz, 3H), 0.97 (t, *J* = 7.2 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 169.4, 168.1, 135.6, 133.2, 132.6, 130.8, 127.4, 120.9, 80.4, 74.7, 65.2, 62.9, 62.1, 20.2, 14.1, 13.5.

**HMRS (ESI)**: Calculated for  $C_{16} H_{21} O_6 N_2 S = [M+H]^+$ : = 369.11148, found: 369.11134 Calculated for  $C_{16} H_{20} O_6 N_2 Na S = [M+H]^+$ : = 391.09343, found: 391.09284 Calculated for  $C_{16} H_{20} O_6 N_2 K S = [M+H]^+$ : = 407.06737, found: 407.06702

**Table S1**: Conditions for optimizations of diastereoselectivity in 3+2 cyclization of *N*-sulfonyl ketimines (1) with  $\alpha$ -silyl imines (9). Diastereoselectivity was determined by integration of peaks in <sup>1</sup>H-NMR spectra of crude reaction mixtures

Condition #	Phosphine	Solvent	T/°C	Scale /	d.r.
				mmol	
1	Triphenylphosphine	DCM	-40 to 21	0.09	n.d.
2	Tri-o-tolylphosphine	Et <sub>2</sub> O	21	0.08	24:10
3	Triphenylphosphine	Et <sub>2</sub> O	21	0.08	19:10
4	Tricyclohexylphosphine	Et <sub>2</sub> O	21	0.08	10:9.2
5	Tri(4-CF₃)-	Et <sub>2</sub> O	21	0.08	22:10
	phenylphosphine				
6	XPhos	Et <sub>2</sub> O	21	0.08	11:10
7	Tri-o-tolylphosphine	MeCN	21	0.08	5:1
8	iPhox	MeCN	21	0.08	100:17
9	(Sa,S)-DTB-Bn-SIPHOX	MeCN	21	0.08	100:23
10	Tri(4-CF₃)-	MeCN	0	0.08	100:16
	phenylphosphine				
11	Tri(4-CF₃)-	MeCN	0	0.3	100:15
	phenylphosphine				



#### 2.2. Vinylogous Addition of Silyl Enol Ether to Ketimines (12a)

The proposed structures were based on evaluation of 1D –NOE NMR experiments (see below).

To 239 mg (1.0 eq, 1.00 mmol) of 3-carbethoxy 1,2 benzothiazole 1,1 dioxide (**1-CO<sub>2</sub>Et**) and AgOAc in dry DCM (3.7 ml) under an Argon atmosphere in an oven-dried Schlenk flask, Silyl Enol Ether (1.6 eq, 312  $\mu$ l, 1.60 mmol) was added at -78°C (Acetone-dry ice bath). The reaction was stirred at same temperature and monitored by TLC. After completion of reaction (10 min), sat. aqueous NaHCO<sub>3</sub> solution was added and stirred for 5 min at 0 °C, then transferred to a separation funnel and 8 ml of Ethyl acetate were added. Layers were separated and the aqueous layer was extracted two more times (8 ml Ethyl Acetate each). The combined organic layers were washed with brine and dried over anhydrous sodium sulfate. After removal of solvent, the residue was purified using silica gel flash column chromatography (22% EA/CyH to 35% EA/CyH) to yield product in 89% yield (300 mg, 0.89 mmol) and 86:14 d.r.

#### (<u>+</u>)Ethyl 3-2-methyl-5-oxo-2,5-dihydrofuran-2-yl)-2,3-dihydrobenzo[d]isothiazole-3carboxylate 1,1-dioxide (*I* or *syn* 12a)

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 7.95 – 7.91 (m, 1H), 7.74 – 7.71 (m, 1H), 7.67 – 7.61 (m, 3H), 6.15 (s, 1H), 5.84 (d, J = 5.7 Hz, 1H), 4.52 – 4.39 (m, 2H), 1.61 (s, 3H), 1.42 (t, J = 7.1 Hz, 3H); <sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 171.8, 167.9, 157.6, 135.8, 133.9, 132.2, 131.6,

127.5, 123.1, 121.7, 89.6, 71.6, 64.9, 20.8, 14.2; **HMRS (ESI)**: Calculated for  $C_{15}H_{16}O_6 N S = [M+H]^+$ : = 338.06928, found: 338.06994

Calculated for  $C_{15} H_{15} O_6 N Na S = [M+H]^+$ : = 360.05123, found: 360.05266

#### I-12a 1D NOE-experiments



Irradiation of methyl group (1.6 ppm) shows NOE with sulfonamide amine N-H (6.15 ppm) and aromatic proton at ~7.6 ppm.





Irradiation of lactone proton in  $\alpha$  carbonyl position (5.8 ppm) shows NOE to aromatic proton at roughly 7.6 ppm. Both observations lead us to conclude that we this is in fact diastereomer in conformation depicted in structure below.

#### (<u>+</u>)-Ethyl 3-2-methyl-5-oxo-2,5-dihydrofuran-2-yl)-2,3-dihydrobenzo[d]isothiazole-3carboxylate 1,1-dioxide (*anti or I-*12a)

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  8.17 (dt, *J* = 8.0, 0.9 Hz, 1H), 7.82 – 7.79 (m, 1H), 7.75 (td, *J* = 7.6, 1.3 Hz, 1H), 7.69 (td, *J* = 7.6, 1.1 Hz, 1H), 7.57 (d, *J* = 5.7 Hz, 1H), 6.20 (d, *J* = 5.6 Hz, 1H), 6.09 (s, 1H), 4.29 – 4.16 (m, 2H), 1.37 (s, 3H), 1.29 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 171.1, 167.7, 157.7, 136.3, 133.9, 132.4, 131.6, 128.7, 123.3, 121.6, 90.4, 72.0, 60.5, 19.6, 13.9.

**HMRS (ESI)**: Calculated for  $C_{15} H_{16} O_6 N S = [M+H]^+$ : = 338.06928, found: 338.06966 Calculated for  $C_{15} H_{15} O_6 N Na S = [M+H]^+$ : = 360.05123, found: 360.05123

#### anti- or I-12a 1D NOE-experiments

![](_page_32_Figure_1.jpeg)

Aromatic proton shows weak NOE of methyl group (1.4 ppm) with aromatic proton at 7.8 ppm, hinting towards conformation depicted below.

![](_page_32_Figure_3.jpeg)

![](_page_33_Figure_0.jpeg)

![](_page_33_Figure_1.jpeg)

![](_page_34_Figure_0.jpeg)

Irradiation of 7<sup>°</sup> (8.15 ppm) shows NOE with not only neighbouring aromatic proton (~7.6 ppm) but also weak effect with methyl group (1.37 ppm).

![](_page_34_Figure_2.jpeg)

#### 2.2.1 Screening conditions for improvement of 12a diastereomeric ratio

Condition #	Catalyst	Solvent	d.r.
1	Ti(O <sup>′</sup> Pr)₄	DCM	1:2
2	Zn(OTf) <sub>2</sub>	DCM	n.d. Complex mixture
3	Cu(OTf) <sub>2</sub>	DCM	3:1
4	Dy(OTf) <sub>3</sub>	DCM	n.d. – Complex mixture
5	AgOAc	Et <sub>2</sub> O	2:1 (f2:f1)
6	AgOAc	THF	10:13
7	AgOAc	DCM +TMEDA	10:6.8
8	AgOAc	DCM	93:7

**Table S1:** Screening conditions to improve d.r. of vinylogous addition to ketimines.

(<u>+</u>)-3-ethyl-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)-5-methyldihydrofuran-2(3H)-one-carbon dioxide (*syn or I-*12b)

![](_page_35_Figure_4.jpeg)

Sulfonamide **syn-12a** (62 mg, 183.8 µmol) and Pd on Carbon (10 wt.%, 22.5 mg, 21.14 µmol) were dissolved in EtOH (1.4 ml) in a 10 ml Schlenk tube and stirred at ambient temperature (21 °C). The vessel was evacuated and refilled with hydrogen. This was repeated two times. The mixture was allowed to come to ambient temperature and stirred under an hydrogen atmosphere for 3 h. Then, reaction mixture was filtered over celite, concentrated and objected to silica gel column chromatography (25% to 35% EA/CyH) to yield 39 mg (0.11 mmol, 63%) of product sulfonamide and recover 28 mg (0.07 mmol, 37%) of starting material.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*) δ 8.02 – 7.99 (m, 1H), 7.81 – 7.78 (m, 1H), 7.69 – 7.65 (m, 2H), 6.05 (s, 1H), 4.44 – 4.32 (m, 2H), 2.51 – 2.45 (m, 1H), 2.16 – 2.11 (m, 1H), 2.07 – 1.98 (m, 2H), 1.52 (s, 3H), 1.37 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 175.9, 168.4, 136.4, 133.7, 132.4, 131.4, 128.3, 121.7, 87.4, 72.7, 64.4, 29.8, 28.0, 23.8, 14.0.

**HMRS (ESI)**: Calculated for  $C_{15} H_{18} O_6 N S = [M+H]^+$ : = 340.08493, found: 340.08524 Calculated for  $C_{15} H_{17} O_6 N Na S = [M+H]^+$ : = 362.06688, found: 362.06717
# 2.3 Folding Pathway

# 2.3.1 Hydrogenolysis of Aziridines

(+)- 4-methyl-3-carbethoxy-3,4-dihydro-2H-benzo[e][1,2]thiazine 1,1-dioxide (13a)



Sulfonamide **2b** (1.0 eq., 40 mg, 0.15 mmol) and Palladium on carbon (10 wt.%, 0.1 eq., 8.0 mg, 0.05 mmol) were dissolved in 1 ml Ethanol in a 10 ml tube equipped with a stirring bar. The vessel was evacuated and refilled with hydrogen using a balloon. This was repeated twice and the reaction was left stirring at 21 °C under an hydrogen atmosphere for 4 h. Then, it was filtered using a syringe filter and concentrated under reduced pressure to give 24 mg (0.09 mmol, 60% yield) product.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*)  $\delta$  7.81 (dq, J = 7.9, 1.4 Hz, 1H), 7.50 (tt, J = 7.6, 1.3 Hz, 1H), 7.41 (tq, J = 7.6, 1.2 Hz, 1H), 7.28 (dd, J = 8.0, 1.2 Hz, 1H), 5.36 – 5.27 (m, 1H), 4.84 (ddd, J = 10.4, 3.7, 1.0 Hz, 1H), 4.32 (qt, J = 7.2, 1.2 Hz, 2H), 3.39 (qd, J = 7.2, 3.7 Hz, 1H), 1.35 (td, J = 7.2, 1.2 Hz, 3H), 1.23 (d, J = 7.2 Hz, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 168.3, 139.2, 136.4, 132.6, 129.4, 128.2, 124.1, 62.4, 57.8, 34.7, 17.5, 14.2.

