

An *in vitro* platform for engineering and harnessing modular polyketide synthases

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Supplementary Methods

Crystallographic structure determination of polyketide products

X-ray crystal structure analysis of venemycin (**1**). Crystals grew as clusters of long, colorless needles by slow evaporation from 90% methanol and 10% water. The data crystal was cut from a larger crystal and had the approximate dimensions of 0.14 x 0.015 x 0.015 mm. The data were collected on an Agilent Technologies SuperNova Dual Source diffractometer using a μ -focus Cu K_{α} radiation source ($\lambda = 1.5418 \text{ \AA}$) with collimating mirror monochromators. A total of 508 frames of data were collected using ω -scans with a scan range of 1° and a counting time of 27.5 s per frame with a detector offset of $\pm 42.7^{\circ}$ and 85 s per frame with a detector offset of 110.8° . The data were collected at 100 K using an Oxford 700 Cryostream low temperature device. Data collection, unit cell refinement, and data reduction were performed using Rigaku Oxford Diffraction's CrysAlisPro V 1.171.40.37a. The structure was solved by direct methods using SHELXT¹ and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for the non-H atoms using SHELXL-2016/6². Structure analysis was aided by use of the programs PLATON³, OLEX2⁴, and WinGX⁵. The hydrogen atoms were calculated in ideal positions with isotropic displacement parameters set to 1.2xUeq of the attached atom (1.5xUeq for methyl hydrogen atoms). The hydrogen atoms on the water molecules were initially assigned by a best fit to the electron density around the oxygen atoms. These hydrogen atoms were subsequently fixed with O-H bond lengths set to 0.84 \AA . However, one of the water molecules, O7, was within H-bonding distance to a symmetry related O7, where one of the H atoms on each was directed towards each other. This required that this H atom, H7B, is disordered. In subsequent refinement models, the site occupancy factor for H7B was fixed at $\frac{1}{2}$ occupancy. Equation 1 was minimized (Equations 1-3). $R_w(F^2)$ refined to 0.2352, with an $R(F)$ of 0.08398 and a goodness of fit, S , of 1.02. Definitions used for calculating $R(F)$, $R_w(F^2)$, and the goodness of fit, S , are provided (Equations 4-6). The data were checked for secondary extinction effects but no correction was necessary. Neutral atom scattering factors and values used to calculate the linear absorption coefficient are from the International Tables for X-ray Crystallography⁶.

X-ray crystal structure analysis of pyrone (**2**). Crystals grew as clusters of colorless plates by slow evaporation from 80% EtOAc, 10% methanol, and 10% water. The data crystal was separated from a cluster of crystals and had the approximate dimensions of 0.20 x 0.11 x 0.04 mm. The data were collected on an Agilent Technologies SuperNova Dual Source diffractometer using a μ -focus Cu K_{α} radiation source ($\lambda = 1.5418 \text{ \AA}$) with collimating mirror monochromators. A total of 1163 frames of data were collected using ω -scans with a scan range of 1° and a counting time of 14.5 s per frame for frames

collected with a detector offset of +/- 42.4° and 47 s per frame with frames collected with a detector offset of 110.8°. The data were collected at 100 K using an Oxford Cryostream low temperature device. Data collection, unit cell refinement, and data reduction were performed using Rigaku Oxford Diffraction's CrysAlisPro V 1.171.40.37a. The structure was solved by direct methods using SHELXT¹ and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for the non-H atoms using SHELXL-2016/6². Structure analysis was aided by use of the programs PLATON³, OLEX2⁴, and WinGX⁵. The hydrogen atoms on the carbon atoms were calculated in ideal positions with isotropic displacement parameters set to 1.2xUeq of the attached atom (1.5xUeq for methyl hydrogen atoms). Equation 1 was minimized (Equations 1, 3, and 7). $R_w(F^2)$ refined to 0.240, with an $R(F)$ of 0.0776 and a goodness of fit, S , of 1.06. Definitions used for calculating $R(F)$, $R_w(F^2)$, and the goodness of fit, S , are provided (Equations 4-6). The data were checked for secondary extinction effects but no correction was necessary. Neutral atom scattering factors and values used to calculate the linear absorption coefficient are from the International Tables for X-ray Crystallography⁶.

X-ray crystal structure analysis of methylvenemycin (**3**). Crystals grew as clusters of thin colorless needles by slow evaporation from 90% methanol and 10% water. The data crystal was separated from a cluster of crystals and had the approximate dimensions of 0.27 x 0.043 x 0.022 mm. The data were collected on an Agilent Technologies SuperNova Dual Source diffractometer using a μ -focus Cu K_α radiation source ($\lambda = 1.5418 \text{ \AA}$) with collimating mirror monochromators. A total of 482 frames of data were collected using ω -scans with a scan range of 1° and a counting time of 54 s per frame for frames collected with a detector offset of +/- 41.6° and 142.5 s per frame with frames collected with a detector offset of 110.7°. The data were collected at 100 K using an Oxford Cryostream low temperature device. Data collection, unit cell refinement, and data reduction were performed using Rigaku Oxford Diffraction's CrysAlisPro V 1.171.40.37a. The structure was solved by direct methods using SHELXT¹ and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for the non-H atoms using SHELXL-2016/6². Structure analysis was aided by use of the programs PLATON³, OLEX2⁴, and WinGX⁵. The hydrogen atoms were calculated in ideal positions with isotropic displacement parameters set to 1.2xUeq of the attached atom (1.5xUeq for methyl hydrogen atoms). The data crystal was twinned. The twin law was determined using PLATON. The twin fraction refined to 0.482(4). Equation 1 was minimized (Equations 1, 3, and 8). $R_w(F^2)$ refined to 0.331, with an $R(F)$ of 0.126 and a goodness of fit, S , of 1.17. Definitions used for calculating $R(F)$, $R_w(F^2)$ and the goodness of fit, S , are provided (Equations 4-6). The data were checked for secondary extinction effects but no correction was necessary. Neutral atom scattering factors and values used to calculate the linear absorption coefficient are from the International Tables for X-ray Crystallography⁶.

X-ray crystal structure analysis of deshydroxyvenemycin (**5**). Crystals grew as clusters of very small, colorless needles by slow evaporation from 90% methanol and 10% water. The data crystal was broken from a cluster and had the approximate dimensions of 0.12 x 0.058 x 0.036 mm. The data were collected on an Agilent Technologies SuperNova Dual Source diffractometer using a μ -focus Cu K_{α} radiation source ($\lambda = 1.5418 \text{ \AA}$) with collimating mirror monochromators. A total of 704 frames of data were collected using ω -scans with a scan range of 1° and a counting time of 41 s per frame with a detector offset of $\pm 42.4^{\circ}$ and 151 s per frame with a detector offset of 110.2° . The data were collected at 100 K using an Oxford 700 Cryostream low temperature device. Data collection, unit cell refinement, and data reduction were performed using Rigaku Oxford Diffraction's CrysAlisPro V 1.171.40.37a. The structure was solved by direct methods using SHELXT¹ and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for the non-H atoms using SHELXL-2016/6². Structure analysis was aided by use of the programs PLATON³, OLEX2⁴, and WinGX⁵. The hydrogen atoms were calculated in ideal positions with isotropic displacement parameters set to 1.2xUeq of the attached atom (1.5xUeq for methyl hydrogen atoms). The phenol ring was disordered by an approximately 180° rotation about the C-C bond connecting the two ring systems. The disorder was modeled using OLEX2. The site occupancy for the major component refined to 79(1)%. Equation **1** was minimized (Equations **1**, **3**, and **9**). $R_w(F^2)$ refined to 0.278, with an R(F) of 0.0923 and a goodness of fit, S, of 1.05. Definitions used for calculating R(F), $R_w(F^2)$ and the goodness of fit, S, are provided (Equations **4-6**). The data were checked for secondary extinction effects but no correction was necessary. Neutral atom scattering factors and values used to calculate the linear absorption coefficient are from the International Tables for X-ray Crystallography⁶.

X-ray crystal structure analysis of deshydroxymethylvenemycin (**6**). Crystals grew as clusters of thin, colorless prisms by slow evaporation from 90% methanol and 10% water. The data crystal had the approximate dimensions of 0.21 x 0.15 x 0.038 mm. The data were collected on an Agilent Technologies SuperNova Dual Source diffractometer using a μ -focus Cu K_{α} radiation source ($\lambda = 1.5418 \text{ \AA}$) with collimating mirror monochromators. A total of 388 frames of data were collected using ω -scans with a scan range of 1° and a counting time of 30.5 s per frame with a detector offset of 0.0° and 91.5 s per frame with a detector offset of 82.6° . The data were collected at 100 K using an Oxford 700 Cryostream low temperature device. Data collection, unit cell refinement and data reduction were performed using Rigaku Oxford Diffraction's CrysAlisPro V 1.171.40.37a. The structure was solved by direct methods using SHELXT¹ and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for the non-H atoms using SHELXL-2016/6². Structure analysis was aided by use of the programs PLATON³, OLEX2⁴, and WinGX⁵. The hydrogen atoms were calculated in ideal positions with

isotropic displacement parameters set to 1.2xUeq of the attached atom (1.5xUeq for methyl hydrogen atoms). Equation 1 was minimized (Equations 1, 3, and 10). $R_w(F_2)$ refined to 0.2472, with an $R(F)$ of 0.0850, and a goodness of fit, S , of 1.04. Definitions used for calculating $R(F)$, $R_w(F^2)$ and the goodness of fit, S , are provided (Equations 4-6). The data were checked for secondary extinction effects but no correction was necessary. Neutral atom scattering factors and values used to calculate the linear absorption coefficient are from the International Tables for X-ray Crystallography⁶.

Equations:

$$\Sigma w(|F_o|^2 - |F_c|^2)^2 \quad (1)$$

$$w = 1/[(\sigma(F_o))^2 + (0.0717*P)^2] \quad (2)$$

$$P = (|F_o|^2 + 2|F_c|^2)/3 \quad (3)$$

$$R_w(F^2) = \{\Sigma w(|F_o|^2 - |F_c|^2)^2 / \Sigma w(|F_o|)^4\}^{1/2} \quad (4)$$

where w is weight given each reflection

$$R(F) = \Sigma(|F_o| - |F_c|) / \Sigma|F_o| \text{ for reflections with } F_o > 4(\sigma(F_o)) \quad (5)$$

