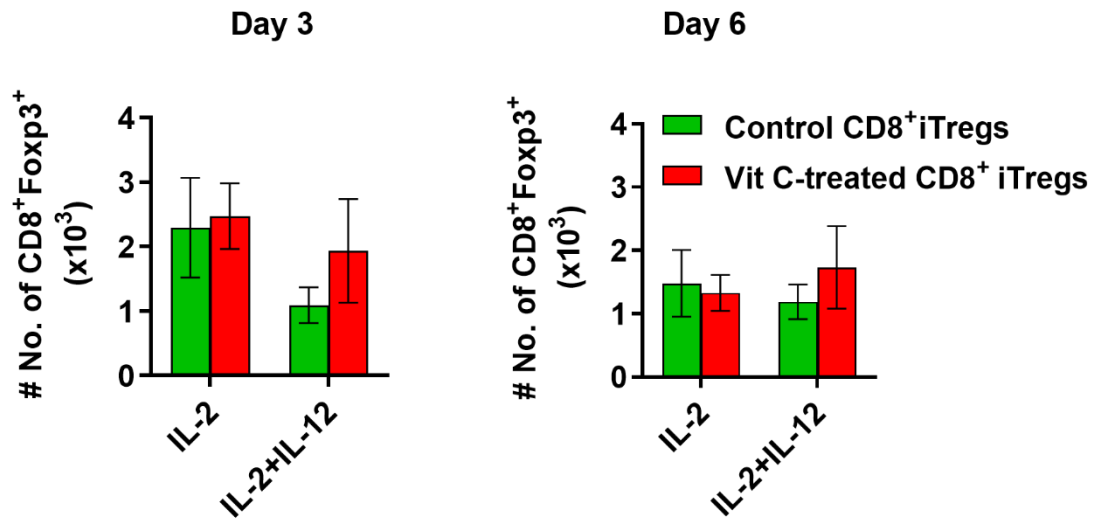


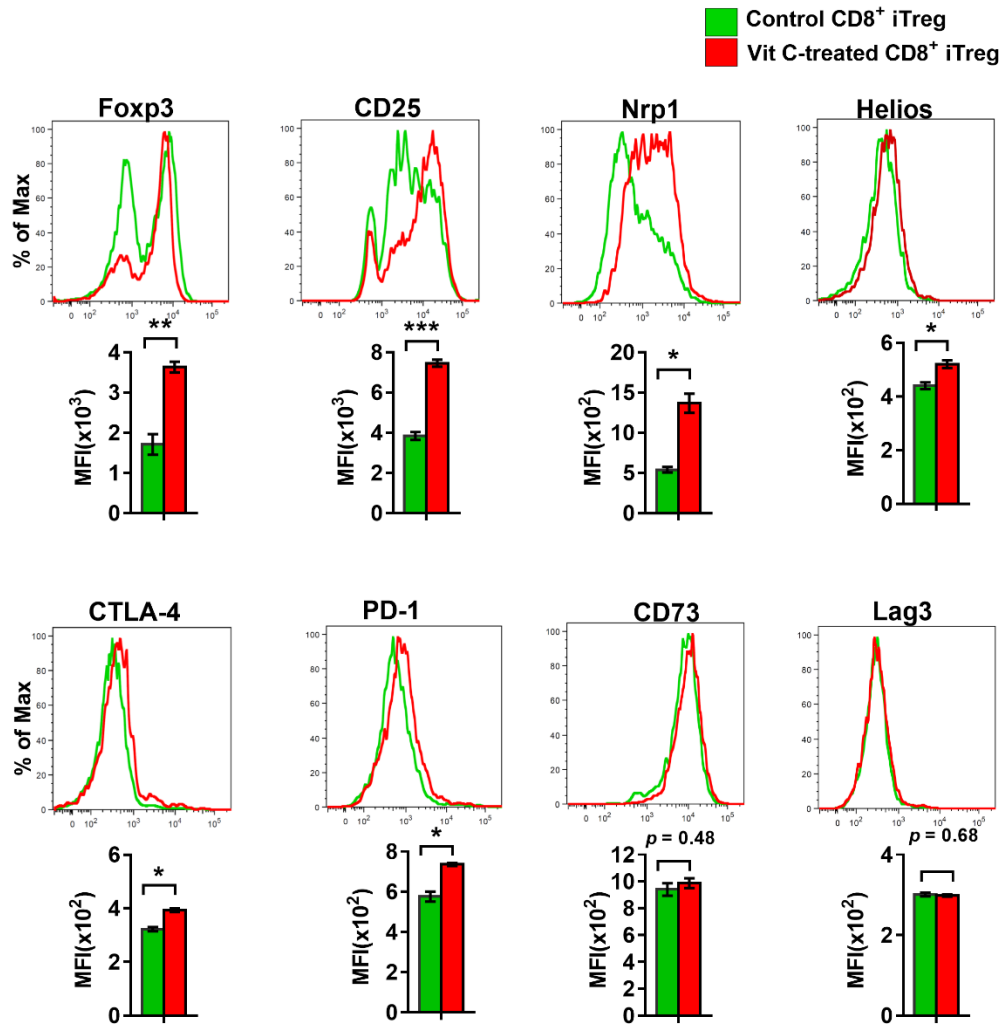
Supplementary Figure 1



**Supplemental Figure 1. Absolute numbers of CD8<sup>+</sup> Foxp3<sup>+</sup> cells after culture for 3 and 6 days.**

On day 5 after CD8<sup>+</sup>iTregs generation, control or Vit C-treated CD8<sup>+</sup> iTregs were harvested, enriched with CD25 microbeads and co-cultured in the presence of IL-2 alone or IL-2+IL-12 recombinant cytokines (n=3/group) for 3 days and 6 days. The absolute number of Foxp3<sup>+</sup> cells was calculated on day 3 or day 6. Data are representative of two independent experiments. *Student's t-test* was used for statistical analysis.