HMRS (ESI): Calculated for  $C_{12} H_{16} O_4 N S [M+H]^+$ : = 270.07946, found: 270.07923



#### NOESY



(<u>+</u>)-syn- 4-Phenyl-3-carbethoxy-3,4-dihydro-2H-benzo[e][1,2]thiazine 1,1-dioxide (13b)



Sulfonamide **2d** (1.0 eq., 17 mg, 51.61  $\mu$ mol) was combined with Pd/C (10 wt.%, 4.4 mg, 4.13  $\mu$ mol) in a 10 ml Schlenk tube and dissolved in 500  $\mu$ l of a Ethanol/Ethyl acetate mixture (2:1 v/v). The reaction vessel was carefully evacuated under stirring and refilled with hydrogen. This was repeated twice and the reaction was allowed to stir at 21 °C under an hydrogen atmosphere. After completion of conversion, the reaction mixture was filtered using a syringe filter and concentrated to give 16 mg (0.05 mmol, quantitative) of desired product.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*)  $\delta$  7.95 – 7.90 (m, 1H), 7.49 – 7.41 (m, 2H), 7.31 – 7.22 (m, 3H), 7.12 (d, *J* = 7.6 Hz, 1H), 6.96 (dd, *J* = 7.3, 2.2 Hz, 2H), 5.11 (d, *J* = 11.7 Hz, 1H), 5.04 (dd, *J* = 11.7, 3.8 Hz [syn], 1H), 4.58 (d, *J* = 3.8 Hz [syn], 1H), 4.15 – 4.03 (m, 2H), 1.16 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 167.1, 137.4, 137.2, 136.7, 132.9, 130.9, 129.2, 129.0, 128.9, 128.4, 124.3, 62.3, 58.8, 46.0, 14.1.

**HMRS (ESI)**: Calculated for C<sub>17</sub> H<sub>18</sub> O<sub>4</sub> N S = [M+H]<sup>+</sup>: = 332.09511, found: 332.09556

# (<u>+</u>)-*syn*- Diethyl 3,4-dihydro-2H-benzo[e][1,2]thiazine-3,4-dicarboxylate 1,1-dioxide (13c)



Sulfonamide **2e** (1.0 eq., 20 mg, 61.47  $\mu$ mol ) and Palladium on carbon (10 wt.%, 0.1 eq., 6.5 mg, 0.01 mmol) were dissolved in 650  $\mu$ l of a 3:1 Ethanol/Ethyl Acetate (v/v) mixture in a 10 ml Schlenk tube equipped with a stirring bar. The vessel was evacuated and refilled with hydrogen using a balloon. This was repeated twice and the reeaction was left stirring at 21 °C under an hydrogen atmosphere. After completion of conversion, filtration using a syringe filter and concentration delivered desired 8 mg (0.02 mmol, 40% yield) of desired sulfonamide as one diastereomer.

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 7.91 – 7.87 (m, 1H), 7.60 – 7.48 (m, 3H), 5.84 (d, J = 12.4 Hz, 1H), 4.85 (dd, J = 12.4, 3.9 Hz [syn], 1H), 4.31 (qd, J = 7.1, 1.1 Hz, 2H), 4.27 (d, J = 3.9 Hz [syn], 1H), 4.17 (qd, J = 7.1, 3.3 Hz, 2H), 1.34 (t, J = 7.1 Hz, 3H), 1.24 (t, J = 7.1 Hz, 3H). <sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 170.8, 168.1, 137.3, 132.8, 130.8, 130.7, 129.9, 125.1, 63.0, 62.8, 56.5, 44.8, 14.4, 14.3.

**HMRS (ESI)**: Calculated for  $C_{14} H_{18} O_6 N S = [M+H]^+$ : = 328.08493, found: 328.08529

(+)-Ethyl (syn)-C4-(pyridin-2-yl)phenylalaninate 1,1-dioxide (13d)



Palladium on carbon (0.1 eq., 6.4 mg, 6.05  $\mu$ mol) and Sulfonamide **2g** (1.0 eq., 20 mg, 60.54  $\mu$ mol) were combined in 600  $\mu$ l Ethanol/Ethyl Acetate (3:1 v/v) in a 10 ml Schlenk tube equipped with a magnetic stirring bar. The vessel was evacuated and refilled with hydrogen. This procedure was repeated 2 times. The mixture was kept stirring for 4 h. Then, it was filtered using a syringe filter and concentrated to give product 6 mg sulfonamide (0.02 mmol, 30%).

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 8.43 (ddd, *J* = 4.9, 1.9, 0.9 Hz, 1H), 7.95 – 7.89 (m, 1H), 7.68 (td, *J* = 7.7, 1.8 Hz, 1H), 7.44 – 7.40 (m, 2H), 7.33 – 7.28 (m, 1H), 7.25 – 7.21 (m, 1H), 7.18 (ddd, *J* = 7.7, 4.9, 1.1 Hz, 1H), 5.12 (dd, *J* = 12.2, 4.1 Hz, 1H), 4.56 (d, *J* = 4.1 Hz, 1H), 4.06 (qd, *J* = 7.1, 2.5 Hz, 2H), 1.08 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 168.0, 159.2, 149.9, 137.7, 137.1, 136.1, 132.4, 130.0, 128.8, 124.9, 123.3, 123.2, 62.0, 58.9, 45.6, 14.1.

**HMRS (ESI)**: Calculated for  $C_{16} H_{17} O_4 N_2 S = [M+H]^+$ : = 333.09035, found: 333.09018

(+)-4-methyl-3-phenyl-3,4-dihydro-2H-benzo[e][1,2]thiazine 1,1-dioxide (14a)



Sulfonamide **2a** (1.0 eq., 20 mg, 73.71  $\mu$ mol) and Palladium on carbon (10 wt.%, 0.1 eq., 7.8 mg, 0.01 mmol) were dissolved in 750  $\mu$ l of a 3:1 (v/v) Ethanol/Ethyl Acetate Mixture in a 10 ml tube equipped with a stirring bar. The vessel was evacuated and refilled with hydrogen using a balloon. This was repeated twice and the reaction was left stirring at 21 °C under an hydrogen atmosphere for 16 h. Then it was filtered using a syringe filter and concentrated under reduced pressure to give 22 mg (0.08 mmol, 69%) of sulfonamide.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.76 (dt, J = 7.8, 1.0 Hz, 1H), 7.65 (td, J = 7.6, 1.1 Hz, 1H), 7.54 (td, J = 7.6, 1.0 Hz, 1H), 7.36 (dt, J = 7.8, 0.9 Hz, 1H), 7.35 – 7.27 (m, 3H), 7.23 – 7.18 (m, 2H), 4.69 (s, 1H), 3.18 (d, J = 13.6 Hz, 1H), 3.03 (d, J = 13.6 Hz, 1H), 1.57 (s, 3H). <sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 144.9, 135.5, 135.3, 133.3, 130.9, 129.5, 128.6, 127.5, 123.5, 121.6, 63.6, 47.8, 26.7.

**HMRS (ESI)**: Calculated for  $C_{15} H_{15} O_2 N S = [M+H]^+$ :  $C_{15} H_{16} O_2 N S = 274.08963$ , found: 274.08965

(+)-3-benzyl-3-methyl-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (14b)



Sulfonamide **2c** (1.0 eq., 20 mg, 0.06 mmol) and Palladium on carbon (10 wt.%, 0.1 eq., 6.4 mg, 0.01 mmol) were dissolved in 600  $\mu$ l Ethanol/Ethyl Acetate (3:1 v/v) in a 10 ml tube equipped with a stirring bar. The vessel was evacuated and refilled with hydrogen using a balloon. This was repeated twice and the reaction was left stirring at 21 °C under an hydrogen atmosphere for 4 h. Then, it was filtered using a syringe filter and concentrated under reduced pressure to give 6 mg (0.02 mmol, 30% yield) product.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.74 – 7.66 (m, 3H), 7.63 (ddd, *J* = 8.2, 7.2, 1.2 Hz, 1H), 7.52 (ddd, *J* = 8.5, 7.7, 1.2 Hz, 2H), 7.42 – 7.37 (m, 2H), 7.35 – 7.30 (m, 1H), 7.24 – 7.15 (m, 3H), 6.94 (dt, *J* = 6.6, 1.6 Hz, 2H), 4.88 (s, 1H), 3.77 (d, *J* = 14.0 Hz, 1H), 3.59 (d, *J* = 14.0 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 143.2, 141.8, 134.7, 134.5, 133.3, 130.6, 129.6, 129.1, 128.8, 128.4, 127.8, 126.4, 125.0, 121.7, 68.8, 46.6

**HMRS (ESI)**: Calculated for  $C_{20}$  H<sub>18</sub> O<sub>2</sub> N S = [M+H]<sup>+</sup>:  $C_{15}$  H<sub>16</sub> O<sub>2</sub> N S = 336.10528, found: 336.10549

(+)-Ethyl 3-benzyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (14c)



Ethyl 1-phenylazirino[1,2-b]benzo[d]isothiazole-7b(1H)-carboxylate 3,3-dioxide **2f** (1.0 eq., 23 mg, 69.83 µmol) was combined with Palladium on Carbon(10 wt.%, 0.1 eq., 7.4 mg, 0.01 mmol) in a 10 ml Schlenk tube and dissolved in 700 µl of a Ethanol/Ethyl acetate mixture (5:1 v/v). The reaction vessel was carefully evacuated under stirring and refilled with hydrogen. This was repeated twice and the reaction was allowed to stir at 21 °C under an hydrogen atmosphere. The reaction was monitored via TLC. After completion of conversion, reaction mixture was filtered using a syringe filter and concentrated to give 12 mg sulfonamide product (0.04 mmol, 66%).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.74 – 7.66 (m, 3H), 7.63 (ddd, *J* = 8.2, 7.2, 1.2 Hz, 1H), 7.52 (ddd, *J* = 8.5, 7.7, 1.2 Hz, 2H), 7.42 – 7.37 (m, 2H), 7.35 – 7.30 (m, 1H), 7.24 – 7.15 (m, 3H), 6.94 (dt, *J* = 6.6, 1.6 Hz, 2H), 4.88 (s, 1H), 3.77 (d, *J* = 14.0 Hz, 1H), 3.59 (d, *J* = 14.0 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 143.6, 141.8, 134.7, 134.5, 133.3, 130.6, 129.6, 129.1, 128.8, 128.4, 127.8, 126.4, 125.0, 121.7, 68.8, 46.6,

**HMRS (ESI)**: Calculated for  $C_{17} H_{18} O_4 N S = [M+H]^+$ : = 332.09511, found: 332.09535 Calculated for  $C_{17} H_{17} O_4 N Na S = [M+H]^+$ : = 354.07705, found: 354.07723

## (+)-3-benzyl-3-(pyridin-2-yl)-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide(14d)



1-phenyl-7b-(pyridin-2-yl)-1,7b-dihydroazirino[1,2-b]benzo[d]isothiazole 3,3-dioxide **2h** (1.0 eq., 10 mg, 29.90  $\mu$ moll) was combined with Pd/C (10 wt.%; 0.15 eq., 4.7 mg, 4.49  $\mu$ mol) and dissolved in 500  $\mu$ l Ethanol/Ethyl Acetate (2:1) in a 10 ml round bottom flask equipped with a magnetic stirring bar and a septum. The mixture was carefully evacuated and refilled with hydrogen (using a hydrogen-filled balloon). This procedure was repeated twice and the reaction was stirred under an hydrogen atmosphere at 21 °C and monitored by TLC. After 15

h, the reaction mixture was filtered. Concentration delivered crude reaction mixture. Silica gel column chromatography yielded 5 mg (0.01 mmol, 50%) of product sulfonamide.