$$S = [\Sigma w(|F_o|^2 - |F_c|^2)^2 / (n - p)]^{1/2} \quad (6)$$

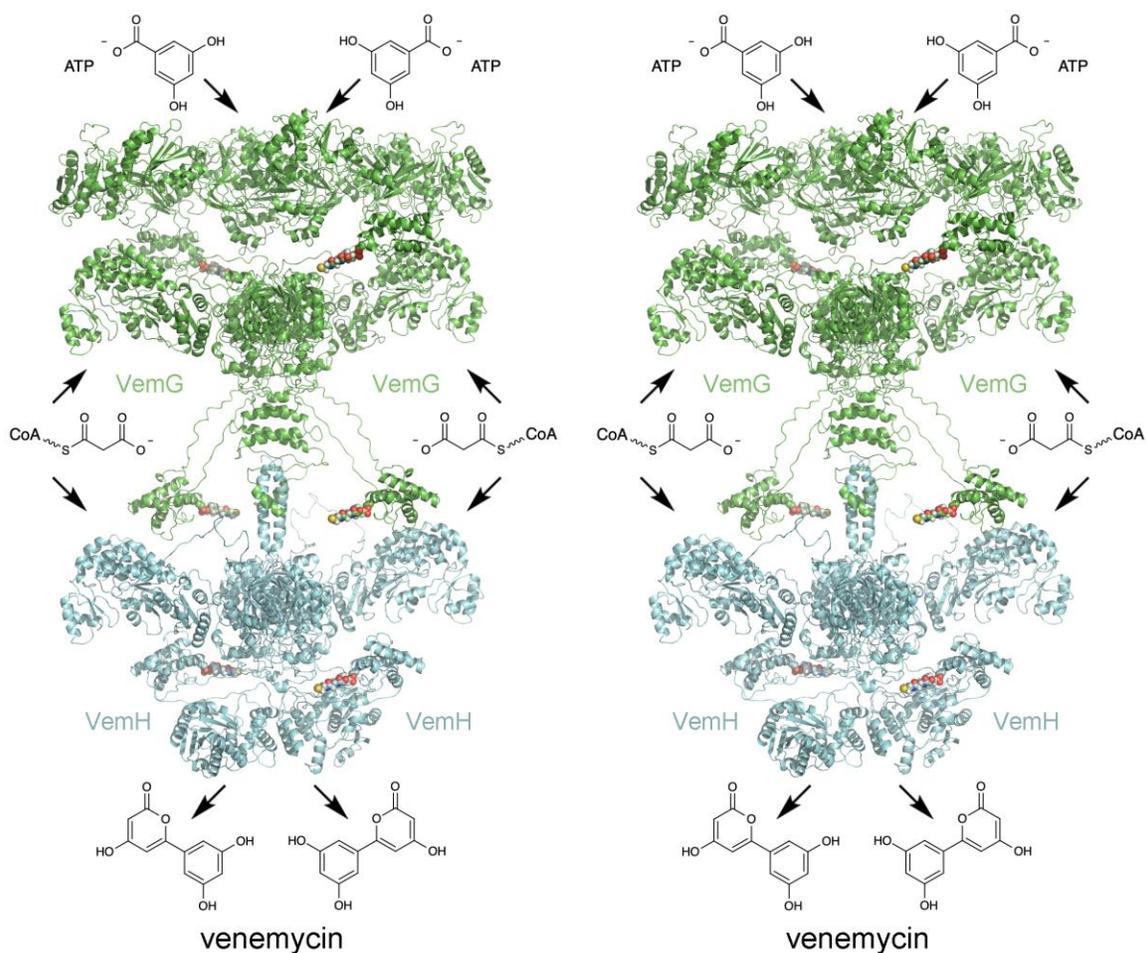
where n is the number of reflections and p is the number of refined parameters

$$w = 1/[(\sigma(F_o))^2 + (0.1135*P)^2 + (0.10739*P)] \quad (7)$$

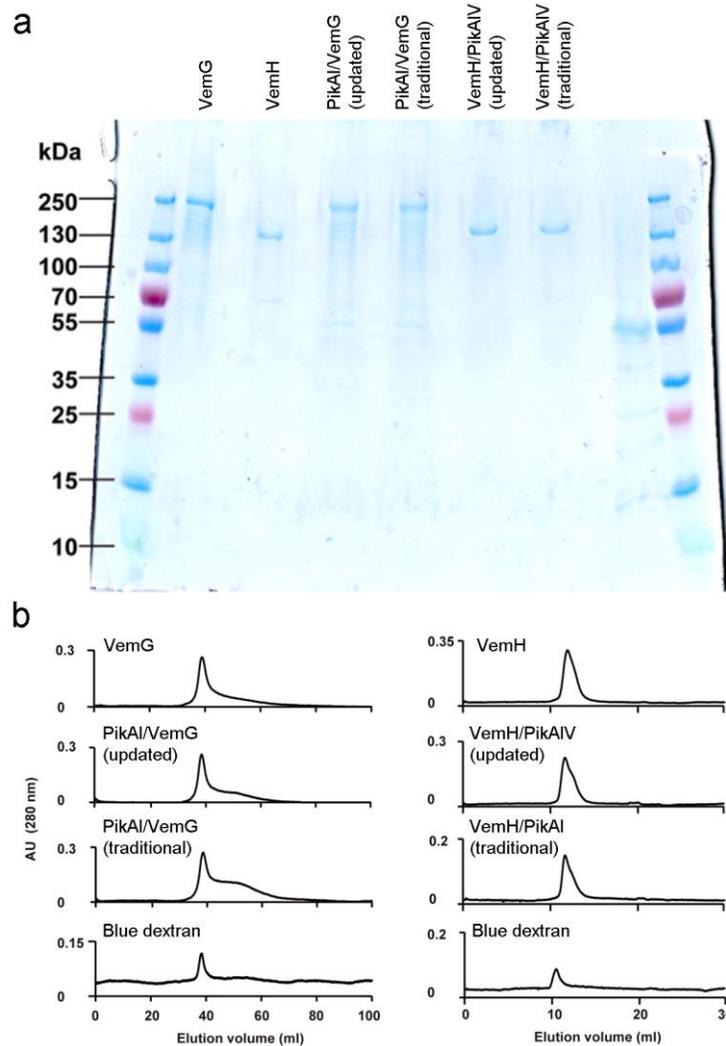
$$w = 1/[(\sigma(F_o)) + (0.12*P)^2] \quad (8)$$

$$w = 1/[(\sigma(F_o))^2 + (0.1208*P)^2] \quad (9)$$

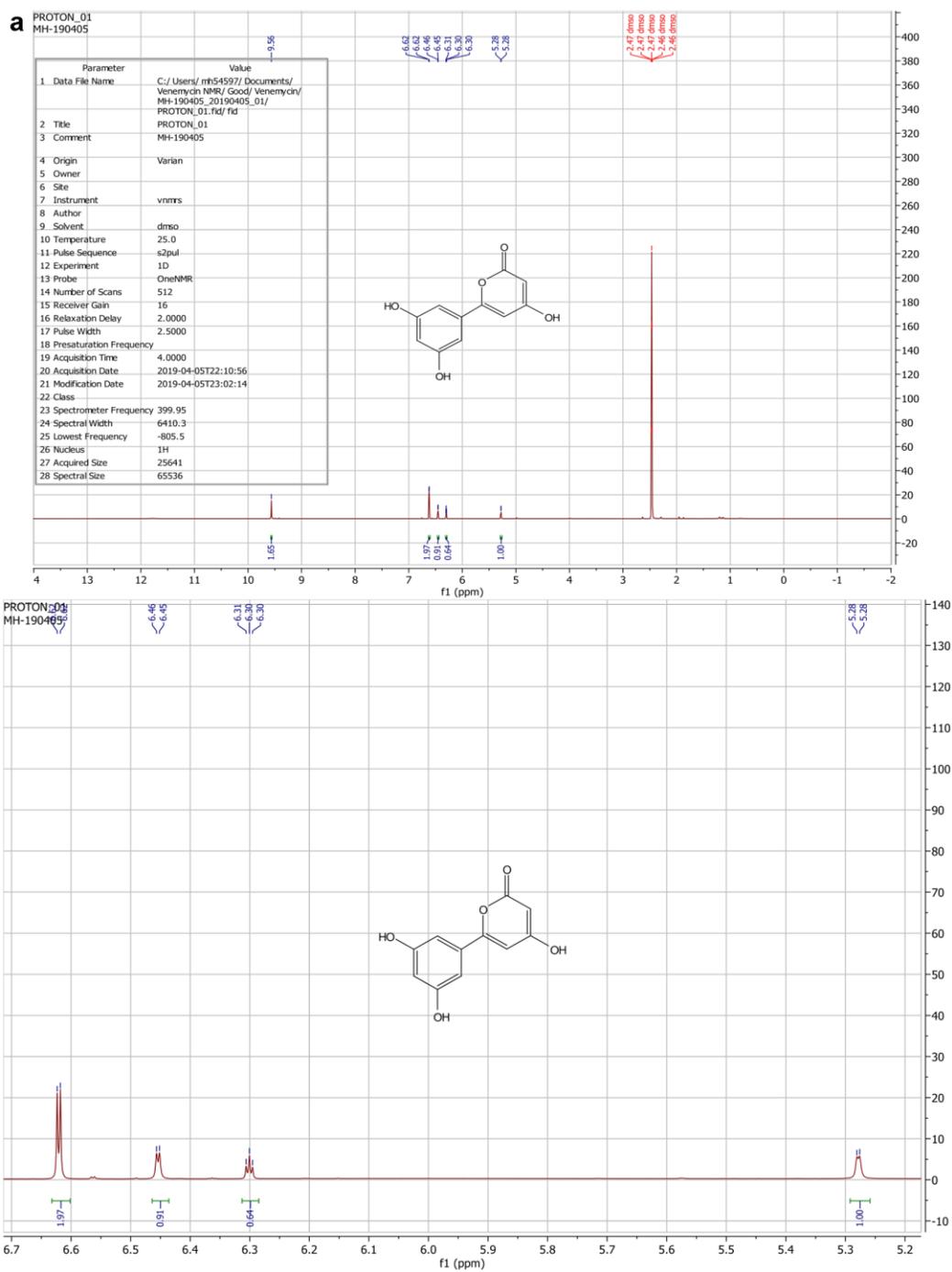
$$w = 1/[(\sigma(F_o))^2 + (0.1318*P)^2 + (1.5455*P)] \quad (10)$$



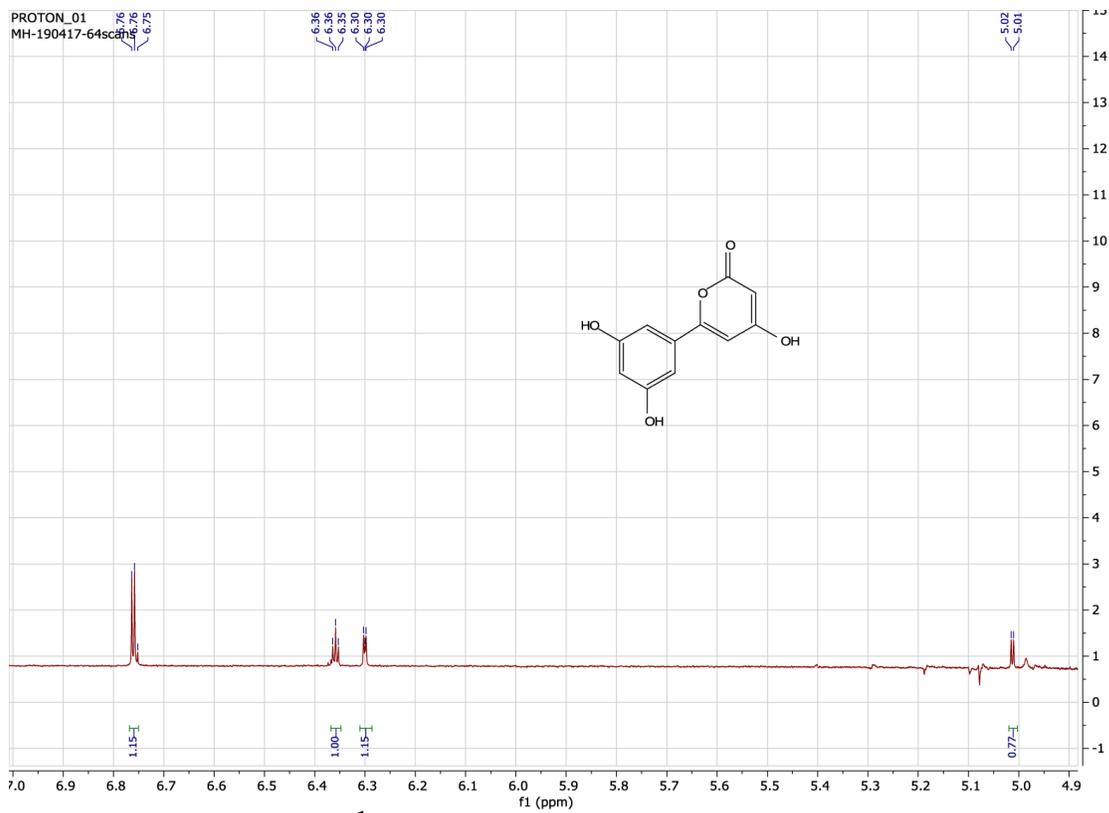
Supplementary Figure 1. All-atom model of the venemycin assembly line. A wall-eyed stereodiamgram shows a model of VemG (green) and VemH (cyan) within the venemycin assembly line. All residues, including those of flexible linkers, are included. The phosphopantetheinyl arms of ACP domains are represented in spheres. Coordinates can be downloaded at http://keatinge-clay.cm.utexas.edu/research/VemGH_model.pdb.



