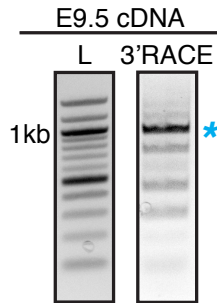


**A**



**B**

**\* Predominant Handlr Isoform \***

**Exon 1**

TCTTCTCTCTGCACTGTCTCTCCTTTGTGCATCTTGGACTCCTGAAGGCCACTCAGAGCATA  
GATGCG

**Exon 2**

CCAGCTTAGTTCTTTGGAGAAGCAGTGTGGAGTCCTAAAAAAGGAGGCTGAGTCTTACCGT  
TGAGGTCGGTGCCGGTAGAGAATGGGAGGGAGTCTGCAGGAGCCAAGCACCCCTAAAGAAG  
AGGAAGAGAGCACGTTGCTGAGATTTGAACAGCGACCTGAGAGGTCAGACTGGACCCAGG  
CCTCCTCAAG

**Exon 3**

AAGGAACGGAGATGGAGATTCTTCCCTTTTCCCGGCCCTTCGGTCCTTCTCCGGAAGAGCT  
ACTCTGGCCAGCTCTGGGCTAGGGGCAGTGGCCGAACAGAAGGGGAAA

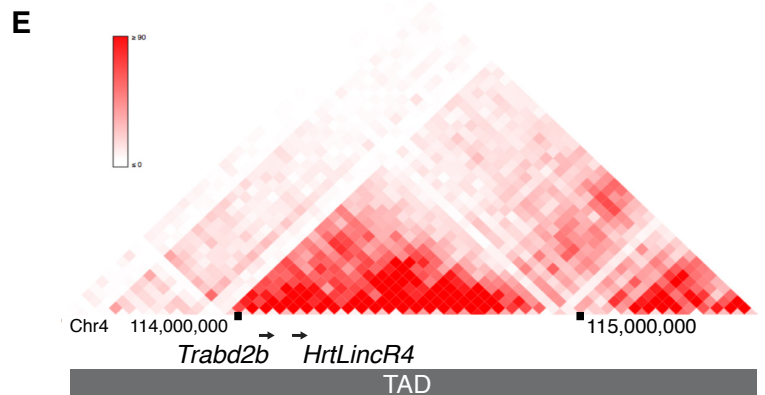
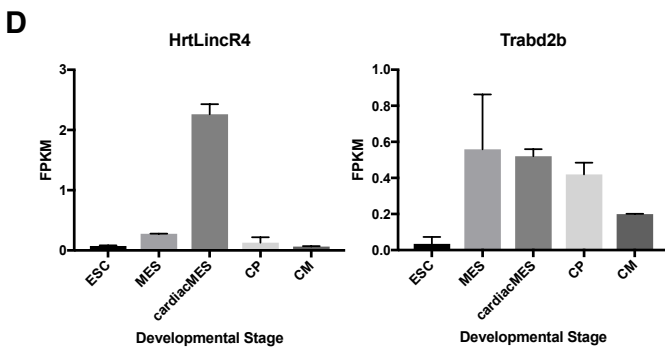
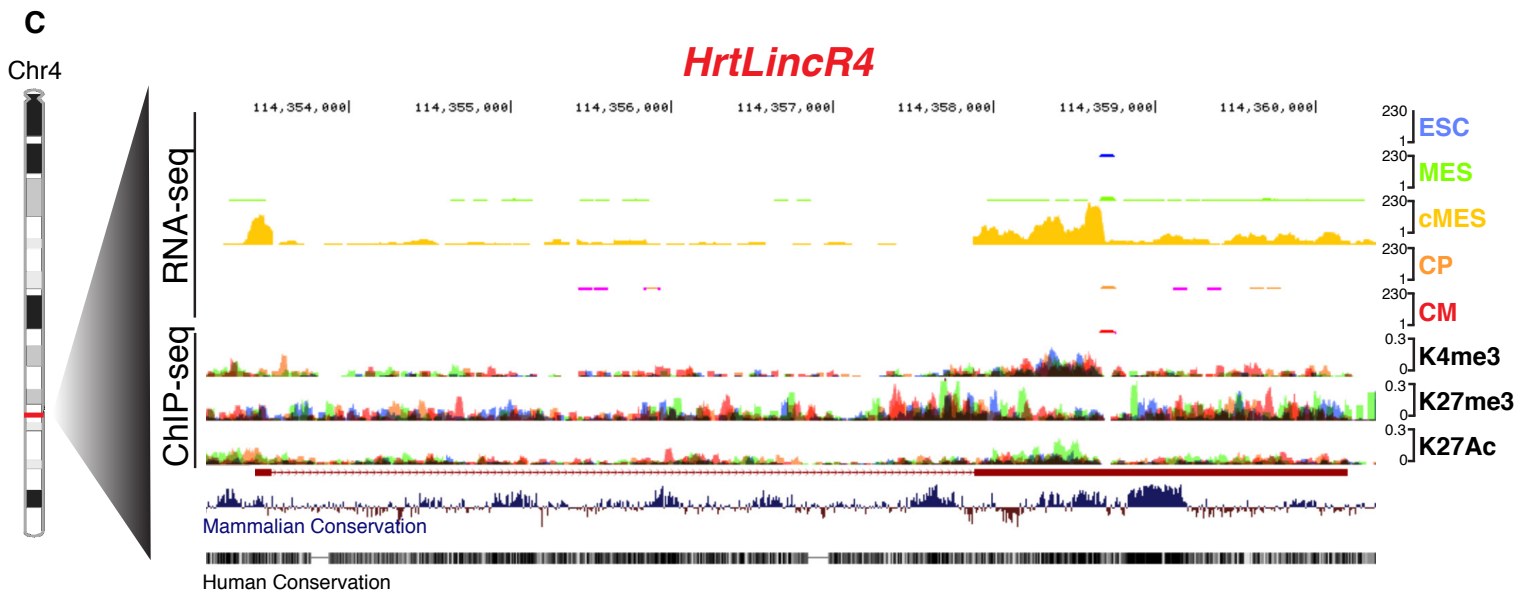
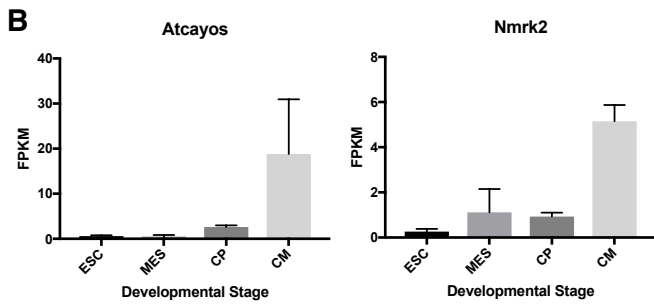
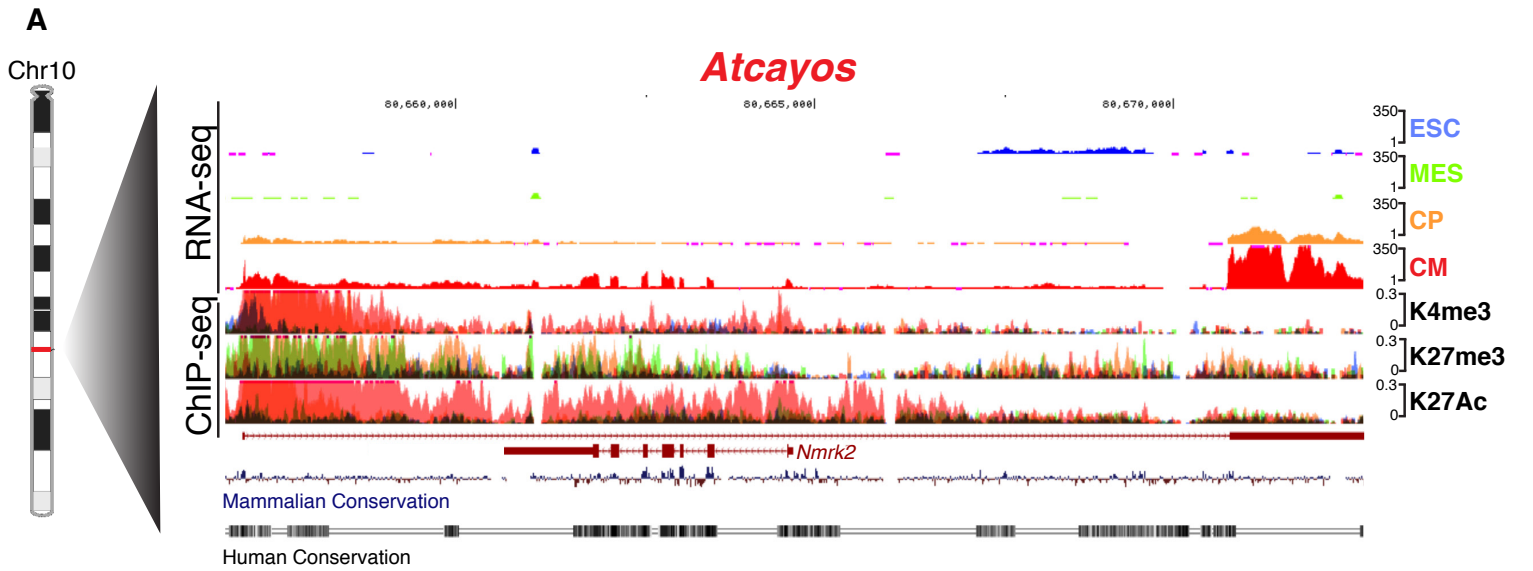
**Exon 4**