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  8.63 (ddd, J = 4.8, 1.8, 1.0 Hz, 1H), 7.97 (dt, J = 8.0, 0.9 Hz, 1H), 7.73 (d, J = 7.76 Hz), 7.70 –7.61 (m, 2H), 7.53 (dd, J = 7.6, 1.0 Hz, 1H), 7.22 (ddd, J = 7.3, 4.8, 1.2 Hz, 1H), 7.20 – 7.13 (m, 3H), 6.99 – 6.94 (m, 2H), 5.83 (s, 1H), 4.10 (d, J = 13.8 Hz, 1H), 3.44 (d, J = 13.8 Hz, 1H).

<sup>13</sup>C NMR (126 MHz, Chloroform-d) δ 158.9, 148.9, 142.5, 137.5, 135.2, 134.5, 133.4, 130.5, 129.8, 128.7, 127.6, 125.5, 123.1, 121.4, 121.0, 69.5, 46.9.

**HMRS (ESI)**: Calculated for  $C_{19} H_{17} O_2 N2 S = [M+H]^+$ : = 337.10053, found: 337.10054

### 2.3.2 Hydrogenolysis of Azetidines 15



General Reaction Scheme

# (<u>+</u>)-Ethyl (E)-2-(1,1-dioxido-5-phenyl-4,5-dihydrobenzo[f][1,2]thiazepin-3(2H)-ylidene)acetate (15a')



Sulfonamide **4a** (1.0 eq., 350 mg, 0.98 mmol) was dissolved in 10 ml Ethyl acetate and 83 mg (10wt.% Pd/C, 8 mol%., 78 µmol) and charged with 8 atm hydrogen in a high pressure cylinder and was stirred for 60 h at 21 °C. Eight drops of glacial AcOH were added and the reaction was placed again under 7.5 atm of hydrogen and stirred at that temperature. After 20 h, almost no conversion could be determined and another 6 mol% of Pd/C were added (by that time, it contained 141 mg, 0.14 eq.,0.13 mmol of Pd/C in total). It was stirred for 18 additional hours. As no further conversion (TLC, U-HPLC-MS analysis) could be observed, reaction mixture was filtered over celite and concentrated. Silica gel column chromatography

(12% to 15% EA/CyH) yielded Products **15a**' and **15a** in 49% (173mg, 0.48 mmol) and 51% (179 mg, 0.50 mmol) yield.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*) δ 11.02 (s, 1H), 7.38 (td, J = 7.5, 1.6 Hz, 1H), 7.34 (td, J = 7.6, 1.4 Hz, 1H), 7.29 (td, J = 7.3, 6.6, 1.2 Hz, 2H), 7.24 – 7.21 (m, 1H), 7.18 – 7.15 (m, 2H), 7.14 (dt, J = 7.6, 1.0 Hz, 1H), 4.93 (s, 1H), 4.59 (dd, J = 10.0, 5.2 Hz, 1H), 4.12 (qd, J = 7.1, 1.9 Hz, 2H), 3.73 (t, J = 12.6 Hz, 1H), 3.24 – 3.17 (m, 1H), 1.23 (t, J = 7.1 Hz, 3H). <sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 168.9, 152.2, 144.4, 139.4, 139.3, 134.1, 132.7, 129.1, 128.3, 127.3, 127.0, 125.8, 96.4, 60.3, 50.8, 38.4, 14.3. **HMRS (ESI)**: Calculated for C<sub>19</sub> H<sub>20</sub> O<sub>4</sub> N S = [M+H]<sup>+</sup>: = 358.11076, found: 358.11134 Calculated for C<sub>19</sub> H<sub>19</sub> O<sub>4</sub> N Na S = [M+H]<sup>+</sup>: = 380.09270, found: 380.09279

# (<u>+</u>)-Ethyl 2-((*syn*)-1,1-dioxido-5-phenyl-2,3,4,5-tetrahydrobenzo[f][1,2]thiazepin-3yl)acetate (15a)



<sup>1</sup>**H NMR** (400 MHz, Chloroform-d) δ 8.09 – 8.04 (m, 1H), 7.45 – 7.37 (m, 2H), 7.34-7.27 (m, 3H), 7.25 – 7.22 (m, 2H), 6.66 – 6.61 (m, 1H), 6.67 – 6.59 (m, 1H), 5.40 (d, *J* = 9.6 Hz, 1H), 5.11 (dd, *J* = 5.8, 2.5 Hz, 1H), 4.41 – 4.31 (m, 1H), 4.21 – 4.08 (m, 2H), 2.79 (dd, *J* = 16.6, 4.9 Hz, 1H), 2.31-2.25 (m, 2H), 1.28 – 1.24 (t, *J* = 7.04 Hz, 3H).

<sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 171.7, 143.4, 142.8, 141.2, 132.8, 129.7, 129.2, 129.1, 127.8, 127.4, 126.8, 61.4, 53.3, 46.6, 39.7, 39.1, 14.3.

**HMRS (ESI)**: Calculated for  $C_{19} H_{22} O_4 N S = [M+H]^+$ : = 360.12641, found: 360.12695

Xray Deposition number at the Cambridge Crystallographic Data Centre: CCDC 1910510



(<u>+</u>)-Tert-butyl 2-((*syn*)-1,1-dioxido-5-phenyl-2,3,4,5-tetrahydrobenzo[f][1,2]thiazepin-3yl)acetate (15b)



59% yield (0.64 mmol, 274 mg)

<sup>1</sup>**H NMR** (600 MHz, Chloroform-d)  $\delta$  8.10 – 8.06 (m, 1H), 7.45 – 7.42 (m, 2H), 7.37 – 7.34 (m, 1H), 7.33 – 7.30 (m, 2H), 7.28 – 7.26 (m, 2H), 6.67 – 6.64 (m, 1H), 5.50 (d, *J* = 9.5 Hz, 1H), 5.13 (d, *J* = 9.9 Hz, 1H), 4.37 – 4.30 (m, 1H), 2.76 (dd, *J* = 16.7, 4.7 Hz, 1H), 2.51 (dd, *J* = 16.7, 4.9 Hz, 1H), 2.36 – 2.25 (m, 2H), 1.46 (s, 9H).

<sup>13</sup>C NMR (151 MHz, Chloroform-d) δ 171.0, 143.3, 142.7, 141.0, 132.5, 129.4, 129.0, 128.9, 127.6, 127.1, 126.6, 53.3, 46.5, 40.4, 38.8, 28.1, 26.9.

**HMRS (ESI)**: Calculated for  $C_{21} H_{26} O_4 N Na S = [M+Na]^+$ : = 411.14748, found: 411.14993

(<u>+</u>)-4-(3-(ethoxycarbonyl)-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)butanoic acid (16)



Pearlman's catalyst (20wt%., 9mol%, 7.2 mg, 0.01 mmol) and **4d** (1.0 eq.,50 mg, 0.12 mmol) were dissolved in a 3:1 (v/v) Methanol/Ethyl Acetate Mixture (1.250) in a 10 ml tube equipped with a stirring bar. The vessel was placed in a high-pressure reactor, which was then sealed and charged with Hydrogen (9 atm). The reactor was placed on a stirring plate and left there for 2 h. Then, reaction mixture was filtered to furnish 37 mg (0.11 mmol, 93% yield) of (+)-4-(3-(ethoxycarbonyl)-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)butanoic acid **16**.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-d)  $\delta$  7.76 (d, J = 7.7 Hz, 1H), 7.72 (d, J = 7.9 Hz, 1H), 7.66 (t, J = 7.7 Hz), 7.60 (dd, J = 11.1, 3.9 Hz, 1H), 5.83 (s, 1H), 4.37 – 4.28 (m, 2H), 2.43 (dt, J = 16.7, 7.0 Hz, 1H), 2.36 (dt, J = 16.7, 7.3 Hz, 1H), 2.29 (ddd, J = 13.7, 11.5, 4.8 Hz, 1H), 2.05 (ddd, J = 13.7, 11.7, 4.8 Hz, 1H), 1.76 – 1.64 (m, 2H), 1.36 (t, J = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 178.3, 169.9, 137.9, 135.4, 133.6, 130.6, 124.9, 121.5, 69.0, 63.8, 39.3, 33.1, 19.7, 14.1.

**HMRS (ESI)**: Calculated for  $C_{14} H_{18} O_6 N S = [M+H]^+$ : = 328.08493, found: 328.08477

## 2.3.2.1 Hydrolysis of Esters 15



# (<u>+</u>)-2-((*syn*)-1,1-dioxido-5-phenyl-2,3,4,5-tetrahydrobenzo[f][1,2]thiazepin-3-yl)acetic acid (17)

### By hydrolysis from Ethyl Ester 15a

Sulfonamide **15a** (1.0 eq. 252 mg, 701.09  $\mu$ mol) was dissolved in 9.16 ml THF/MeCN 4:1 v/v mixture. Then, 2.92 ml water and aqueous 2 M NaOH (974  $\mu$ l) was added to the solution. The resulting mixture was stirred at ambient temperature stirred for 19 hours. The reaction was quenched by addition of 1 M HCI (2.103 ml, 2.10 mmol) and concentrated under reduced pressure. Silica gel column chromatography (20% to 40% EA/CyH to 10% EtOH/EA) delivered 217 mg (0.65 mmol, 93%) free acid as a white solid.

## By hydrolysis of tert-butyl ester 15b

Tert-butyl Ester **15b** (1.0 eq., 145 mg, 374.20  $\mu$ mol) was dissolved in 3.2 ml THF/1 M aq. HCl (3:1 v/v) in a 30 ml microwave tube. The vessel was sealed and irradiated at 100 °C/250 W for 45 min using a microwave synthesizer. The reaction was quenched by addition of aqueous 2 M NaOH and extracted using Ethyl acetate ( 3 x 5 ml). Concentration and silica gel column chromatography (20% to 40% to 10% EtOH/EA) delivered free acid (24 mg, 0.07 mmol, 19%) delivered free acid (24 mg, 0.21 mmol, 56%).

