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## Supplementary Materials for

### **UPR<sup>ER</sup> promotes lipophagy independent of chaperones to extend life span**

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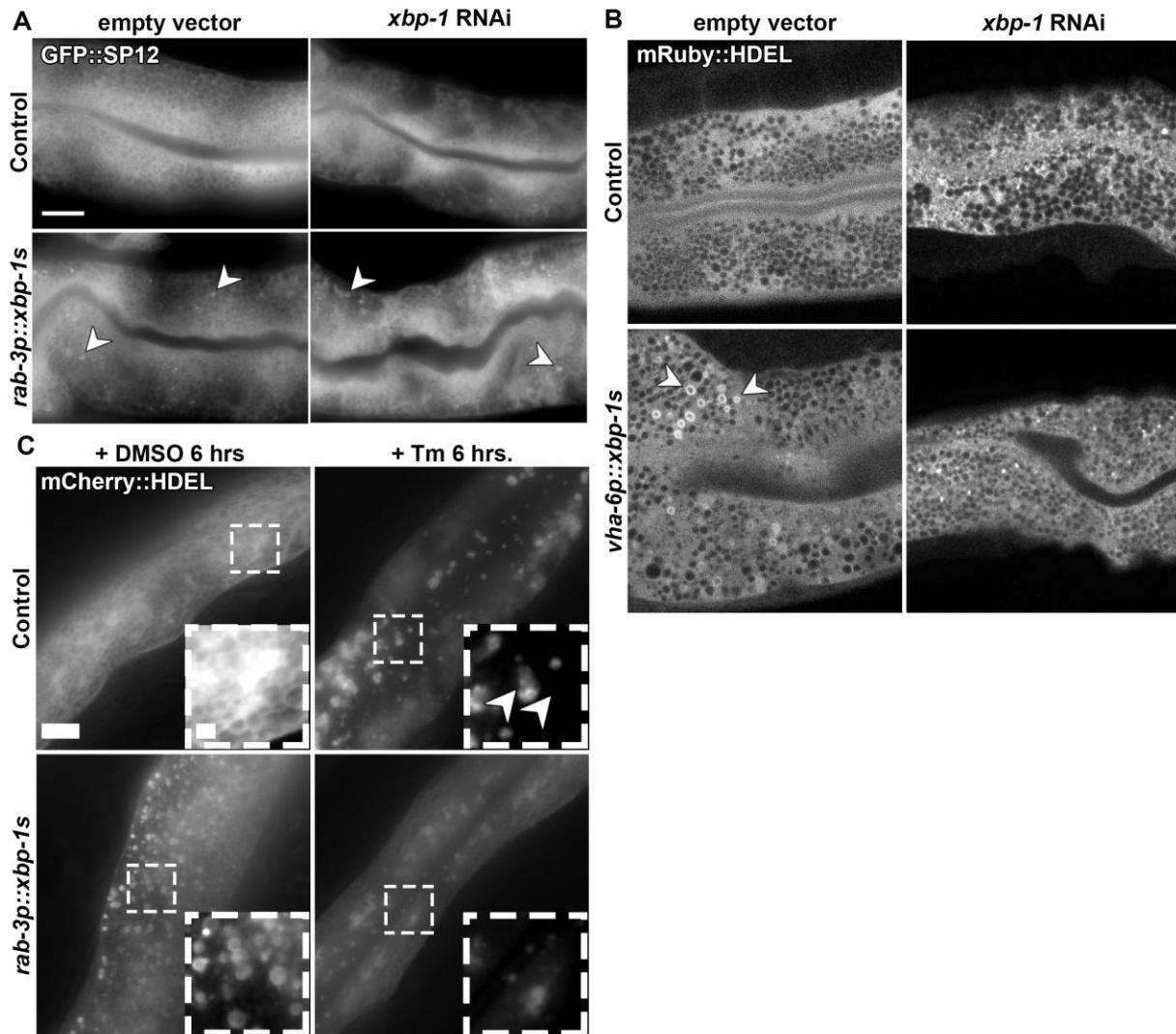
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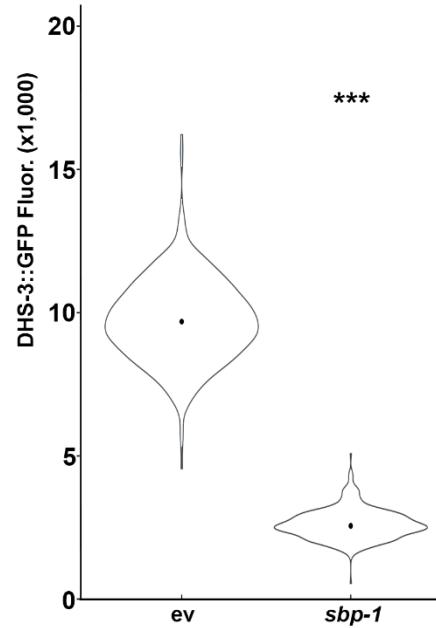
## Supplemental Information

