

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

X-ray experiments experiments were performed on custom software for SSRL beamline 7-1 and ALS beamline 5.0.3

Data analysis

Additionally, Applied published crystallography tools:

Applied published crystallography tools:

Data reduction - XDS (Versions January 30, 2009, December 31, 2001, and July 4, 2012)

Data scaling - XSCALE (Versions January 30, 2009, December 31, 2001, and July 4, 2012)

Refinement - CCP4 program suite, Refmac5 (Version 5.5.0109 and 5.6.0117)

Validation - Molprobit

Validation - wwPDB validation service

Statistical tools

SAS 9.4, SAS Institute Inc., Cary, NC, USA

Molecular simulation software

FREAD ref. 42 Open source code for structure

Maestro ref. 44 Proprietary Structure based design application from Schrodinger

AmberTools 1.4 ref. 45 Open source code for building and analysing molecular dynamics data

NAMD 2.7 ref. 47 Open source molecular dynamics application

ACEMD ref. 48 Proprietary molecular dynamics application from Acelera

Plumed 1.3 ref. 49 Open source molecular dynamics application

GROMACS ref. 52 Open source molecular dynamics application

Biacore software

Biacore T100 BIAevaluation software version 1.1

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The authors declare that [the/all other] data supporting the findings of this study are available within the paper [and its supplementary information files].

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

- Sample size: No sample size calculation was made in the current study. Sample sizes for animal studies based on standard protocols to enable statistic validity as shown in the text. Where n=1 data is used, data in the published experiment is supported by a number of experiments on compounds with the same mode of action not described in the text.
- Data exclusions: No data excluded except in animal experiments where data only excluded as a consequence of technical issues e.g. blood contamination in lavage fluid
- Replication: Where applied, all attempts at replication were successful and are included in the data analyses
- Randomization: Cages of animals were randomized into groups at the start of each study.
- Blinding: Investigators were not blinded to group allocation, but arthritis scores were independently checked

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Antibodies |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Animals and other organisms |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |

Methods

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

Antibodies

Antibodies used

List of antibodies used in publication.

Immunoprecipitation: anti-TNFR1 (polyclonal goat R&D AB-225-PB): Human TNF RI/TNFRSF1A Antibody
Control (polyclonal goat R&D AB-108-C): Normal Goat IgG Control

Western blots:

anti-RIP1 (BD-610459 1:250 dilution); detection with anti-mouse-HRP (Cell Signalling -7076 1:2000 dilution) : full name Mouse

Anti-RIP Clone 38/RIP
 anti-pNFkB (Cell Signalling -3033 1:1000 dilution); detection with anti-rabbit HRP (Cell Signalling-7074 1:2000 dilution) : full name Phospho-NF- κ B p65 (Ser536) (93H1) Rabbit mAb
 anti-GAPDH Cell Signalling- 5174 1:4000 dilution); detection with anti-rabbit HRP (Cell Signalling-7074 1:2000 dilution) : full name GAPDH (D16H11) XP® Rabbit mAb

Validation

All antibodies confirmed to be applicable for methods, as detailed in data sheets provided by commercial supplier

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)

HEK-Blue™ CD40L SEAP (secreted embryonic alkaline phosphatase) reporter cell line (InvivoGen, #hkb-CD40)
 Jurkat (Clone 6E-1 , human acute T cell lymphoma suspension cell line – ATCC - TIB-152)
 L929 cells (ECACC, catalogue #85011425)

Authentication

Authentication included in supplier data sheets

Mycoplasma contamination

All cell lines were negative for mycoplasma

Commonly misidentified lines
(See [ICLAC](#) register)

None used

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals

Male Balb/c mice aged 6 -8 weeks were used in all in vivo studies

Wild animals

The study did not involve wild animals

Field-collected samples

The study did not involve samples collected from the field

Ethics oversight

All in vivo studies were reviewed by an internal Ethical Review Body (ERB) and conducted in accordance with the Animals (Scientific Procedures) Act 1986, EU Directive 2010/63/EU.

Note that full information on the approval of the study protocol must also be provided in the manuscript.