<sup>1</sup>**H NMR** (500 MHz, Acetonitrile-*d*<sub>3</sub>) δ 7.99 – 7.95 (m, 1H), 7.43 (t, J = 7.5 Hz, 2H), 7.38 – 7.32 (m, 2H), 7.29 – 7.25 (m, 2H), 6.63 – 6.59 (m, 1H), 5.51 (d, J = 9.9 Hz, 1H), 5.05 (d, J = 10.6 Hz, 1H), 4.36-4.27 (m, 1H), 2.59 (dd, J = 16.1, 5.6 Hz, 1H), 2.51 (dd, J = 16.1, 7.8 Hz, 1H), 2.32 (d, J = 14.0, 1.5 Hz,1H), 2.08 (dd, J = 14.0, 2.4 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, CD<sub>3</sub>CN) δ 172.3, 144.5, 143.9, 142.3, 133.5, 130.1, 129.8, 129.8, 128.1, 127.9, 127.5, 54.4, 47.3, 40.2, 40.1. **HMRS (ESI)**: Calculated for C<sub>17</sub> H<sub>18</sub> O<sub>4</sub> N S = [M+H]<sup>+</sup>: = 332.09511, found: 332.09543

#### 2.3.2.2 Synthesis of Amides 18a-c



(<u>+</u>)-2-((*syn*)-1,1-dioxido-5-phenyl-2,3,4,5-tetrahydrobenzo[f][1,2]thiazepin-3-yl)-N,Ndiethylacetamide (18a)



To a solution of 40 mg of carboxylic acid **17** (1.0 eq.,120.70 µmol) in chloroform (0.9 ml) was added oxalyl chloride (1.15 eq., 238 µl, 5% solution in DCM). The vial was heated gently and dissolved after a few minutes. It was then heated at 60 °C overnight. The reaction was allowed to cool to room temperature and then concentrated under reduced pressure to remove any excess oxalyl chloride. Then, the reaction mixture was redissolved in THF (0.9 ml) and diethylamine (4.0 eq., 50 µl, 483 µmol) was added dropwise at 0 °C. After completion of conversion, reaction mixture was concentrated under reduced pressure, dried *in vacuo* and purified by preparative HPLC (C18, 25% to 80% MeCN/H<sub>2</sub>O (+0.1% v/v HCOOH)) to yield 27 mg (0.07 mmol, 58%).

<sup>1</sup>**H NMR** (600 MHz, Chloroform-*d*) δ 8.08 – 8.02 (m, 1H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.34 – 7.31 (m, 1H), 7.27 – 7.23 (m, 4H), 6.64 – 6.59 (m, 1H), 5.09 (d, *J* = 10.8 Hz, 1H), 4.36 – 4.29 (m, 1H), 3.38 (dt, *J* = 14.1, 7.0 Hz, 1H), 3.32-3.20 (m, 3H), 2.80 (dd, *J* = 16.7, 4.5 Hz, 1H), 2.68 – 2.58 (m, 2H), 2.14 (ddd, *J* = 13.9, 2.3, 1.0 Hz, 1H), 1.17 (t, *J* = 7.1 Hz, 3H), 1.10 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (151 MHz, Chloroform-d) δ 170.0, 143.8, 142.8, 141.3, 132.5, 129.5, 129.1, 129.0, 127.9, 127.2, 126.6, 54.0, 46.9, 42.3, 40.5, 38.7, 37.3, 14.3, 13.1.

**HMRS (ESI)**: Calculated for  $C_{21} H_{27} O_3 N S = [M+H]^+$ : = 387.17369, found: 387.17375 Calculated for  $C_{21} H_{26} O_3 N Na S = [M+H]^+$ : = 409.15563, found: 409.15560

(<u>+</u>)-2-((*syn*)-1,1-dioxido-5-phenyl-2,3,4,5-tetrahydrobenzo[f][1,2]thiazepin-3-yl)-1morpholinoethan-1-one (18b)



To a solution of Acid **17** (1.0 eq., 25 mg, 76  $\mu$ mol) in chloroform (1 ml) was added thionyl chloride (1.20 eq., 7.5  $\mu$ l, 90  $\mu$ mol, as 5% solution in CHCI<sub>3</sub>). The vial was heated gently and dissolved after a few minutes. It was then heated at 40° C for 16 h. The reaction was allowed to cool to room temperature and then concentrated under reduced pressure to remove any excess thionyl chloride. Crude acyl chloride was then dissolved in 500  $\mu$ l THF in an oven-dried Schlenk tube equipped with a magnetic stirring bar under Argon atmosphere. Then, 5.0 eq. morpholine (29  $\mu$ l, 0.34 mmol) was added and the reaction was kept stirring overnight. Then, it was concentrated, dried and purified by preparative HPLC to deliver 14 mg (7.49  $\mu$ mol, 10% yield) of product sulfonamide.

Purification by preparative HPLC (C18, 35% to 90% MeCN/H<sub>2</sub>O (+0.1% v/v TFA))

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  8.08 – 8.02 (m, 1H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.35 – 7.30 (m, 1H), 7.29 – 7.23 (m, 4H), 6.65 – 6.61 (m, 1H), 6.17 (d, *J* = 9.6 Hz, 1H), 5.07 (d, *J* = 10.8 Hz, 1H), 4.37 (m, 1H), 3.77 – 3.61 (m, 5H), 3.53 – 3.40 (m, 3H), 2.79 (dd, *J* = 16.7, 4.8 Hz, 1H), 2.70 – 2.55 (m, 2H), 2.16 (d, *J* = 12.7 Hz, 1H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 167.3, 141.6, 140.7, 139.2, 130.5, 127.5, 127.1, 127.0, 125.8, 125.2, 124.6, 64.8, 64.5, 51.7, 45.0, 44.0, 39.8, 36.9, 35.4.

**HMRS (ESI)**: Calculated for  $C_{21} H_{25} O_4 N_2 S = [M+H]^+$ : = 401.15218, found: 401.15295 Calculated for  $C_{21} H_{24} O_4 N_2 Na S = [M+H]^+$ : = 423.13490 found: 423.13396

# (<u>+</u>)-2-((syn)-1,1-dioxido-5-phenyl-2,3,4,5-tetrahydrobenzo[f][1,2]thiazepin-3-yl)-Npropylacetamide (18c)



To a solution of Acid **17** (1.0 eq., 25 mg, 76  $\mu$ mol) in chloroform (1 ml) was added thionyl chloride (1.20 eq., 7.5  $\mu$ l, 90  $\mu$ mol, as 5% solution in CHCl<sub>3</sub>). The vial was heated gently and dissolved after a few minutes. It was then heated at 40° C for 16 h . The reaction was allowed to cool to room temperature and then concentrated under reduced pressure to remove any excess thionyl chloride. Crude acyl chloride was then dissolved in 500  $\mu$ l THF in an oven-dried

Schlenk tube equipped with a magnetic stirring bar under Argon atmosphere. Then, 5.0 eq. n-propylamine (28  $\mu$ I, 0.34 mmol) was added and the reaction was kept stirring overnight. Then, it was concentrated, dried and purified by preparative HPLC to deliver 14 mg (0.04 mmol, 50% yield) of product sulfonamide.

Purification by preparative HPLC (C18, 35% to 90% MeCN/H<sub>2</sub>O (+0.1% v/v TFA))

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  8.04 (dd, J = 5.5, 2.0 Hz, 1H), 7.40 (t, J = 7.5 Hz, 2H), 7.35 – 7.30 (m, 1H), 7.29 – 7.27 (m, 1H), 7.24 (ddd, J = 8.0, 3.5, 2.0 Hz, 3H), 6.63 – 6.58 (m, 1H), 6.31 (s, 1H), 5.59 (d, J = 6.0 Hz, 1H), 5.09 (d, J = 10.6 Hz, 1H), 4.30 (s, 1H), 3.23 – 3.13 (m, 2H), 2.76 (dd, J = 15.6, 4.3 Hz, 1H), 2.41 – 2.32 (m, 2H), 2.23 – 2.17 (m, 1H), 1.52 (h, J = 7.1 Hz, 2H), 0.92 (t, J = 7.4 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 170.8, 143.6, 142.6, 141.2, 132.5, 129.5, 129.1, 127.8, 127.2, 126.7, 53.7, 46.5, 41.4, 40.5, 38.7, 22.9, 11.5

**HMRS (ESI)**: Calculated for  $C_{20} H_{25} O_3 N_2 S = [M+H]^+$ : = 373.15804, found: 373.15774 Calculated for  $C_{20} H_{24} O_3 N_2 Na S = [M+H]^+$ : = 395.13998 found: 395.13907

# 2.3.3 Synthesis of fused Pyrrolines by Reduction



(<u>+</u>)-Ethyl 9b-phenyl-1,2,3,9b-tetrahydrobenzo[d]pyrrolo[1,2-b]isothiazole-1-carboxylate 5,5-dioxide (19a)



To 23 mg Pyrroline **5a** (1.0 eq., 0.06 mmol) in 1.5 ml of a Methanol/Ethyl Acetate Mixture (5:1) was added 3.2 mg Pearlman's Catalyst (20wt.%, 4mol%, 2.26 µmol,) in a Schlenk tube. The mixture was cooled to 0°C, degassed and refilled with hydrogen, again degassed, once more refilled with hydrogen using a balloon and stirred at rt.

After 40 min, TLC (Ethyl Acetate/CyH 1:3) showed complete conversion of starting material (rf= 0.33) to a product (rf = 0.24). The mixture was filtered through a syringe filter and the

solvent was removed under reduced pressure to obtain crude product, which was purified using silica gel column chromatography (14%-20% EA/CH) to yield **12a** in 95% yield (22 mg, 0.06 mmol).

<sup>1</sup>**H NMR** (600 MHz, Chloroform-*d*) δ 8.05 (d, *J* = 7.9 Hz, 1H), 7.73 – 7.67 (m, 2H), 7.59 – 7.53 (m, 3H), 7.33– 7.27 (m, 2H), 7.27 – 7.21 (m, 1H), 3.94-3.73 (m, 4H), 3.60 (t, *J* =7.6 Hz, 1H), 2.49 (m, 1H), 2.15 (m, 1H), 1.0 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (151 MHz, Chloroform-d) δ 170.2, 143.6, 139.1, 135.9, 133.1, 129.7, 128.2, 128.1, 126.8, 125.6, 121.7, 78.2, 61.3, 56.7, 48.4, 28.9, 13.7

**HMRS (ESI)**: Calculated for  $C_{19} H_{20} O_4 N S = [M+H]^+$ : = 358.11076, found: 358.11092 Calculated for  $C_{19} H_{19} O_4 N Na S = [M+H]^+$ : = 380.09270, found: 380.09282

## 19a NOESY





Structure of lowest energy conformation of substrate hints that addition leading to antipyrrolidine may be preferred, as the most probable structure (deduced via NOE interactions) suggests.