### Supplemental Figures



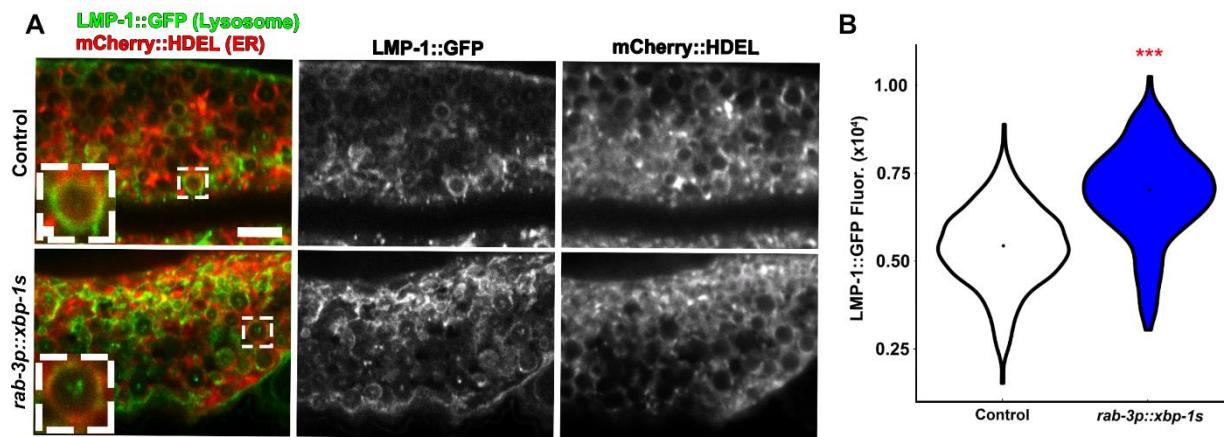
**Fig. S1. ER remodeling of the intestine through UPR<sup>ER</sup> activation requires neuronal nonautonomous signaling.** (A) ER morphology in intestine of control and neuronal *xbp-1s* animals. An alternate ER transmembrane marker (VS25: *vha-6p::GFP::C34B2.10 (SP12)*) is used here to demonstrate that phenotypes depicted are not an artifact of luminal ER fluorophores. While the ER remodeling is much more subtle using this marker, clear spherical structures can be detected as marked by arrowheads. Animals were grown from hatch on ev or *xbp-1* RNAi and live-imaged using a compound microscope. Scale = 10  $\mu$ m. (B) Representative micrographs of ER morphology (mRuby::HDEL) in intestine of control and intestinal *xbp-1s* (*vha-6p::xbp-1s*) animals. ER remodeling is depicted by arrowheads. (C) Representative compound micrographs of ER (mCherry::HDEL) with and without 6 hours treatment with 25 ng/ $\mu$ L tunicamycin. Briefly, animals were treated in tunicamycin in M9 liquid spinning at 20°C or with 1% DMSO as a control. Drugs were washed out and animals were live-imaged immediately

on a compound microscope. Scale bar is 10 $\mu$ m. For A-C, animals are imaged at late Day 2 of adulthood.