GTTTTAGGTTTTTGCCTGTTGCTCTTAGGGATCCAGGGGCAGAGTTTTGCCTTAGGTGCTG  
AAG

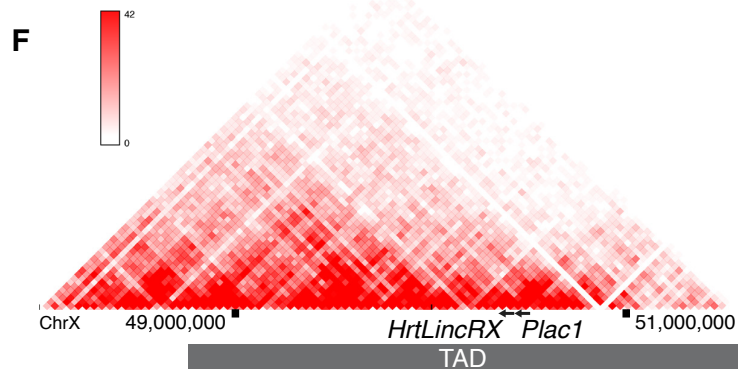
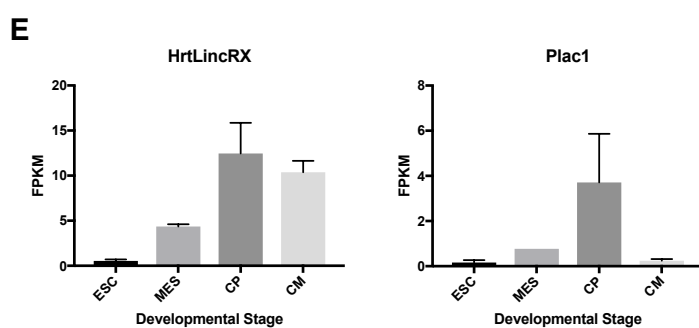
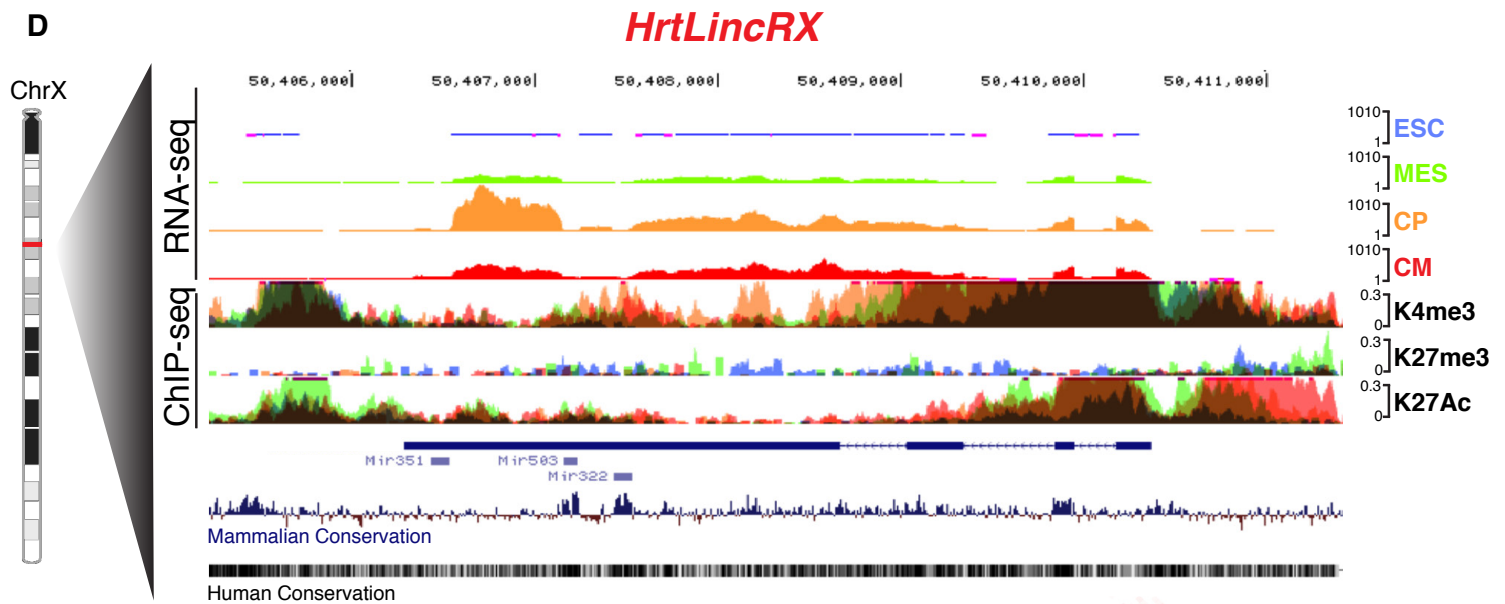
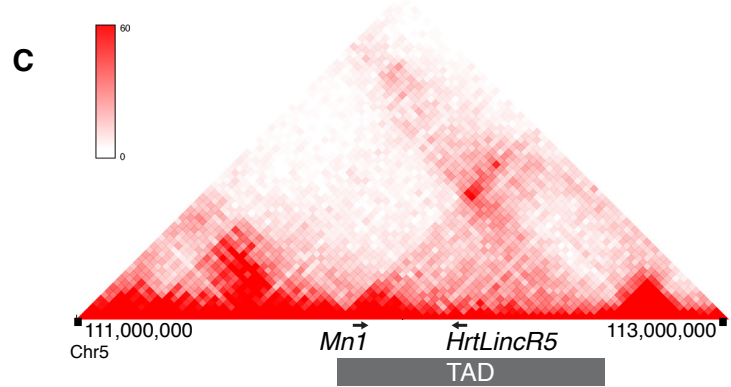
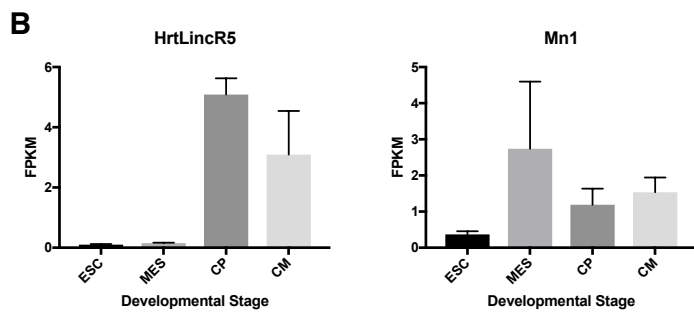
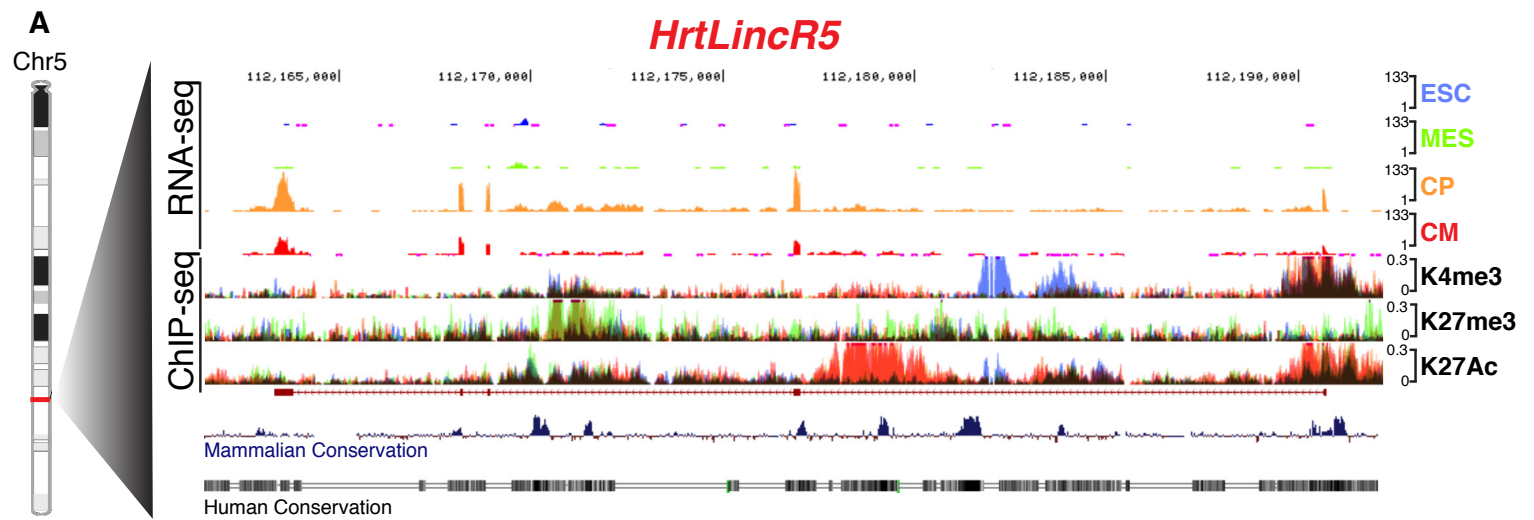
**Exon 5**