(<u>+</u>)-9b-(ethoxycarbonyl)-1,2,3,9b-tetrahydrobenzo[d]pyrrolo[1,2-b]isothiazole-1carboxylic acid 5,5-dioxide (19b)



Structure assigned in analogy to 19a.

Pearlman's catalyst (10mol%, 8.4 mg, 0,01 mmol) and **5b** (1.0 eq., 50.0 mg, 0.12 mmol) were dissolved in a 3:1 Methanol/Ethyl Acetate (3:1 v/v) mixture in a 10 ml glass tube equipped with a stirring bar. The open vessel was placed in a high-pressure reactor, which was then sealed and charged with Hydrogen (9 atm). The reactor was placed on a stirring plate and left there for 3 h. Then, reaction mixture was filtered using a syringe filter. Concentration of filtrate delivered **19b** in 92% yield (36 mg, 0.11 mmol).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.94 (d, *J* = 7.8 Hz, 1H), 7.77 (d, *J* = 7.65 Hz, 1H), 7.68 (td, *J* = 7.6, 1.3 Hz, 1H), 7.63 (dd, *J* = 7.6, 1.1 Hz, 1H), 4.21 (m, 2H), 3.89 (dt, *J* = 10.7, 7.6 Hz, 1H), 3.68 (ddd, *J* = 10.7, 7.6, 4.6 Hz, 1H), 3.27 (t, *J* = 8.4 Hz, 1H), 2.67 (m, 1H), 2.42 (m, 1H), 1.26 (t, *J* = 7.1 Hz, 2H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 175.7, 168.7, 136.7, 135.1, 133.7, 130.9, 126.4, 121.5, 63.0, 60.6, 53.7, 46.5, 30.1, 13.9.

**HMRS (ESI)**: Calculated for  $C_{14} H_{16} O_6 N S = [M+H]^+$ : = 326.06928, found: 326.06963 Calculated for  $C_{14} H_{15} O_6 N Na S = [M+H]^+$ : = 348.05123, found: 348.05164

### 2.3.4 Ring modulation reaction of cycloadduct 8a

## (+)-3-(aminomethyl)-2-methyl-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (20)



Sulfonamide **8a** (1.0 eq., 55 mg, 0.18 mmol) was dissolved in 1.8 ml in a 10 ml glass vial equipped with a magnetic stirring bar. Pd/C (10 wt.%, 0.1 eq., 19.5 mg, 0.02 mmol) was added and the open vial was placed in a high-pressure reactor, whichw as charged with 7 atm of hydrogen. The reactor was placed on a stirring plate and the reaction was stirred at ambient temperature for 16 h. Then, it was filtered over celite and concentrated. Purification by preparative HPLC (C18, 10% to 50% MeCN/H<sub>2</sub>O (+0.1% v/v TFA))) delivered unstable Product **20** in 87% yield (34 mg, 0.16 mmol).

<sup>1</sup>**H NMR** (500 MHz, Acetonitrile- $d_3$ )  $\delta$  7.75 – 7.73 (d, J = 8.1 Hz, 1H), 7.67 – 7.65 (m, 1H), 7.57 (t, J = 7.9 Hz, 2H), 4.77 (dd, J = 8.8, 4.9 Hz, 1H), 2.70 (dd, J = 12.6, 4.9 Hz, 1H), 2.54 (dd, J = 12.6, 8.8 Hz, 1H), 2.28 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CD<sub>3</sub>CN) δ 139.8, 136.6, 133.6, 130.2, 125.7, 121.3, 118.0, 64.1, 55.8, 45.4.

**HMRS (ESI)**: Calculated for  $C_9 H_{13} O_2 N_2 S = [M+H]^+$ : = 213.06922, found: 213.06943





Crude sulfonamide **20** (1.0 eq.,33 mg, 109.86  $\mu$ mol) was dissolved in 1.0 ml MeOH in a 10 ml glass vial equipped with a magnetic stirring bar. Pd/C (10 wt.%, 0.1 eq., 19.5 mg, 0.02 mmol) was added and the open vial was placed in a high-pressure reactor, which was charged with 7 atm of hydrogen. The reactor was placed on a stirring plate and the reaction was stirred at ambient temperature for 16 h. Then, it was filtered over celite and concentrated. It was dissolved in 2 ml dry acetonitrile and triethylamine (2.30 eq., 35  $\mu$ l, 252.7  $\mu$ mol) and CDI (1.15 eq., 20.5 mg, 0.13 mmol) were added and the reaction stirred at 21° C for 16 h. Then, it was

stopped by addition of 1 M aqueous HCl (50  $\mu$ l) and concentrated. The residue was purified by preparative HPLC (C18, 10% to 60% MeCN/H<sub>2</sub>O (+0.1% v/v TFA)) to yield 6 mg (23%, 0.03 mmol) of **21**.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.84 (d, *J* = 7.8, 1H), 7.71 (td, *J* = 7.6, 1.0 Hz, 1H), 7.62 (t, *J* = 7.7, 1H), 7.42 (dq, *J* = 7.8, 0.7 Hz, 1H), 5.40 (dd, *J* = 9.3, 4.5 Hz, 1H), 4.01 (t, *J* = 9.3 Hz, 1H), 3.67 (dd, *J* = 9.1, 4.5 Hz, 1H), 2.89 (s, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 152.5, 136.9, 135.3, 132.9, 129.5, 122.5, 121.2, 53.9, 49.3, 29.9.

**HMRS (ESI)**: Calculated for  $C_{10} H_{11} O_3 N_2 S = [M+H]^+$ : = 239.04849, found: 239.04863 Calculated for  $C_{10} H_{11} O_3 N_2 Na S = [M+Na]^+$ : = 261.03043, found: 261.03012

### 2.3.5 Synthesis of tetrahydropyridones 22a-d



### (+)-7,8,10,10a-tetrahydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5-dioxide (22a)



Enamide **6a** (1.0 eq., 195 mg, 0.83 mmol) was combined with Pd/C (10wt%, 0.1 eq., 88 mg, 82.89 µmol) and dissolved in 16 ml EtOH/Ethyl Acetate 1:1 (v/v). The mixture was stirred in a high-pressure reactor 18 h under 8 atm of hydrogen pressure. Then, reaction mixture was filtered over celite and concentrated. The main component of reaction mixture was alcohol. Crude reaction mixture was dissolved in 22 ml DCM and solid sodium bicarbonate (6.25 eq., 435 mg, 0.01 mol) was added. The reaction mixture was cooled down to 0° C (ice-water bath) and DMP (2.5 eq., 879 mg, 2.07 mmol) was carefully added portionwise over 10 min under a stream of Argon. The reaction was monitored via TLC. After completion of conversion (3 h) 50 ml of aqueous sodium thiosulfate solution was added and the reaction mixture was transferred to a separation funnel. Layers were separated and the aqueous layer was extracted twice

more with 40 ml DCM each. Combined organic layers were washed with brine (25 ml each) and dried over anhydrous sodium sulfate. Concentration delivered crude product ,which was objected to silica gel flash column chromatography (29% to 35% EA/CyH) to yield tetrahydropyridin-4one **22a** in 59% yield (117 mg, 0.49 mmol).

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.89 – 7.85 (m, 1H), 7.67 (td, *J* = 7.6, 1.3 Hz, 1H), 7.63 – 7.58 (m, 1H), 7.41 – 7.34 (m, 1H), 4.66 (dd, *J* = 12.0, 3.7 Hz, 1H), 4.19 (ddd, *J* = 13.3, 7.3, 2.1 Hz, 1H), 3.40 (ddd, *J* = 13.2, 12.0, 3.7 Hz, 1H), 2.97 (ddd, *J* = 14.1, 3.7, 1.7 Hz, 1H), 2.83 (dddd, *J* = 14.8, 12.0, 7.3, 0.8 Hz, 1H), 2.63 – 2.54 (m, 2H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 204.4, 136.6, 134.5, 133.3, 129.9, 123.3, 121.6, 58.1, 45.9, 39.4, 38.4.

**HMRS (ESI)**: Calculated for  $C_{11} H_{12} O_3 N S = [M+H]^+$ : = 238.05324 found: 238.05324

(<u>+</u>)-10a-phenyl-7,8,10,10a-tetrahydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9-one 5,5dioxide (22b)



Tricyclic sulfonamide **6b** (307 mg, 986.01 µmol) and Pd on Carbon (10 wt%,74 mg, 69.02 µmol) were dissolved in Ethanol (18 ml) in a 100 ml Schlenk flask equipped with a magnetic stirring bar. The mixture stirred at ambient temperature with a balloon filled with hydrogen attached. The reaction was monitored by TLC. After 3 h, TLC analysis had implicated complete conversion of starting material and the mixture was filtered over celite and solvent was evaporated under reduced pressure to give crude reaction product, which was purified by silica gel column chromatography (18% to 32% EA/CyH) to give product sulfonamide **22b** in 90% yield (278 mg, 0.89 mmol).

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.92 – 7.89 (m, 1H), 7.59 – 7.51 (m, 2H), 7.44 – 7.29 (m, 6H), 7.03 – 6.98 (m, 1H), 4.18 (ddd, *J* = 14.7, 7.6, 1.5 Hz, 1H), 3.55 (dd, *J* = 14.4, 1.6 Hz, 1H), 3.13 (ddd, *J* = 14.7, 12.3, 3.8 Hz, 1H), 2.96 – 2.84 (m, 3H), 2.34 – 2.25 (m, 1H).