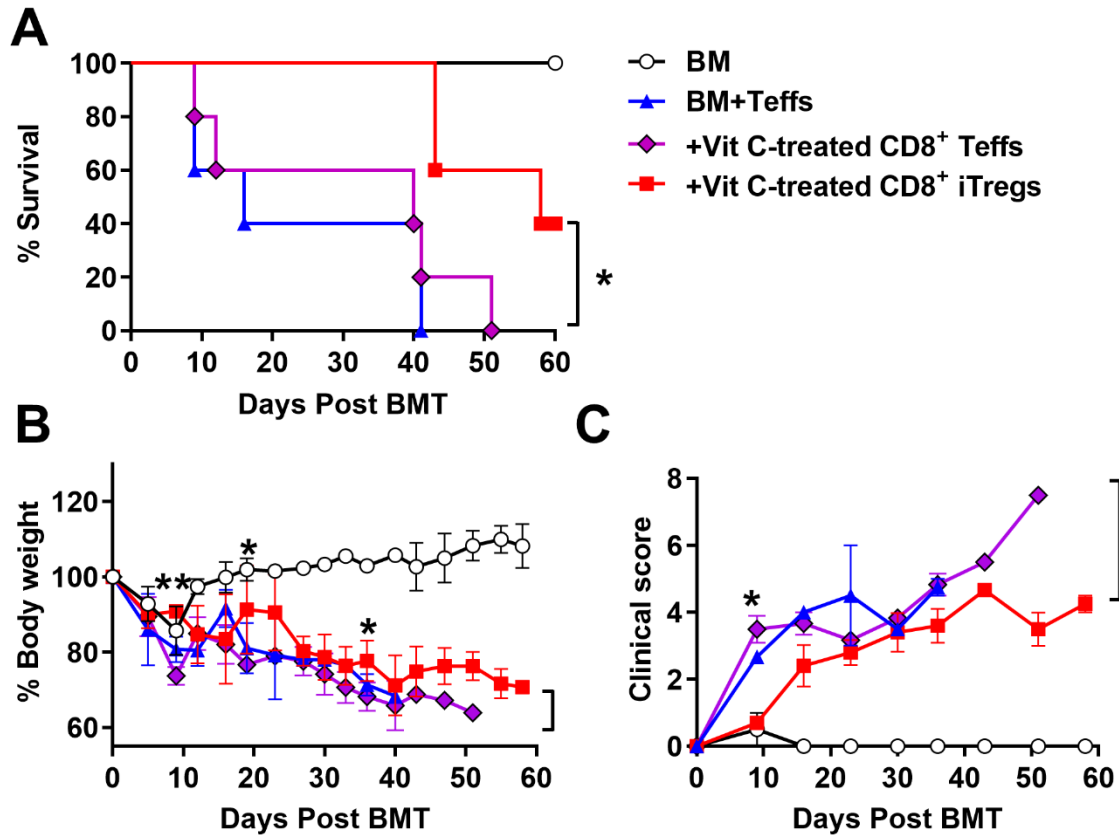
Supplementary Figure 2



**Supplemental Figure 2. Treg canonical makers expressions of control or Vitamin C-treated CD8<sup>+</sup> iTregs on day 5 after *in vitro* generation.**

On day 5 after *in vitro* CD8<sup>+</sup>iTregs generation, control or VitC-treated CD8<sup>+</sup> iTregs were harvested and analyzed for Foxp3, CD25, Nrp1, Helios, CTLA-4, PD-1, CD73, and Lag3 expression. Data are representative of three independent experiments. *Student's t-test* was used for statistical analysis. \*p ≤ 0.05, \*\*p ≤ 0.01 and, \*\*\*p ≤ 0.001.