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AGAAAACAGTTCTAATTCAAAGATGCTGGATCTGGCTTCCTTTAAAGCCTGAAGTCGCTGAA  
GTGGCCATAGA ACTATGGGGTCCAGAGGACTGCCAGTCTCAGCCAGCCTTTAGGCTGGAGA  
GAGGCTGTGGGCCTGCTTGGGGACTAAAATGGATGAATAAGAAACCTGTAAGAAATGTAAAC  
AGATGGAATGGCAAGGAGGAGAGCGCTGGAGATTGTTTCAGGTGTGAGCTGAGGTTTCTGA  
GTTTGTACTGGGGATGCCTTTGCACACAGTGAGGTGCCCCCTCCCTAAGTGGCTCTTGTTTT  
GCACAAACAGCCCCACCCTGCGCACACACTGTGGGTCTCAGTAGGGACTGAGGGTGTAGG  
TGCTCAGGTTTTGGGGTGGTGGTGAAGGGGCACTCAGAATCAACAAGACAGGGCGGGGA  
AAATCAAATCAACCAAGGAACCAGCCTTCCATTTCTGAAAAGCCGCGCTCTGNNCCCGTG  
CTTCAGTTCTGAAATACGAATAAATGGNGTCAGATCCCGTACTACAAAAAAAAAAAAAAAAA

**Figure S1. Predominant *Handlr* transcript *in vivo*.** A. Electrophoresed *Handlr* 3' RACE PCR products from E9.5 mouse cDNA. B. Sanger sequence of predominant RNA transcript, polyA tail highlighted; T in place of U due to DNA sequencing; arrow, location of 3'RACE *Handlr*-specific primer.



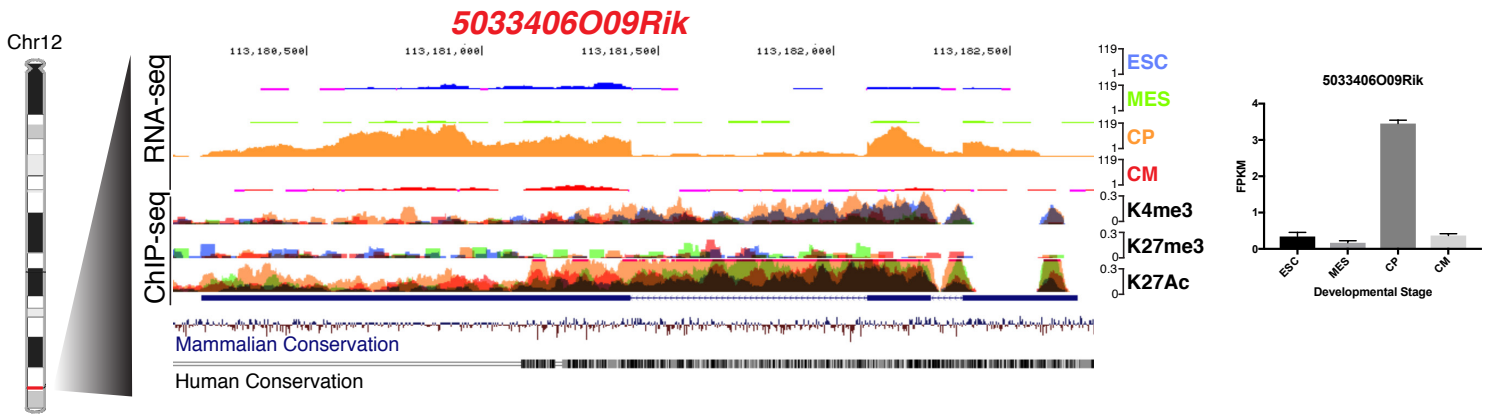
**Figure S2. Genomic characterization of *Atcayos* and *HrtLincR4* *in vitro*.** A. UCSC Genome Browser tracks of *Atcayos* and *Nmrk2* RNA-seq and overlaid histone H3 ChIP-seq at ESC, MES, CP, and CM stages of *in vitro* differentiation. B. Quantified expression of *Atcayos* and *Nmrk2* at each differentiation stage. C. UCSC Genome Browser tracks of *HrtLincR4* RNA-seq and overlaid histone H3 ChIP-seq during cardiac differentiation *in vitro*; D. Quantified expression of *HrtlincR4* and *Trabd2b* at each differentiation stage. E. 3D Genome Browser Hi-C heatmap of chromosome interactions around *HrtlincR4* and *Trabd2b* loci. ESC, embryonic stem cell, MES, mesoderm, CP, cardiac progenitor, CM, cardiomyocyte; blue, ESC, green, MES, orange, CP, red, CM; K4me3, histone H3 lysine 4 trimethylation; K27me3, histone H3 lysine 27 trimethylation; K27Ac, histone H3 lysine 27 acetylation; TAD, topologically associated domain. Ensembl annotations in red. Data presented as mean +/- S.E.M.