<sup>13</sup>**C NMR** (101 MHz, Chloroform-d) δ 204.5, 142.8, 137.8, 133.8, 132.7, 129.8, 129.5, 129.1, 127.5, 124.0, 121.8, 69.1, 49.6, 39.1, 36.1. **HMRS (ESI)**: Calculated for  $C_{17}$  H<sub>16</sub> O<sub>3</sub> N<sub>2</sub> S = [M+H]<sup>+</sup>: = 314.08454, found: 314.08470

# (<u>+</u>)-10a-(pyridin-2-yl)-7,8,10,10a-tetrahydro-9H-benzo[4,5]isothiazolo[2,3-a]pyridin-9one 5,5-dioxide (22c)



Sulfonamide **6c** (1.0 eq., 220 mg, 704.35 µmol) and Pd/C (0.12 eq., 90 mg, 84.52 µmol) were combined in a 35 ml reaction tube equipped with a magnetic stirring bar. 7 ml of Ethyl Acetate/EtOH (15:7 v/v) were added and the vessel was placed in a high-pressure reactor. The reactor was charged with 7 atm of hydrogen pressure and stirred for 16 h at 22 °C. Then reaction mixture was filtered over celite and concentrated. Silica gel column chromatography (19 to 23% EA/CyH) delivered desired product in 24% yield (54 mg, 0.17 mmol) and recovered starting material. Corresponding alcohol or other side products could not be detected.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*)  $\delta$  8.67 – 8.62 (m, 1H), 7.92 – 7.88 (m, 1H), 7.65 (td, *J* = 7.7, 1.8 Hz, 1H), 7.60 – 7.59 (m, 2H), 7.31 (ddt, *J* = 10.2, 8.0, 0.8 Hz, 2H), 7.26 – 7.24 (m, 1H), 4.20 (ddd, *J* = 14.3, 7.4, 2.4 Hz, 1H), 4.00 (dd, *J* = 14.5, 1.6 Hz, 1H), 3.25 (ddd, *J* = 14.3, 11.4, 4.1 Hz, 1H), 2.80 (dddd, *J* = 15.4, 11.4, 7.4, 0.8 Hz, 1H), 2.70 (dd, *J* = 14.4, 0.8 Hz, 1H), 2.38 (dddd, *J* = 15.4, 4.0, 2.4, 1.5 Hz, 1H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 201.8, 155.4, 147.3, 138.8, 135.9, 131.8, 130.9, 128.1, 122.1, 121.6, 119.7, 119.3, 67.7, 47.2, 36.6, 34.3.

**HMRS (ESI)**: Calculated for  $C_{16} H_{15} O_3 N_2 S = [M+H]^+$ : = 315.07979 found: 315.07962

(<u>+</u>)-ethyl 9-oxo-7,8,9,10-tetrahydro-10aH-benzo[4,5]isothiazolo[2,3-a]pyridine-10acarboxylate 5,5-dioxide (22d)



Sulfonyl enamide **6d** (1.0 eq., 150 mg, 0.49 mmol) was dissolved in 9ml EtOH and 26 mg (5mol%, 10wt.%content ,24.40  $\mu$ mol) of Pd/C was added to the mixture. The reaction vessel (35 ml tube containing magnetic stirring bar) was placed in a high-pressure reactor that was charged with 7 bars of hydrogen and placed on a stirring plate. The reaction was stirred at 20° C and monitored by TLC.

The crude reaction mixture was filtered over celite, concentrated and the residue objected to silica gel flash column chromatography (25 to 28 to 32 % EA/CyH) to yield tetrahydropyridin-4one **22c** in 97% yield (146 mg 0.47 mmol).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.91 – 7.85 (m, 1H), 7.73 – 7.63 (m, 2H), 7.58 – 7.53 (m, 1H), 4.30 – 4.17 (m, 3H), 3.62 (ddd, *J* = 14.0, 11.6, 4.1 Hz, 1H), 3.36 (dd, *J* = 14.4, 1.6 Hz, 1H), 2.76 (dddd, *J* = 15.0, 11.6, 7.6, 0.7 Hz, 1H), 2.62 (dd, *J* = 14.5, 0.7 Hz, 1H), 2.51 (ddt, *J* = 15.0, 4.0, 1.9 Hz, 1H), 1.24 (t, *J* = 7.1 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 202.7, 168.1, 135.9, 133.7, 133.5, 130.8, 123.8, 121.9, 68.3, 63.5, 48.9, 38.8, 37.1, 14.1.

**HMRS (ESI)**: Calculated for  $C_{14} H_{16} O_5 N S = [M+H]^+$ : = 310.07437 found: 310.07468

2.3.6 Synthesis of tricyclic sulfonamides 23a-e via reductive amination



(<u>+</u>)-Ethyl 9-morpholino-7,8,9,10-tetrahydro-10aH-benzo[4,5]isothiazolo[2,3-a]pyridine-10a-carboxylate 5,5-dioxide (23a)



Morpholine (1.2 eq., 6.7  $\mu$ l, 0.08 mmol) was added dropwise via syringe to a solution of **22d** (1.0 eq., 20 mg, 64.7  $\mu$ mol) with 2-Ethylhexanoic acid (1.25 eq., 12.9  $\mu$ l, 0.08 mmol) in DCM (0.6 ml) and stirred for 20 min. Then, in situ generated NaBH(OEh)<sub>3</sub> (1.25 eq., 12.9  $\mu$ l, 0.08 mmol) was added and the reaction stirred overnight. Then, solvents were removed and the residue purified by silica gel column chromatography (30% to 45% EA/CyH) to yield 16 mg (0.04 mmol, 65%) of amine product.

<sup>1</sup>**H NMR** (400 MHz, Chloroform-*d*) δ 7.81 (d, *J* = 7.7 Hz, 1H), 7.75 (d, *J* = 7.6 Hz, 1H), 7.66 – 7.54 (m, 2H), 4.27 (dq, *J* = 10.8, 7.3 Hz, 1H), 4.16 (dq, *J* = 10.8, 7.1 Hz, 1H), 3.80 (ddd, *J* = 14.0, 4.9, 2.0 Hz, 1H), 3.75 – 3.62 (m, 4H), 3.50 (td, *J* = 13.7, 2.6 Hz, 1H), 3.15 (ddd, *J* = 14.2, 3.3, 2.0 Hz, 1H), 2.66 (s, 2H), 2.50 (p, *J* = 3.1 Hz, 1H), 2.40 (s, 2H), 2.04 (dt, *J* = 15.1, 2.6 Hz, 1H), 1.88 – 1.75 (m, 1H), 1.65 – 1.58 (m, 2H), 1.28 (t, *J* = 7.2 Hz, 3H).

<sup>13</sup>**C NMR** (101 MHz, Chloroform-d) δ 170.4, 137.5, 133.3, 133.2, 130.2, 124.4, 121.5, 67.1, 64.2, 62.5, 56.7, 51.4, 36.6, 33.9, 25.5, 14.1.

**HMRS (ESI)**: Calculated for  $C_{18} H_{25} O_5 N_2 S = [M+H]^+$ : = 381.14787 found: 381.14779 Calculated for  $C_{18} H_{24} O_5 N_2 Na S = [M+H]^+$ : = 403.12981 found: 403.12960 Calculated for  $C_{18} H_{24} O_5 N_2 K S = [M+H]^+$ : = 419.10375 found: 419.10375



NOE experiment shows NOE between Morpholine H and CH<sub>2</sub>-group of ethyl ester.

# (<u>+</u>)-9-morpholino-8,9,10,10a-tetrahydro-7H-benzo[4,5]isothiazolo[2,3-a]pyridine 5,5dioxide (23b)



**22a** (1.0 eq., 16 mg, 67.43 µmol), morpholine (1.25 eq., 145 µl, 84.3 µmol, as 5% v/v solution in 1,2-DCE) and powdered 4 A molecular sieves were combined in dry 1,2 DCE (800 µl) in a two-necked round bottom flask under an Argon atmosphere and stirred for 12 h at ambient temperature. Then, sodium triacetoxyborohydride (1.4 eq., 20 mg, 94.40 µmol) and glacial acetic acid (1.05 eq., 4 µl, 70.80 µmol) were added and the reaction stirred at ambient temperature overnight. Then, it was concentrated in vacuo and the residue purified (C18, 10% to 38% MeCN/H<sub>2</sub>O (+0.1% v/v TFA)) yield 7.0 mg (0.02 mmol, 34%) of product sulfonamide.

<sup>1</sup>**H NMR** (700 MHz, Chloroform-*d*)  $\delta$  7.81 (d, *J* = 7.7 Hz, 1H), 7.65 (t, *J* = 7.4 Hz, 1H), 7.57 (t, *J* = 7.6 Hz, 1H), 7.47 (d, *J* = 7.7 Hz, 1H), 5.06 (dd, *J* = 9.4, 4.1 Hz, 1H), 4.02 (d, *J* = 4.7 Hz, 4H), 3.61 (ddt, *J* = 25.0, 13.2, 7.3 Hz, 2H), 3.33 – 2.99 (m, 6H), 2.69 (dt, *J* = 14.2, 5.3 Hz, 1H), 2.28 (h, *J* = 5.6, 4.3 Hz, 1H), 2.25 – 2.15 (m, 3H).

<sup>13</sup>**C NMR** (176 MHz, Chloroform-d) δ 137.5, 134.8, 133.6, 130.0, 124.2, 122.1, 64.3, 60.1, 54.5, 50.4, 36.9, 29.6, 23.3.

**HMRS (ESI)**: Calculated for  $C_{15} H_{21} O_3 N_2 S = [M+H]^+$ : = 381.12674 found: 381.12676



In NOESY experiment, no NOE between benzylic proton and the other tertiary proton can be established. For the other amines, in contrast, such an NOE could be found. Consequently, we suggest that we isolated above depicted stereoisomer.

(<u>+</u>)-9-(propylamino)-8,9,10,10a-tetrahydro-7H-benzo[4,5]isothiazolo[2,3-a]pyridine 5,5dioxide (23c)



**22a** (1.0 eq., 28 mg, 110.93 µmol), n-propylamine (1.25 eq., 11 µl, 138.66 µmol) and powdered 4 A molecular sieves were combined in dry 1,2 DCE (800 µl) in a 10 ml oven-dried Schlenk tube under an Argon atmosphere and sodium triacetoxyborohydride (1.4 eq., 33 mg, 155.30 µmol) and glacial acetic acid (1.05 eq., 6.6 µl, 116.47 µmol) were added and the reaction stirred at ambient temperature overnight. The reaction was concentrated and the crude reaction material was purified by preparative HPLC (C18, 3% to 7% MeCN/H<sub>2</sub>O (+0.1% v/v TFA)) Lyophilization then delivered product amine **23c** (9.4 mg, 0.03 mmol, 30%) and alcohol side product **24** (11 mg, 0.05 mmol, 41%).