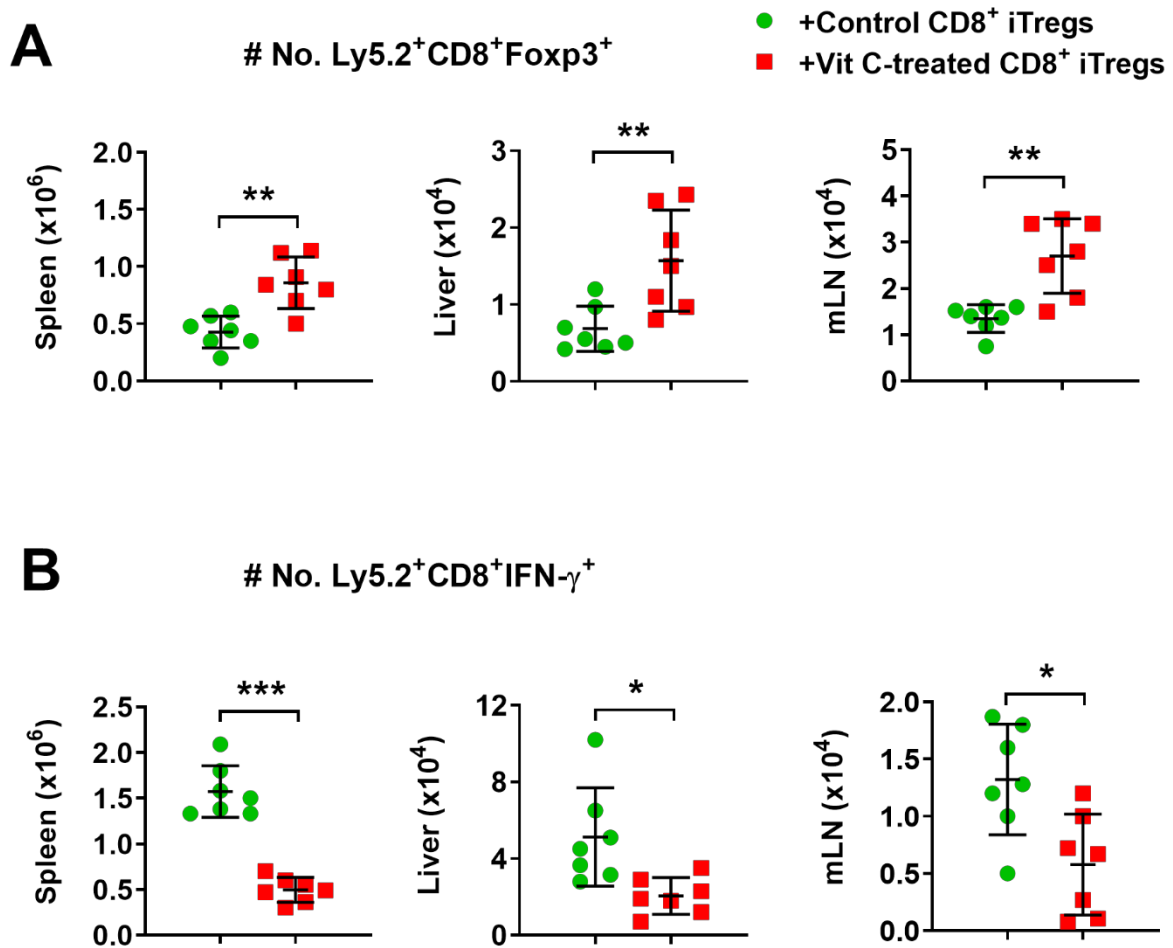
Supplementary Figure 3



**Supplemental Figure 3. Effect of Vitamin C-treated CD8<sup>+</sup> effector T cells on aGVHD.**

Lethally irradiated BALB/c mice were transplanted with  $5 \times 10^6$  BM plus either  $1 \times 10^6$  Vit C-treated CD8<sup>+</sup>Teffs or CD8<sup>+</sup>iTregs. Three days later,  $0.7 \times 10^6$  CD25-depleted T-cells were i.v injected to induce aGVHD. Recipients were monitored for survival rate (A), body weight loss (B), and GVHD clinical signs (C) until day 60 (n=5/group). Log-rank (Mantel-Cox) test was used for statistical analysis of the survival. *Student's t-test* was used for statistic of body weight and GVHD clinical scores. \*p ≤ 0.05 and \*\*p ≤ 0.01.

