

Supplementary Material

1 Supplementary Figures and Tables

1.1 Supplementary Figures

a CLUSTAL O(1.2.4) multiple sequence alignment



Supplementary Figure 1. (a) Multiple sequence alignment in Clustal Omega [53] of β thymosins (Tb4, Tb10 and Tb15) amino acid sequences with the help of BLAST online tool (Sequences were taken from Uniprot database). (b) Analysis of antibodies' specificity against T β 4 (Table S1. List of

antibodies). The A375 cells were transfected with plasmids encoding T β 4, T β 10 or T β 15 tagged with 3xHA and upon fixation immunostained with anti-HA and two anti-T β 4 antibodies. Scale bar = 20 μ m. SC: antibodies from Santa Cruz Biotechnology Inc. MM: antibodies from Merck-Millipore. (c) Immunocytochemical analysis of A375-sh*TMSB4X* and A375-shscramble clones with anti-T β 4 antibodies (SC). Scale bar = 20.



Supplementary Figure 2. Quantitative RT-PCR amplification curves for *HPRT1*, *TMSB4X*, *TMSB10* and *TMSB15* obtained from cDNA of WM1341D (a) and A375 (b) cells.



Focal adhesion

Supplementary Figure 3. Visualization of vinculin and $3xHA-T\beta4$, $3xHA-T\beta10$ or $3xHA-T\beta15$ respectively in WM1341D and A375 cells transfected with pLVX-IRES-puro- $3xHA-T\beta4$, pLVX-IRES-puro- $3xHA-T\beta10$ or pLVX-IRES-puro- $3xHA-T\beta15$. Arrows point at focal adhesions. Scale bar = $20 \ \mu m$.



Supplementary Figure 4. (a) Negative controls for PLA assay. (b) Visualization of vinculin, ILK, FAK, a parvin, Pinch 2 and integrin aVb3 in WM1341D and A375 cells. The cells were additionally stained with antibodies recognizing all isoactins (clones AC40 or C11) or fluorescently labeled phalloidin. Arrows point at focal adhesions. Scale bar = $20 \mu m$.



Supplementary Figure 5. (**a-b**) Analysis of vinculin level in WM1341D and A375 cell lysates: (**a**) a representative immunoblot and Ponceau S staining of the whole membrane, (**b**) densitometrical analysis (n=6).



Supplementary Figure 6. Analysis of FAs' formation in tested cell lines: Visualization of a-parvin and F-actin in WM1341D and A375 cells fixed 24 h after seeding the cells (**a**). Estimation of FAs' number (**b**) and size of FA's (**c**). **** denotes significant difference between marked groups. Arrows point at focal adhesions. Scale bar = $20 \mu m$.



Supplementary Figure 7. Analysis of cells' size (Forward scatter - FSC). Cells were analyzed with a BD FACS Calibur Flow Cytometry System using BD CellQuest Pro Software; cells were washed three times with cation-free PBS supplemented with 2 mM EDTA and then transferred to a FACS tube (n=3). Cells were gated on single living cells. The number of analysed cells in every experiment was 10000. a) Histogram showing distribution of cells' size in WM1341D and A375 cell lines. b) Histogram statistic.



Supplementary Figure 8. qRT-PCR analysis of *TMSB4X* (a) *TMSB10* (b) and *TMSB15* (c) expression level in A375-shscr and A375-shTMSB4X clones.



Supplementary Figure 9. Analysis of FAs' formation in tested cell lines: Visualization of a-parvin and F-actin in A375 cells with lowered *TMSB4X* expression. (a) Estimation of FAs' number (b) and size of FA's (c). ****denotes significant difference between marked groups. Arrows point at focal adhesions. Scale bar = $20 \mu m$.



Supplementary Figure 10. Single cell trajectory in 2D space with quantitative parameters defining directionality. Directionality ratio from the start point (P1) to the end-point (PN) of the cell trajectory is calculated by dividing the displacement (distance to origin, DTO) and total length of trajectory (total distance, TD).



Supplementary Figure 11. Analysis of Matrigel[®] coated wells after scratching. Cells and surfaces of wells were immunostained using anti-laminin antibodies and counterstained with fluorescently-labeled phalloidin. Wound Maker did not remove Matrigel[®] from surfaces of wells. Scale bar = 200 μ m.



