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Supplemental Information

DNAJC14 Ameliorates Inner Ear

Degeneration in the DFNB4 Mouse Model

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Supplementary Table S1. Antibodies used in this study.

Name	Catalog number (Manufacturer)	Dilution (Use)
Primary Antibody		
Pendrin	G-19 (Santa Cruz)	1:1000 (WB)
Aldolase	N-15 (Santa Cruz)	1:1000 (WB)
3xHA-DNAJC14	F-7 (Santa Cruz)	1:1000 (WB)
Myc-Hsc70	1B5 (Abcam)	1:1000 (WB)
HA-Arf1	F-7 (Santa Cruz)	1:1000 (WB)
anti-pendrin	2-hR2 (ABfrontier)	1:1000 (WB)
anti- β -Actin	sc-47778 (Santa Cruz)	1:2000 (WB)
anti-DNAJC14	ab121535 (Abcam)	1:1000 (WB), 1:200 (IF)
anti-Flag	F3165 (Sigma)	1:2000 (WB), 1:100 (IF)
anti-DAPI	D1306 (Thermo)	1:5000 (ICC), 1:10000 (IHC)
anti-KCNJ10	APC-035 (Alomone Labs)	1:100 (IF)
Secondary Antibody		
anti-mouse, HRP conjugated	G-21040 (Thermo)	1:1000 (WB)
anti-rabbit, HRP conjugated	32460 (Thermo)	1:1000 (WB)
Alexa 488, anti-mouse	A11001 (Invitrogen)	1:1000 (IF)
Alexa 488, anti-rabbit	A11008 (Invitrogen)	1:1000 (IF)
Alexa 568, anti-rabbit	A11036 (Invitrogen)	1:1000 (IF)

IF, immunofluorescence; WB, western blotting; IHC, immunohistochemistry; ICC, immunocytochemistry