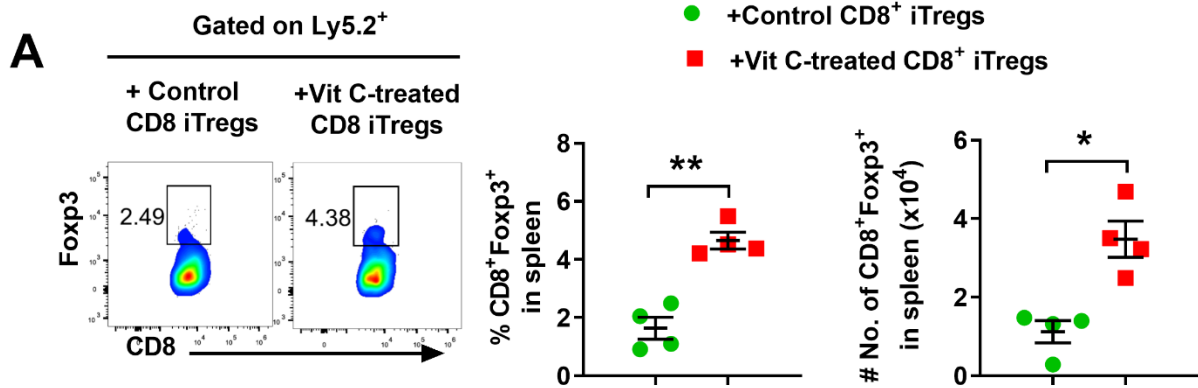
Supplementary Figure 4



Supplemental Figure 4. Absolute numbers of control and Vit C CD8<sup>+</sup> iTregs (Ly5.2<sup>+</sup>) 7 days after being transferred in the recipients.

Lethally irradiated BALB/c mice were transferred with  $5 \times 10^6$  Rag1<sup>-/-</sup> BM and  $1 \times 10^6$  CD8<sup>+</sup> iTregs (Ly5.2<sup>+</sup>). Three days later,  $2 \times 10^6$  CD25-depleted T-cells from C57BL/6 (Ly5.1<sup>+</sup>) congenic mice were i.v. injected to induce GVHD. At day 7 after allo-BMT, spleen, liver and mesenteric lymph nodes (mLNs) were harvested and analyzed. (A) Absolute numbers of transferred control or Vit C-treated CD8<sup>+</sup>Tregs that expressed Foxp3 in recipient spleen, liver or mLN are shown. (B) Absolute numbers of transferred control or Vit C-treated CD8<sup>+</sup>Tregs that secreted IFN- $\gamma$  are shown. Data are combined from two independent experiments (n=7/group). Student's *t*-test was used for statistical analysis. \**p*  $\leq$  0.05, \*\**p*  $\leq$  0.01 and \*\*\**p*  $\leq$  0.001.

