

SUPPLEMENTAL MATERIAL

Ventricular myocyte cell model

The base cell model used was the human ventricular myocyte model by O'Hara et al ¹ with the endocardial control parameters. To address a well-known tissue propagation issue, the I_{Na} m-gate steady-state activation curve was left-shifted by 10 mV. To simulate the different congenital LQTS subtypes, the conductance of I_{Ks} or I_{Kr} was completely blocked in the case of LQT1 and LQT2 respectively, and the conductance of I_{NaL} was increased in LQT3. To simulate the bulk heterogeneity, the conductances of I_{Kr} in LQT1, I_{Ks} in LQT2, and I_{NaL} in LQT3 were adjusted to create a repolarization gradient. The $I_{Ca,L}$ steady-state activation and inactivation curves were taken from either the original O'Hara et al formulation or another human measurement from Li et al ², parameterized in the form of $d_{\infty} = 1/(1 + \exp\left[\frac{-(V+d_{\infty,V})}{d_{\infty,k}}\right])$ and $f_{\infty} = 1/(1 + \exp\left[\frac{V+f_{\infty,V}}{f_{\infty,k}}\right])$.

The parameters for LQT1 were: $G_{Ks} = 0$ mS/ μ F, $G_{Kr,1} = 0.025$ mS/ μ F, $G_{Kr,2} = 0.017$ mS/ μ F, $d_{\infty,V} = 3.940$ mV, $d_{\infty,k} = 4.230$ mV, $f_{\infty,V} = 19.58$ mV, $f_{\infty,k} = 3.696$ mV.

The parameters for LQT2 were: $G_{Kr} = 0$ mS/ μ F, $G_{Ks,1} = 0.050$ mS/ μ F, $G_{Ks,2} = 0.030$ mS/ μ F, $d_{\infty,V} = 4.8$ mV, $d_{\infty,k} = 6.2$ mV, $f_{\infty,V} = 28.5$ mV, $f_{\infty,k} = 7.8$ mV.

The parameters for LQT3 were: $G_{NaL,1} = 0.050$ mS/ μ F, $G_{NaL,2} = 0.066$ mS/ μ F, $d_{\infty,V} = 4.8$ mV, $d_{\infty,k} = 6.2$ mV, $f_{\infty,V} = 28.5$ mV, $f_{\infty,k} = 7.8$ mV.

Anatomical ventricle modeling

The anatomical geometry of the ventricle model and fiber direction field were obtained from previous work by Ten Tusscher et al ³, consisting of ~1.7 million voxels of 0.5 mm size, incorporating a Purkinje network (Fig.S1).

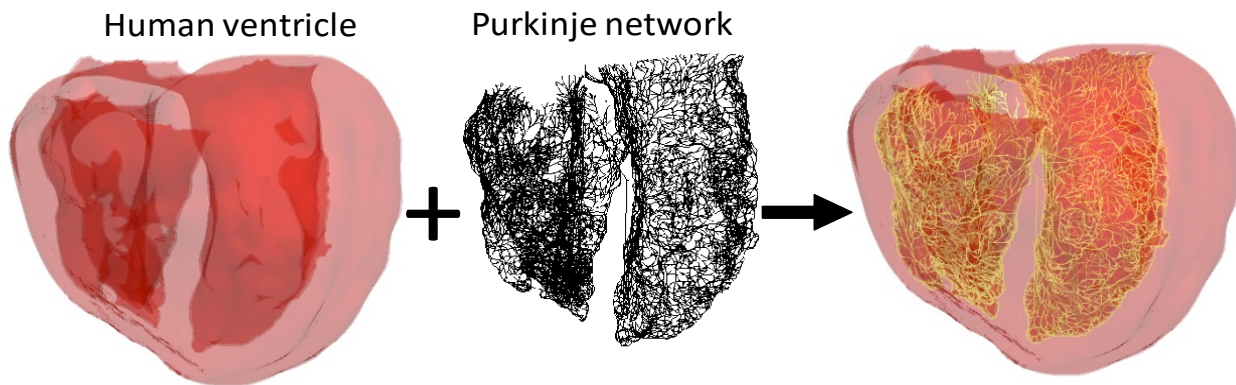


Figure S1. Human ventricle myocardium model coupled with the Purkinje network model. The Purkinje network starts at the simulated AV node where sinus beats are initiated, is grown onto the endocardial surface.