Figure S1

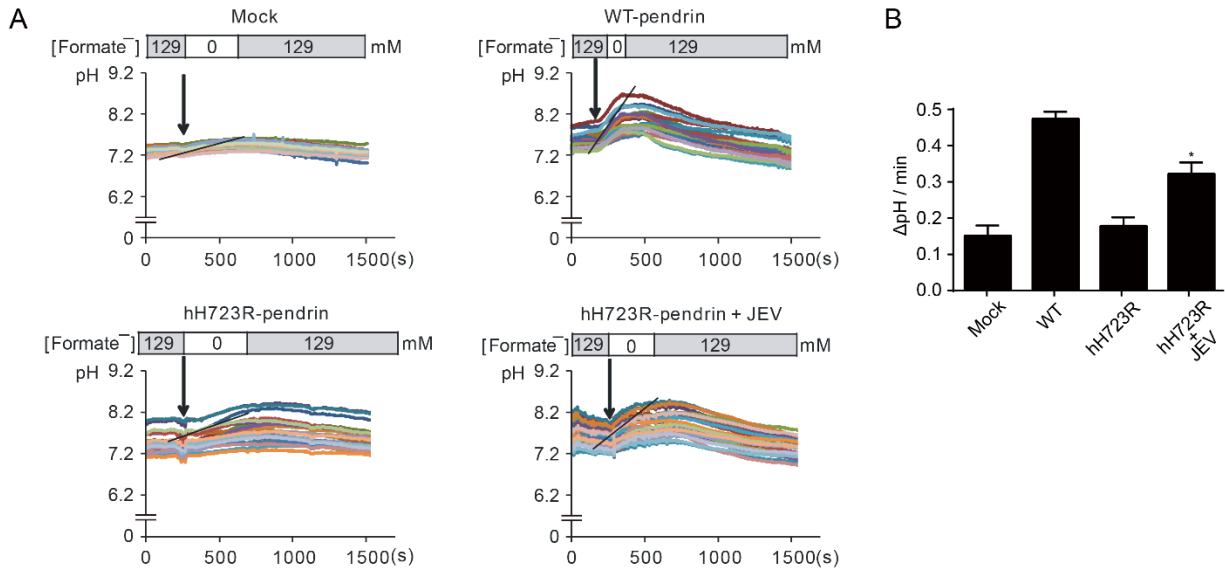


Figure S1. HCO₂⁻/HCO₃⁻ exchange activity by activation of DNAJC14. (A) Formate/Bicarbonate (HCO₂⁻/HCO₃⁻) exchange activity was measured by recording the pH-sensitive fluorescent probe BCECF in PANC-1 mock cells, and in wild-type (WT)- and human H723R (hH723R)-pendrin stably expressing cells with/without Japanese encephalitis virus (JEV, 3x10⁶ pfu) to activate DNAJC14, as detailed in the Methods. The quantification of multiple experiments is depicted in (B). The HCO₂⁻/HCO₃⁻ exchange activity significantly increased in cells expressing hH723R-pendrin with JEV treatment (n = 4 in each group). *, p < 0.05.