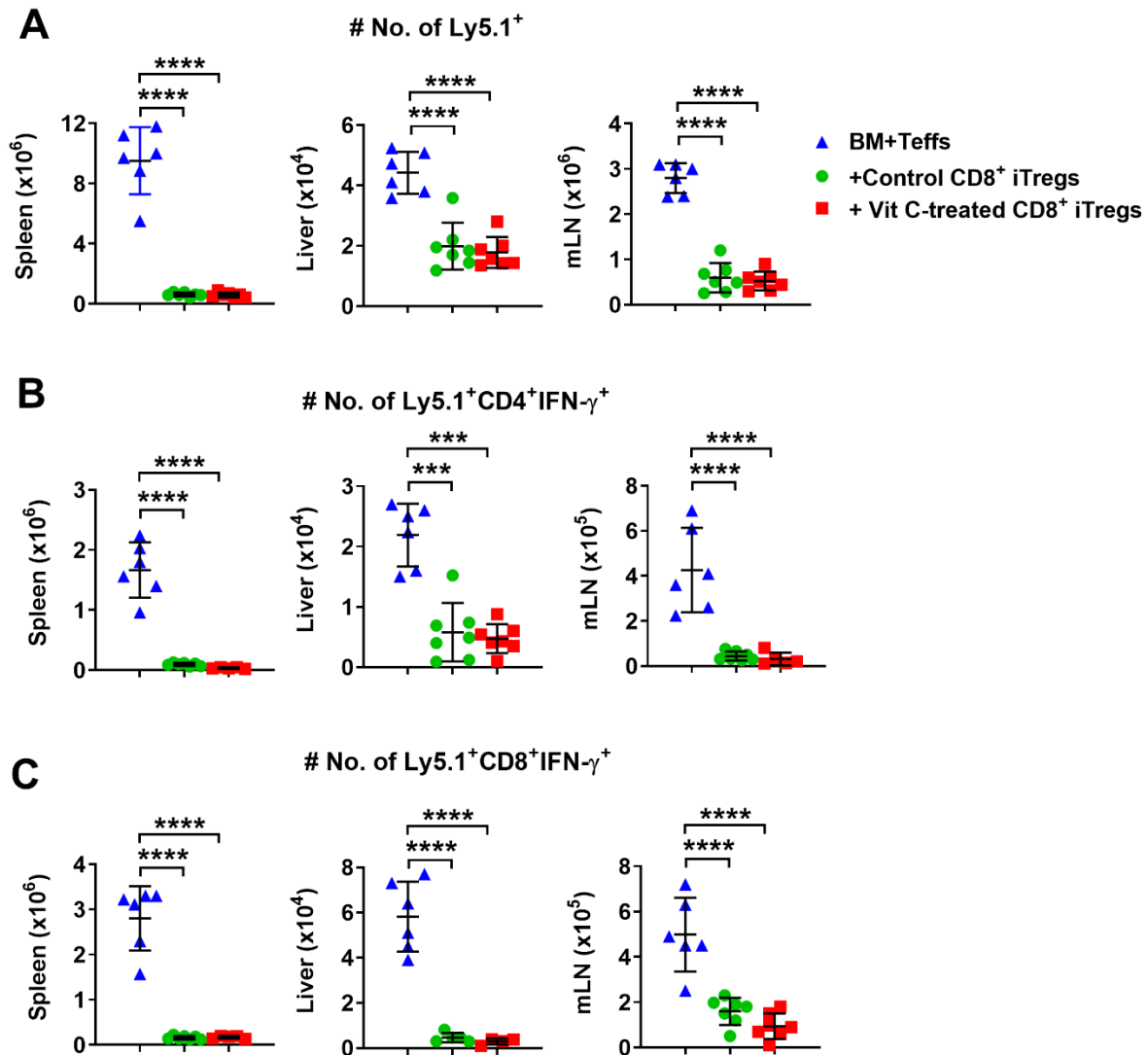
Supplementary Figure 5



**Supplemental Figure 5. Percentages and absolute numbers of transferred control or Vit C-treated CD8<sup>+</sup> iTregs that maintained Foxp3 expression 14 days in the recipients.**

Lethally irradiated BALB/c mice were transferred with  $5 \times 10^6$  Rag1<sup>-/-</sup> BM and  $1 \times 10^6$  CD8<sup>+</sup> iTregs (Ly5.2<sup>+</sup>). Three days later,  $0.7 \times 10^6$  CD25-depleted T-cells from C57BL/6 (Ly5.1<sup>+</sup>) congenic mice were i.v injected to induce GVHD. On day 14 after allo-BMT, the recipient spleens were harvested and analyzed. (A) Frequencies and absolute numbers of Foxp3 (Ly5.2<sup>+</sup>CD8<sup>+</sup>Foxp3<sup>+</sup>) retention among transferred control or Vit C-treated CD8<sup>+</sup> iTregs are shown (n=4/group). *Student's t-test* was used for statistical analysis. \*p ≤ 0.05 and \*\*p ≤ 0.01.