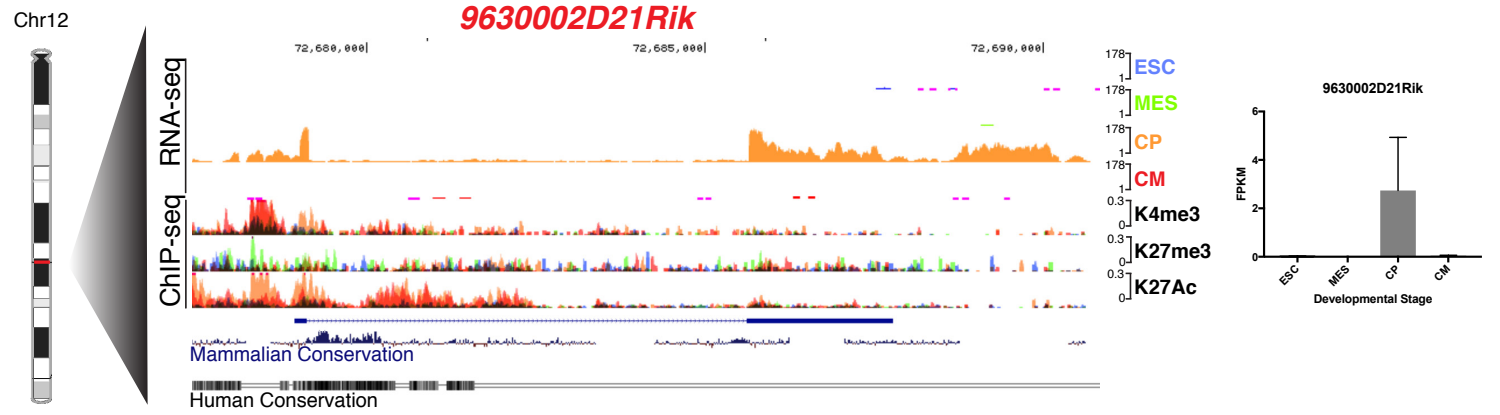
**Figure S3. Genomic characterization of *HrtLincR5* and *HrtLincRX* *in vitro*. A.**

UCSC Genome Browser tracks of *HrtLincR5* RNA-seq and overlaid histone H3 ChIP-seq during cardiac differentiation *in vitro*. Ensembl annotation in red. B. Quantified expression of *HrtlincR5* and *Mn1* at each differentiation stage. C. 3D Genome Browser Hi-C heatmap (of chromosome interactions around *HrtlincR5* and *Mn1* loci. D. Genome Browser tracks of *HrtLincRX* RNA-seq and overlaid histone H3 ChIP-seq during cardiac differentiation *in vitro*; E. Quantified expression of *HrtLincRX* and *Plac1* at each differentiation stage. F. 3D Genome Browser Hi-C heatmap of chromosome interactions around *HrtlincRX* and *Plac1* loci. ESC, embryonic stem cell, MES, mesoderm, cMES, cardiac mesoderm; CP, cardiac progenitor, CM, cardiomyocyte; blue, ESC; green, MES; orange, CP; red, CM; K4me3, histone H3 lysine 4 trimethylation; K27me3, histone H3 lysine 27 trimethylation; K27Ac, histone H3 lysine 27 acetylation; RefSeq annotation in blue. RefSeq annotations, including 3' miRNA cluster, in blue. Data presented as mean +/- S.E.M.