Figure S2

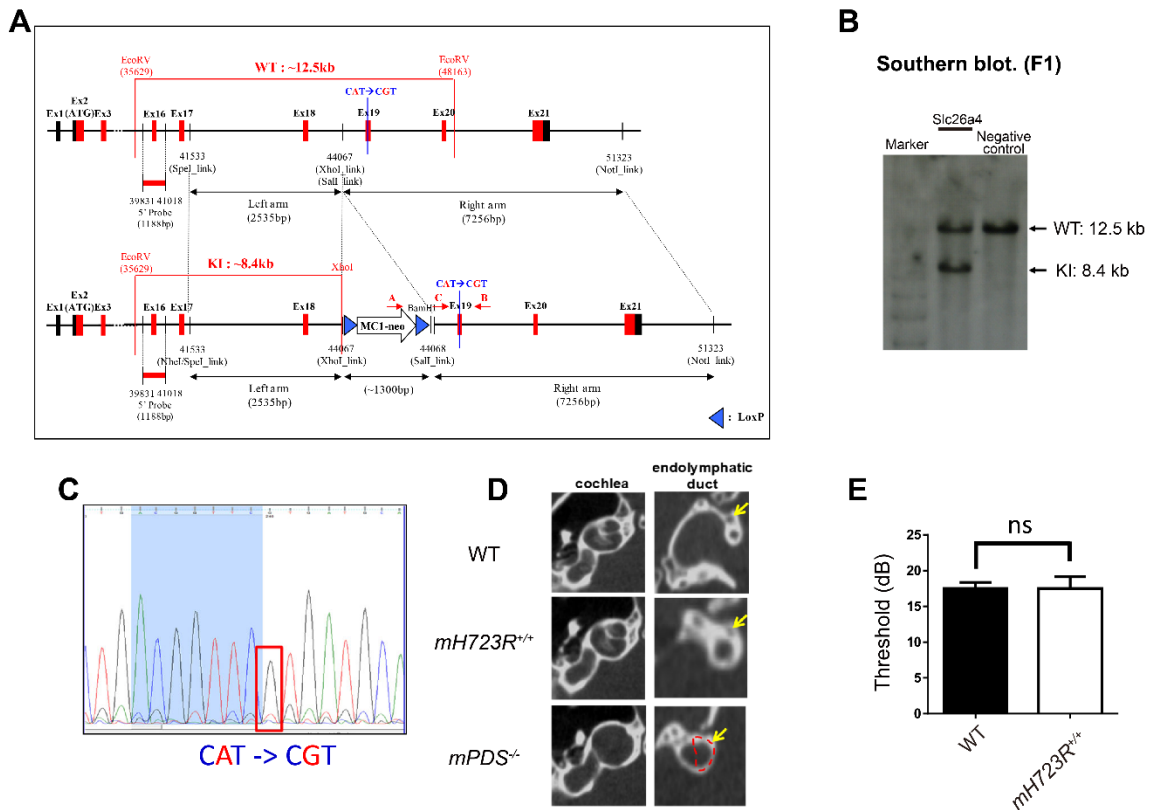


Figure S2. Targeting scheme of the H723R knock-in mouse model. (A) A Bac clone containing *slc26a4* genomic region was used to construct the targeting vector. The expected size of the restriction enzyme fragment of the wild-type (WT) and knock-in (KI) was ~12.5 kb and ~8.4 kb, respectively. The loxP-flanked neomycin resistance gene (neo) was used as a selection marker during embryonic stem (ES) cell culture. (B) The *slc26a4* knock-in (KI) was identified with Southern blotting. (C) Sanger sequencing of the target region revealed that the c.2168A>G point mutation was present in the founder mouse. (D) Micro-computed tomogram of the inner ear. While *slc26a4* knock-out mice (*mPDS^{-/-}*) showed a dilated endolymphatic duct and cochlear hydrops, WT and mouse p.H723R KI homozygote [*mH723R^{+/+}*] showed normal cochlea and endolymphatic duct. (E) In the auditory brainstem response, both WT and *mH723R^{+/+}* mice showed normal hearing thresholds (n = 13 in each group). ns, not significant.