### Mixture of diastereomers, ~10:9 syn:anti (see NOESY)

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.77 (d, J = 7.7 Hz, 1H, *major*), 7.72 (d, J = 7.7 Hz, 1H, *minor*), 7.60 (td, J = 7.6, 1.1 Hz, 1H), 7.55 – 7.50 (m, 2H), 7.47 – 7.38 (m, 3H), 5.15 (dd, J = 12.2, 3.1 Hz, 1H, minor), 4.25 (dd, J = 11.7, 2.9 Hz, 1H, major), 3.99 (ddd, J = 14.4, 12.8, 3.4 Hz, 1H), 3.91 (ddd, J = 13.5, 4.9, 2.0 Hz, 1H), 3.77 (ddd, J = 14.2, 5.6, 2.2 Hz, 1H), 3.55 (t, J = 3.5 Hz, 1H, minor), 3.42 (tt, J = 12.1, 3.6 Hz, 1H, major), 3.14 – 2.86 (m, 7H), 2.32 (d, J = 12.5 Hz, 1H, major), 2.19 (d, J = 14.9 Hz, 1H), 2.10 (ddd, J = 18.7, 9.2, 4.2 Hz, 1H), 2.06 (s, 0H), 1.89 (q, J = 7.8 Hz, 2H), 1.81 (dd, J = 13.5, 10.0 Hz, 1H), 0.94 (td, J = 7.4, 3.5 Hz, 6H). <sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 137.3, 136.4, 134.9, 134.6, 133.3, 133.2, 129.9, 129.6, 123.9, 123.4, 121.6, 121.4, 56.9, 54.8, 52.8, 49.0, 46.0, 37.9, 35.0, 32.6, 32.1, 26.5, 24.5, 20.0, 19.6, 11.4, 11.4.

**HMRS (ESI)**: Calculated for  $C_{14} H_{21} O_2 N_2 S = [M+H]^+$ : = 281.13183 found: 281.13189





(<u>+</u>)-9-hydroxy-8,9,10,10a-tetrahydro-7H-benzo[4,5]isothiazolo[2,3-a]pyridine 5,5dioxide (24)



rel. stereochemistry

This product was isolated from reductive amination reaction towards **23c** in significant amount (see procedure above). Only by preparative HPLC, the product mixture could be purified and we found quite some amount of alcohol **24**.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.80 (dd, J = 7.9, 1.1 Hz, 1H), 7.61 (td, J = 7.6, 1.2 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.38 (dd, J = 7.6, 1.0 Hz, 1H), 4.31 (dd, J = 11.9, 3.1 Hz, 1H), 4.00 – 3.88 (m, 2H), 3.08 (td, J = 13.1, 3.0 Hz, 1H), 2.54 (dddd, J = 12.2, 4.6, 3.2, 1.8 Hz, 1H), 2.10 (dq, J = 12.5, 2.4 Hz, 1H), 1.68 (tdd, J = 12.7, 11.2, 5.1 Hz, 1H), 1.45 (q, J = 11.8 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 137.5, 135.2, 132.9, 129.5, 123.1, 121.6, 68.4, 57.5, 39.2, 37.9, 33.4.

**HMRS (ESI)**: Calculated for  $C_{11} H_{14} O_3 N S = [M+H]^+$ : = 240.06889 found: 240.06871





NOE

(<u>+</u>)-9-morpholino-10a-phenyl-8,9,10,10a-tetrahydro-7H-benzo[4,5]isothiazolo[2,3a]pyridine 5,5-dioxide (23d)



**22b** (1.0 eq., 25 mg, 71.8 µmol), morpholine (1.25 eq., 8 µl, 89.8 µmol) and powdered 4 A molecular sieves were combined in dry 1,2 DCE (600 µl) in a two-necked round bottom flask under an Argon atmosphere and sodium triacetoxyborohydride (1.4 eq., 21 mg, 100.5 µmol) and glacial acetic acid (1.05 eq., 4.3 µl, 75.4 µmol) were added and the reaction stirred at ambient temperature. After 16 h additional 2.3 eq. of morpholine (14 µl, 165.14 µmol) and an additional 1.4 eq. of sodium acetoxyborohydride (21 mg, 100.52 µmol) were added to the mixture. After 6 h of additional stirring, the reaction was quenched by addition of 3 ml sat. aq. ammonium chloride solution. The mixture was transferred to a separation funnel, diluted with water (15 ml) and extracted with 3 x 4 ml ethyl acetate. Combined organic layers were washed with brine and dried over anhydrous sodium sulfate. Concentration yielded crude product which was purified by preparative HPLC (10 to 38% Acetonitrile/water (+0.1 % HCOOH).) Lyophilization then delivered product (7.7 mg, 0.02 mmol, 28%).

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*) δ 7.71 (d, *J* = 7.8 Hz, 1H), 7.60 – 7.55 (m, 2H), 7.54 – 7.47 (m, 2H), 7.44 (ddd, *J* = 8.1, 6.8, 1.5 Hz, 1H), 7.27 (dd, *J* = 8.5, 6.9 Hz, 2H), 7.20 (d, *J* = 6.2 Hz, 2H), 3.89 (ddd, *J* = 15.4, 8.6, 1.4 Hz, 1H), 3.82 (s, 4H), 3.20 (ddd, *J* = 15.3, 9.9, 7.3 Hz, 1H), 2.97 – 2.59 (m, 2H), 2.54 (tdd, *J* = 12.0, 8.4, 2.5 Hz, 1H), 2.43 (dq, *J* = 16.6, 8.5, 7.8 Hz, 1H), 1.84 (dt, *J* = 12.5, 8.3 Hz, 1H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 142.7, 141.4, 134.1, 132.6, 129.9, 129.1, 128.3, 125.3, 124.6, 121.4, 69.0, 64.6, 57.9, 49.6, 35.3, 35.2, 23.4.

**HMRS (ESI)**: Calculated for  $C_{21} H_{25} O_3 N_2 S = [M+H]^+$ : = 385.15804 found: 385.15842





(<u>+</u>)-10a-phenyl-9-(propylamino)-8,9,10,10a-tetrahydro-7H-benzo[4,5]isothiazolo[2,3a]pyridine 5,5-dioxide (23e)



**22b** (1.0 eq., 25 mg, 71.80 µmol), n-propylamine (1.25 eq., 7.43 µl, 89.75 µmol) and powdered 4 A molecular sieves were combined in dry 1,2 DCE (600 µl) in a two-necked round bottom flask under an Argon atmosphere and sodium triacetoxyborohydride (1.4 eq., 21 mg, 100.52 µmol) and glacial acetic acid (1.05 eq., 4.312 µl, 75.39 µmol) were added and the reaction stirred at ambient temperature. After 16 h, additional 2.5 eq. of n-propylamine (15 µl, 179.50 µmol) and an additional 1.4 eq. of sodium acetoxyborohydride (21 mg, 100.52 µmol) were added to the mixture. After 1 d of additional stirring, reaction was concentrated under reduced pressure and the residue was purified by preparative HPLC (C18, 10% to 38% MeCN/H<sub>2</sub>O (+0.1% v/v TFA)) to yield 5 mg (20%, 14.03 µmol) of product amine.

<sup>1</sup>**H NMR** (500 MHz, Chloroform-*d*)  $\delta$  7.77 (dd, *J* = 7.7, 1.2 Hz, 1H), 7.61 (dd, *J* = 7.8, 1.6 Hz, 2H), 7.53 (td, *J* = 7.6, 1.5 Hz, 1H), 7.48 (td, *J* = 7.5, 1.0 Hz, 1H), 7.43 (d, *J* = 7.8 Hz, 1H), 7.34 (t, *J* = 7.5 Hz, 2H), 7.28 (d, *J* = 7.2 Hz, 1H), 3.92 – 3.85 (m, 1H), 3.30 – 3.21 (m, 1H), 2.90 (q, *J* = 9.7, 8.2 Hz, 3H), 2.70 – 2.51 (m, 2H), 2.48 – 2.36 (m, 1H), 1.78 (dd, *J* = 13.3, 6.6 Hz, 1H), 1.56 – 1.43 (m, 2H), 0.79 (t, *J* = 7.4 Hz, 3H).

<sup>13</sup>**C NMR** (126 MHz, Chloroform-d) δ 142.1, 133.8, 132.5, 129.8, 129.3 (3 C), 128.5, 125.4, 124.5, 121.5, 57.5, 50.0, 47.6, 37.9, 37.1, 34.4, 25.8, 11.4.

**HMRS (ESI)**: Calculated for  $C_{20} H_{25} O_2 N_2 S = [M+H]^+$ : = 357.16313 found: 357.16270





# Cheminformatic Evaluation of compound Collection Distribution of compound collection over chemical space (PMI)

Nelson's group developed the LLAMA platform which allows scientists to evaluate compounds according to their suitability for medicinal chemistry research after different parameters.<sup>[11]</sup> They also offer a principal moment of inertia (PMI) plot. Making use of this feature we could generate Supp. Figure 1 by overlay of different compounds. Position of different members of compound collection displays, in how far its three-dimensional structure resembles either rod (top left), sphere (top right) or disk (bottom).



**Figure S1**:PMI plot of Compound collection. "+" denotes averaged center position in PMI plot.<sup>[11]</sup> Black squares show relative positions of compounds in comparison to commonly known bioactive sulfonamides (green, see also examples in Figure 1 of paper). Aziridines and azetidines, in particular are denoted by blue and light-blue squares, respectively. Azetidines are highlighted in margenta while their ring-expansion and derivatization products are depicted as orange squares.

## 3.2 Predicted molecular properties of compounds



**Figure S2:** 3D Scatter Plot of clogp, tPSA and molweight as parameters that may be beneficial for bioavailability.

The figure shows synthesized compounds as dots according to their projective molecular properties: topological polar surface area (<sup>*t*</sup>PSA, should be less than 140 or 90, if availability to central nervous system is required), clogp (should be roughly between -1 and 4), both displaying a degree of how well compounds pass through plasma membrane and molecular weight, which should be generally less than 500 Da. All of compounds fit these requirements.
## 4. Biological Evaluation

## 4.1 Supporting Figures



**Figure S3:** Activity (induction) of benzosulfonamides in the cell painting assay. Compounds were subjected to Cell painting assay performed in U2OS cells at a concentration of 10  $\mu$ M. The induction is a measure of biological activity and is determined as the ratio of the number of parameters significantly different from the control vs. the total number of parameters.



Figure S4. Structures of fenbendazole<sup>[13]</sup> (a), tubulexin A<sup>[14]</sup> (b) and the PLK1 inhibitor<sup>[15]</sup> (c).



Figure S5. Fingerprint comparison for 5b (10  $\mu$ M) with the fingerprint for a PLK1 inhibitor (PLK1 inh. 2  $\mu$ M).



**Figure S6.** Benzosulfonamides 4a and 5b lead to accumulation of round cells. U2OS cells were treated with the compounds or DMSO followed by live-cell imaging. Images of cell after 24 h of treatment with 4a, 5b or DMSO as a control are shown. Scale bar:  $300 \mu m$ .



Figure S7. Benzosulfonamides 4a and 5b lead to accumulation of phospho-histone 3positive cells. U2OS cells were treated with the compounds or DMSO for 24 h prior to staining for phospho-histone 3 as a marker for mitotic arrest and DNA using DAPI. Scale bar: 50  $\mu$ m.

