1 Supplemental Figure Legends

2 Figure S1. Validation of CD11c Itgax-DTR chimeras. (A) Representative flow plots of

3 chimerism with Itgax DTR and host cells in uninfected, untreated chimeras. (B) Quantification

4 of chimerism with Itgax DTR and host cells in uninfected, untreated chimeras. (C)

5 Representative flow plots and quantification of total monocytes and monocyte subsets in the lung

6 10 days post infection showing lack of depletion of monocytes in DTx treated animals. (D)

7 Example staining of D14 post-infection DTx- and PBS-treated chimeras, showing CD69 and

8 CD103 staining on flu-specific CD8 T cells in the lung. (E) Number of FluNP-specific CD8 T

9 cells in the extra-vascular compartment of the lung (LEV), CD69+ CD103+ cells of the extra-

10 vascular compartment (LRM) and airways (BAL) on day 14 post infection.

Figure S2. Gating strategy for pulmonary APCs. (A)Representative flow plots and gating strategy for the identification of extravascular pulmonary APC subsets of C57BL/6J mice intranasally infected with x31 influenza virus. Discrimination of vascular versus extra vascular cells via i.v. staining with anti-CD45.2 was performed in each experiment as shown. plots are representative of 3 independent experiments with 5 mice used in each experiment.

16 Figure S3. Number of APCs in the lung of WT and CCR2-/- mice 10 days post infection

17 with x31 influenza virus. (A)Representative flow plots of APCs present in WT and CCR2-/-

18 mouse lungs on day 10 post-infection. (B) Number of extra-vascular classical and non-classical

19 monocytes (left graph) or DC subsets (right graph) in the lungs of WT and CCR2-/- mice on day

- 20 10 post-infection. All graphs error bars are S.E.M. Data are representative of 3 independent
- experiments with 5 mice per time point. * p < 0.05 n.s. p > 0.05(two-tailed Student's *t*-test).

22 Figure S4. Depletion of non-classical monocytes in CX3CR1-DTR mice has no effect upon

23 lung T_{RM} generation following influenza infection. (A) Graph and representative flow plots of

1	the frequency of total lymphocytes in the lung and spleen derived from donor bone marrow
2	versus host lymphocytes 6 weeks after chimera production. Gated to exclude CD45- singlet
3	lymphocytes. (B) Graph and representative flow plots of the frequency of classical and non-
4	classical monocytes in the lung and spleen of chimeras treated with DTx or PBS 10 days post
5	infection. Plots are gated on MHC-II lo, Ly6g-, CD11b+, CD11c- singlet lymphocytes.(C) Graph
6	of the number of OT-I CD8 T cells day 45 post infection in PBS and DTx treated mice in the
7	lung extra-vascular compartment (LEV), bronchoalveolar lavage (BAL), and spleen, and graph
8	of the number of CD69+ CD103+ OT-I CD8 T cells day 45 post infection in PBS and DTx
9	treated mice in the lung-extra vascular compartment and bronchoalveolar lavage. All error bars
10	are S.E.M. * p<0.05 n.s. p>0.05(two-tailed Student's <i>t</i> -test) Data are representative of two
11	independent experiments.

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Figure S5. Phenotype of OT-I T cells after three days of in vitro stimulation with lung APC 13 subsets. (A) Representative flow plots of CD127 staining on CD8 OT-I T cells following three 14 days of *in vitro* culture with OVA peptide pulsed monocytes of indicated subtypes. (B) 15 Frequencies of CD127 positivity among highly divided CD8 OT-I T cells following three days of 16 in vitro culture with OVA peptide pulsed monocytes of indicated subtypes. Error bars SEM, data 17 represents 3 experiments using 3 technical replicates *in vitro* each. (C) Representative flow plots 18 of CD103+ staining on CD8 OT-I T cells following three days of in vitro culture with OVA 19 peptide pulsed monocytes of indicated subtypes. (D) Representative flow plots of CD69+ 20 staining on CD8 OT-I T cells following three days of in vitro culture with OVA peptide pulsed 21

22 monocytes of indicated subtypes.

d

CD69

CD45.1 (Host)







DTx



PBS



D14 Post Infection









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