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Supplemental Information

Differential Functions of Splicing Factors

in Mammary Transformation and

Breast Cancer Metastasis

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SUPPLEMENTAL FIGURES



Figure S1. SFs exhibit different alteration patterns in human breast tumors and are associated with different breast cancer subtypes (related to Figure 1).

(A) Type of SF alterations detected in TCGA human breast tumors (n=960), normalized to total SF alterations for each gene. % of tumors that exhibit an alteration is shown for each of the following categories: i) mRNA upregulation (Z-score \geq 2.0), ii) both mRNA upregulation and DNA amplification, iii) DNA amplification only, iv) mRNA downregulation (Z-score \leq 2.0), v) both mRNA downregulation and DNA deletion, vi) DNA deletion only, and vi) mutation.

(B) SF expression in breast tumors classified into subtypes using the PAM50 signature is compared to 'normal' breast tissues (n=99). SF expression in each subtype is compared to the combination of all other subtypes (t-test; $***P<1.10^{-10}$, **P<0.00001, *P<0.01). Only SFs that exhibit differences between subtypes are shown.

(C) Graphical representation of SF alterations in TCGA TNBC (n=82). Copy number and expression changes are assessed by DNAand RNA-seq. Individual genes are represented as rows, and individual patients as columns. Alterations in breast cancer genes *TP53*, *MYC*, *ERBB2* and *BRCA1* are in the lower panel.



Figure S2. TCGA breast tumors with high SF levels exhibit alterations in alternatively spliced isoforms (related to Figure 1). (A) TCGA human breast tumors are classified based on *SF*-expression into two groups: *SF*-high (Z-score>1.5) or *SF*-low (Z-score ≤ 0) for each listed SF. The number of samples in each category is indicated.

(B) Number of DSEs detected by RNA-seq in *SF-high vs. SF-low* TCGA breast tumors in at least 10 tumor samples ($|\Delta PSI| \ge 10\%$; FDR<5%; *P*<0.01) sorted by AS event type (CA: cassette exon; MXE: mutually exclusive exon; RI: retained intron; A5'SS: alternative 5' splice site; A3'SS -alternative 3' splice site).

(C) Number of *SF-high* tumor samples in which DSEs are detected, plotted for each SF. % of DSEs detected in \geq 80 tumors is indicated. D) Skipped (Δ PSI \leq -10%) and included (Δ PSI \geq 10%) DSEs in SF-high *vs.* SF-low TCGA breast tumor samples reproducibly detected in \geq 10 tumor samples were plotted by Δ PSI values. % of skipped and included DSEs are indicated.



Figure S3. Stable overexpression of specific SFs in human mammary epithelial MCF-10A cells promotes early changes in mammary acinar morphology, size and proliferation (related to Figure 2).

(A) SF mRNA expression in SF-OE stable MCF-10A cell lines as measured by RT-QPCR using specific primers and normalized to control MCF-10A cells (n=3 biological replicates; mean±SD).

(B) Protein expression of T7-tagged SFs stably overexpressed in MCF-10A cells as detected by western blot using a T7 mouse antibody, with a β -catenin loading control. Stars indicate nonspecific bands on a longer exposure.

(C) Representative brightfield images of acini size and morphology in control and SF-OE MCF-10A cells from day 1 to 8 (Scale bar: 50 µm).

(D) Acinar sizes of control and SF-OE acini from day 1 to 8 (n=3 biological replicates; >100 acini per experiment; Mann-Whitney ***P < 0.0001; mean±SD.

(E) Proliferation of control and SF-OE acini is assessed by immunostaining with a Ki-67 antibody quantified as % of Ki-67-positive in day-8 acini ($n\geq3$ biological replicates; >50 acini per experiment).



Figure S4. SF-OE in human mammary MCF-10A cells leads to differential splicing and gene expression (related to Figure 3). **(A-D)** Similarities and differences in differentially spliced events (DSEs).

(A) Skipped ($\Delta PSI \le -10\%$) and included ($\Delta PSI \ge 10\%$) DSEs in SF-OE vs. control MCF-10A day-8 acini plotted by ΔPSI values for each SF and for each AS event type (CA: cassette exons; A3'SS: alternative 3' splice site; A5'SS: alternative 5' splice site; MXE: mutually exclusive exon; RI- retained intron. % skipped and included DSEs are indicated.