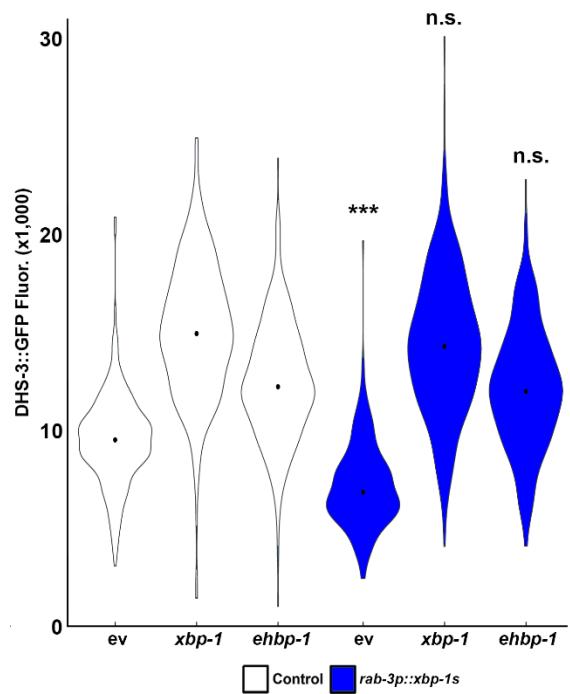


**Fig. S2. LAMPro technology can be reliably used to quantify LDs using DHS-3::GFP.**

Quantification of lipid droplets (LDs) (via DHS-3::GFP fluorescence) in animals grown on ev or *sbp-1* RNAi from hatch. *sbp-1* is used as a control for decreased lipids. \*\*\* = p < 0.001 using non-parametric Mann-Whitney testing. Plots are representative of three biological replicates and n = 921 (ev), 435 (*sbp-1*). Animals were imaged at late Day 2 of adulthood.