The partial differential equation governing membrane voltage V in the anatomical model is:

$$\frac{\partial V}{\partial t} = -I_{ion}/C_m + \sum_i \sum_j \frac{\partial}{\partial x_i} D_{ij} \frac{\partial}{\partial x_j} V$$

where $C_m = 1 \mu\text{F}/\text{cm}^2$ is the membrane capacitance, I_{ion} is the total ionic currents of the cell, and D_{ij} is the 3D conductivity tensor describing the anisotropy of voltage propagation corresponding to the fiber direction field, with i, j as indices for the 3 cartesian dimensions. Numerical integration was performed using with a finite difference forward Euler method and an operating splitting and time adaptive algorithm ⁴ with the time step (Δt) varying between 0.01 ms and 0.1 ms. A bulk heterogeneity in the RV was set using the parameters described above (Fig.S2).

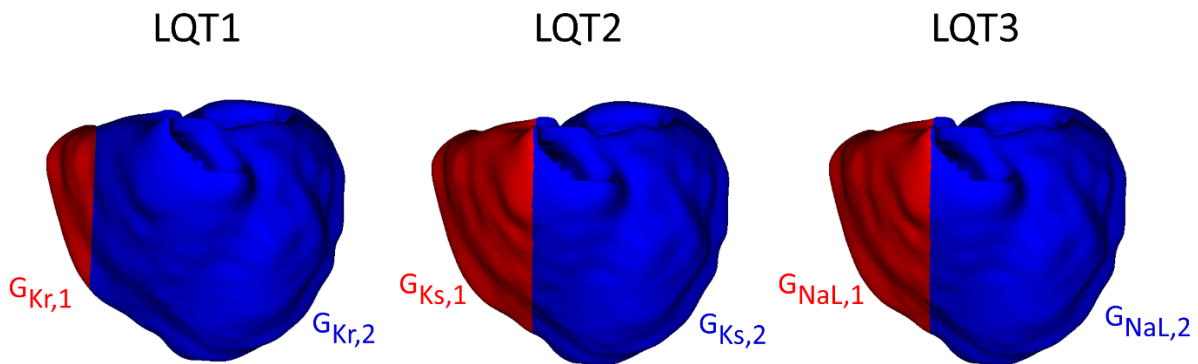


Figure S2. Bulk heterogeneities for each LQTS subtype. The spatial heterogeneity is created using two different conductance regions in G_{K_r} for LQT1, G_{K_s} for LQT2, and G_{NaL} for LQT3. All other parameters are homogeneous across the ventricle for each subtype. The smaller $G_{K_r,1}$ region used in LQT1 was to avoid 2:1 block when demonstrating arrhythmogenesis at fast pacing.

Purkinje network modeling

We tailored our model to provide for a physiological activation sequence, ECG properties, and robust forward and retrograde conduction at the Purkinje-myocyte junctions (PMJs). The human Purkinje action potential model by Stewart et al ⁵ was used for our Purkinje network. The Purkinje network was modeled as a connected set of 1D cables with the diffusion constant set to result in a conduction velocity of ~ 4 m/s. The primary tree of the Purkinje network began at a straight His-bundle which then branched into the right bundle branch and the left bundle branch (including the left anterior and posterior fascicles). An AV-node delay was simulated by a very slow diffusion constant at the beginning of the His-bundle. The rest of the network was generated using a fractal surface mapping method developed by Sahli Costabal et al ⁶, mapped to the RV and LV endocardial surfaces. The terminal Purkinje fibers were allowed to form loops as

consistent with experimental data ^{7,8}. The final full structure (Fig.S1) consisted of 58,705 computational cell nodes with $\Delta x = 0.5$ mm. A PMJ was created between each Purkinje cell node to the closest ventricular myocyte cell node, modeled as a passive Ohm's law conductance. The relative capacitance ratio between the Purkinje and ventricular myocyte computational nodes was set to 1.0 which allowed for robust forward and retrograde conduction across the PMJ.

The differential equation governing Purkinje cell voltage V_P is:

$$\frac{dV_P}{dt} = -\frac{I_{ion,P}}{C_{m,P}} + \sum_i D_P \frac{V_i - V_P}{\Delta x^2} + I_{PMJ}/C_{m,P}$$

with

$$I_{PMJ} = \begin{cases} G_{PMJ}(V - V_P), & \text{cells with a PMJ} \\ 0, & \text{cells without a PMJ} \end{cases}$$

where $C_{m,Purkinje}$ is the capacitance of the Purkinje cell node, $I_{ion,Purkinje}$ is the total ionic currents of the Purkinje cell, $D_{Purkinje} = 0.0300$ cm²/ms is the diffusion constant, $G_{PMJ} = 0.05$ mS/ μ F. The ventricular myocyte that belongs to the PMJ also receives an equal and opposite I_{PMJ} . Numerical integration was performed using a finite difference forward Euler method with operator splitting and a fixed time step of $\Delta t = 0.01$ ms.