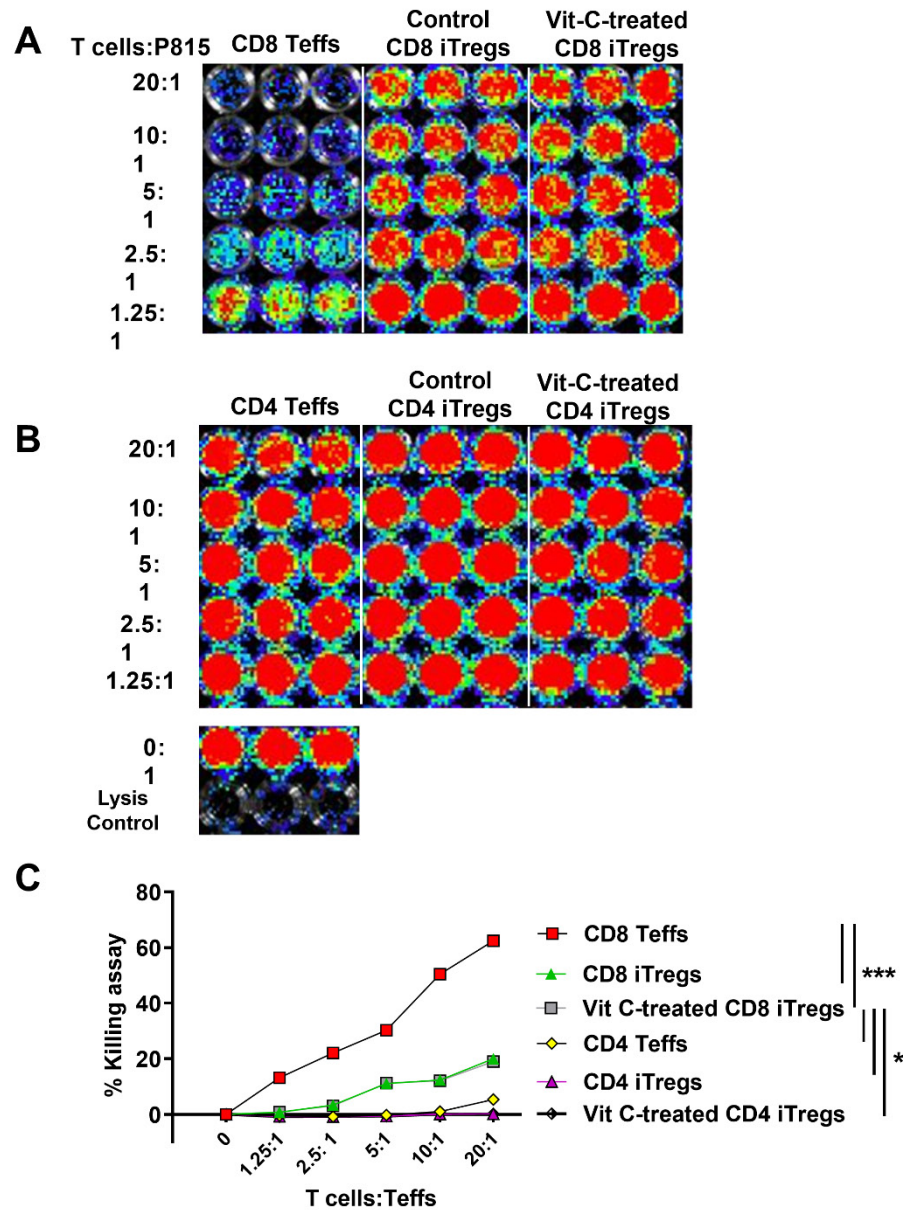
Supplementary Figure 6



**Supplemental Figure 6. Absolute numbers of T effector cells that produced IFN- $\gamma$  in the recipients.**

Lethally irradiated BALB/c mice were transplanted with  $5 \times 10^6$  Rag1<sup>-/-</sup> BM and  $1 \times 10^6$  of either control or Vit C-treated CD8<sup>+</sup> iTregs (Ly5.2<sup>+</sup>). Three days later,  $2 \times 10^6$  CD25-depleted T-cells from C57BL/6 (Ly5.1<sup>+</sup>) congenic mice were i.v injected to induce GVHD. On day 7 after allo-BMT, spleen, liver and mesenteric lymph nodes (mLNs) were harvested and analyzed. (A) Absolute numbers of total Teff cells (Ly5.1<sup>+</sup>) from spleen, liver and mLN are shown. (B-C) Absolute numbers of IFN- $\gamma$ -producing either CD4<sup>+</sup> (B) or CD8<sup>+</sup> Teff cells (C) are shown. Data are combined from two independent experiments (n=7/group). One-way ANOVA was used for statistical analysis. \*\*\*p  $\leq$  0.001 and \*\*\*\*p  $\leq$  0.0001.