Supplementary Figure 12. Evaluation of T β 4 role in A375 clones with decreased T β 4 level. Representative pictures of cells migrating collectively on plastic recorded over 72h for A375-*shTMSB4X* clones (n=3).



Supplementary Figure 13. Evaluation of T β 4 role in A375 clones with decreased T β 4 level. Representative pictures of cells migrating collectively on MatrigelTM recorded over 72h for A375-*shTMSB4X* clones (n=3).



Supplementary Figure 14. Visualization of β -catenin, N-cadherin and ZO-1 in WM1341D and A375 cells seeded at different densities. The cells were additionally stained with fluorescently labeled phalloidin. Arrows point at cellular protrusions involved in cell movement such as invadopodia and lamellipodia. Scale bar = 100 μ m or 20 μ m.



Supplementary Figure 15. qRT-PCR analysis of *SNAI1* (a) *VIM* (b) expression level in A375-*shscr* and A375-*shTMSB4X* clones.

1.2 Supplementary Tables

Antibody	Company	Immunocyt. /PLA	WB
rabbit anti-Tβ4 (FL-44)	Santa Cruz Biotechnology Inc.	1:50	-
rabbit anti-cortactin (H-191)	Santa Cruz Biotechnology Inc.	1:50	1:200
rabbit anti-Tks-5 (M-300)	Santa Cruz Biotechnology Inc.	1:50	1:200
goat anti-HA (Y-11)	Santa Cruz Biotechnology Inc.	1:50	1:200
rabbit anti-FAK (C-903)	Santa Cruz Biotechnology Inc.	1:50	1:200
mouse anti- $\alpha V\beta 3$	Merck-Millipore	1:100	-
rabbit anti-Tβ4	Merck-Millipore	1:100	1:1000
rabbit anti-pan laminin	Sigma-Aldrich	1:60	-
rabbit anti-ILK	Sigma-Aldrich	1:100	1:1000
rabbit anti-total actin C11	Sigma-Aldrich	1:100	-
mouse anti-total actin AC40	Sigma-Aldrich	1:100	-
rabbit anti-α-parvin	Cell Signaling	1:400	-
rabbit anti-vimentin (D21H3)	Cell Signaling	-	1:1000
rabbit anti-b-catenin (D10A8)	Cell Signaling	-	1:1000
rabbit anti-SNAI2 (C19G7)	Cell Signaling	-	1:1000
rabbit anti-SNAI1 (C15D3)	Cell Signaling	-	1:1000
rabbit anti-ZO-1 (D7D12)	Cell Signaling	-	1:1000
rabbit anti-ZEB1 (D80D3)	Cell Signaling	-	1:1000
mouse anti-ZO-1 (1A12)	Thermo Fisher Scientific	1:200	-
mouse anti-vinculin (V284)	AbD Serotec (Bio- Rad)	1:100	1:1000
HRP-conjugated anti-rabbit	Cell Signaling	-	1:4000
HRP-conjugated anti-mouse	Cell Signaling	-	1:4000
donkey anti-goat-Alexa Fluor® 488	Invitrogen	1:200	-
donkey anti-mouse-Alexa Fluor® 488	Invitrogen	1:200	-
donkey anti-mouse-Alexa Fluor® 568	Invitrogen	1:200	-
donkey anti-mouse-Alexa Fluor® 647	Invitrogen	1:200	-
donkey anti-rabbit-Alexa Fluor® 488	Invitrogen	1:200	-
donkey anti-rabbit-Alexa Fluor® 568	Invitrogen	1:200	-

Supplementary Table 1. List of antibodies used in the study.