1D and 2D tissue models

The partial differential equation governing voltage for the 1D cable is

$$\frac{\partial V}{\partial t} = -I_{ion}/C_m + D \frac{\partial^2 V}{\partial x^2}$$

where V is the membrane voltage, $C_m = 1$ μ F/cm² is the membrane capacitance, $D = 0.0005$ cm²/ms is the diffusion constant (proportional to gap junction conductance), and $\Delta x = 0.15$ mm.

For 2D tissue, the equation for voltage is

$$\frac{\partial V}{\partial t} = -I_{ion}/C_m + D \left(\frac{\partial^2 V}{\partial x^2} + \frac{\partial^2 V}{\partial y^2} \right)$$

where V is the membrane voltage, $C_m = 1$ μ F/cm² is the membrane capacitance, $D = 0.0001$ cm²/ms is the diffusion constant (proportional to gap junction conductance), and $\Delta x = 0.15$ mm.

Numerical integration was performed using a finite difference forward Euler method and an operating splitting and time adaptive algorithm with the time step (Δt) varying between 0.01 ms and 0.1 ms.

Pseudo-ECG computation

Pseudo ECGs were computed using the following formula ^{9, 10}

$$\Phi_e(x', y', z') \sim \iiint (-\nabla V) \cdot \left[\nabla \frac{1}{r} \right] dx dy dz$$

where ∇V is the spatial gradient of the membrane voltage and $r = [(x - x')^2 + (y - y')^2 + (z - z')^2]^{1/2}$ represents the distance from a source point (x, y, z) to a field point (x', y', z') , with the integral performed over all heart cells.

For the human ventricle simulations, we calculated a pseudo-ECG for V1 to V6 (see Fig.S3 for lead placement). A bandpass filter of 0.1-150 Hz was applied to each recording.

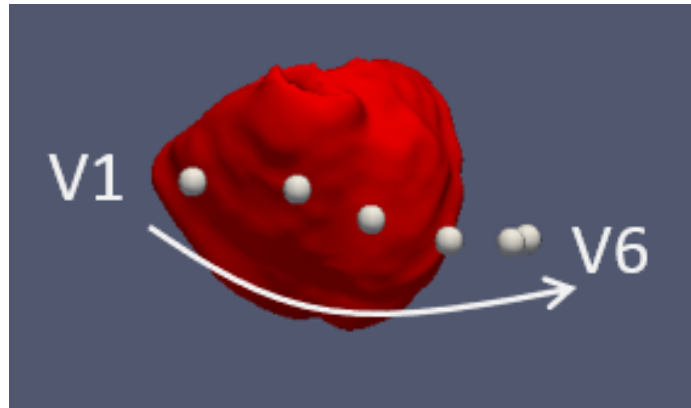


Figure S3. Lead placement for pseudo-ECG computation.

Visualization methods

Single cell, 1D, and 2D results were visualized in Python using the Matplotlib library ¹¹. ECGs were also plotted in Python using the Matplotlib library. Anatomical heart results were visualized in 3D using the VisIt software tool (Lawrence Livermore National Laboratory) ¹².