#### 4.2 Experimental section - Biology

#### Reagents

DAPI (4',6-Diamidin-2-phenylindol, #10236276001), anti-tubulin FITC (Cat# F2168;) and Hoechst 33342 (Cat. No. B2261-25mg) were purchased from Sigma Aldrich. Anti-phosphohistone3 antibody was obtained from Abcam (Cat# ab5176). Tubulin purified from porcine brain was purchased from Cytoskeleton, Inc (#T240). Human bone osteosarcoma epithelial *cells* U2OS were obtained from CLS (Cat# 300364; RRID: CVCL\_0042). DMEM, sodium pyruvate and non-essential amino acid obtained from PAN Biotech, and fetal bovine serum (FBS) was purchased from Gibco. Mito Tracker Deep Red (Cat. No. M22426), Phalloidin (A12381), Concanavalin A (Cat. No. C11252), WGA-Alexa594 conjugate (Cat. No. W11262) and µl/ml SYTO 14 solution (Cat. No. S7576) were obtained from Thermo Fisher Scientific.

#### Cell culture

U2OS cells were maintained in DMEM medium supplemented with 10 % fetal bovine serum (FBS) at 37 °C and 5% CO<sub>2</sub> in a humidified atmosphere. Cells were regularly assayed for mycoplasma contamination and were always confirmed to be free of mycoplasma.

#### **Cell painting assay**

The cell painting assay<sup>[12]</sup> was carried out as described by and Bray *et al.*<sup>[17]</sup> with some modifications<sup>[18]</sup>. Initially, 5 µl U2OS medium were added to each well of a 384-well plate (PerkinElmer CellCarrier-384 Ultra). Subsequently, 1600 U2OS cells were seeded per well in 20 µl medium. The plate was incubated for 10 min at the room temperature prior to incubation for 4 h at 37 °C. Compounds were then added with the Echo 520 acoustic dispenser (Labcyte) and cells were incubation for 20 h at 37 °C. Subsequently, mitochondria were stained with Mito Tracker Deep Red for 30 min in the dark at 37 °C. Cells were fixed using 3.7 % formaldehyde (in PBS) for 20 min in the dark at 37 °C prior to permeabilization Triton X-100

for 15 min 37 °C in the dark. After three washing steps, cells were stained with Alexa Fluor 594 Phalloidin, Concanavalin A Alexa Fluor 488, Hoechst 33342, WGA-Alexa594 conjugate and SYTO 14. The plate was incubated for 30 min at 37 °C in the dark and washed three times with PBS. After the final washing step, the PBS was not aspirated. The plates were sealed and centrifuged for 1 min at 500 rpm.

The plates were prepared in triplicates with shifted layouts to reduce plate effects and imaged using a Micro XL High-Content Screening System (Molecular Devices) in 5 channels (DAPI: Ex350-400/ Em410-480; FITC: Ex470-500/ Em510-540; Spectrum Gold: Ex520-545/ Em560-585; TxRed: Ex535-585/ Em600-650; Cy5: Ex605-650/ Em670-715) with 9 sites per well and 20x magnification (binning 2).



Figure S8: Image generation from Cell painting Assay.

The generated images were processed with the CellProfiler package (https://cellprofiler.org/) on a computing cluster of the Max Planck Society to extract 1716 cell features (parameters). The data was then further aggregated as medians per well (9 sites -> 1 well), then over the three replicates.

Further analysis was performed with custom Python (https://www.python.org/) scripts using the Pandas (https://pandas.pydata.org/) and Dask (https://dask.org/) data processing libraries (separate publication to follow).

From the total set of 1716 parameters a subset of highly reproducible and robust parameters was determined using the procedure described by Woehrmann et al.<sup>[19]</sup> in the following way: Two biological repeats of one plate containing reference compounds were analysed. For every parameter, its full profile over each whole plate was calculated. If the profiles from the two repeats showed a similarity >= 0.8 (see below), the parameter was added to the set.

This procedure was only performed once and resulted in a set of 579 robust parameters out of the total of 1716 that was used for all further analyses.

# Determination of reproducible Parameters

1716	Determined by CellProfiler	
ŧ	Keep parameters that have a minimum correlation of 0.80 between repeats for all cpds.	
579	Final set of relevant parameters. Used for all further analyses	

Figure S9: Determination of reproducible parameters in cell painting assay.

To determine the phenotypic profiles for each test compound Z-scores were then calculated for each parameter as how many times the Median Absolute Deviation (MAD) of the controls the measured parameter value of a test compound deviates from the Median of the controls:



$$z - score = \frac{value_{meas.} - Median_{Controls}}{MAD_{Controls}}$$

The phenotypic compound profile is then determined as the list of z-scores of all parameters for one compound.

In addition to the phenotypic profile, an induction value was determined for each compound as the fraction of significantly changed parameters, in percent:

Induction  $[\%] = \frac{number of parameters with abs. values > 3}{total number of parameters}$ 

Similarities of phenotypic profiles were calculated from the correlation distances between two profiles

(https://docs.scipy.org/doc/scipy/reference/generated/scipy.spatial.distance.correlation.html;

Similarity = 1 - Correlation Distance) and the compounds with the most similar profiles were determined from a set of 3000 reference compounds that was also measured in the assay.

An example for two compounds with highly similar profiles (96% similarity):



An example for two compounds with low similarity profiles (0% similarity):



Each colored band represents one Z-score of a parameter.

#### Immunocytochemistry

U2OS cells were seeded per well in a 96-well plate and incubated overnight. Cells were treated with compounds or DMSO as a control for 24 hours. Cells were then fixed using 3.7% paraformaldehyde in phosphate-buffered saline. PBS and permeabilized with 0.1% Triton X-100 (in PBS) prior to staining with DAPI to visualize DNA and anti-tubulin-FITC antibody or anti-phosphor-Histone3 antibody antibodies overnight at 4 C°. Images were acquired using Observer Z1 (Carl Zeiss, Germany) using 20X and 40 X objectives (LD Plan-Neofluar). Automated image analysis to quantify the percentage of phospho-histone3-positive cells was performed using the DAPI stain to assess the total number of cells and the software MetaMorph. Using GraphPad Prism 5, one-way analysis of variance (ANOVA) plus T-test were performed to check the significance of data represented in Figure 1f (\*\*\*, P-value < 0.0001).

#### In vitro tubulin polymerization assay:

*In vitro* tubulin polymerization assay was performed as described previously.<sup>[14]</sup> Briefly, porcine  $\alpha/\beta$ -tubulin was diluted in a general buffer containing 80 mM PIPES (pH 6.9), 2 mM MgCl<sub>2</sub> and 0.5 mM EGTA. Next, 10  $\mu$ M of  $\alpha/\beta$ -tubulin was added to a solution containing MgCl<sub>2</sub> and glutamate with a final concentration of 0.88  $\mu$ M and 0.8 mM, respectively, and added to a 96-well plate. Afterwards, compounds at a final concentration 20  $\mu$ M were added to the tubulin solution and incubated at room temperature for 20 min. The plate was then incubated on ice for another 20 min prior to addition of GTP to a final concentration of 500  $\mu$ M. Tubulin polymerization was monitored for 60 min by means of turbidity measurements at 340 nm.

#### Real-time live-cell analysis

Cell growth was monitored by means of real-time live-cell analysis using the IncuCyte S3 (Essen Bioscience). For this, 4000 U2OS cells were seeded per well in 96-well plate and incubated overnight. The medium was then exchanged for fresh medium that contained the compounds or DMSO as a control. Cells were incubated for 48 h and cell growth were imaged

every two hours using in the bright-field mode. Cell confluence was quantified as a measure of cell growth using the IncuCyte S3 2019A software.

# 5. X-Ray Structures of Compounds 15a and syn-10

$$O_{NH}$$
  
 $Ph$   
 $CO_2Et$   
15a

# Crystal data and structure refinement for 15a.

Identification code	pbca		
Empirical formula	$C_{19}H_{21}NO_4S$		
Formula weight	359.43		
Temperature/K	100		
Crystal system	orthorhombic		
Space group	Pbca		
a/Å	16.4445(14)		
b/Å	9.7793(8)		
c/Å	21.9616(18)		
$\alpha/^{\circ}$	90		
β/°	90		
$\gamma^{/\circ}$	90		
Volume/Å <sup>3</sup>	3531.8(5)		
Z	8		
$\rho_{calc}g/cm^3$	1.352		
$\mu/mm^{-1}$	0.207		
F(000)	1520.0		
Crystal size/mm <sup>3</sup>	$0.277\times0.116\times0.059$		
Radiation	MoKa ( $\lambda = 0.71073$ )		
$2\Theta$ range for data collection/° 6.104 to 53.998			
Index ranges	$-21 \le h \le 21,  -12 \le k \le 12,  -28 \le l \le 28$		
Reflections collected	82730		
Independent reflections	$3852 [R_{int} = 0.0453, R_{sigma} = 0.0160]$		
Data/restraints/parameters	3852/0/251		
Goodness-of-fit on F <sup>2</sup>	1.075		
Final R indexes [I>= $2\sigma$ (I)]	$R_1 = 0.0417, wR_2 = 0.0946$		
Final R indexes [all data]	$R_1 = 0.0493, wR_2 = 0.0985$		
Largest diff. peak/hole / e Å <sup>-3</sup> 0.49/-0.50			



Identification code	C0076_0m
Empirical formula	$C_{16}H_{20}N_2O_6S$
Formula weight	368.40
Temperature/K	100.0
Crystal system	monoclinic
Space group	P21/n
a/Å	7.7558(3)
b/Å	10.0904(4)
c/Å	22.1316(10)
$\alpha/^{\circ}$	90
β/°	98.937(2)
γ/°	90
Volume/Å <sup>3</sup>	1710.97(12)
Z	4
$\rho_{calc}g/cm^3$	1.430
$\mu/\text{mm}^{-1}$	0.225
F(000)	776.0
Crystal size/mm <sup>3</sup>	$0.937 \times 0.388 \times 0.274$
Radiation	MoKa ( $\lambda = 0.71073$ )
$2\Theta$ range for data collection/°	5.494 to 77.996
Index ranges	$\text{-13} \leq h \leq \text{13},  \text{-17} \leq k \leq \text{17},  \text{-39} \leq \text{I} \leq \text{39}$
Reflections collected	83416
Independent reflections	9831 [ $R_{int} = 0.0298$ , $R_{sigma} = 0.0188$ ]
Data/restraints/parameters	9831/0/234
Goodness-of-fit on F <sup>2</sup>	1.094
Final R indexes [I>= $2\sigma$ (I)]	$R_1 = 0.0294, wR_2 = 0.0828$
Final R indexes [all data]	$R_1 = 0.0317, wR_2 = 0.0848$
Largest diff. peak/hole / e Å $^{\text{-}3}$	0.68/-0.43

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## 5. Copies of NMR Spectra
























































































S127




























































S157




























































S187











S192











S197









S202