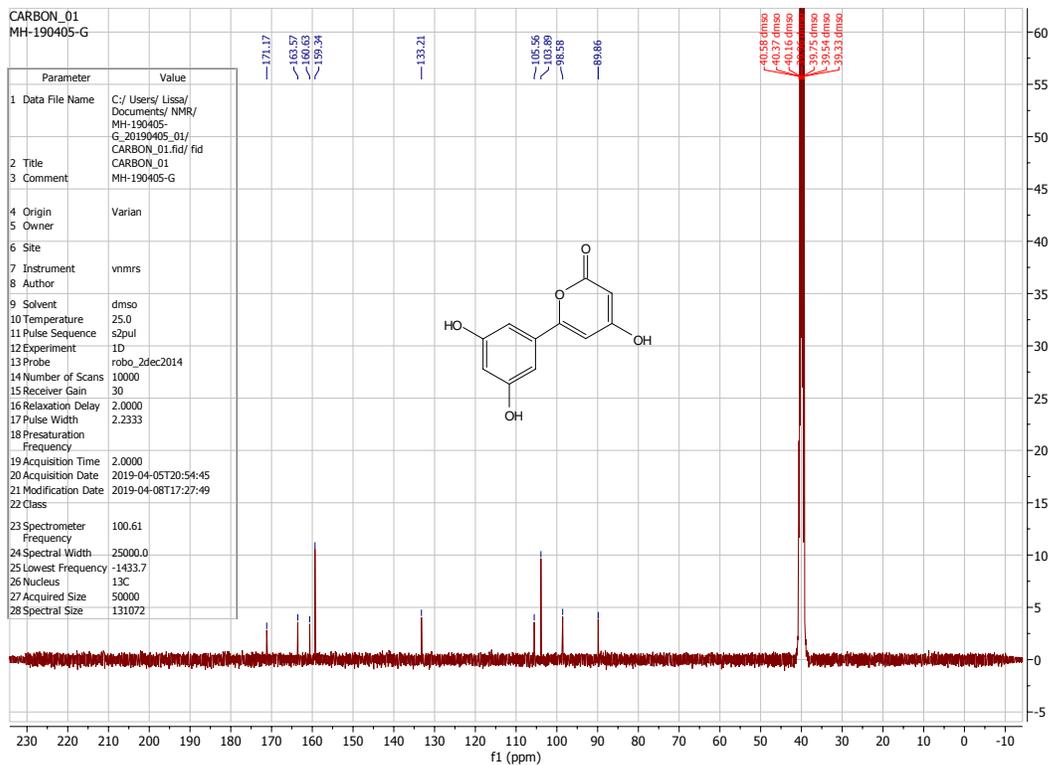
Supplementary Figure 2. Analysis of protein quality. a) SDS-PAGE gel of VemG, VemH, hybrids, and MatB. Lane 1 - VemG, 232 kDa; lane 2 - VemH, 140 kDa; lane 3 - PikAI/VemG (updated), 233 kDa; lane 4 - PikAI/VemG (traditional), 233 kDa; lane 5 - VemH/PikAIV (updated), 144 kDa; lane 6 - VemH/PikAIV (traditional), 143 kDa; lane 7, MatB (53 kDa). PageRuler Plus Prestained Protein Ladder, 4-20% Tris-glycine gel (Thermo Fisher Scientific). b) Size exclusion chromatography. VemG and PikAI/VemG hybrids were analyzed by Sephacryl S-300 HR column equilibrated with size exclusion buffer (50 mM potassium phosphate, 150 mM NaCl, 1 mM TCEP, 10% v/v glycerol, pH 7.5). VemH and VemH/PikAIV hybrids were analyzed by Superdex 200 Increase 16/600 GL column equilibrated with size exclusion buffer. Blue dextran indicates the void volumes for both columns.



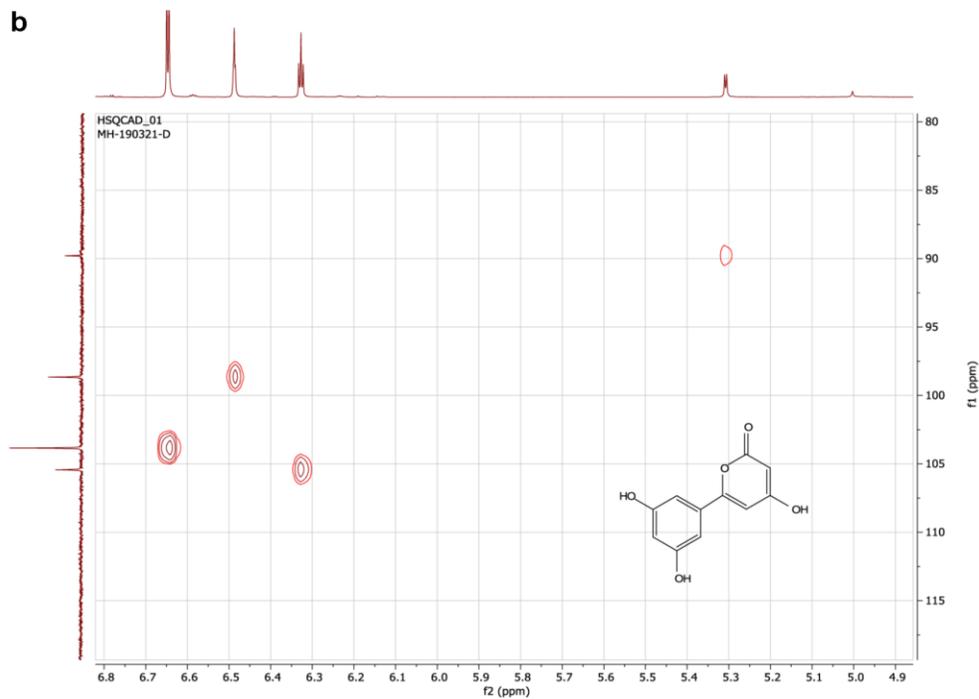
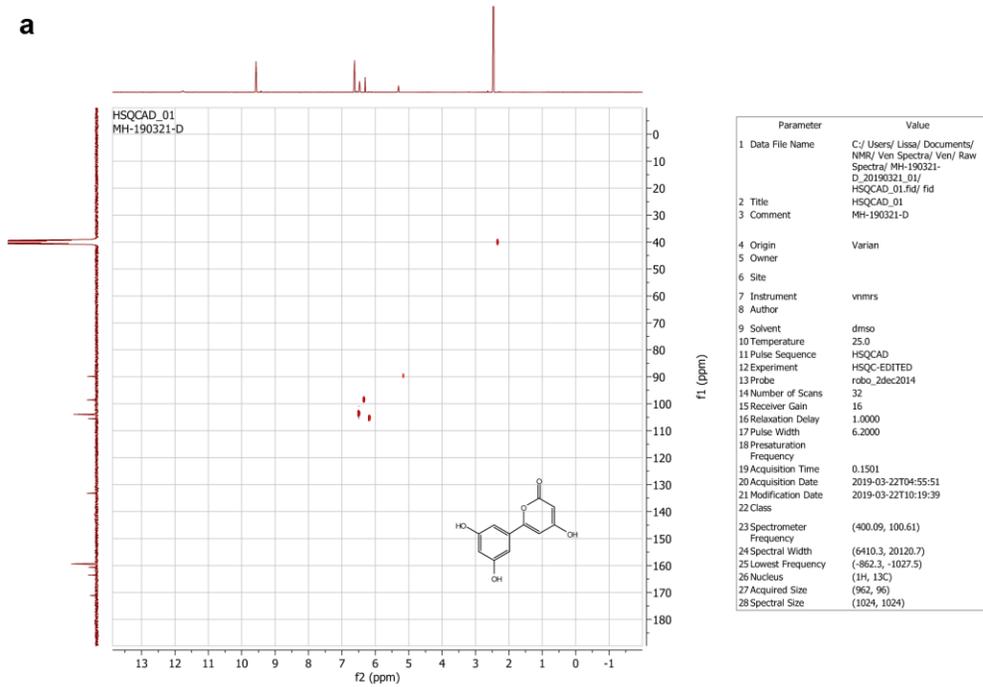
Supplementary Figure 3. ^1H NMR of venemycin (1) in DMSO- d_6 . a) Full spectrum. b) Expanded region.



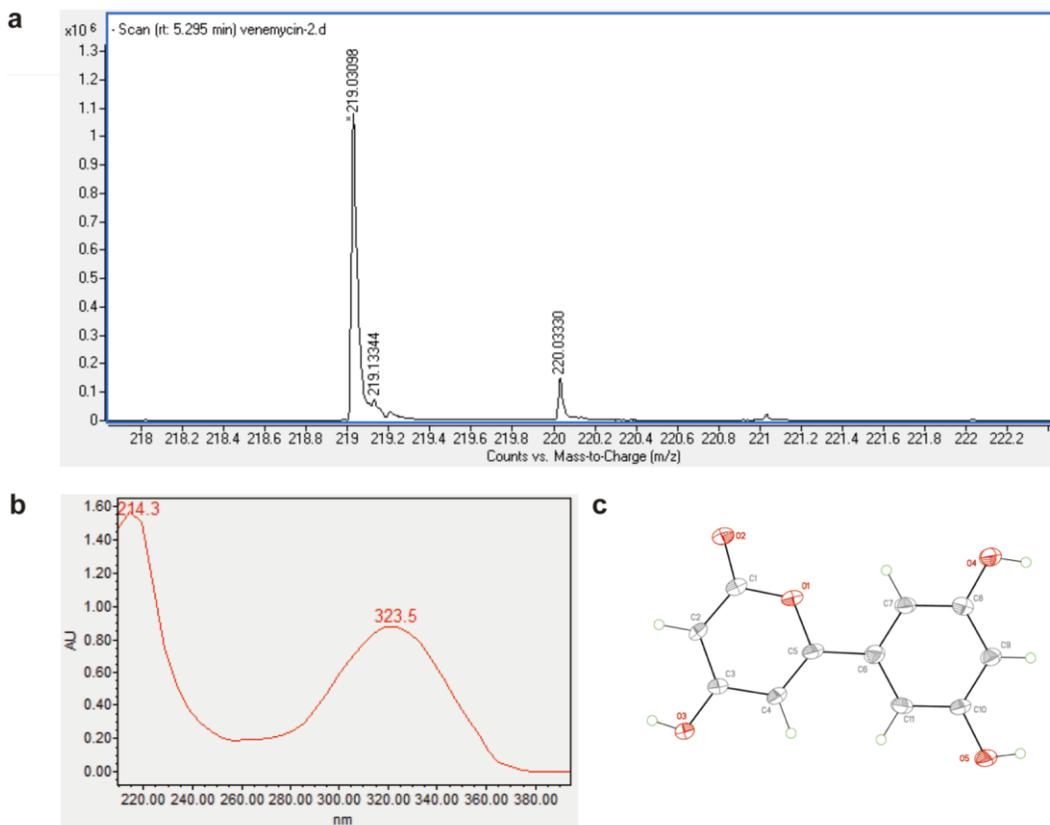
Supplementary Figure 4. ^1H NMR of venemycin (1) in D_2O .



