Fig S1



Fig S1. Egr-1 deficiency impairs proinflammatory cytokine mRNA transcription but has differential effects on chemokine mRNA transcription during *P. aeruginosa* lung infection. Wild-type (+/+) and Egr-1-deficient (-/-) mice were intranasally infected with 1 × 10⁹ CFU/mouse of *P. aeruginosa* 8821 for 4 h, 24 h or an equivalent volume of saline as a control (NT). The total RNA extracted from lungs was reverse transcribed to cDNA and subjected to real-time quantitative PCR for *IL-1β* (A), *IL-6* (B), *TNF* (C), *MIP2* (D), *KC* (E), *LIX* (F), *IP-10* (G) and *RANTES* (H) gene expression. The gene expression was normalized to housekeeping control gene HPRT (n = 3 ± SEM, *p < 0.05).



Fig S2. Egr-1-deficient BMMs display impaired proinflammatory cytokine and chemokine production following *P. aeruginosa* infection. Wild-type (+/+) and Egr-1-deficent (-/-) BMMs were infected with *P. aeruginosa* 8821 at a MOI of 10 for 1 h, 2 h, 4 h or left untreated (NT). Total RNA isolated from these cells was reverse transcribed to cDNA and subjected to real-time quantitative PCR for *IL-1* β (A), *IL-6* (B), *TNF* (C), *MIP2* (D), *KC* (E), *LIX* (F), IP-10 (G) and RANTES (H) gene expression. The gene expression was normalized to housekeeping control gene HPRT (n = 3 ± SEM, *p < 0.05, ***p < 0.001, ****p < 0.0001).

Fig S3

PAO1-infected Macrophages



Fig S3. Egr-1-deficient BMMs display impaired proinflammatory cytokine and chemokine production following *P. aeruginosa* PAO1 infection. Wild-type (+/+) and Egr-1-deficient (-/-) BMMs were infected with *P. aeruginosa* PAO1 at a MOI of 10 for 3 h, 6 h, 12 h or left untreated (NT). Cell supernatants were collected for the determination of IL-1 β (A), IL-6 (B), TNF (C), IL-12 (D), IL-17 (E), MIP2 (F), KC (G), LIX (H), IP-10 (I) and RANTES (J) secretion by ELISA (n = $3 \pm SEM$, *p < 0.05, **p < 0.01, ***p < 0.001, N.D. = not detected).

Fig S4

В A С D **30 _ Egr-1+/+** Egr-1-/-Egr-1 +/+ Egr-1+/+ 20000-Egr-1 +/+
Egr-1 -/-10000 8000 3000 02 10-12 (bg/mL) 15000 3000 JNL (bd/ml) 1000 IL-1ß (pg/mL) IL-6 (pg/mL) 6000 10000 4000 5000 1000 2000 NT NT 12 h 12 h 3 h 6 h 3 h NT 3 h 6 h 12 h Е F G Н 1.0 Egr-1+/+ 25 Egr-1 +/+ Egr-1 -/-10000 Egr-1 +/+ 5000-Egr-1 +/+ Egr-1 -/-0.8 20 8000 4000 IL-17 (pg/mL) (Jm/6d) XIJ MIP2 (pg/mL) 0.6 6000 KC (pg/ml) 3000 N.D. 0.4 4000 2000 0.2 2000 1000 0.0 0 ŃТ 6 h 12 h n 3 h NT 3 h 6'n 12 h NT ŃТ 12 h 3 h 6 h 6 h I J Egr-1 +/+ 5000 2500 Egr-1 4000 RANTES (pg/mL) 2000 IP-10 (pg/mL) 3000 1500 2000 1000 1000 500 0 NT 3h

PAO1-infected dendritic cells

Fig S4. Egr-1-deficient BMDCs have increased LIX production following *P. aeruginosa* PAO1 infection. Wild-type (+/+) and Egr-1-deficient (-/-) BMDCs were infected with *P. aeruginosa* PAO1 at a MOI of 10 for 3 h, 6 h, 12 h or left untreated (NT). Cell supernatants were collected for the determination of IL-1 β (A), IL-6 (B), TNF (C), IL-12 (D), IL-17 (E), MIP2 (F), KC (G), LIX (H), IP-10 (I) and RANTES (J) secretion by ELISA. (n = 3 ± SEM, *p < 0.05, **p < 0.01, ***p < 0.001, ***p < 0.0001, N.D. = not detected).



Fig S5. Egr-1-deficient BMMs display diminished NF-κB and NFAT activation during *P. aeruginosa* infection. Wild-type (+/+) and Egr-1-deficient (-/-) BMMs were infected with *P. aeruginosa* 8821 at a MOI of 10 for 20 min, 1 h, 3 h or left untreated (NT). Nuclear proteins were extracted and subjected to EMSA by incubation with ³²P-labeled NF-κB (A) and NFAT (C) DNA probes. Data are representative of three individual experiments. Densitometry analysis was performed for NF-κB (B) and NFAT (D) activities, and data are expressed as fold change data versus wild-type untreated BMMs (n = ± 3 SEM, *p < 0.05).



Fig S6. Egr-1 deficiency has no effect on MAPK activation in macrophages during *P. aeruginosa* infection. Wild-type (+/+) and Egr-1-deficient (-/-) BMMs were infected with *P. aeruginosa* 8821 at a MOI of 10 for 1 h, 2 h, 4 h, 6 h or left untreated (NT). Cell lysates were subjected to Western blot analysis for determining phospho- and total ERK, p38 and JNK, as well as actin as a loading control. Blots are representative of three independent experiments (A). Densitometry analysis of phosphorylated ERK (B), p38 (C) and JNK (D) was normalized to their total protein respectively (n = $3 \pm SEM$).



Fig S7. Egr-1 deficiency leads to increased nitric oxide production in neutrophils and macrophages in response to *P. aeruginosa* PAO1 infection. Wild-type (+/+) and Egr-1-deficient (-/-) neutrophils and BMMs were infected with *P. aeruginosa* PAO1 for 6 h or or left untreated (NT). The NO₂⁻ levels were tested in cell lysates (A, C) and supernatants (B, D) at 6 h (n = $3 \pm SEM$, *p < 0.05, **p < 0.01).



Fig S8. Egr-1 deficiency has no impact on macrophage apoptosis during *P. aeruginosa* **infection.** Wild-type (+/+) and Egr-1-deficient (-/-) BMMs were infected with *P. aeruginosa* 8821 at a MOI of 10 for 2 h or left untreated (NT). The cells were stained with FITC Annexin V and 7-AAD, and a total 2×10^4 cells from each sample was analyzed on a flow cytometer. The density plots for wild-type (A and B) and Egr-1-deficient (C and D) macrophages were represented of three individual experiments. The bar graphs for the data of cell death (E) and apoptosis (F) were presented as percentage (n = 3).