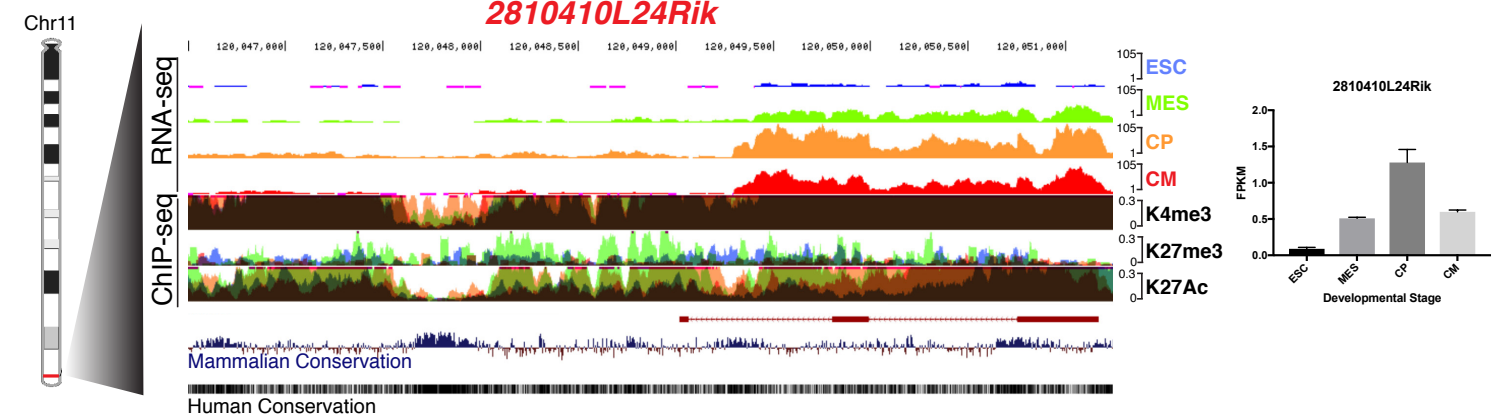
**A**



**B**



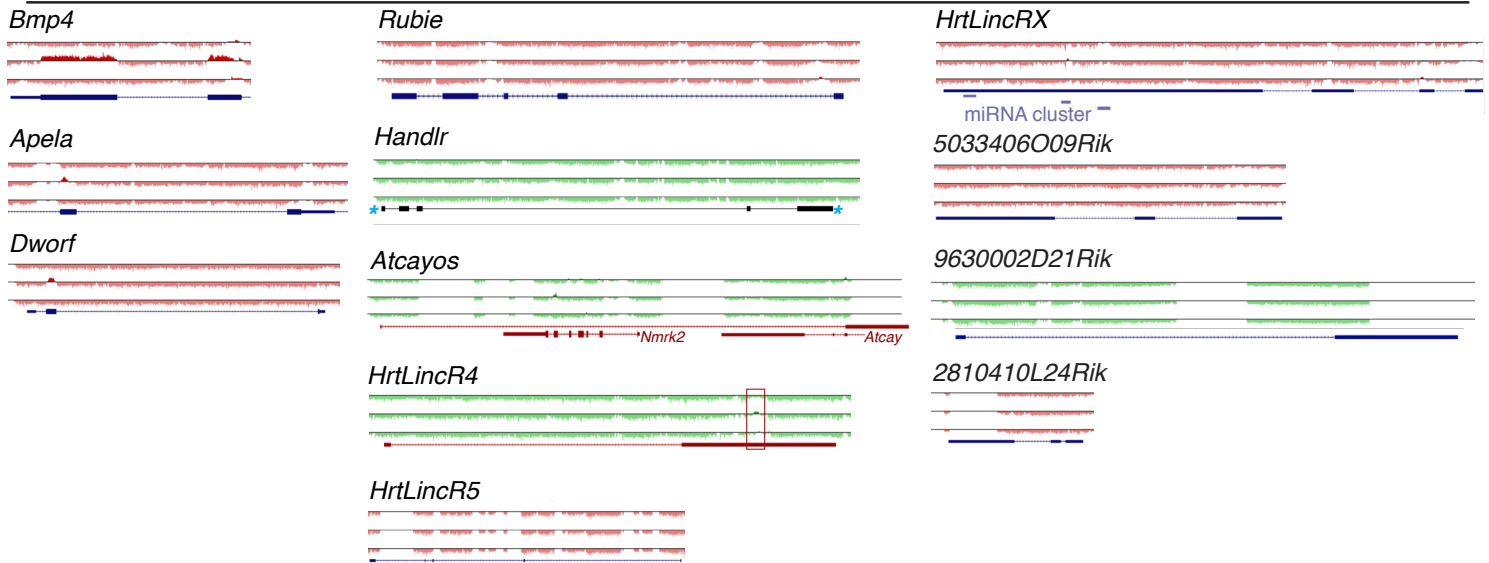
**C**



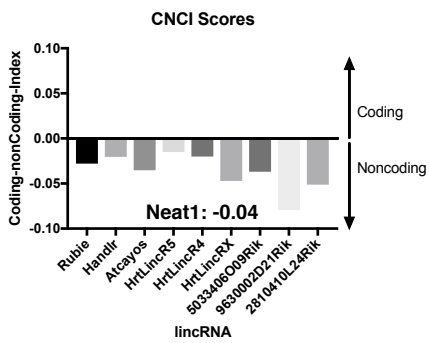
**Figure S4. Genomic characterization of *5033406O09Rik*, *9630002D21Rik*, and *2810410L24Rik in vitro*.** A. UCSC Genome Browser tracks of *5033406O09Rik* RNA-seq and overlaid histone H3 ChIP-seq during cardiac differentiation *in vitro*, as well as quantified expression at each differentiation stage. B. UCSC Genome Browser tracks of *9630002D21Rik* RNA-seq and overlaid histone H3 ChIP-seq during cardiac differentiation *in vitro*, as well as quantified expression at each differentiation stage. C. UCSC Genome Browser tracks of *2810410L24Rik* RNA-seq and overlaid histone H3 ChIP-seq during cardiac differentiation *in vitro*, as well as quantified expression at each differentiation stage. ESC, embryonic stem cell, MES, mesoderm, cMES, cardiac mesoderm; CP, cardiac progenitor, CM, cardiomyocyte; blue, ESC; green, MES; orange, CP; red, CM; K4me3, histone H3 lysine 4 trimethylation; K27me3, histone H3 lysine 27 trimethylation; K27Ac, histone H3 lysine 27 acetylation; Ensembl annotation in red, RefSeq annotations in blue. Data presented as mean +/- S.E.M.