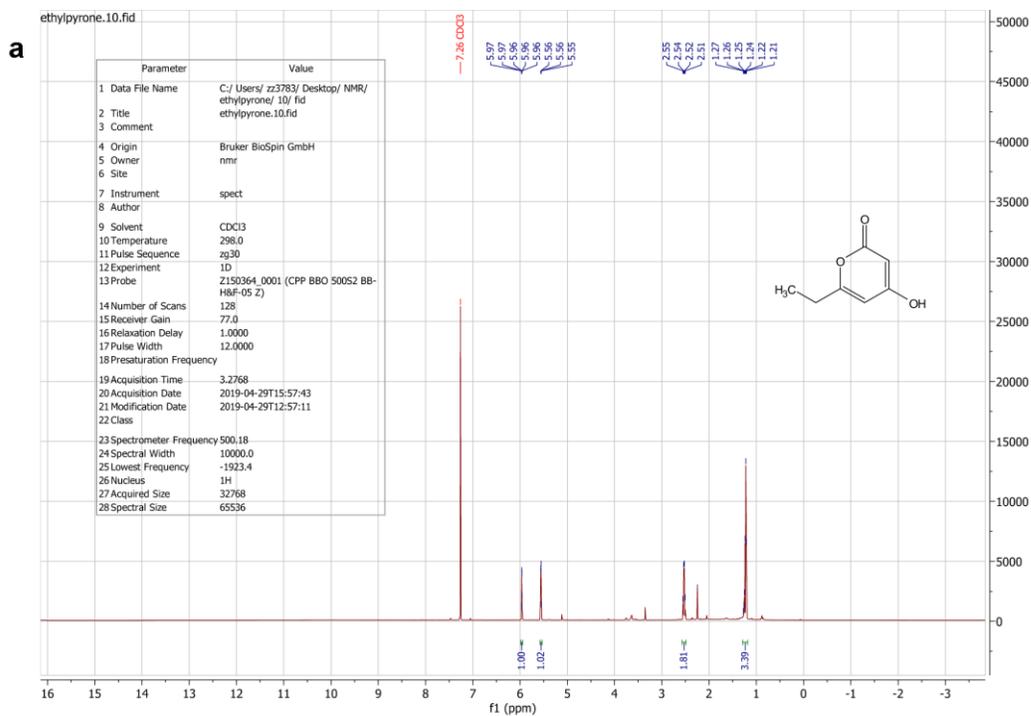
Supplementary Figure 5. ^{13}C NMR of venemycin (1) in DMSO-d_6 .

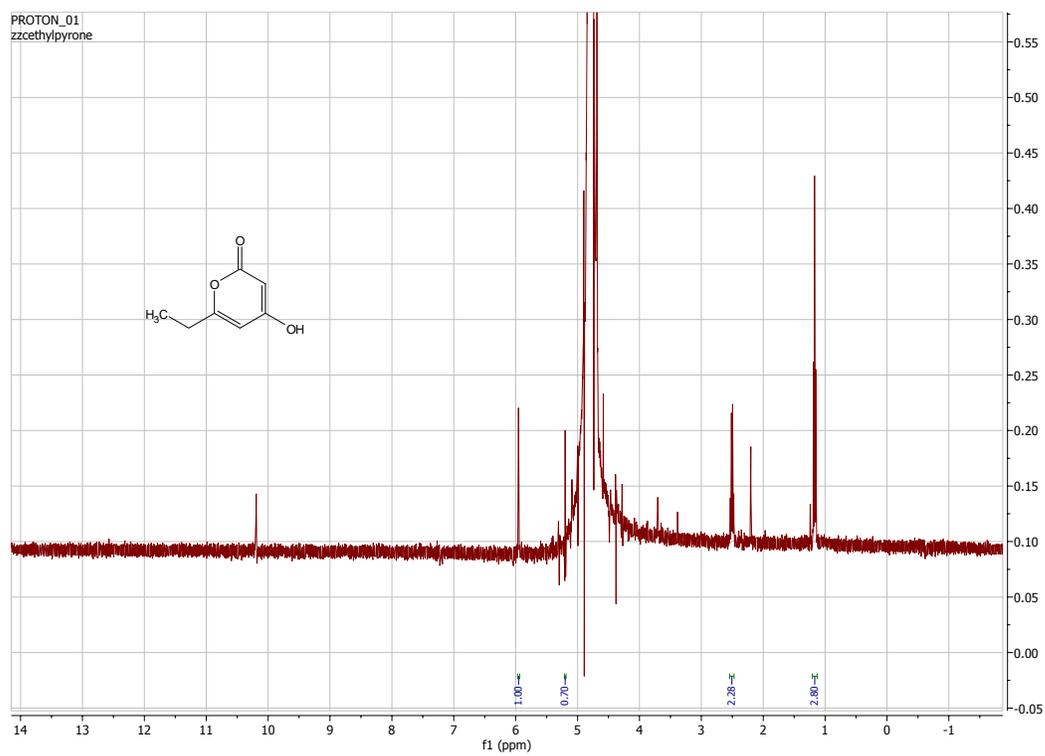


Supplementary Figure 6. ^1H - ^{13}C -HSQC of venemycin (1) in DMSO- d_6 . a) Full spectrum. b) Expanded region.

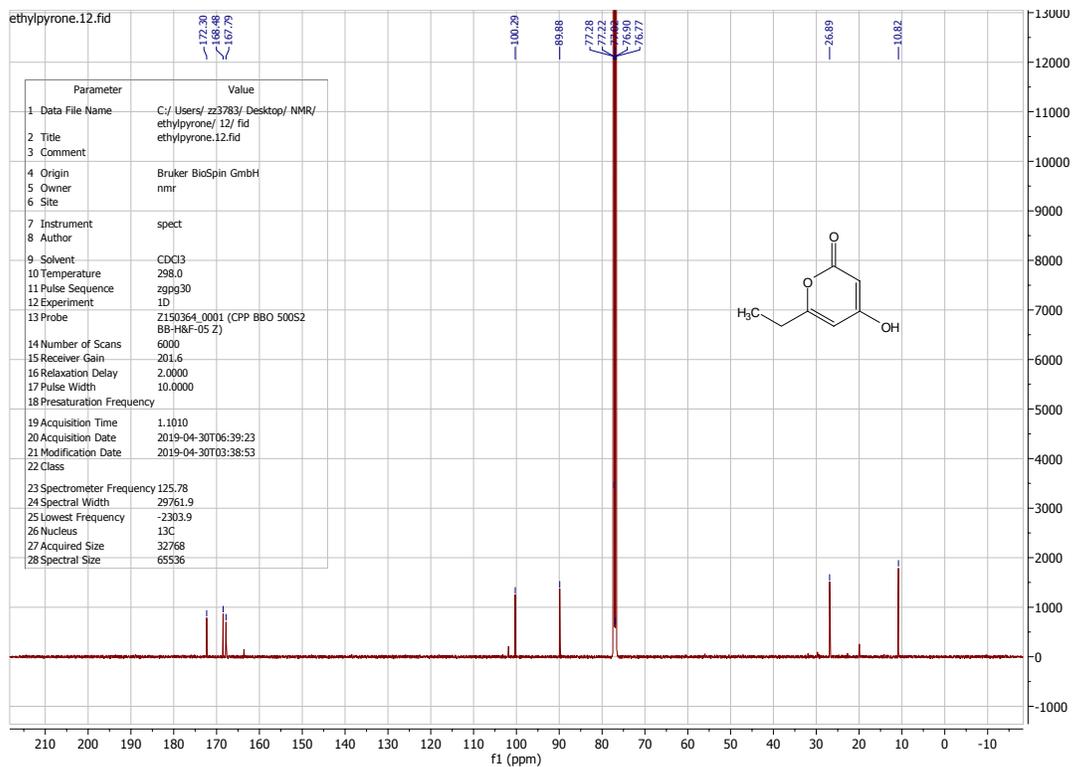


Supplementary Figure 7. Additional characterization of venemycin (1). **a)** High resolution MS. $[M-H]^-$ 219.0299 m/z calculated, 219.0310 m/z found . **b)** Absorbance spectrum. **c)** X-ray crystal structure.

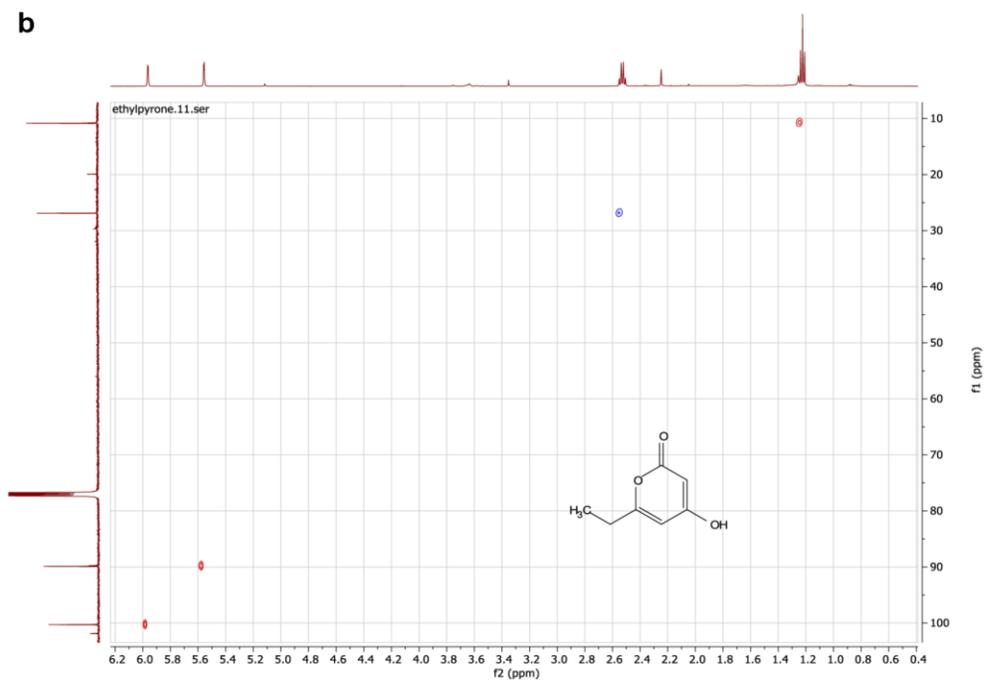
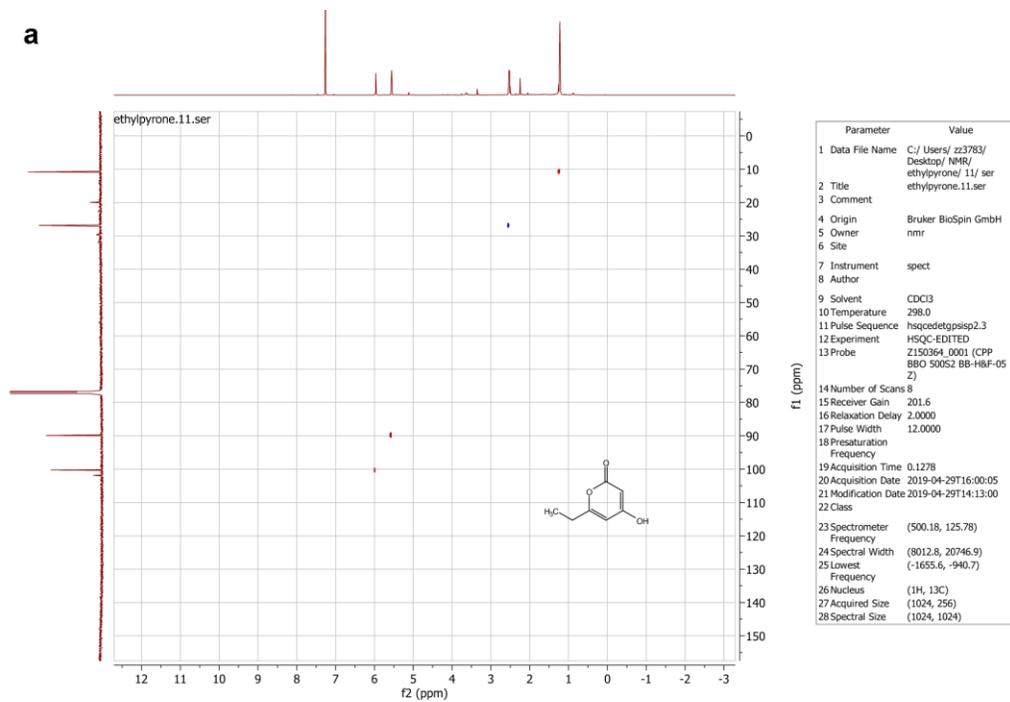