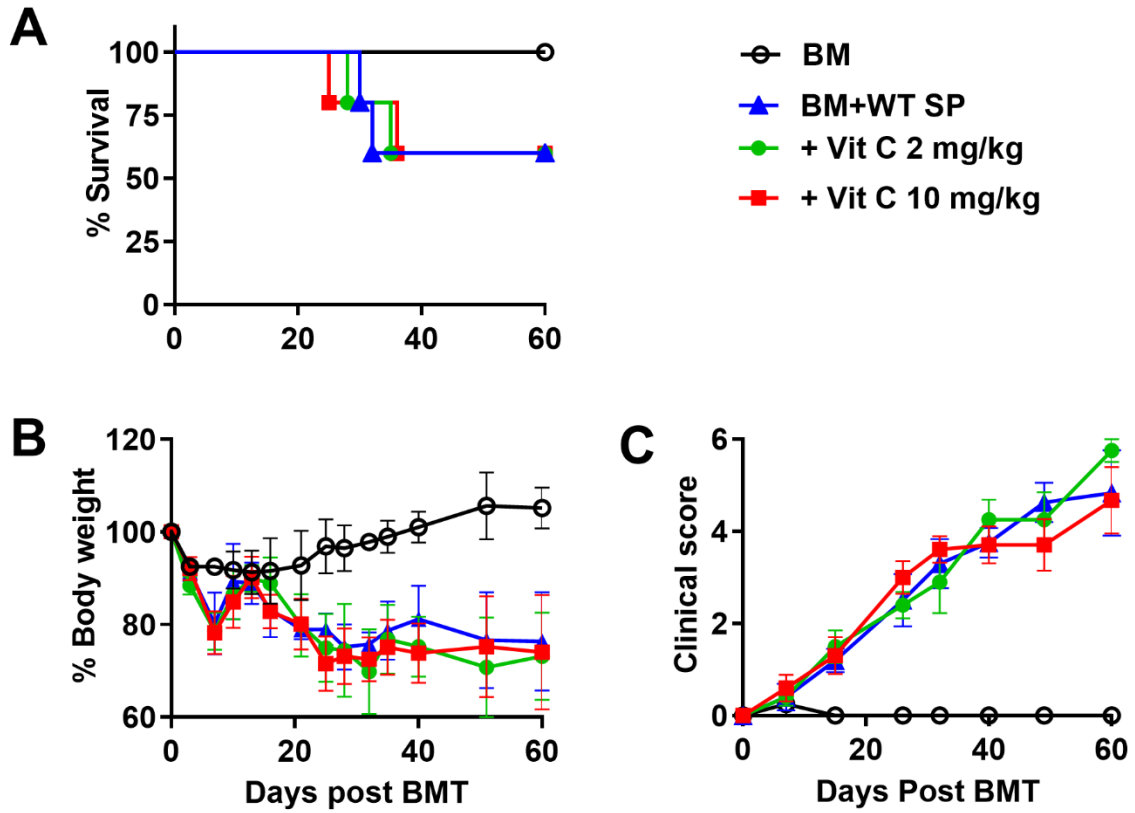
Supplementary Figure 7



**Supplemental Figure 7. Cytotoxic activity of control or VitC-treated CD8<sup>+</sup> iTregs against P815 expressing luciferase.**

Allogeneic CD4 or CD8 Teffs or control CD8<sup>+</sup>iTregs or VitC-treated CD8<sup>+</sup>iTregs (effector;E) were co-cultured with P815-expressing luciferase (T cell : target ratio) 20:1, 10:1, 5:1, 2.5:1 and 1.25:1 at 37°C for 4 hrs. Each ratio was seeded in triplicate wells. After incubation, luciferin was added into the culture. Bioluminescent imaging data were analyzed and quantified using living imager software. (A-B) Luminescence images of 96-well plates show viable P815 tumor cells in red signal. P815-expressing luciferase incubated with lysis buffer was used as a positive control represented lysis activity. (C) Graph depicting the percentages of cytotoxic activity in different E:T ratio are shown (n=3/group). One-way ANOVA was used for statistical analysis. \*p ≤ 0.05 and \*\*\*p ≤ 0.001.