**A**

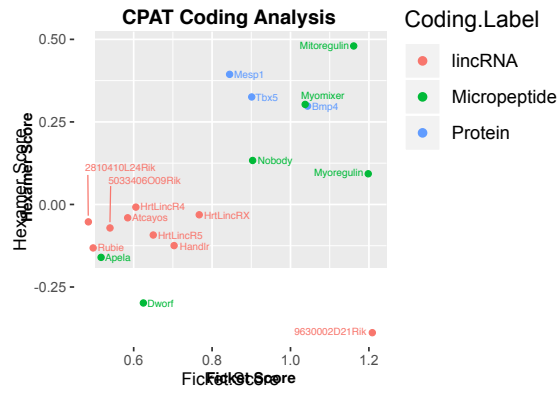
PhyloCSF



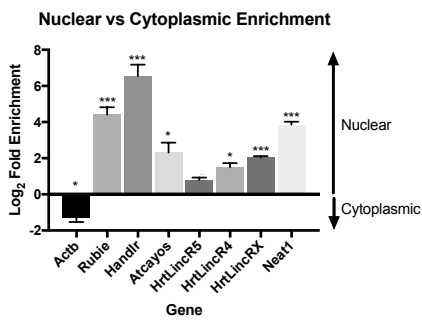
**B**



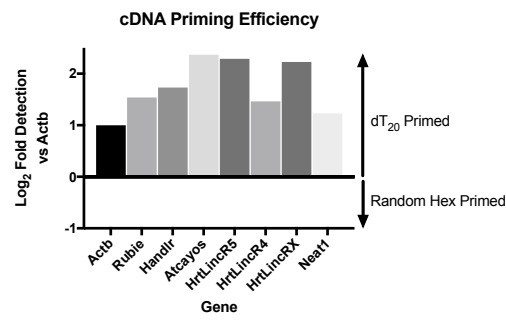
**C**



**D**

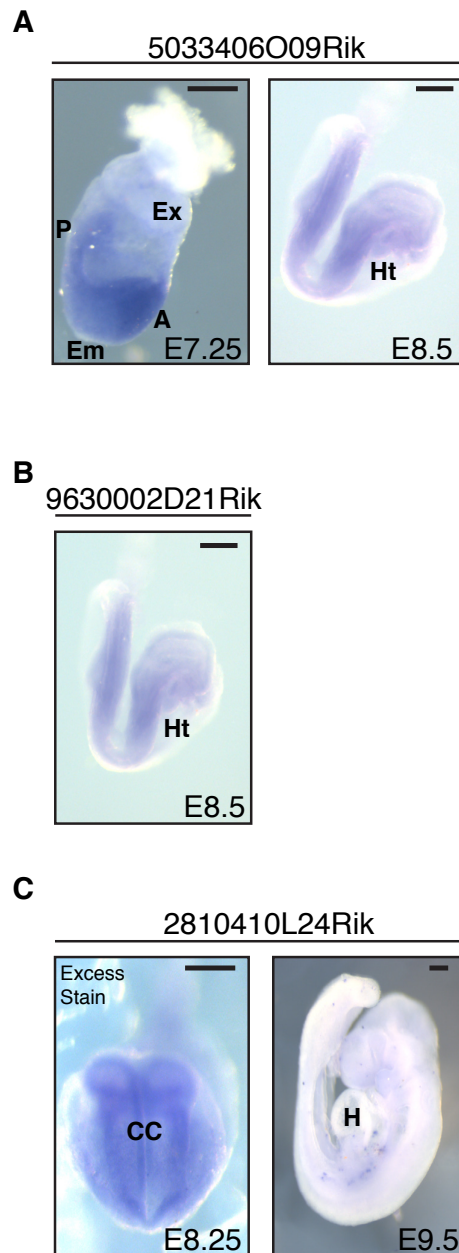


**E**

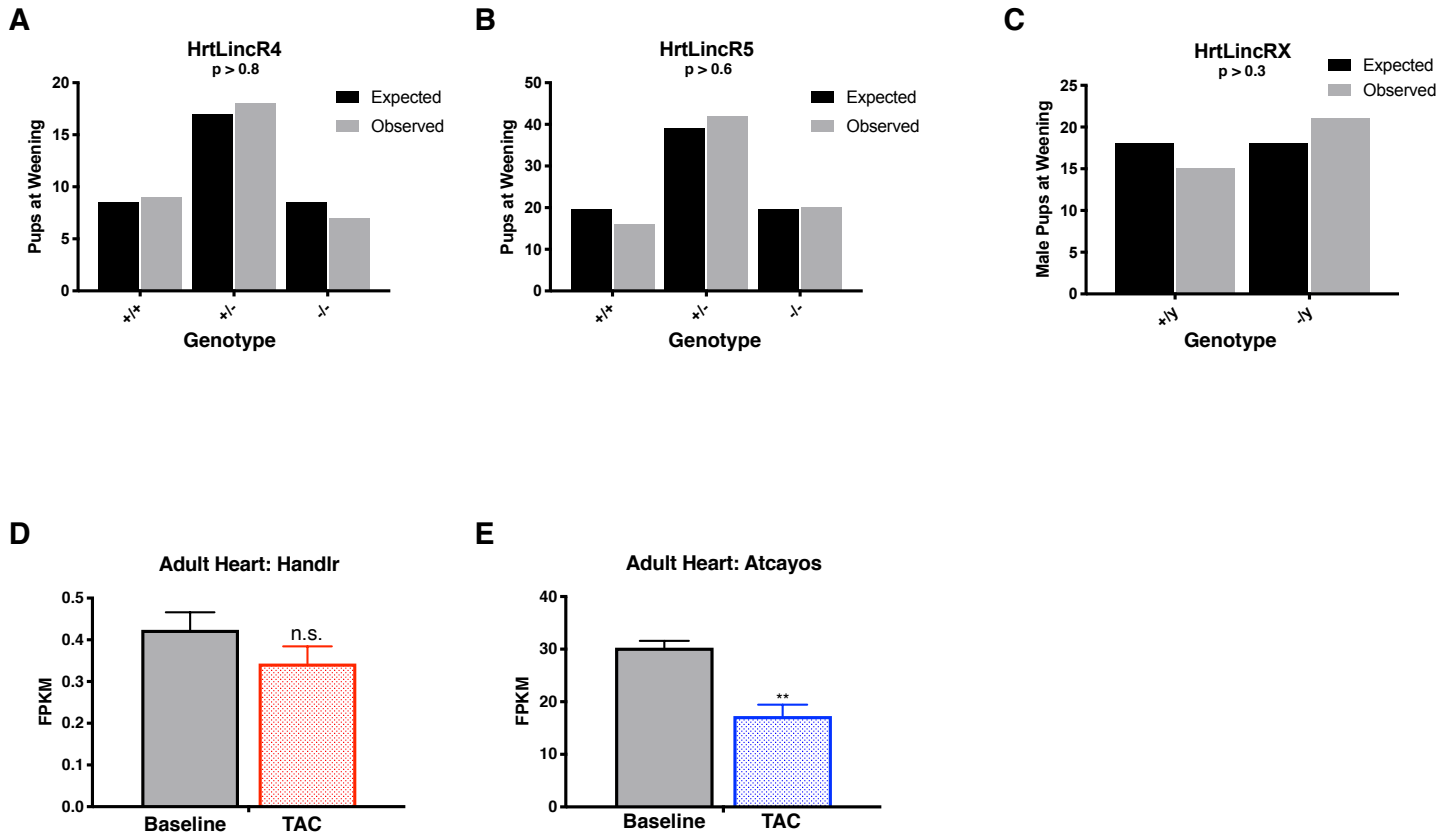




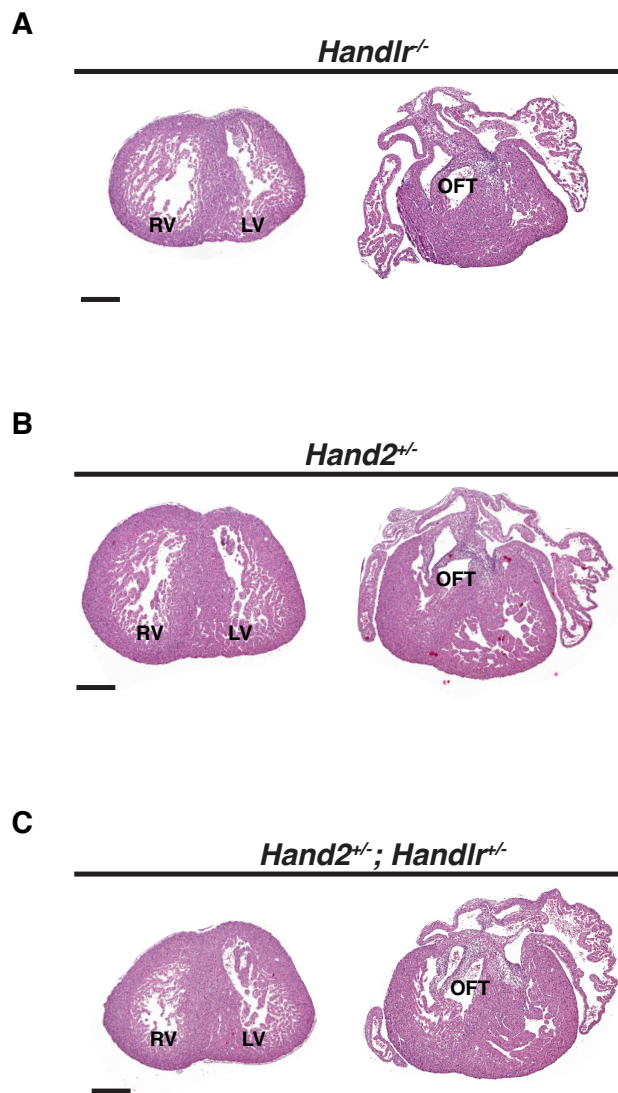
**Figure S5. Molecular characterization of lincRNA cohort.** A. UCSC Genome Browser tracks of PhyloCSF codon scores for all three frames of known protein coding gene *Bmp4*, micropeptide coding genes *Apela* and *Dwarf*, and lincRNA cohort; red box, potential 28 amino acid-coding open reading frame in *HrtLincR4*; scale, -15 to +15; positive score indicates higher coding potential; green, (+) strand; red, (-) strand. B. Coding-non-Coding-Index (CNCI) scores for lincRNA cohort compared to known lincRNA *Neat1*. C. Comparison of CPAT algorithm calculations of Fickett and hexamer scores for known protein coding genes, micropeptide coding genes, and our lincRNA cohort. D. Nuclear vs cytoplasmic enrichment of lincRNA cohort compared to *Actb* and known nuclear-enriched lincRNA *Neat1*; \*,  $p < 0.05$ ; \*\*\*,  $p < 0.005$ ; n.s., not significant; Student's 2-tailed t-test. E. Efficiency of RT-qPCR amplification from dT<sub>20</sub>- or random hexamer-primed cDNA for lincRNA cohort compared to *Actb* and known polyadenylated lincRNA *Neat1*. Data presented as mean  $\pm$  S.E.M.



**Figure S6. lincRNA expression patterns *in vivo*.** A. *in situ* hybridization staining for 5033406O09Rik at E7.25 and E8.5. B. *in situ* hybridization staining for 9630002D21Rik at E8.5. C. *in situ* hybridization staining for 2810410L24Rik at E8.25 and E9.5. A, anterior; Em, embryonic region; Ex, extraembryonic region; P, posterior; CC, cardiac crescent; Ht, heart tube; H, heart.



