

# 1 **Supplementary Results**

## 2 ***Simulated species responses to media variation recapitulate experimental results***

3 We ran several sets of simulations with the same set of 61 initial species compositions  
4 as the main 10-species dataset but with small amounts of stochastic noise added to the  
5 nutrient inflow. Specifically, inflow concentrations for each compound in each simulation  
6 were sampled from a normal distribution with a mean equal to the compound's original  
7 inflow rate and a standard deviation set to a particular fraction of the mean (Methods).  
8 We then examined whether the differences in simulated species growth across  
9 simulations with large media fluctuations (8-10%) recapitulated experimental  
10 observations from Faith *et al.* (1). Overall, we found agreement in several observed  
11 trends. First, Faith *et al.* observed that casein abundance was strongly associated with  
12 the total community biomass (presumably, due to limiting amino acid or nitrogen  
13 concentrations). In our simulations, the total amount of amino acids is indeed positively  
14 associated with total biomass (Spearman  $\rho=0.25$ ,  $p=0.005$ ). Moreover, we fit linear  
15 regression models of total biomass across all simulations based on inflow metabolite  
16 levels, and found that concentrations of both amino acids and carbohydrates explain  
17 most of the variation in this quantity in our simulations (adjusted  $R^2$  of 0.97), with other  
18 nutrients adding only negligible effects, similar to the results of Faith *et al.* Faith *et al.*  
19 further observed that while most species increase their growth rate in the presence of  
20 higher protein; *E. rectale*, *D. piger* and *M. formatexigens* are negatively associated with  
21 the amount of casein in the mouse diet. In our simulations, a model predicting growth  
22 rate based on amino acid levels explains no variation in *E. rectale* and only 8% in *M.*

23 *formatexigens*, in contrast to other taxa, for which amino acids explained up to 54% of  
24 growth rate variation. Additionally, the initial growth rates for all species were positively  
25 correlated (Spearman,  $p < 0.1$ ) with an average of 2.7 different amino acid compounds  
26 (range 1 to 6), while *E. rectale* and *M. formatexigens* were each only correlated with a  
27 single amino acid, *L*-cysteinyglycine. Lastly, our simulations also recapitulate  
28 differences in carbohydrate use: Faith *et al.* observed preferential expansion of *B.*  
29 *ovatus* and *B. thetaiotaomicron* on a high-starch diet compared to a high-sugar diet. In  
30 our simulations, the growth rate of all species was associated with the amount of  
31 available simple sugars, but only *B. ovatus*, *B. thetaiotaomicron*, and *E. rectale* were  
32 significantly correlated with the quantity of starch in the inflow (Spearman rho  
33 coefficients of 0.5, 0.53, 0.53 respectively, all  $p < 10^{-9}$ ). These findings suggest that our  
34 simulation framework successfully encapsulates at least some of the nutrient limitations  
35 shaping the growth dynamics of this model community.

### 36 ***Analysis of an alternative definition of contribution values based on steady-state*** 37 ***fluxes***

38 Our contribution value metric attributes metabolite variance to each species depending  
39 on its cumulative metabolite uptake or secretion over the entire simulation, rather than  
40 its arrived-at steady-state metabolite flux at the time of “sampling”. To assess the  
41 impacts of this choice, we calculated contribution values using an alternative definition  
42 based solely on steady-state fluxes. Specifically, we calculated the contribution of each  
43 species to the metabolite flux at the final time point of simulations run for 144 hours and  
44 for 1440 hours, using the same 61 initial species compositions as the main 10-species  
45 dataset and identical nutrient inflow. Under this definition, steady-state contribution

46 values explain the variation in metabolite flux rate at the time of sampling, rather than  
47 the accumulated variation in metabolite concentrations (cumulative contribution values).  
48 We compared these alternative steady-state contributions to the original set of  
49 cumulative contribution values at both time points, finding that they are highly similar.  
50 Specifically, in our original dataset of simulations run for 144 hours, the Pearson  
51 correlation between steady-state and cumulative contribution values for each metabolite  
52 was on average 0.99 (minimum of 0.75, median of 0.999 across all metabolites). Only 6  
53 of the 520 analyzed species-metabolite pairs differ in contributor status between the two  
54 definitions: 4 pairs are key cumulative contributors but not steady-state contributors, and  
55 2 pairs are the reverse. The AUC for detection of steady-state contributors is 0.710  
56 (compared with 0.717 for cumulative contributors). These differences were even more  
57 negligible for simulations run for longer durations: in a dataset of simulations run for  
58 1440 hours, the average metabolite-level correlation between steady-state and  
59 cumulative contribution values was 0.9999 (minimum 0.9997, median of 0.99999).  
60 These results indicate that for these simulations, differences in species composition and  
61 metabolic activity early in the simulation are *not* the predominant factor in the observed  
62 discrepancy between species-metabolite correlations and true key contributors to  
63 metabolic variation.