(B) Gene Set Enrichment Analysis for DSEs detected in SF-OE MCF-10A day-8 acini showing the top 10 Hallmark gene sets.

(C) Shared DSGs in SF-OE MCF-10A day-8 acini for each indicated SF pair. Bubble size is proportional to the number of shared DSGs and color indicates *P*-value. See also Table S3G.

(**D**) % of DSEs that affect a region containing an annotated protein domain (*i.e.*, IPRO ID) for both unique and shared DSEs in SRSF4-, SRSF6- and TRA2 β -OE MCF-10A day-8 acini. Fisher's exact test *P*-value is indicated. See also Table S3H.

(E-G) Similarities and differences in differentially expressed genes (DEGs).

(E)Volcano plots depicting log_2 fold gene expression changes (log_2 FC) and significance as $-log_{10}$ (q-value) for each in SF-OE vs. control MCF-10A day-8 acini. Significant genes are shown in blue, non-significant in grey. See also Table S4.

(F) Overlap in DEGs in SF-OE MCF-10A day-8 acini for each indicated SF pair. Bubble size is proportional to the number of shared differentially expressed genes and color indicates *P*-value. See also Table S4.

(G) Gene Set Enrichment Analysis for DEGs detected in SRSF4,-SRSF6-, and TRA2 β -OE MCF-10A day-8 acini showing the top biological functions. See also Table S4.



Figure S5. SF levels affect cell migration and invasion (related to Figure 5).

(A-C) SF-OE promotes different types of cell movement in human mammary MCF-10A cells.

(A) Representative images of intra-acinar cell movement of control or SF-OE MCF-10A day-4 acini imaged during an 18h period by live cell microscopy. Cells moving abnormally outside or inside the acini are indicated by arrowheads (scale bar: 50µm).

(B-C) Quantification of control or SF-OE MCF-10A cell migration in a wound-healing assay (B), or in transwell migration assays (C). The plot shows the distribution and the median (horizontal line) for each condition ($n\geq4$ biological replicates; t-test ****P*<0.0001, ***P*<0.005, **P*<0.05).

(D-J) SF levels affect cell invasion in TNBC MDA-MB231 cells.

(D) Relative cell number measured by MTT assay at days 1 to 4 in CTLsh or SFsh MDA-MB231 cell lines +/-DOX (n=3 biological replicates; mean +/-SD).

(E) Cell doubling time as calculated from (D). (n=3 biological replicates; mean \pm SD; t-test, n.s. not significant).

(F-G) Migration of CTLsh or SFsh MDA-MB231 cells +/-DOX in 2D wound-healing (F) or 2D transwell assay (G) ($n \ge 3$; t-test, **P < 0.005, *P < 0.05).

(H-I) Representative images of CTLsh or SFsh MDA-MB231 cells, grown in 3D +/-DOX, imaged at day 8 with brightfield (H) and fluorescence (I) (scale bar: 100µm). Cell expressed DsRed once the shRNA is induced.

(J) Maximal projection reconstruction of representative Z-stack fluorescent confocal images of TRA2 β sh2 MDA-MB231 cells, grown in 3D +/-DOX, imaged at day 8 for GFP and DsRed expression (scale bar: 100 μ m). Cells that integrate the shRNA-containing TRMPV plasmid constitutively express GFP, as well as DsRed once the shRNA is expressed.



Figure S6. TRA2β-KD and TRA2β-regulated spliced isoforms affect cell invasion in TNBC cells (related to Figure 5 and 6).

(A-C) TRA2β-KD affects cell invasion and spliced isoforms in SUM159PT TNBC cells.

(A) TRA2 β protein expression in TRA2 β sh2 SUM159PT cells +/-DOX compared to control. TRA2 β levels are quantified 72h after -1X DOX treatment by western blotting using a TRA2 β -specific antibody and normalized to actin. % TRA2 β expression +DOX is normalized to -DOX (n=3; mean±SD; *P<0.05).