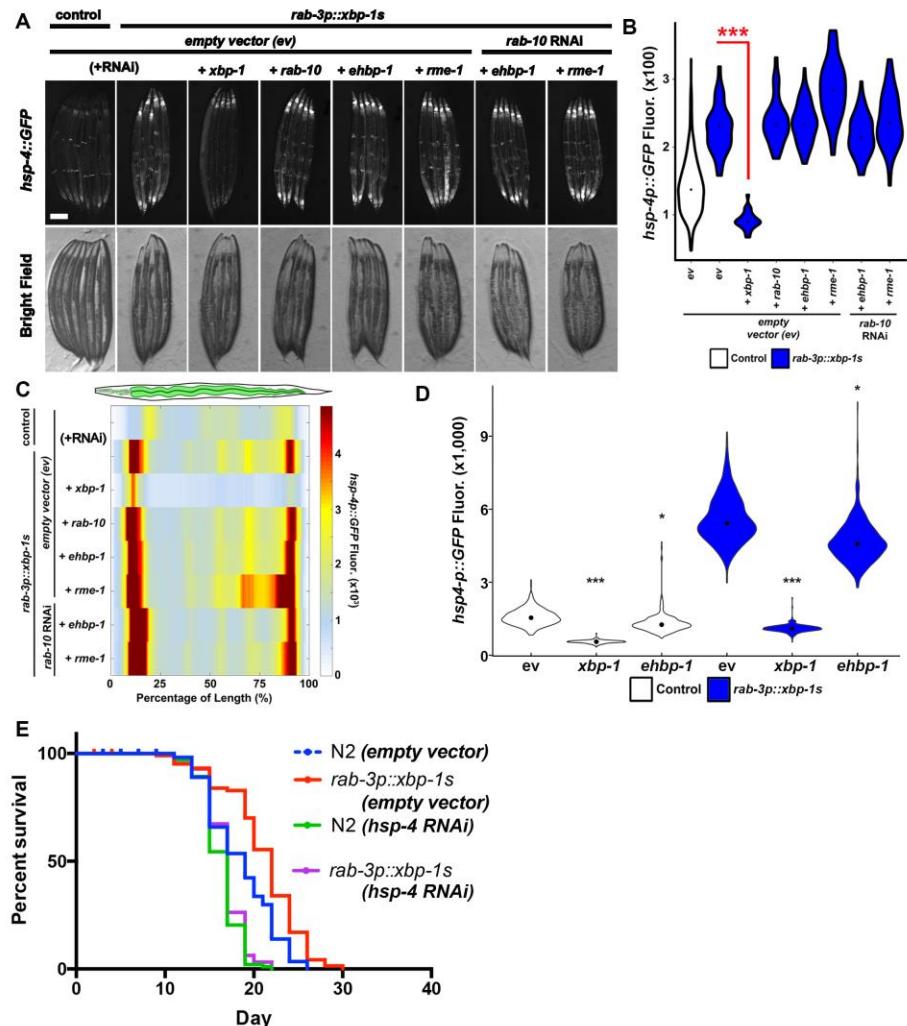


**Fig. S3. ER remodeling is coincident with more intestinal lysosomes in neuronal *xbp-1s* animals.** (A) Confocal micrographs of intestinal ER (*vha-6p::mCherry::HDEL*) and lysosomes (*vha-6p::lmp-1::GFP*), in fixed animals, with and without *rab-3p::xbp-1s*. Scale = 5  $\mu$ m (and 0.5  $\mu$ m for inset). (B) Quantification of lysosomes (via LMP-1::GFP) in control and *rab-3p::xbp-1s* animals. \*\*\* =  $p < 0.001$  using non-parametric Mann-Whitney testing.  $n > 50$  animals per condition. For A-B, animals are imaged at late Day2 of adulthood.



**Fig. S4. Lipid depletion in neuronal *xbp-1s* is dependent on *xbp-1* and *ehbp-1*.**

Quantification of LDs (via DHS-3::GFP) in control and *rab-3p::xbp-1s* animals grown on ev, *xbp-1*, or *ehbp-1* RNAi from hatch. \*\*\* =  $p < 0.001$ , n.s. =  $p > 0.10$  using non-parametric Mann-Whitney testing. Plots are representative of four biological replicates and  $n = 320\text{-}484$  animals per strain. Animals are imaged at late Day 2 of adulthood.



**Fig. S5. Knockdown of lipophagic components does not affect chaperone induction by *xbp-1s*.** (A) Fluorescent light micrographs of *hsp-4p::GFP* UPR<sup>ER</sup> reporter animals, imaged as Day 2 adults, and treated with the indicated RNAi(s) from hatch. Scale = 100  $\mu$ m. (B) Quantification of *hsp-4p::GFP* fluorescence in control (white) and *rab-3p::xbp-1s* (blue) nematodes, using a Biosorter, following various RNAi treatments. \*\*\* = p < 0.001 by Mann-Whitney. Wild-type controls on *xbp-1* and *ehbp-1* RNAis can be found in panel (d). (C) Median profiles of GFP fluorescence of the *hsp-4p::GFP* animals, under various RNAi treatments, in a *rab-3p::xbp-1s* background, following alignment. Plots are representative of three biological replicates and derived from n = 28-71 per strain. (D) Quantification of *hsp-4p::GFP* fluorescence in control (white) and *rab-3p::xbp-1s* (blue) nematodes, using a Biosorter, following various RNAi treatments. \*\*\* = p < 0.001, \* = p < 0.10 using non-parametric Mann-Whitney testing. p-values are in comparison to their corresponding ev controls. Data is representative of two independent trials with n = 224-573 per strain. (E) Lifespan data for wild-type N2 animals or neuronal *xbp-1s* (*rab-3p::xbp-1s*) animals treated with EV or *hsp-4* RNAi from hatch. RNAi knockdown of *hsp-4* is sufficient to eliminate the lifespan extension normally seen in neuronal

*xbp-1s* animals. See **table S1** for lifespan statistics. For A-D, animals are imaged at Day 1 of adulthood.