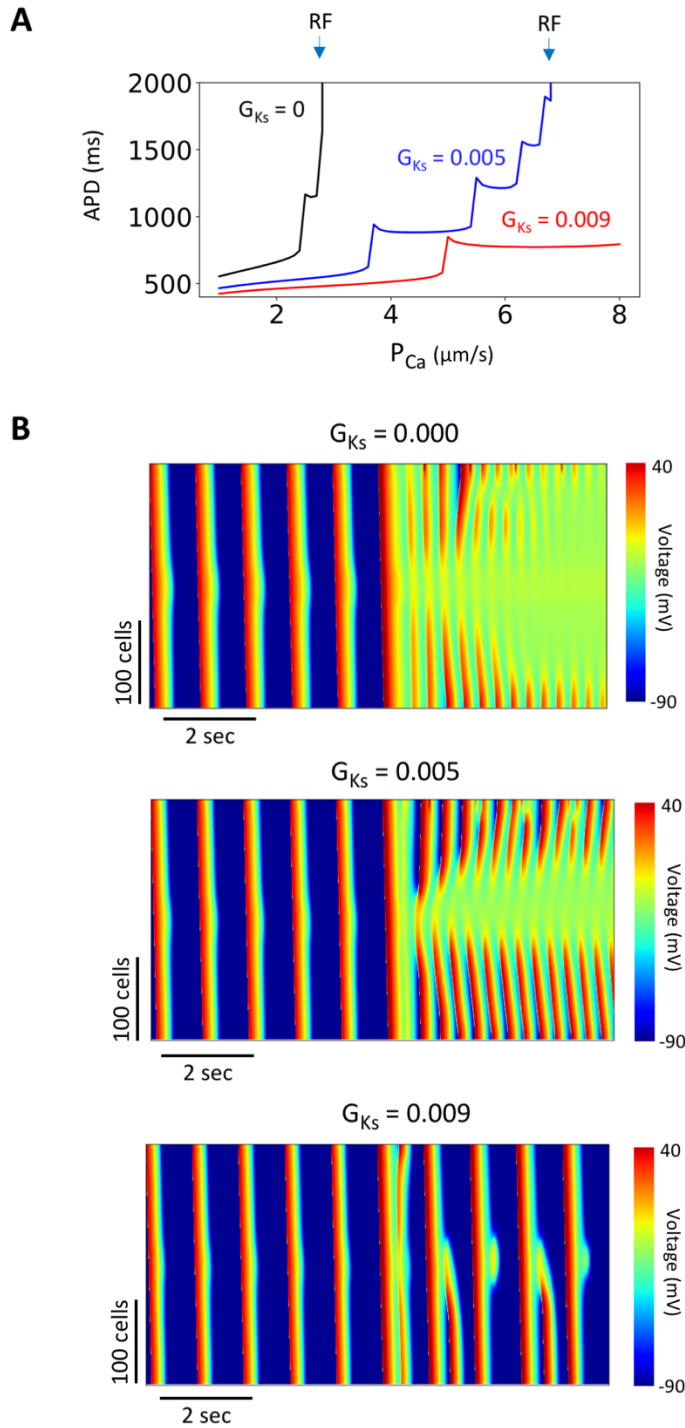


Figure S4. Effect of adding I_{Ks} back to LQT1. A) Single cell APD versus P_{Ca} for LQT1 ($G_{Ks}=0$ mS/ μ F) as in Fig.6A but with varying amounts of I_{Ks} added back. B) Linescans of voltage in a 1D cable for $P_{Ca} = 4.1$ μ m/s at PCL=1000 ms as in Fig.6C but with varying amounts of I_{Ks} added back.

Movie Captions

Movie 1. Spontaneous initiation of arrhythmia in LQT2. Three view movie including ventricle voltage maps (left), wavefronts (colored red, middle), and Purkinje network voltage maps (right). The ECG strip and time marker are shown below. P_{Ca} ramp protocol simulation with $P_{Ca,H} = 2.8 \mu\text{m/s}$ (corresponding to the 6th ECG trace in Fig.2).

Movie 2. 2D simulation of spontaneous focal arrhythmias from the R-from-T mechanism. Voltage map from the 2D tissue corresponding to Fig.3D.

Movie 3. 2D simulation of spontaneous reentrant arrhythmias from the R-from-T mechanism. Voltage map from the 2D tissue corresponding to Fig.3E.

Movie 4. Spontaneous initiation of arrhythmia in LQT3. Three view movie including ventricle voltage maps (left), wavefronts (colored red, middle), and Purkinje network voltage maps (right). The ECG strip and time marker are shown below. P_{Ca} ramp protocol simulation with $P_{Ca,H} = 3.2 \mu\text{m/s}$ (corresponding to the 6th ECG trace in Fig.4).

Movie 5. Spontaneous initiation of arrhythmia in LQT1. Three view movie including ventricle voltage maps (left), wavefronts (colored red, middle), and Purkinje network voltage maps (right). The ECG strip and time marker are shown below. P_{Ca} ramp protocol simulation with $P_{Ca,H} = 3.1 \mu\text{m/s}$ at 60 bpm (corresponding to the 4th ECG trace in Fig.5).

Movie 6. Schematic diagram distinguishing the “R-to-T” and “R-from-T” mechanisms of arrhythmia initiation. Animated version of Figure 8 in the main text.

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