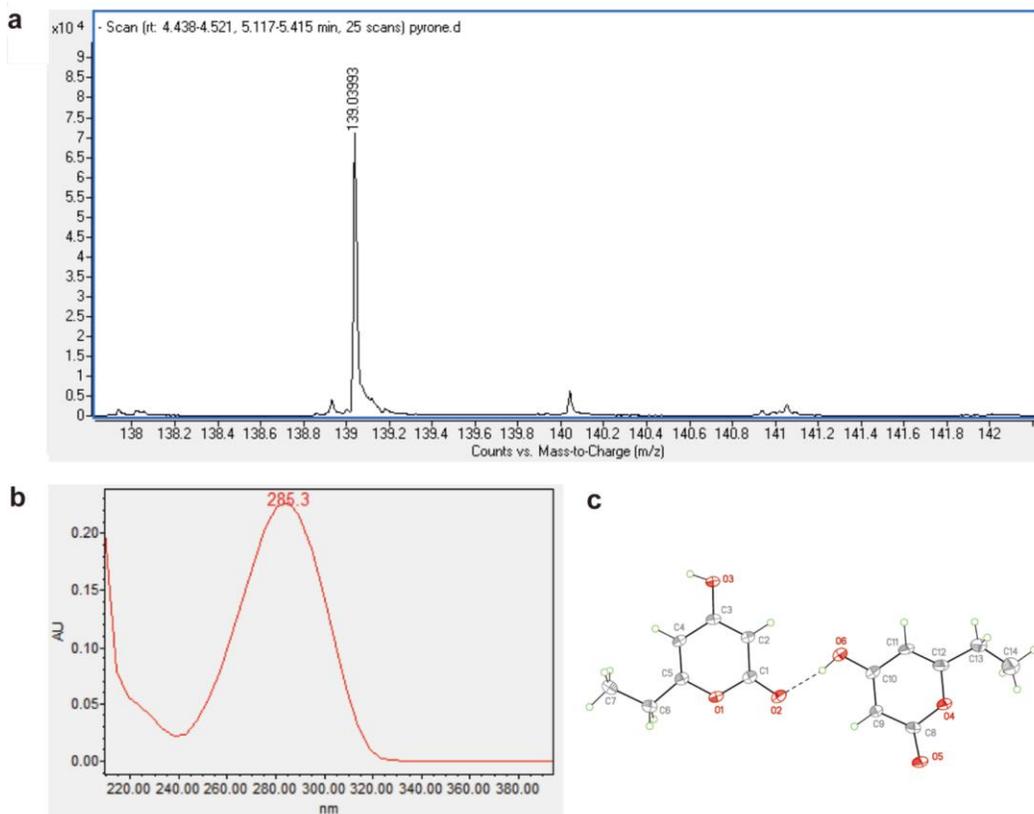
Supplementary Figure 9. ^1H NMR of pyrone (2) in D_2O .



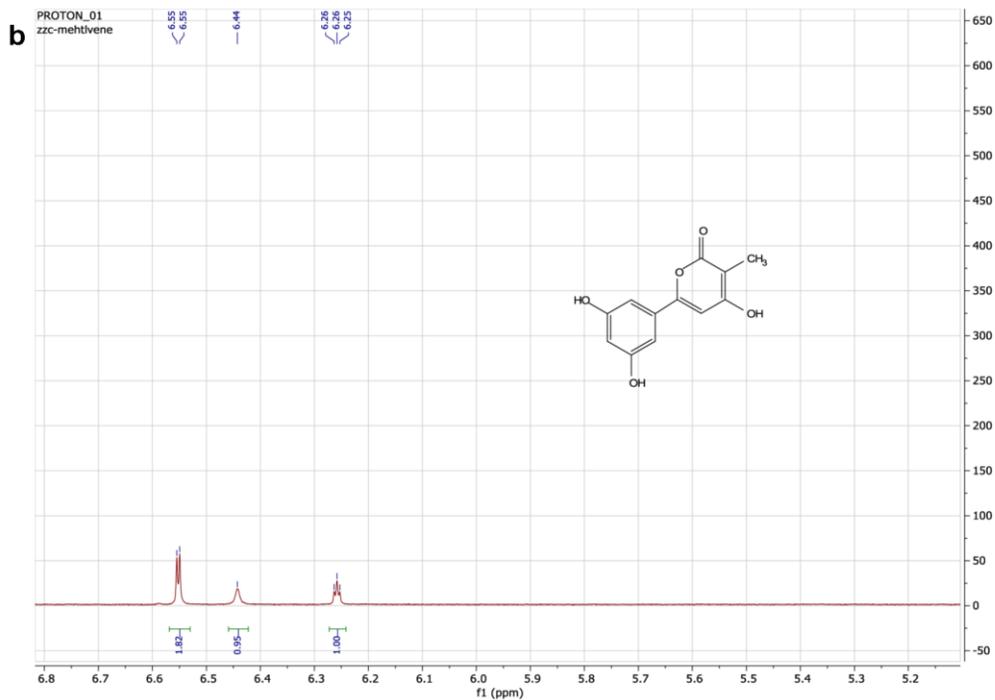
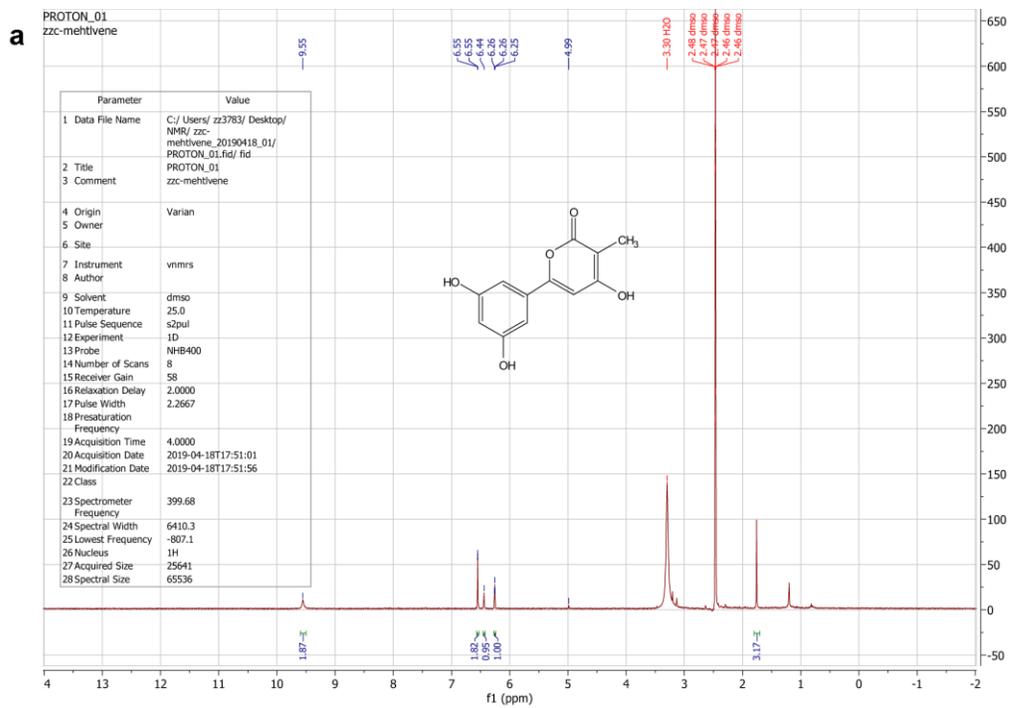
Supplementary Figure 10. ¹³C NMR of pyrone (2) in CDCl₃.



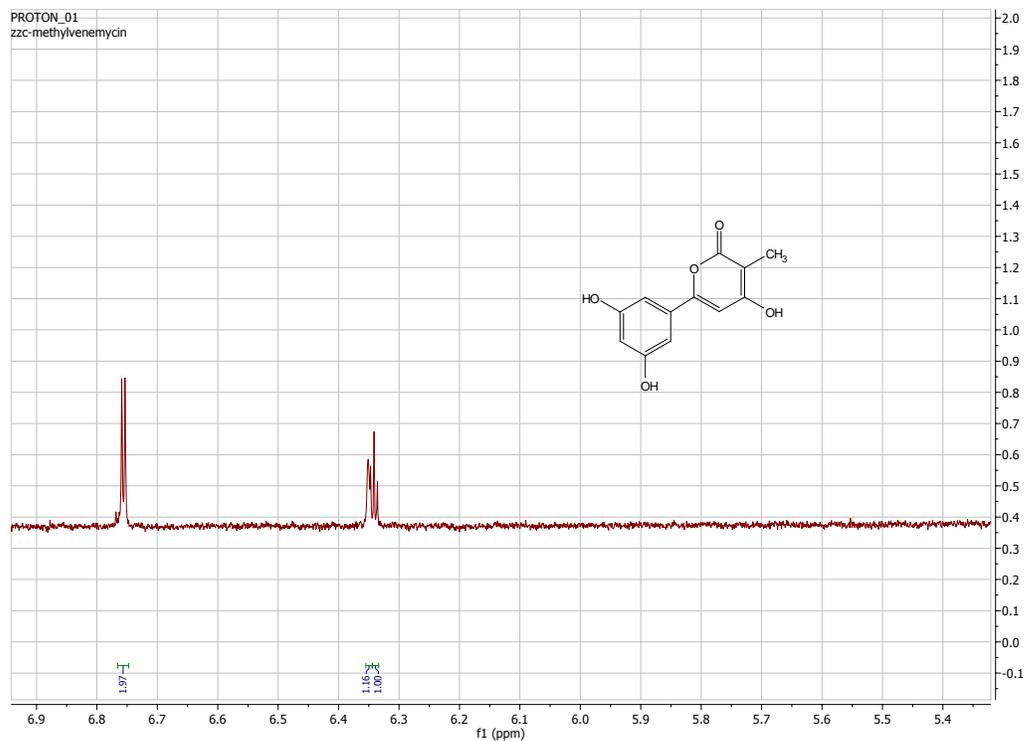
Supplementary Figure 11. ^1H - ^{13}C -HSQC of pyrone (2) in CDCl_3 . a) Full spectrum. b) Expanded region.