Supplementary Figure 8

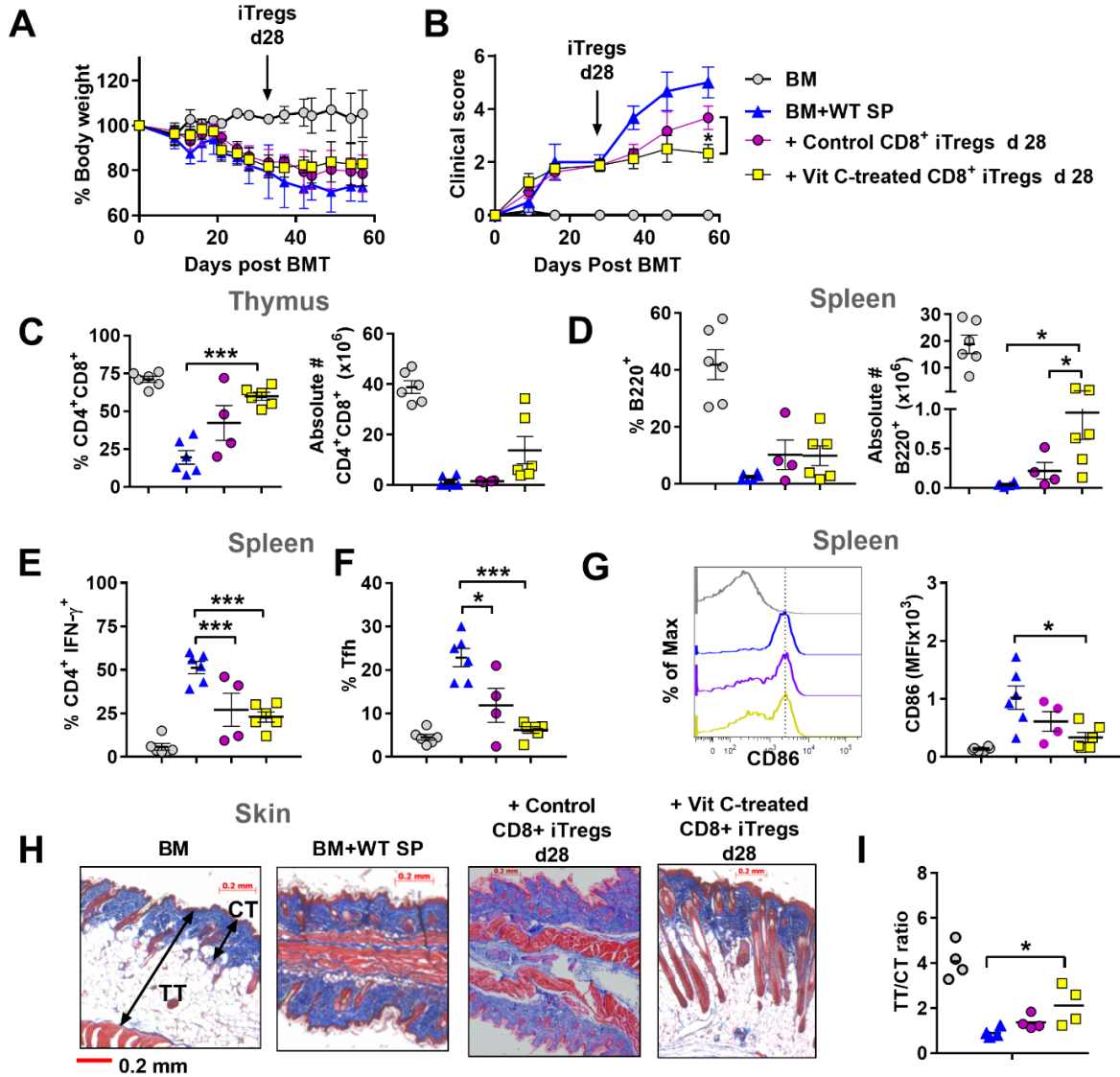


**Supplemental Figure 8. The effect of oral supplement with Vitamin C on cGVHD.**

Lethally irradiated BALB/c recipients were transplanted with TCD-BM ( $5 \times 10^6$ /mouse) and  $0.5 \times 10^6$ /mouse of whole splenocytes from WT B6. The recipients were daily oral gavaged with 2 or 10 mg/kg of Vit C for 21 days. Survival rate (A), body weight loss (B), and GVHD clinical signs (C) were monitored until day 60 ( $n=5$ /group). Log-rank (Mantel-Cox) test was used for statistical analysis of mice survival. *Student's t-test* was used for statistic of body weight loss and GVHD clinical scores.



Supplementary Figure 9



**Supplement Figure 9. Vitamin C-treated CD8<sup>+</sup> iTreg treatment reverses established chronic GVHD.**

(A-B) Lethally irradiated BALB/c mice were transplanted with TCD-BM  $5 \times 10^6$ /mouse and  $0.35 \times 10^6$ /mouse of whole splenocytes from WT B6 donors. The recipients without iTreg transfer serve as a cGVHD control group (WT SP). Tregs were transferred  $0.35 \times 10^6$  /mouse at day 28 post-BMT (d28). All recipient mice were monitored for survival rate (data not shown), body weight (A) and cGVHD clinical scores (B). Experiment was ended on day 50-55 and recipient thymus and spleen were analyzed. Percentages and total number of CD4<sup>+</sup>CD8<sup>+</sup> in thymus (C) and B-cell reconstitution (D) in spleen are shown on gated donor cells. (E-F) Percentages of CD4<sup>+</sup> IFN- $\gamma$ <sup>+</sup> and Tfh (CD4<sup>+</sup>Foxp3<sup>+</sup>PD<sup>hi</sup>CXCR5<sup>+</sup>) are shown in spleen. (G) Histogram and percentage of Mean Fluorescent Intensity (MFI) of CD86 expression are shown. (H) Recipient skin was stained by Masson's Trichrome stain (X5 magnification). (I) Collagen deposition in skin was calculated by dividing total thickness (TT, from epidermis to sub-cutaneous muscle layer) by collagen thickness (CT). Data are combined from two independent experiments (n = 4-6/group). One-way ANOVA was used for statistical analysis. \*p  $\leq$  0.05 and \*\*\*p  $\leq$  0.001.