Primer	Sequence	Amplicon size (nt)	Tm (°C)
<i>TMSB4X_</i> f	5'→caaccatgtctgacaaacc	96	60
<i>TMSB4X_</i> r	5'→aaggcagtggatttttctct		60
<i>TM</i> SB10_f	5'→ccagacatgggggaaat	94	60
<i>TM</i> SB10_r	5'→caatggtctctttggtcg		60
<i>TMSB15</i> _f	5'→atgagtgataagccagacttg	122	60
<i>TMSB15</i> _r	5'→cactctttctcttgctggatag		60
<i>VIM_</i> f	5`→aaagtccgcacattcgagca	482	60
<i>VIM_</i> r	5`→ggtggacgtagtcacgtagc		60
<i>SNAI</i> 1_f	5'→accccaatcggaagcctaac	151	60
<i>SNAI</i> 1_r	5'→tcccagatgagcattggcag		60
<i>HPRT1_</i> f	5'→gaccagtcaacaggggacat	165	60
<i>HPRT1_</i> r	5'→gcttgcgaccttgaccatct		60

Supplementary Table 2. List of qPCR primers used in the study.

Vector	Primers sequences	Description
pLVX-IRES-puro	5' → tagaggatctatttccggtgaattcaccatgtctga caaaccc 5' → taggggggggggggggggggggggggggggggggggg	From pLVX-IRES-tdTomato-FlagAkt1 plasmid, a gift from Eva Gonzalez (Addgene plasmid # 64831) (Kajno et al., 2015).there were removed sequnces encoding FlagAkt1, tdTomato, part of IRES and WPRE sequences by <i>EcoR</i> I and <i>Kpn</i> I restriction enzymes. Next we reconstructed IRES and WPRE sequences and simultaneously cloned cDNA encoding puromycin resistance gene amplified from the SHC016 vector (Sigma- Aldrich).
pLVX-IRES-puro- 3xHA-Tβ4	5' → tagaggatctatttccggtgaattcaccatggatta cccatacg 5' → cactcatggtgaattcagaagcgtaatctg 5' → ttctgaattcaccatgagtgataagccag 5' → taggggggggggggggggggggggggggggggggggg	As a backbone vector was used pLVX-IRES- puro , which generation is described above. Between <i>EcoRI</i> and <i>BamHI</i> restriction sites there were cloned sequences coding 3xHA tag amplified from p3xHA-C1 plasmid (Müller et al., 2012) and cDNA encoding T β 4 amplified from cDNA of the A375 cells.
pLVX-IRES-puro- 3xHA-Tβ10	5' → tagaggatctatttccggtgaattcaccatggatta cccatacg 5' → cagacatggtgaattcagaagcgtaatctg 5' → ttctgaattcaccatgtctgacaaaccc 5' → taggggggggggggggggggggggggggggggggggg	As a backbone vector was used pLVX-IRES- puro , which generation is described above. Between <i>EcoRI</i> and <i>BamHI</i> restriction sites there were cloned sequences encoding 3xHA tag amplified from p3xHA-C1 plasmid (Müller et al., 2012) and cDNA encoding T β 10 amplified from cDNA of the A375 cells.
pLVX-IRES-puro- 3xHA-Tβ15	5' → tagaggatctatttccggtgaattcaccatggatta cccatacg 5' → ctgccatggtgaattcagaagcgtaatctg 5' → ttctgaattcaccatggcagacaaaccag 5' → taggggggggggggggggggggggggggggggggggg	As a backbone vector was used pLVX-IRES- puro , which generation is described above. Between <i>EcoRI</i> and <i>BamHI</i> restriction sites there were cloned sequences encoding 3xHA tag amplified from p3xHA-C1 plasmid (Müller et al., 2012) and cDNA encoding T β 15 amplified from cDNA of the A375 cells.

Supplementary Table 3. List of clonning vectors used in the study.

Number	Sequence			
TMSB4X MISSION shRNA				
TRCN0000158860	ccgggagaaattcgataagtcgaaactcgagtttcgacttatcgaatttctcttttttg			
TRCN0000219784	ccggatgtctgacaaacccgatatgctcgagcatatcgggtttgtcagacattttttg			
TRCN0000163494	ccgggaagacagagacgcaagagaactcgagttetettgcgtetetgtettettttttg			
TRCN0000164355	ccggcgattgaacaggagaagcaagctcgagcttgcttctcctgttcaatcgttttttg			
TRCN0000219785	ccggacaagcattgccttcttatttctcgagaaataagaaggcaatgcttgttttttg			
Non-Target shRNA				
SHC016	ccgggcgcgatagcgctaataatttctcgagaaattattagcgctatcgcgcttttt			

Supplementary Table 4. List of silencing sequence used in the study.