**Figure S7. Viability and phenotypic effects after lincRNA knockout.** A. Offspring recovered at weaning from *HrtLincR4*<sup>+/-</sup> x *HrtLincR4*<sup>+/-</sup> cross vs expected Mendelian ratios. B. Offspring recovered at weaning from *HrtLincR5*<sup>+/-</sup> x *HrtLincR5*<sup>+/-</sup> cross vs expected Mendelian ratios. C. Male offspring recovered at weaning from *HrtLincRX*<sup>+/-</sup> x *HrtLincRX*<sup>+/-</sup> cross vs expected Mendelian ratios; p-values in panels A-C derived from Chi-Square test. D. RNA-seq expression of Handlr in adult heart before and after TAC. E. RNA-seq expression of Atcayos in adult heart before and after TAC from Duan et al, 2017. \*\*, p < 0.01, Student's 2-tailed t-test; TAC, transverse aortic constriction; WT, wild type.



**Figure S8. Effect of *Handlr* ablation on heart development at E15.5.** A-C. Oblique transverse hematoxylin and eosin histological sections of cardiac ventricular and OFT morphogenesis, respectively, at E15.5. A. Representative *Handlr*<sup>-/-</sup> morphology. B. Representative *Hand2*<sup>+/-</sup> morphology. C. Representative *Hand2*<sup>+/-</sup>; *Handlr*<sup>+/-</sup> morphology. RV, right ventricle; LV, left ventricle; OFT, outflow tract; WT, wild type; scale bar, 300 μm.

Target	Forward Primer	Reverse Primer	Product Size (bp)
<i>Handlr</i>	CTGTGGGTCTCAGTAGGGAC	TTTTGATTTTCCCCGCCCTG	101
<i>Rubie</i>	CTGCTTCCTCCTCTTCGTGT	CCAAATGCTTCAGTTCCCT	90
<i>Atcayos</i>	CCCAGTCGTTGAAGAGGAGT	TTCACTTCCCACCCCATCTC	126
<i>HrtLincR4</i>	TCTACAACCACAGTCAGCCC	CTGAGATATCCCCGCCTGAG	113
<i>HrtLincR5</i>	CCAGCAGCACCAACCAATA	AGACGGTGGACCCTTTTCTT	94
<i>HrtLincRX</i>	CTTAAGGAGTGGGGCTGTTAT	AATGATCCAGTGCTAACATATGC	78
<i>Hand2</i>	CAGCTACATCGCCTACCTCA	CTCCTCTTTCACGTCGGTCT	103
<i>Bmp4</i>	AGCCAACACTGTGAGGAGTT	GGATGCTGCTGAGGTTGAAG	104
<i>Nmrk2</i>	GAAGACCACCCTGACCAACA	CCCAGTGTAAAGCCGTCC	122
<i>Trabd2b</i>	TCGAGGACAGCACTACGATC	CTGGAGTTGGTGTGAGGACT	101
<i>Mn1</i>	TGCCAGAACATGATTGCCAG	GCTCAGTTTCTCTTCCCT	90
<i>Plac1</i>	TACCAGTTTCACTACCGCGT	ACAGGACACGGGAATCACAT	135
<i>Neat1</i>	CTGCACTGTAGATCGGGACC	CTTTCCCCAACACCCACAAG	117

**Table S1. RT-qPCR primers.** Primers utilized to amplify gene or lincRNA cDNA. Oligomers listed in 5'- to 3'- orientation.

Target	Fwd Primer	Rev Primer
<i>Handlr</i>	GACTTGGATCTGTCTCGCCT	TCTGTCCCCGGAGTGTAGTA
<i>Rubie</i>	CTCCCTGCTTCCTCCTCTTC	GTCCTGGTCATGGTCTGTCA
<i>Atcayos</i>	GAGATGGGGTGGGAAGTGAA	ACCTTTACCTGCTGAGCCAT
<i>HrtLincR4</i>	ACCGAGAGACAGCAGTTGAA	CCGCGAGTCCCTTTGTAATG
<i>HrtLincR5</i>	TATGGGGATGGGAACTCAGC	AGGATGAACGGAGAAGCACA
<i>HrtLincRX</i>	ACCGCTCCAACCCGATTC	GCTCCCACTGTTGCTTTTCA
<i>5033406O09Rik</i>	CCCACTCTCTGCTGATCCAA	AGCCCCGTTTCTTCTTCTGA
<i>9630002D21Rik</i>	AGGATGAACGGAGAAGCACA	GCCTTCTTCCTCCTCTGGTT
<i>2810410L24Rik</i>	CTCACATCCTGTTCTGGGGT	AACATCCCGTGCTGTAAAGC

**Table S2. Primers for *in situ* hybridization probe generation.** Oligomers listed in 5'- to 3'- orientation.

Target	Left Cut	Right Cut
<i>Handlr</i>	AATGATGGTCATCCACCG TTAAAAAGAAGGCCCGG	CCTAATAAGATGCCTGAC CCTTCTCATTGAAACCCC
<i>Rubie</i>	GGGCTAAATAACCCATTA	GATTAATTCATGCCCGG
<i>Atcayos</i>	CAACCCGCTTTAACTCC CTCATAACTTGGGGTGTC	GACCAGCATCAGAGTCAC ACCCGCCTGGTGAGCCGA
<i>HrtLincR4</i>	GGTTCAGGCAGGAATAC	GGGGACAGCCTTTACAA
<i>HrtLincR5</i>	GTGATAGACCACTTCTTG	GCGCCCATCCGTATGC
<i>HrtLincRX</i>	GGCCATTTTGACCCGTCG	GGCGGTCCACTTTAGGGG

**Table S3. tru-sgRNA crRNA oligomers.** Cas9-targeting oligomers cloned into pX330 for lincRNA knockout. Sequences are truncated by 2-3 nucleotides to diminish off-target effects. Oligomers listed in 5'- to 3'- orientation. Genomic PAM sequence located immediately 3'- to each target sequence. Left and right cuts relative to *mus musculus* mm9 assembly.

Locus	Fwd Primer	Rev Primer	WT band size (bp)	KO band size (bp)
<i>Handlr</i>	TGGATCCAACCTAACCTAATGCTT	ACGCGCAGGAAAGGTAAAT CCGCATTAGTTTCCCTTCCC	600	400
<i>Rubie</i>	CCTGGTTGCCGTCATTAAGT	GTGTATGTGGGCAGAGAGGT AGTCACTGTTTCGCATGAGA	617	680
<i>Atcayos</i>	TGACGACAGCTGATAGGGC	TGCTTGCCCTAGTATGGTCA AGTTCATCTTCAAGGCCCT	443	280
<i>HrtLincR4</i>	ATCTTGGGGCCATCTCCAAA	GACAATTGGCTGTGGTGGTT TTTCTCTGGACCCGTGTGAG	448	500
<i>HrtLincR5</i>	GTAAAGGCTCTTGTCACAGGG	TTTTCTTGCAACTCCCAGC TAACTAGAGGGAGGGGAGGG	255	180
<i>HrtLincRX</i>	AGGAGAGAGAAAAGCCGTGT	CGCGGTGCAATGAAAGACTA GGGCCACTCATTTGTAAGCA	784	855
<i>Bmp4 (loxP)</i>	GAGCTAAGTTTTGCTGGTTTGC	GCCCATGAGCTTTTCTGAGA	200	250 (loxP)
<i>Cre Transgene</i>	ATGCTTCTGTCCGTTTGCCG	CCTGTTTTGCACGTTACCG	280	
<i>Hand2 (loxP)</i>	AAAGAGTCAGTGGTTCTCATCC GAAGTTCCTATTCTCTAGAAA	AATCTAACCCAACCCCTCGC	600	634 (loxP) 300 (KO)

**Table S4. Genotyping primers.** 3-primer genotyping oligomers used for embryo genotyping. Wild type (WT) and knockout (KO) alleles amplified independently, resulting in variable gDNA amplified products of variable lengths (larger band sizes in KO reactions do not represent DNA insertions).