**Table S1. Statistics for all life-span data.**

Supplemental Table S1. Statistical Analyses of Lifespan Data					
Corresponding Figure	Strain, Treatment	Median Lifespan	# Deaths/# Total	% Change in Median Lifespan	p-value
		[days]			Log-rank (Mantel-Cox)
2f	N2, vector RNAi, 1% DMSO	22	95/120	n/a	n/a
	<i>rab-3p::xbp-1s</i> , vector RNAi, 1% DMSO	26	94/120	1.2	<0.0001
	N2, <i>xbp-1</i> RNAi, 1% DMSO	14	92/120	0.6	<0.0001
	<i>rab-3p::xbp-1s</i> , <i>xbp-1</i> RNAi, 1% DMSO	15	95/120	0.7	<0.0001
	N2, <i>ehbp-1</i> RNAi, 1% DMSO	16	92/120	0.7	<0.0001
	<i>rab-3p::xbp-1s</i> , <i>ehbp-1</i> RNAi, 1% DMSO	15	96/120	0.7	<0.0001
2g	N2, vector RNAi, 25ng/ $\mu$ L tunicamycin	14	97/120	n/a	n/a
	<i>rab-3p::xbp-1s</i> , vector RNAi, 25ng/ $\mu$ L tunicamycin	18	109/120	1.3	<0.0001
	N2, <i>xbp-1</i> RNAi, 25ng/ $\mu$ L tunicamycin	12	98/120	0.9	<0.0001
	<i>rab-3p::xbp-1s</i> , <i>xbp-1</i> RNAi, 25ng/ $\mu$ L tunicamycin	12	101/120	0.9	<0.0001
	N2, <i>ehbp-1</i> RNAi, 25ng/ $\mu$ L tunicamycin	14	111/120	1.0	0.2271
	<i>rab-3p::xbp-1s</i> , <i>ehbp-1</i> RNAi, 25ng/ $\mu$ L tunicamycin	14	111/120	1.0	0.8208
S5e	N2, vector RNAi	19	100/120	n/a	n/a
	<i>rab-3p::xbp-1s</i> , vector RNAi	22	88/120	1.2	<0.0001
	N2, <i>hsp-4</i> RNAi	17	102/120	0.9	<0.0001
	<i>rab-3p::xbp-1s</i> , <i>hsp-4</i> RNAi	17	96/120	0.9	<0.0001
3d	N2, vector RNAi	17	101/120	n/a	n/a
	<i>vha-6p::ehbp-1</i> , vector RNAi	21	98/120	1.2	0.0221

**Table S2. Organisms and strains used.**

BACTERIAL AND VIRUS STRAINS		
OP50	CGC	N/A
HT115	CGC	N/A
DH5α	Invitrogen	
EXPERIMENTAL MODELS: ORGANISMS/STRAINS		
<i>C. elegans</i> : Bristol (N2) strain as wild type (WT)	CGC	N2
<i>C. elegans</i> : AGD927: uthls270 [rab-3p::Xbp-1s, myo-2p::tdTomato]	(Taylor and Dillin, 2013)	N/A
<i>C. elegans</i> : AGD928: uthls270[rab-3p::Xbp-1s, myo-2p::tdTomato]; zcls4[hsp-4p::GFP]	(Taylor and Dillin, 2013)	N/A
<i>C. elegans</i> : AGD1289: N2, uthls368[rab-3p::HSF-1 FL, myo-2p::tomato]	(Douglas et al., 2015)	N/A
<i>C. elegans</i> : AGD1376: hjSi158[vha-6p::SEL-1(1-79)::mCherry::HDEL]; uthls270[rab-3p::Xbp-1s, myo-2p::tdTomato]	This study	N/A
<i>C. elegans</i> : AGD1415: pwls23[vit-2::GFP]	This study	N/A
<i>C. elegans</i> : AGD1771: N2, uthls464[rgf-1p::Xbp-1s, myo-2p::tdTomato]	This study	N/A
<i>C. elegans</i> : AGD2048: dhs-3p::dhs-3::GFP; hjSi158 [vha-6p::SEL-1(1-79)::mCherry::HDELlet-858 3' UTR]	This study	N/A
<i>C. elegans</i> : AGD2065: dhs-3p::dhs-3::GFP; hjSi158 [vha-6p::SEL-1(1-79)::mCherry::HDELlet-858 3' UTR]; uthls270[rab-3p::Xbp-1s, myo-2p::tdTomato]	This study	N/A
<i>C. elegans</i> : AGD2129: pwls23[vit-2::GFP]; uthls270 [rab-3p::Xbp-1s, myo-2p::tdTomato]	This study	N/A
<i>C. elegans</i> : AGD2192: unc-119(ed3) III; uthSi60[vha-6p::ERss::mRuby::HDEL::unc-54 3'UTR cb-unc-119(+)] IV	This study	N/A
<i>C. elegans</i> : AGD2230: uthls488[vha-6p::xbp-1s, myo-2p::BFP]; unc-119(ed3) III; uthSi60[vha-6p::ERss::mRuby::HDEL::unc-54 3'UTR cb-unc-119(+)] IV	This study	N/A
<i>C. elegans</i> : AGD2267: uthls270 [rab-3p::xbp-1s, myo-2p::tdTomato]; unc-119(ed3)III; uthSi60 [vha-6p::Erss::mRuby::HDEL::unc-54 3'UTR cb-unc-119(+)]	This study	N/A
<i>C. elegans</i> : AGD2349: uthls270[rab-3p::Xbp-1s, myo-2p::tdTomato]; vha-6p::GFP::C34B2.10(SP12)+unc119(+); unc-119(ed3) III; uthSi60[vha-6p::ERss::mRuby::HDEL::unc-54 3'UTR cb-unc-119(+)] IV	This study	N/A
<i>C. elegans</i> : AGD2363: N2, uthEx911[vha-6p::ehbp-1::unc-54 3'UTR; myo-2p::GFP]	This study	N/A
<i>C. elegans</i> : AGD2364: N2, uthEx912[vha-6p::ehbp-1::unc-54 3'UTR; myo-2p::GFP]	This study	N/A