64

#### 65 ***Analysis of an alternative definition of key taxon-metabolite pairs***

66 For most analyses, we defined the key taxonomic contributors for a particular metabolite  
67 as those species with the highest positive contribution values, or those that are  
68 responsible for the observed pattern of variation in a metabolite. However, an  
69 alternative goal could be to detect all microbes that substantially impact levels of a given

70 metabolite across samples, regardless of whether their effects are ultimately reflected in  
71 the observed concentrations. To this end, we defined *key player* species as those with a  
72 contribution value, either positive or negative, greater in magnitude than 20% of the total  
73 contribution magnitude. This resulted in 91 species-metabolite key player pairs,  
74 including 65 of the previously defined 'positive' key contributor pairs but also 26 players  
75 with negative contributions, which were distributed similarly across metabolites and  
76 species (Figure S5, panels A-B). Examining how well these key players were detected  
77 by a correlation-based analysis, we found similar performance as for key contributors  
78 (Figure S5, panels C-G), including a comparable positive predictive value (31.9%) and  
79 AUC (0.73).

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### 81 ***Effects of simulation length and Vmax parameter on correlation results***

82 We assessed the sensitivity of our correlation results to the parameters used in our  
83 simulations. Specifically, we first evaluated the effect of the duration of simulations on  
84 our results. We ran additional simulations of the same 61 10-species compositions for  
85 5,760 time points (or 1,440 hours), and calculated contribution values and correlation  
86 coefficients at 22 intermediate time points starting at 36 hours (Figure S7). Species  
87 compositions and metabolite concentrations became increasingly less variable with  
88 longer simulation time, converging towards similar steady states dominated in  
89 abundance by 5 of the 10 species (Figure S7A-B). Correspondingly, the number of key  
90 contributors decreased with increasing simulation length, from 121 contributors across  
91 all 52 analyzed metabolites at 36 hours, to 75 at 1,440 hours (Figure S7B-C). The  
92 number of significantly correlated species-metabolite pairs, however, increased from  
93 179 to 375 over the same datasets, detecting contributors with higher sensitivity but

94 lower specificity (Figure S7D). Ultimately, the AUC and positive predictive value both  
95 decrease slightly with increasing simulation length, with the AUC shifting from 0.67 to  
96 0.73 and positive predictive value from 39.7% to 18.4% (Figure S7E). This transition  
97 occurs sharply initially before reaching an inflection point and beginning to stabilize  
98 around 144 hours, the length of time chosen for our main analysis.

99

100 We also generated additional datasets with the same initial species compositions but  
101 with widely varied values for the universal  $V_{max}$  parameter, which was set to 20 in the  
102 main set of analyses. Changing this parameter had very minimal impact on both the  
103 simulation abundance profiles and the results of correlation analysis (Figure S7F-H).  
104 The AUC for the identification of key contributors was not associated with the value of  
105 the  $V_{max}$  parameter, and only ranged from 0.70 to 0.72.

106

107 ***Features distinguishing true key contributors from false positives among***  
108 ***correlated pairs***

109 We constructed additional regression models to assess whether there are features that  
110 can distinguish true key contributors from false positives among all correlated species-  
111 metabolite pairs. We fit regression models to similarly assess whether species and/or  
112 metabolite identity are indicative of whether a correlated species-metabolite pair  
113 represents a true or false positive relationship. We found that species identity ( $p =$   
114 0.047), but not metabolite identity, was predictive of key contributor status among  
115 correlated pairs. This is unsurprising given that the number of key contributions from  
116 each species varied widely, while all metabolites have at least one key contributor.

117

118 ***Additional effects of inflow fluctuations on contribution and correlation profiles***

119 We assessed whether the addition of external metabolite fluctuations impacted the  
120 profile of species contributing to each metabolite. For most metabolites (28 out of 52,  
121 including 12 out of 14 non-inflow metabolites), the top microbial contributor did not  
122 change across all levels of fluctuation. However, for many inflow metabolites, the large  
123 external fluctuations can result in a switch of contribution value sign. In these cases,  
124 activity by a microbe that contributed to variation in a constant-inflow setting instead has  
125 a mitigating impact, resulting in a negative contribution. Of the 65 key contributors to  
126 variation in inflow metabolites in the original dataset, 34 (52%) had a negative  
127 contribution value in at least one simulation run with external fluctuations. This  
128 observation highlights that our definition of key contributors is context-dependent,  
129 identifying the entities primarily responsible for the observed variation.

130

131 We also examined whether the detection of the 14 variable metabolites not present in  
132 the nutrient inflow was affected by random fluctuations in inflow metabolites. Variation in  
133 8 of these metabolites was significantly positively correlated with variation in the  
134 surrounding inflow (Spearman rho,  $p < 0.01$ ), suggesting that their synthesis fluxes were  
135 affected by changes in microbial growth or nutrient usage that resulted from  
136 environmental shifts. Correlation analysis tended to identify key contributors for these  
137 metabolites with slightly higher specificity and lower sensitivity as inflow fluctuations  
138 increased (Figure S8).

139 **Supplementary References**

- 140 1. Faith JJ, McNulty NP, Rey FE, Gordon JI. 2011. Predicting a Human Gut  
141 Microbiota's Response to Diet in Gnotobiotic Mice. *Science* 333:101–104.

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