Time Point	<i>Handlr<sup>+/+</sup></i>					<i>Handlr<sup>-/-</sup></i>				
	Baseline	Week 1	Week 4	Week 6	Week 8	Baseline	Week 1	Week 4	Week 6	Week 8
ENDOarea;d (mm <sup>2</sup> )	16.21±0.71	16.26±1.19	16.65±1.42	16.19±1.58	17.35±1.63	14.00±0.70	14.63±0.29	15.39±1.16	15.65±0.49	16.05±1.09
ENDOarea;s (mm <sup>2</sup> )	10.07±0.45	11.82±1.15	12.59±1.10	12.30±1.17	13.35±1.36	7.51±0.53	9.72±0.35	10.68±1.25	11.38±0.27	12.45±1.04
IVS;d (mm)	0.64±0.04	1.02±0.05	0.90±0.06	1.12±0.06	1.20±0.08	0.63±0.04	0.93±0.04	1.13±0.04	1.05±0.07	1.29±0.06
IVS;s (mm)	0.96±0.04	1.38±0.04	1.16±0.08	1.25±0.05	1.39±0.05	0.91±0.02	1.28±0.07	1.33±0.06	1.28±0.05	1.46±0.11
LVID;d (mm)	4.54±0.10	4.38±0.16	4.65±0.19	4.58±0.24	4.68±0.19	4.19±0.09	4.25±0.04	4.49±0.18	4.57±0.06	4.43±0.13
LVPW;d (mm)	0.66±0.05	1.00±0.05	1.03±0.11	1.22±0.07	1.13±0.04	0.61±0.04	0.89±0.09	0.93±0.09	1.03±0.05	1.11±0.05
LVPW;s (mm)	0.82±0.06	0.97±0.06	1.26±0.10	1.52±0.07	1.37±0.07	0.87±0.05	1.09±0.12	1.32±0.22	1.37±0.12	1.25±0.06
Doppler (mm/s)		-3665±206					-3671±209			
HR (bpm)	511.23±16.33	513.64±11.34	512.55±28.10	506.11±26.13	545.55±29.02	470.08±12.68	493.20±21.93	492.34±24.46	476.12±30.90	498.91±19.11
Body Weight (g)	28.30±0.76	26.70±0.41	28.57±0.70	29.87±0.73	31.00±0.53	28.70±1.49	27.96±1.74	28.44±1.19	30.00±1.52	29.60±1.03
LV Mass (mg)	110.68±6.06	191.11±17.75	198.85±27.89	252.14±24.46	257.60±24.23	90.33±4.89	156.06±12.16	201.37±11.15	209.86±11.13	245.80±19.90
FAC (%)	37.78±1.55	27.93±3.16	24.25±2.52	23.85±1.96	23.38±1.37	46.50±1.70	33.50±2.68	31.45±4.94	27.06±2.83	22.50±3.65

**Table S5. *Handlr* KO TAC echocardiography.** Echocardiographic measurements in 12-20 week-old litter-matched adult males at baseline and after transverse aortic constriction (TAC). d, diastolic; s, systolic; IVS, interventricular septum; LVID, left ventricular internal diameter; LVPW; left ventricular posterior wall; HR, heart rate; LV, left ventricle; FAC, fractional area contractility. Data presented as mean ± S.E.M. N = 4-9 for all measurements.

Time Point	Atcayos <sup>+/+</sup>					Atcayos <sup>-/-</sup>				
	Baseline	Week 1	Week 4	Week 6	Week 8	Baseline	Week 1	Week 4	Week 6	Week 8
ENDOarea;d (mm <sup>2</sup> )	12.95±/0.65	15.66±/1.10	14.89±/0.89	15.09±/1.57	16.15±/1.51	14.95±/1.70	15.21±/0.94	14.44±/0.97	14.17±/1.26	15.74±/1.20
ENDOarea;s (mm <sup>2</sup> )	8.11±/0.43	10.15±/0.94	10.71±/0.84	11.02±/1.52	12.47±/1.38	8.90±/0.86	9.95±/0.81	10.22±/0.87	10.65±/1.25	12.27±/1.30
IVS;d (mm)	0.77±/0.03	1.05±/0.06	1.08±/0.05	1.03±/0.04	1.08±/0.05	0.62±/0.04	0.90±/0.09	1.01±/0.04	0.99±/0.05	1.14±/0.07
IVS;s (mm)	1.01±/0.06	1.42±/0.07	1.32±/0.06	1.30±/0.04	1.39±/0.06	0.83±/0.04	1.33±/0.05	1.25±/0.07	1.18±/0.05	1.31±/0.05
LVID;d (mm)	4.01±/0.09	4.32±/0.15	4.47±/0.14	4.47±/0.19	4.45±/0.20	4.25±/0.09	4.22±/0.15	4.33±/0.16	4.24±/0.17	4.52±/0.16
LVPW;d (mm)	0.82±/0.04	1.05±/0.05	1.09±/0.04	1.17±/0.07	1.21±/0.06	0.60±/0.04	0.93±/0.05	1.14±/0.04	1.21±/0.09	1.12±/0.07
LVPW;s (mm)	0.99±/0.06	1.24±/0.06	1.36±/0.06	1.36±/0.03	1.37±/0.05	0.77±/0.05	1.14±/0.07	1.35±/0.06	1.35±/0.08	1.33±/0.05
Doppler (mm/s)		-3457±/174					-3509±/210			
HR (bpm)	485.01±/22.77	513.39±/13.83	520.69±/15.65	510.88±/18.73	538.68±/13.28	451.29±/34.10	515.49±/7.01	555.15±/17.93	513.05±/19.75	527.46±/29.90
Body Weight (g)	28.37±/0.77	27.40±/0.51	28.50±/0.74	29.31±/0.59	29.44±/0.47	27.36±/0.81	26.41±/0.63	27.26±/0.74	28.31±/0.82	29.29±/0.84
LV Mass (mg)	117.88±/7.48	195.39±/16.13	216.98±/14.07	221.22±/19.53	234.12±/20.87	91.61±/3.64	154.58±/11.35	202.07±/11.93	203.23±/18.00	234.41±/22.46
FAC (%)	37.41±/0.66	35.60±/1.93	29.18±/2.09	27.98±/2.40	23.47±/2.09	40.13±/1.73	34.66±/2.63	29.59±/2.08	25.66±/3.29	22.75±/3.03

**Table S6. Atcayos KO TAC echocardiography.** Echocardiographic measurements in 12-20 week-old litter-matched adult males at baseline and after transverse aortic constriction (TAC). d, diastolic; s, systolic; IVS, interventricular septum; LVID, left ventricular internal diameter; LVPW; left ventricular posterior wall; HR, heart rate; LV, left ventricle; FAC, fractional area contractility. Data presented as mean ±/ S.E.M. N = 4-9 for all measurements.