<i>C. elegans</i> : AGD2365: N2, uthEx911[vha-6p::ehbp-1::unc-54 3'UTR; myo-2p::GFP]; unc-119(ed3) III; uthSi60[vha-6p::ERss::mRuby::HDEL::unc-54 3'UTR cb-unc-119(+)] IV	This study	N/A
<i>C. elegans</i> : AGD2366: N2, uthEx911[vha-6p::ehbp-1::unc-54 3'UTR; myo-2p::GFP]; dhs-3p::dhs-3::GFP; hjSi158[vha-6p::SEL-1(1-79)::mCherry::HDELlet-858 3' UTR]	This study	N/A
<i>C. elegans</i> : AGD2367: N2, uthEx911[vha-6p::ehbp-1::unc-54 3'UTR; myo-2p::GFP]; zcls4(hsp-4p::GFP)V	This study	N/A
<i>C. elegans</i> : EG6703: unc-119(ed3) III; cxTi10816 IV; oxEx1582	CGC	EG6703
<i>C. elegans</i> : SJ4005: zcls4(hsp-4p::GFP)V	(Taylor and Dillin, 2013)	SJ4005
<i>C. elegans</i> : VS25: vha-6p::GFP::C34B2.10(SP12)+unc119(+); unc-119(ed3) III	CGC	VS25
<i>C. elegans</i> : VS30: hjSi158[vha-6p::SEL-1(1-79)::mCherry::HDELlet-858 3' UTR]	CGC	VS30
OLIGONUCLEOTIDES – ALL OLIGOS ORDERED FROM IDT		
Primer Name	Primer Sequence	Primer Purpose
JRD260 F_all_xbp1s_vectors	TAGCATTCTGTAGA ATTCCAAGTGAGC	forward primer to clone vector side of pJRD16
JRD262 R_my03_vector_xbp1s	ctagatggatctagtggtc gtggg	reverse primer to clone vector side of pJRD16
JRD255 R_all_xbp1s_inserts	GCGCTCAGTTGGAA ATTCTACGAATGC TATTAATTTGGAA GAAATTTAGGTGCG ATTG	Reverse primer to clone xbp-1s insert for pJRD16
JRD257 F_xbp1s_insert_my03	ctagatggatctagtggtc gtggg	Forward primer to clone xbp-1s insert for pJRD16
gg288F	ATAGCATTCTGTAG CGAATGCGTATTG TaccggaccagtcAAC TCTCTTATCAAGG AAAAC	Forward primer to PCR ERSS::mRuby for pGIL122
gg22R	aagtcaaggcacgggc gcgagatgTTAagttc gtcatgTCCTCCTCC AAGTCCAG	Reverse primer to PCR ERSS::mRuby for pGIL122
gg153F	catctcgcccccgtg	Forward primer to PCR PCR vector for pGIL122