Figure S3

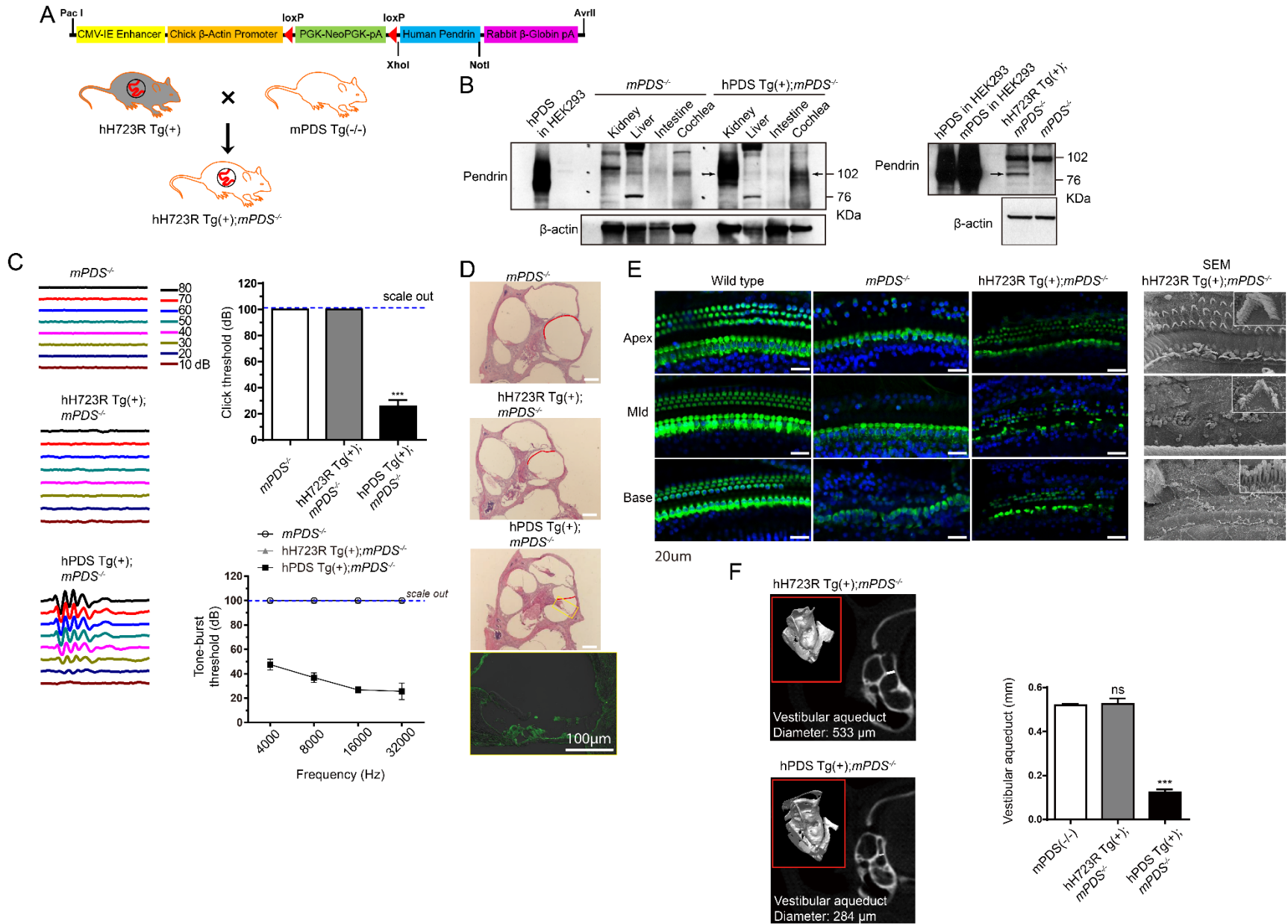


Figure S3. Generation of a human H723R-pendrin mouse model mimicking deafness in human DFNB4.

(A) Construct of the human p.H723R *SLC26A4* transgene in the pCB vector backbone. The wild-type (WT) or the p.H723R *SLC26A4* gene is Cre-inducible. The human WT-pendrin (hPDS) and H723R-pendrin (hH723R) transgenic mice [Tg(+)] were crossed with pendrin knock-out [*mPDS*^{-/-}] mice to generate hH723R Tg(+);*mPDS*^{-/-} mice, in which only ectopic hH723R-pendrin was expressed without expression of mouse pendrin. To induce transgene expression in the inner ear, we crossed this strain with Pax2-Cre mice. (B) When WT- or hH723R-Tg(+) mice were crossed with Pax2-Cre, expression of the *SLC26A4* gene was observed in the inner ear and kidney (black arrows), but not in the other organs. (C) Auditory brainstem response by click and tone-burst stimuli for *mPDS*^{-/-}, hH723R(+);*mPDS*^{-/-}, and hPDS Tg(+);*mPDS*^{-/-} mice. *mPDS*^{-/-} and hH723R Tg(+);*mPDS*^{-/-} mice were deaf, whereas the hearing function of hPDS Tg(+);*mPDS*^{-/-} mice was nearly normalized (n = 8 mice in each group). (D) H&E staining of the inner ear showing that *mPDS*^{-/-} and hH723R Tg(+);*mPDS*^{-/-} mice had dilatation of the scala media, whereas hPDS Tg(+);*mPDS*^{-/-} mice had normal sized scala media without endolymphatic hydrops. Immunostaining of the inner ear (lower panel) showed ectopic pendrin expression (anti-pendrin antibody, green) in the hair cells, supporting cells, inner limbus, lateral wall, and stria vascularis (Pax2-Cre-dependent). Scale bar, 20 μm. (E) Immunofluorescence and surface electron microscopy (SEM) images of the organ of Corti labeled with phalloidin (green) and DAPI (blue) in wild-type, *mPDS*^{-/-}, and hH723R Tg(+);*mPDS*^{-/-} mice. Note that the outer hair cells showed better survival in hH723R Tg(+);*mPDS*^{-/-} than in *mPDS*^{-/-} mice. Magnification of electron microscope is x2000 [inset: x30000 (upper and middle), and x35000 (lower)]. Scale bars, 20 μm. (F) Micro-computed tomogram was performed to compare the size of vestibular aqueduct. hH723R Tg(+);*mPDS*^{-/-} mice had enlarged vestibular aqueduct, which was comparable to *mPDS*^{-/-} mice. In contrast, hPDS Tg(+);*mPDS*^{-/-} showed significantly normalized size of vestibular aqueduct when compared to that of *mPDS*^{-/-} (n = 6–9 in each group). ns, not significant; ***, p < 0.001.