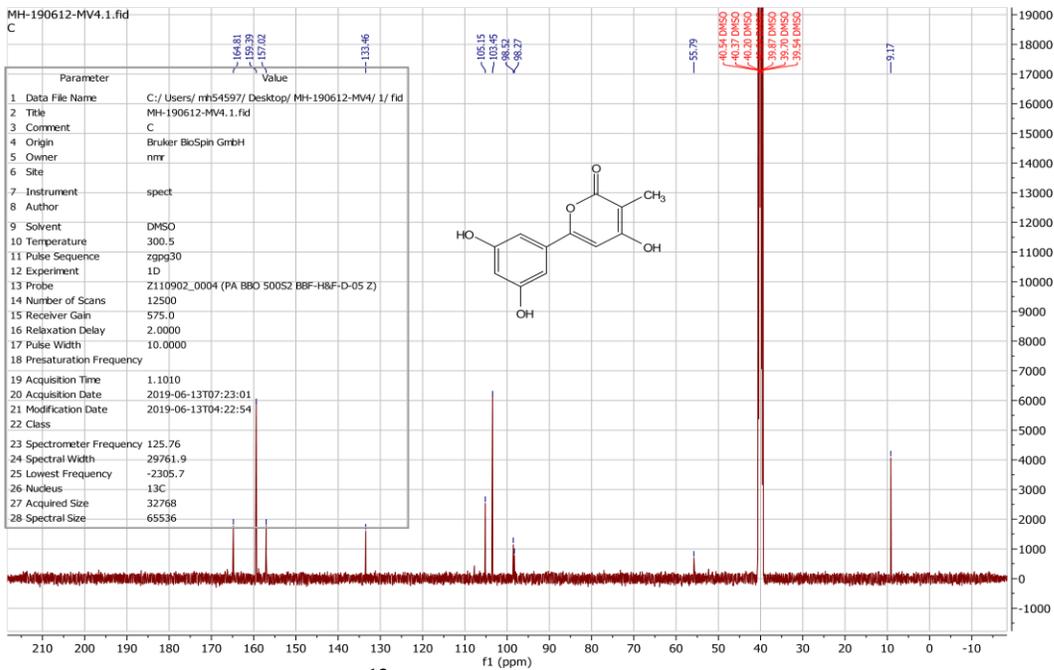
Supplementary Figure 12. Additional characterization of pyrone (2). a) High resolution MS. $[M-H]^-$ 139.0401 m/z calculated, 139.0399 m/z found. b) Absorbance spectrum. c) X-ray crystal structure.



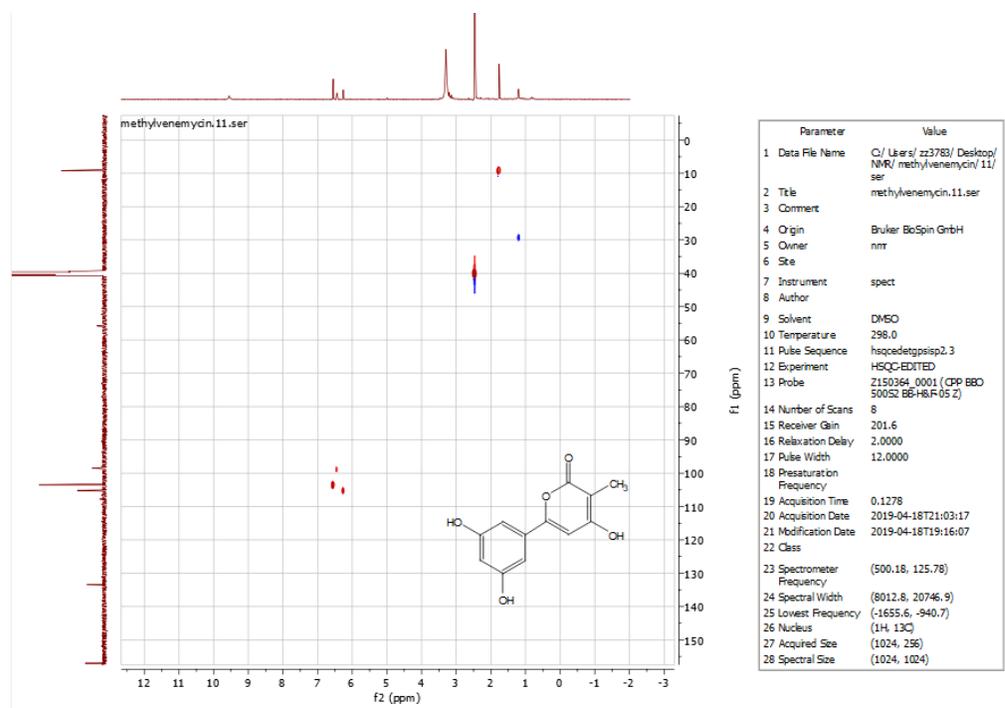
Supplementary Figure 13. ¹H NMR of methylvenemycin (3) in DMSO-d₆. a) Full spectrum. b) Expanded region.



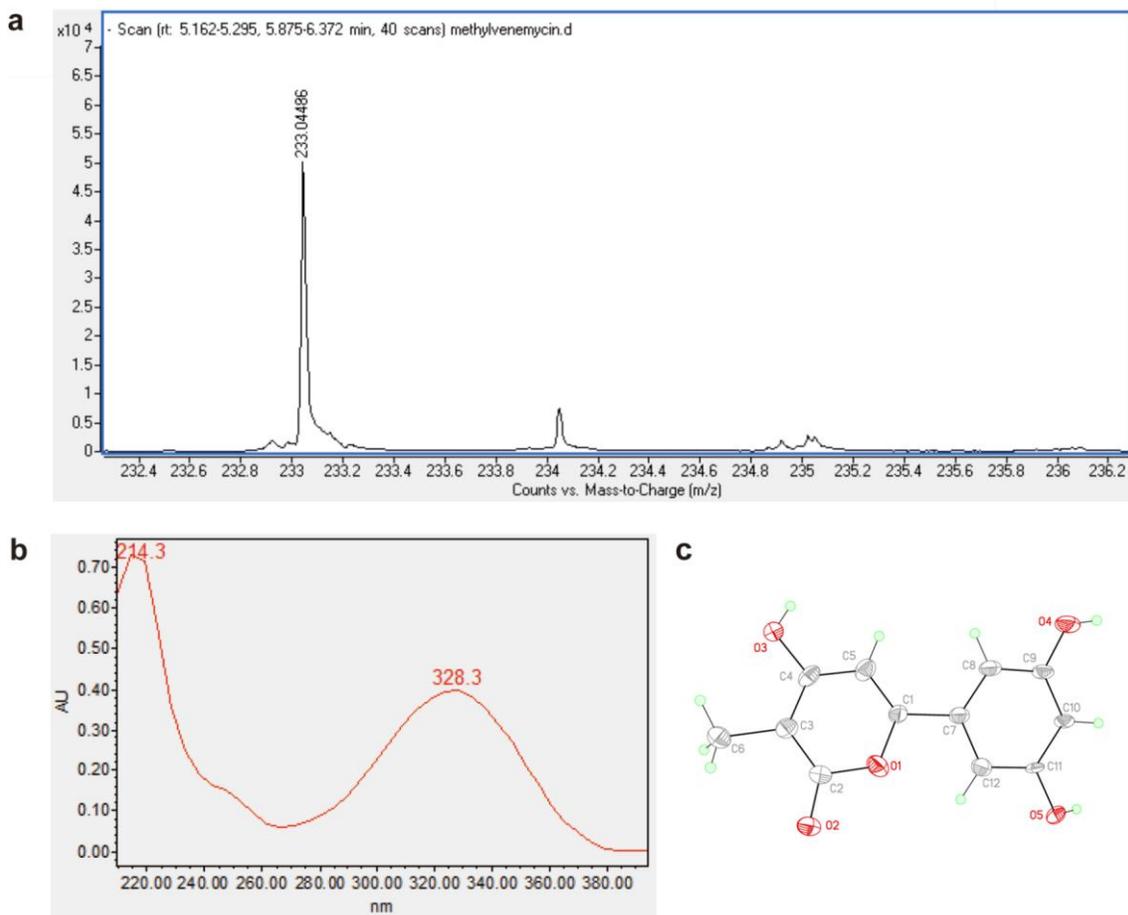
Supplementary Figure 14. ^1H NMR of methylvenemycin (3) in D_2O .



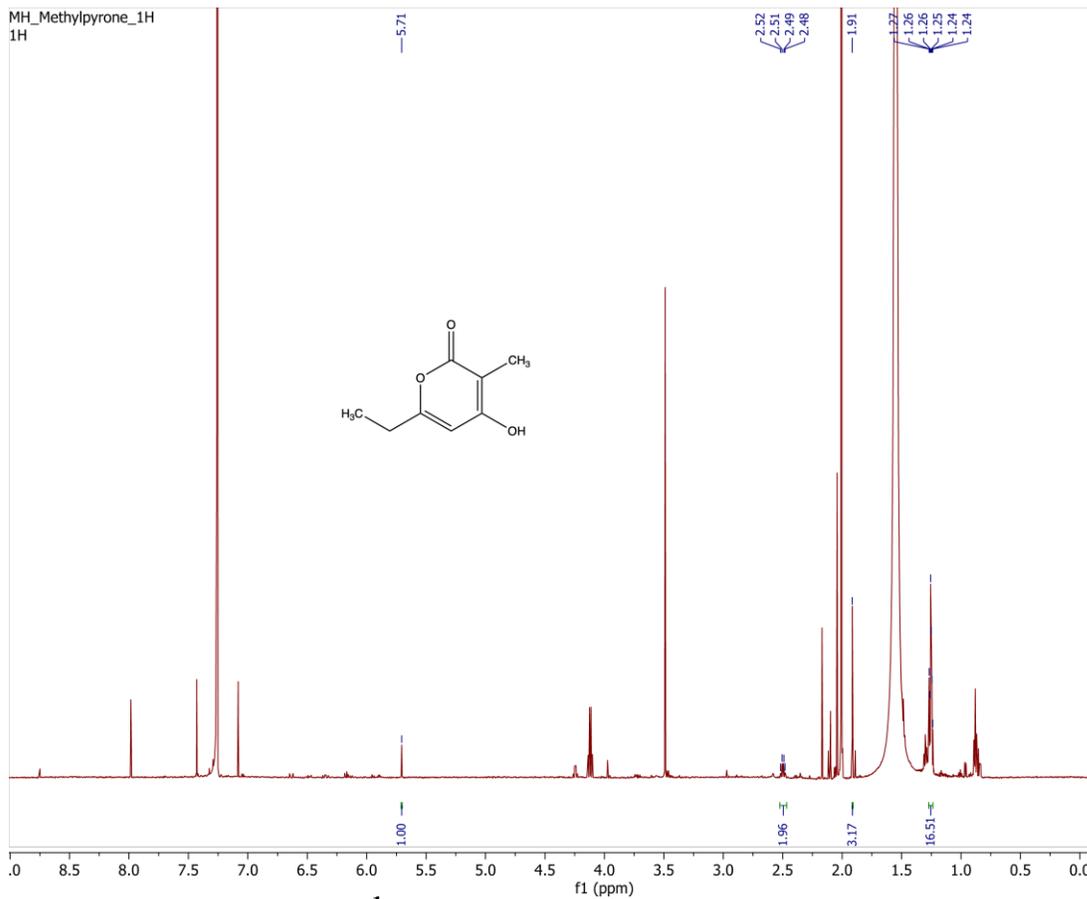
Supplementary Figure 15. ¹³C NMR of methylvenemycin (3) in DMSO-d₆.