gg45R	ttttatgggtttggtaggttt ag	Reverse primer to PCR vector for pGIL122
SM8 vha6p-ehbp F1	cctaccaaaaaccataaa aatggcggtattctcg cg	Forward primer to PCR ehbp-1 for pSM1
SM9 vha-6p-ehbp R1	ATTATCAACTTCTA CTCACtcttttcgaatg aattttgctgaaca	Reverse primer to PCR ehbp-1 for pSM1
SM10 vha6-ehbp1 F2	attcattcgaaaaaaaagaG TGAGTAGAACGTTG ATAATTGAAAGT G	Forward primer to PCR vector for pSM1
SM11 vha6-ehbp1 R2	cgacgaagaataccgc catttttatgggtttggtag gttttagtc	Reverse primer to PCR vector for pSM1
RECOMBINANT DNA		
pCFJ356: Integration vector for EG6703 MOSCI: contains Right xcTi10816, cb-unc-119(+), and Left cxTi10816	Addgene	34871
pCFJ601: MOS-1 transposase	Addgene	34874
pCFJ90: myo-2p::mCherry::unc-54 3' UTR	Addgene	19327
pCFJ104: myo-3p::mCherry::unc-54 3' UTR	Addgene	19328
pGH8: rab-3p::mCherry::unc-54 3' UTR	Addgene	19359
pEK1: myo-2p::GFP::unc-54 3' UTR	This study	N/A
pEK2: myo-2p::tdtomato::unc-54 3' UTR	This study	N/A
pRT5: pAD1 rgef-1p::xbp-1s cDNA::unc-54 3'UTR	(Taylor and Dillin, 2013)	N/A
pSM1: vha-6p::ehbp-1::ehbp-1 3'UTR	This study	N/A
pGIL122: pCFJ356::vha-6p::hsp-4 ERSS::mRuby::HDEL::unc-54 3' UTR	This study	N/A

### Sequences used for cloning

#### pSM1 (cloned into pCFJ90 backbone replacing myo-2p::mCherry with vha-6p::ehbp-1 sequence using Gibson)

vha-6p:

gcatgtacttataggcgctacgcattaccaggactgaaacaatggagttgagccataatgtactaaaaattttaatgcgtttac  
aaaaatattttcagatctcgagatcatgaaaactatcaaacacgcgcgcctggagcaatcgagtcgtccccaatattcagggtgt  
atatgcataatgttttagactacattcggtaagtgcacttcaggatataactgttaattttaaattcagcgcaacgcggaggaaag  
tctgtatcgtttaacaaaatacataatcgatctcgatctcacggagatattgcgcagatgcgtcaacgttgcgcgtgtatccaa  
gcacagaactgcattaagtataactatttactcgatcttgcacatagggtttaaatcatattatgcatattatcatattcaatgca  
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cgagaccaactaatagcatgacaaggcatttcaattgcgcctggagcgcaattgggtttattcgaaaatcgcacattctgtttccccata  
atataaaaattccaggacgatataattacattctcacaatattgcattacagacaccgacaaagaatctccacctgatatgaaac  
aatgagccaacaatgttatctgtattgccaccaccacattccatgtcattcgtatataattgtttcaattgaatattgcaggtatatcgaa

atggaaactgttaaggcctcatcttcaatttcataacatcatccatcattccagagcagctccggccacacaaaaatttgtggcggtctgat  
attgataatcgacttcttgacgtgcctgacggagcagcaaagcgagcactgataagacaatgaagaactaaaaattgtctcggtt  
ttcagtcttagttctgcagcacatttttgtttctcctatttccgcatttcctaactttctgatgtccattcaaatgattttgttataaaaattgttt  
aatttcaggcgactaaaacacctaccaaaaaccataaaaaa

ehbp-1:

### ehbp-1 3' UTR (predicted):

tacgcttcaacgttattttactcaattaaaactacgggtcttccttctgaaaacaagttcccccagtgtcccttgttatcttcatca  
tttttctcccccataatttactctttccgtttttggcaaattttaggagttaatcaaaccatcaaatttctatctttcatctctatgttat  
attttgtattcagtttcttcgtgcacacaatccaaatgcttcagttcaataaatcaattcagtttcaatgtatgttcagcaa  
aaattcattcgaaaaaaga

**pGIL122 (cloned into pCFJ356 multiple cloning site using Gibson):**

vha-6p:

## hsp-4 ERSS:

ATGAAAGTTTCTCGCTGATTCTGATAGCATTGTAGCGAATGCGTATTGT

mRuby:

AACTCTTATCAAGGAAAACATGCGCATGAAGGTTCTTGAGGGATCTGTTAACGGAC  
ATCAGTTCAAGTGCAGTGGAGAAGGAGAAGGAAATCCATACATGGTATGTGATACATGGAA  
ATAGGCACATTCAATTAAATATTGTATTTCAAGGAACTCAAACATGCGCATCAAGGTTAT  
CGAAGGAGGACCACCTCCATTGCTTCGACATCCTGCTACCTCTTCATGTACGGATCT  
CGCACTTCAATTAAAGTATCCAAGGAAATTCCAGATTCTTCAAGCAGTCTTCCCAGAAGG  
ATTCACTTGGGAGCGCGTTACCCGCTACGAGGATGGAGGAGTTATCACCGTTATGCAAGA  
TACCTCCCTTGAGGGATGGATGCCCTGTTACCAAGCCTCAAGTTCGTGGAGGTTAACCTCCCCA

TCTAACGGAGCTGTTATGCAGAAGAAGACTAAGGGATGGGAACCAAACACTGAAATGATGT  
ACCCAGCTGATGGAGGACTTCGCGGATACACCCACATGGCTCTTAAGGTTGATGGAGGAG  
GACACCTTCTTGCTCTTCGTTACCACCTACCGCTCTAAGAAGACTGTTGGAAATATTAAG  
ATGCCAGGAATCCATGCTGTCGATCACCGTCTTGAGCGTCTTGAGGAGTCTGATAACGAGA  
TGTTCGTTGTTCAACGTGAGCAGCCTGCTGCTAAGTCGCTGGACTTGGAGGAGGA

unc-54 3'UTR:

AACTCTCTTATCAAGGAAAACATGCGCATGAAGGTTGTTCTGAGGGATCTGTTAACGGAC  
ATCAGTTCAAGTGCAGTGGAGAAGGAGAAGGAAATCCATACATGGTATGTGATACATGGAA  
ATAGGCACATTCAATTAAATATTGTATTTCAAGGAAACTCAAACATGCGCATCAAGGTTAT  
CGAAGGAGGACCACCTCCATTGCTTGCACATCCTGCTACCTCTTCATGTACGGATCT  
CGCACTTTCAATTAAAGTATCCAAGGAAATCCAGATTCTCAAGCAGTCTTCCCAGAAGG  
ATTCACTTGGGAGCGCGTACCCGCTACGAGGATGGAGGAGTTATCACCGTTATGCAAGA  
TACCTCCCTTGAGGATGGATGCCTGTTACCACGCTCAAGTCGTTGGAGTTAACTTCCA  
TCTAACGGAGCTGTTATGCAGAAGAAGACTAAGGGATGGGAACCAAACACTGAAATGATGT  
ACCCAGCTGATGGAGGACTTCGCGGATACACCCACATGGCTCTTAAGGTTGATGGAGGAG  
GACACCTTCTTGCTCTTCGTTACCACCTACCGCTCTAAGAAGACTGTTGGAAATATTAAG  
ATGCCAGGAATCCATGCTGTCGATCACCGTCTTGAGCGTCTTGAGGAGTCTGATAACGAGA  
TGTTCGTTGTTCAACGTGAGCAGCCTGCTGCTAAGTCGCTGGACTTGGAGGAGGA