(B) Maximal projection reconstruction of representative Z-stack fluorescent confocal images of TRA2 β sh2 and control SUM159PT cells, grown in 3D +/-DOX, imaged at day 5 for GFP and DsRed expression (scale bar: 500 μ m). Maximal projection images are composed of 30-35 Z-stack images spaced every 55 μ m. At least 15 fields are imaged per condition. Cells that integrate the shRNA-containing TRMPV plasmid constitutively express GFP, as well as DsRed once the shRNA is expressed. Control cells are stained with calcein-AM live-cell dye.

(C) RT-PCR validations of selected DSEs in TRA2 β sh2 SUM159PT and control cells ±DOX, as in Figure 6D. (n=3; mean±SD; t-test, ***P<0.0005; *P<0.005; *P<0.005; n.s. not significant).

(E-G) TRA2β-regulated isoforms of CCDC88C and TANK affect cell invasion and morphogenesis of MDA-MB231 TNBC cells.

(E,F) DOX-inducible shRNAs targeting mRNA isoforms of TANK+ex3 (E) or CCDC88C+ex26 (F), decrease the expression of exoncontaining isoforms in 3D MDA-MB231 cells grown on day 8 as detected by RT-PCR, and compared to TRA2 β sh2. A representative gel is shown, along with isoform structures. PSI values for all samples and Δ PSI for significant samples are calculated from RT-PCR for +DOX vs. -DOX (n=3; mean±SD; t-test, *P<0.05).

(G) Maximal projection reconstruction of representative Z-stack fluorescent confocal images of TANK+ex3sh or CCDC88C+ex26sh MDA-MB231 cells grown in 3D, ±DOX, imaged at day 6 for GFP and DsRed expression (scale bar: 200µm), and compared to TRA2 β sh2 MDA-MB231 cells. Maximal projection images are composed of 30-35 Z-stack images spaced every 55 µm. At least 15 fields are imaged per condition. Cells constitutively express GFP, whereas DsRed is expressed with the shRNA is activated.



Figure S7. TRA2β-KD decreases metastatic burden in TNBC metastasis models (related to Figure 7).

(A-J) TRA2β-KD decreases metastatic burden in a sponteanous metastasis model of TNBC.

(A) TRA2β levels are assessed by western blot in primary tumors from animals injected with CTLsh or TRA2βsh2 MDA-MB231 cells ±DOX. DOX induces shRNA expression.

(B,C) Primary tumor size is measured at necropsy in CTLsh and TRA2βsh2 tumors (B) and representative H&E-stained tumor sections are shown (scale bar: 1mm) (C) at 8 weeks post-injection.

(D) % of animals that exhibited macro-metastases at different organ sites in animals injected with CTLsh and TRA2 β sh2 MDA-MB231 cells ±DOX is assessed at necropsy at 8 weeks post-injection.

(E-J) Lung (E-G) and liver (H-J) metastasis in animals injected with CTLsh or TRA2 β sh2 MDA-MB231 cells ±DOX are scored through the whole organ using multiple H&E sections evenly distributed every 2mm. The number of metastatic foci per mm² (E, H) and the average size of metastatic foci (F, I) are quantified (n≥4; t-test ***P*<0.001, **P*<0.01, n.s. not significant). Representative H&E-stained frozen tumor sections are shown (scale bar: 1mm) (G, J).

(K-N) TRA2β-KD decreases lung tumors formation in an intravenous experimental metastasis model of TNBC.

(**K**) MDA-MB231 TRA2βsh2 cells are injected into the tail vein of NSG mice; metastases are monitored by bioluminescence imaging and histopathology.

(L) Bioluminescence detection of metastasis in mice injected with MDA-MB231 TRA2 β sh2 cells ±DOX at 6 weeks post-injection. DOX induces shRNA expression.

(M) Representative H&E pictures from lung sections of mice injected with MDA-MB231 TRA2 β sh2 cells \pm DOX at 6 weeks post-injection (scale bar: 2 mm). Metastases are scored through the whole organ using sections evenly distributed every 2mm and quantified in D.

(N) Quantification of metastasis burden (% of metastatic organ area relative to the whole organ area) in mice injected with MDA-MB231 TRA2 β sh2 cells +/-DOX at 6 weeks post-injection (n≥4; t-test, *P<0.02).