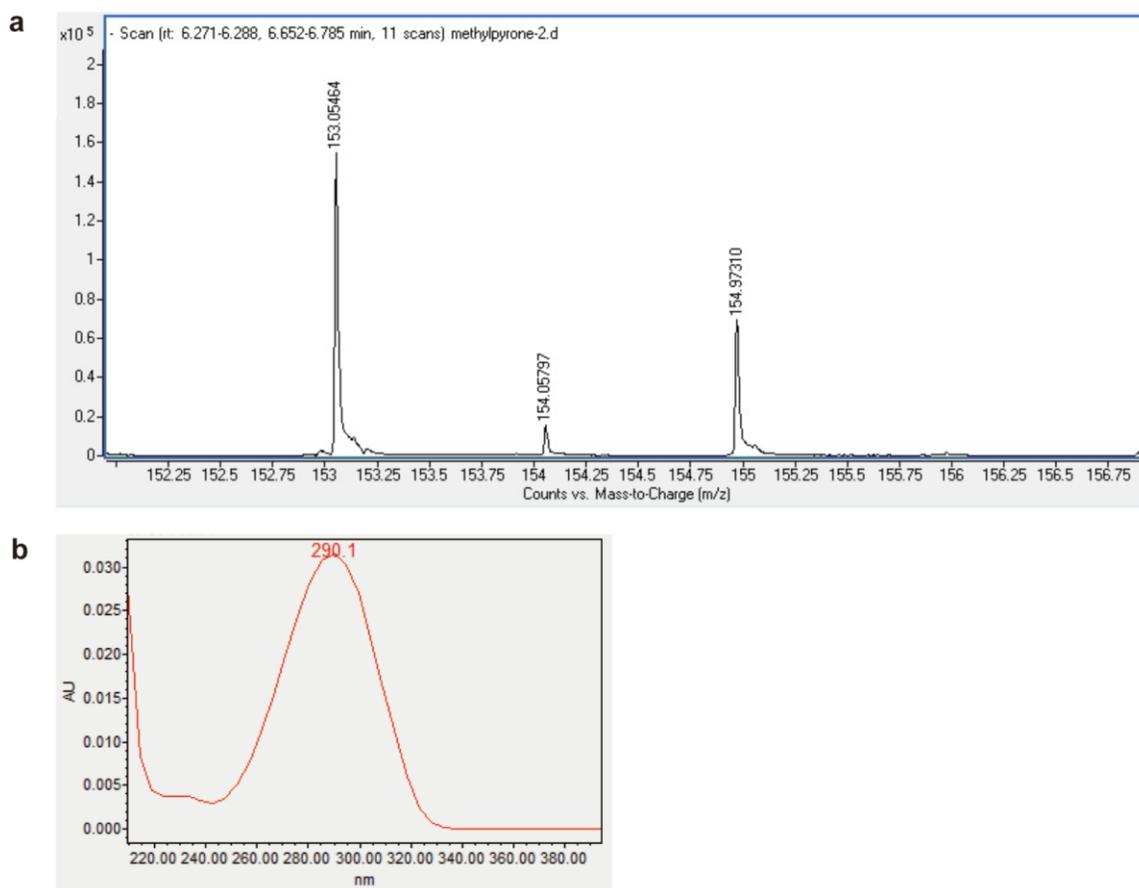
Supplementary Figure 16. ^1H - ^{13}C -HSQC of methylvenemycin (3) in DMSO- d_6 .



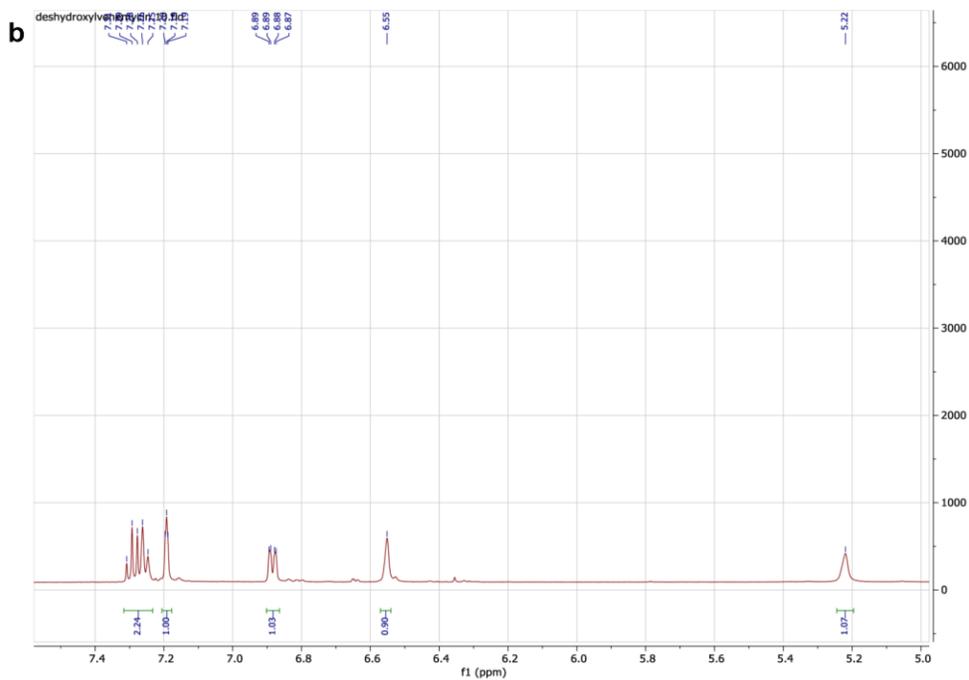
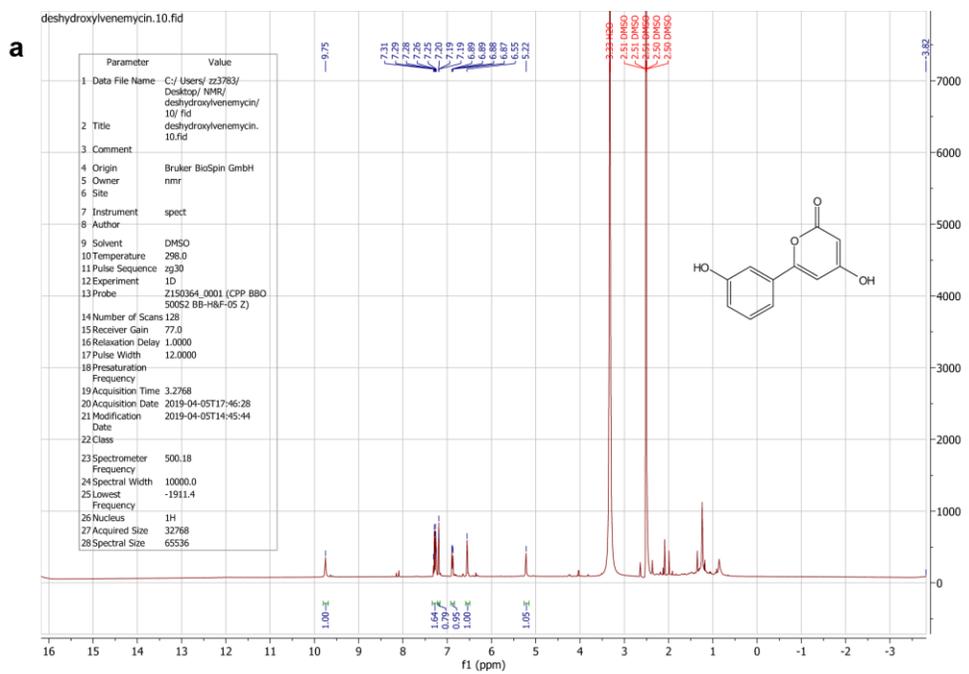
Supplementary Figure 17. Additional characterization of methylvenemycin (3). **a)** High resolution MS. 233.0455 m/z calculated, 233.0449 m/z found. **b)** Absorbance spectrum. **c)** X-ray crystal structure.



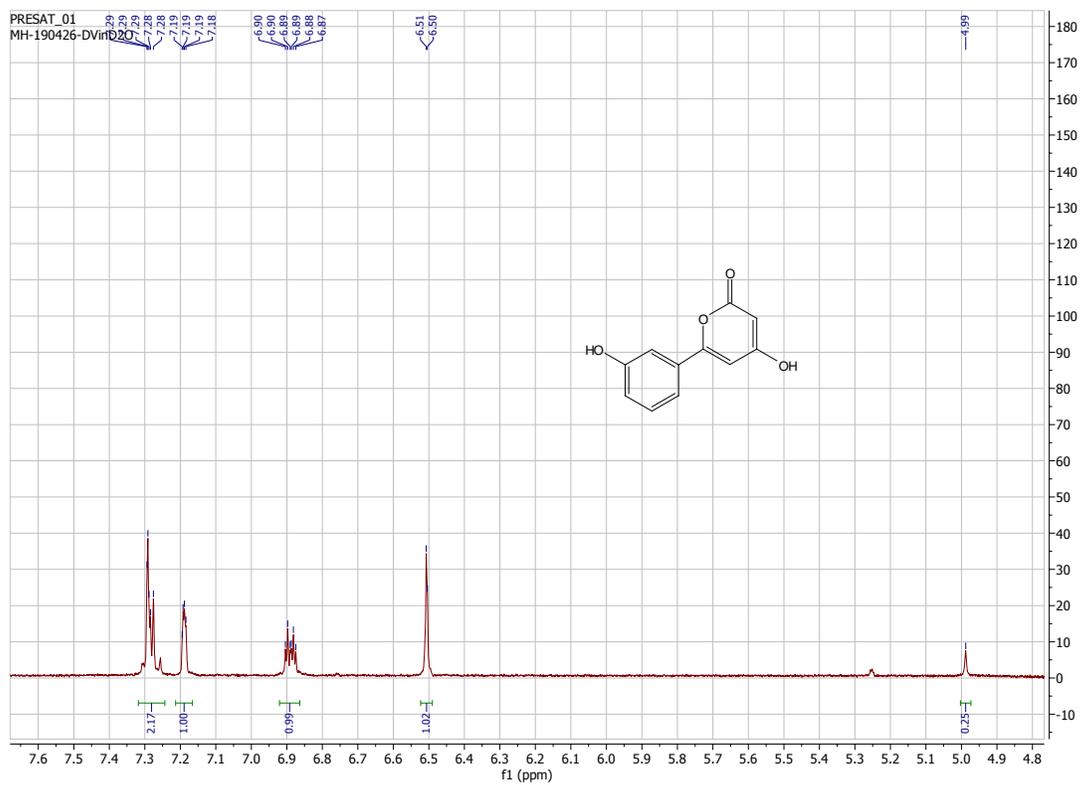
Supplementary Figure 18. ^1H NMR of methylpyrone (4) in CDCl_3 .



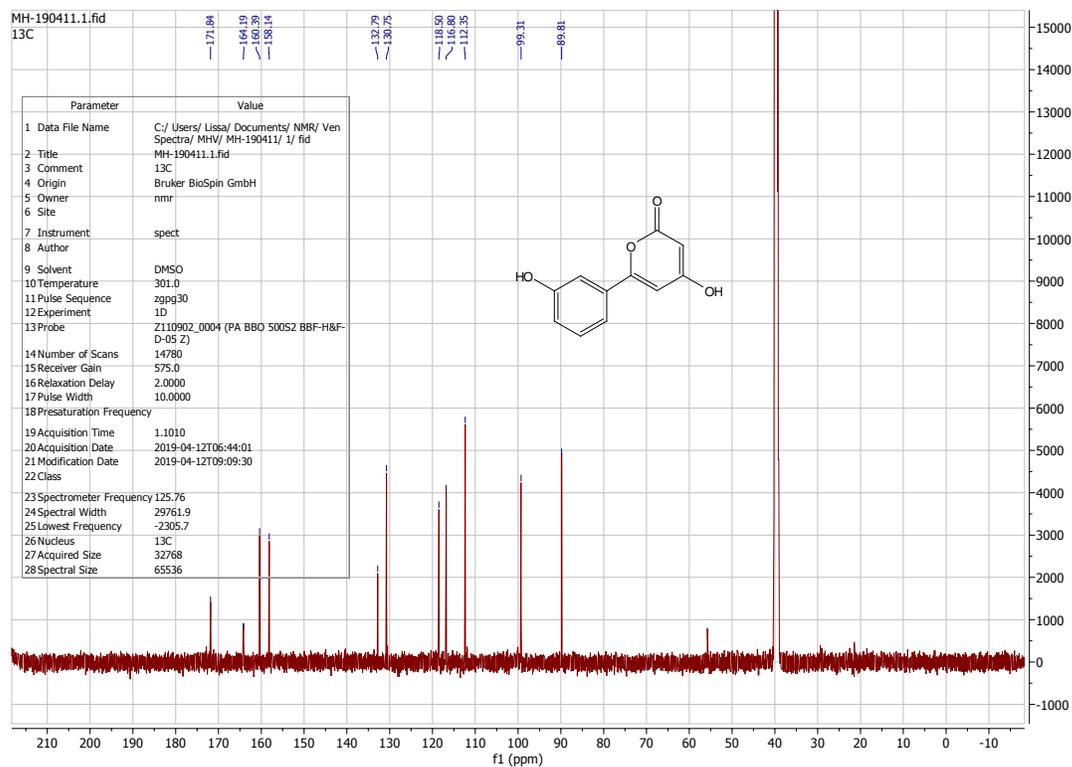
Supplementary Figure 19. Additional characterization of methylpyrone (4). a) High resolution MS. 153.0557 m/z calculated, 153.0546 m/z found. b) Absorbance spectrum.



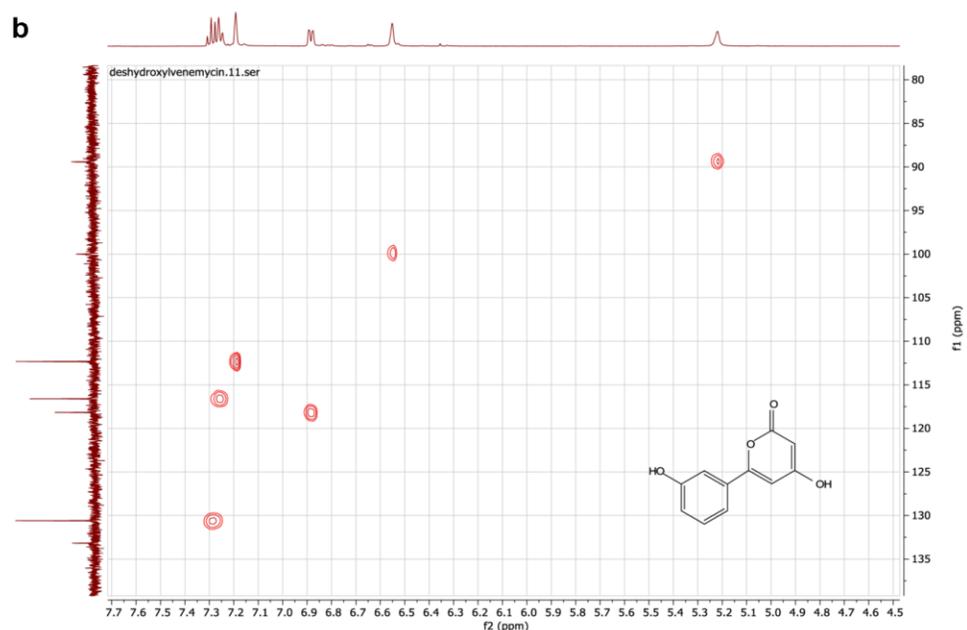
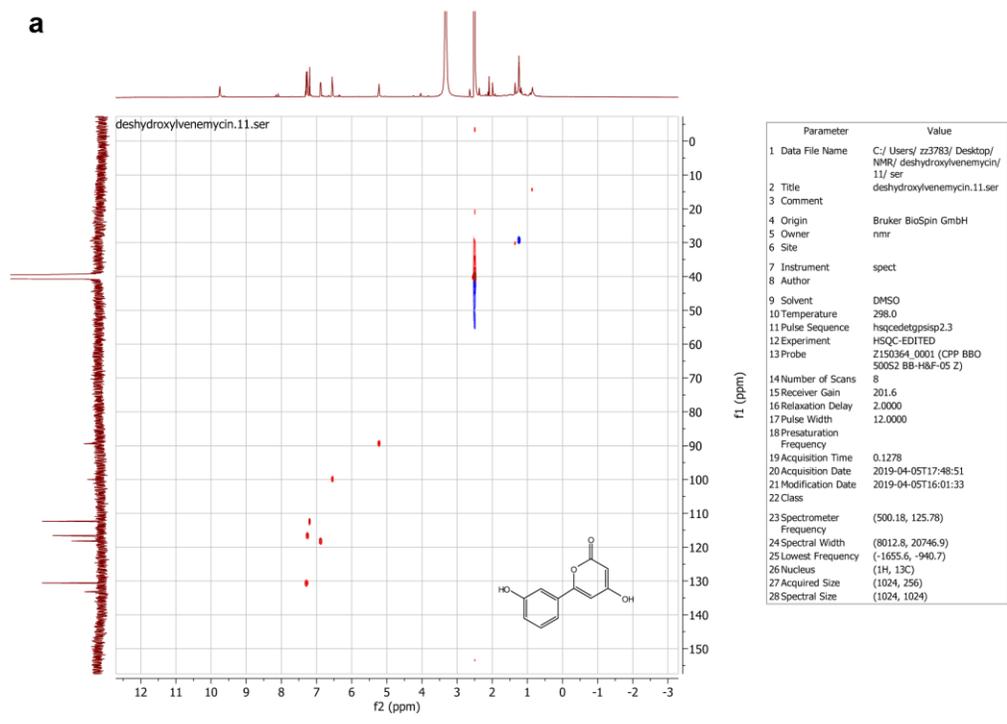
Supplementary Figure 20. ^1H NMR of deshydroxyvenemycin (5) in DMSO- d_6 . **a)** Full spectrum. **b)** Expanded region.



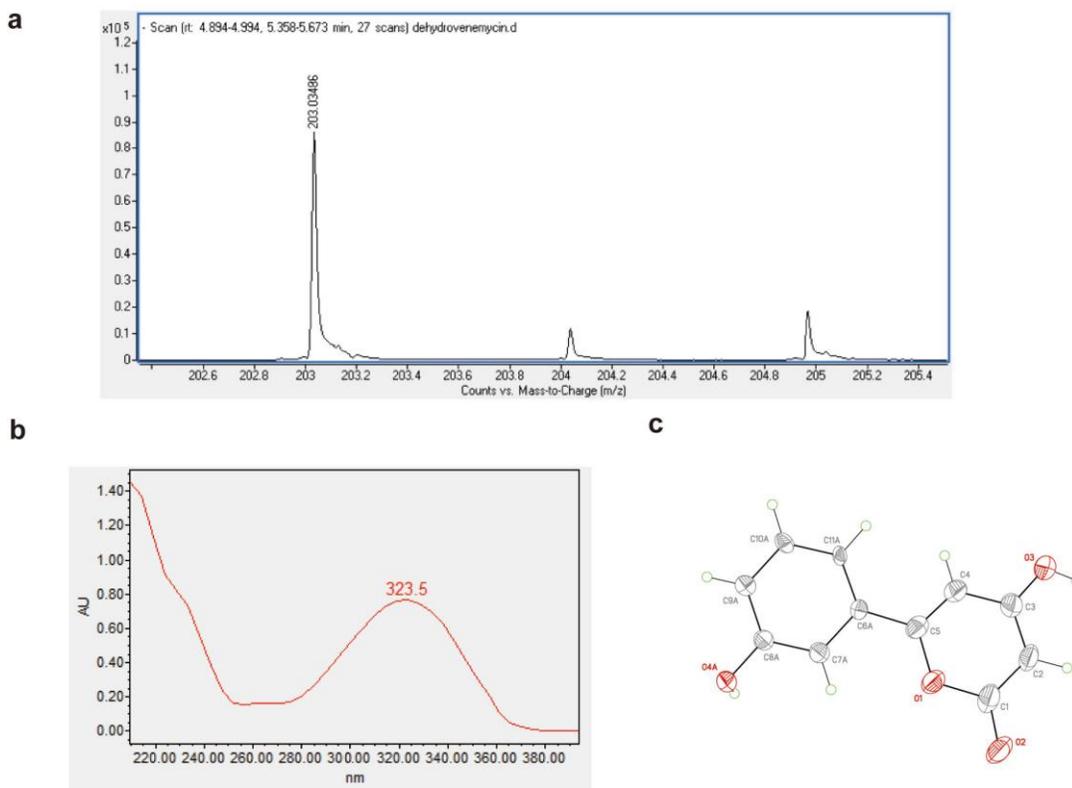
Supplementary Figure 21. ¹H NMR of deshydroxyvenemycin (5) in D₂O.



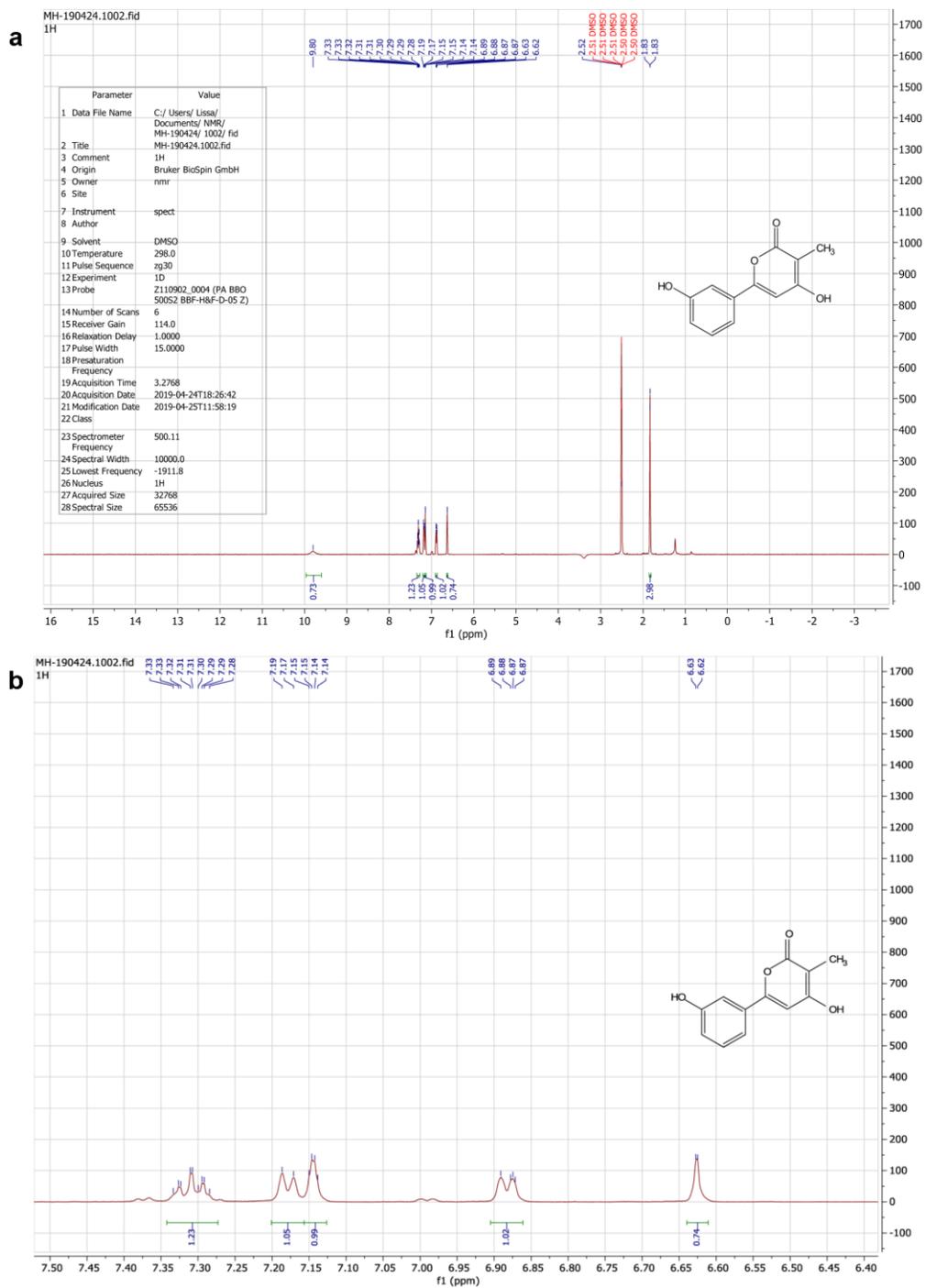
Supplementary Figure 22. ^{13}C NMR of deshydroxyvenemycin (5) in DMSO-d_6 .



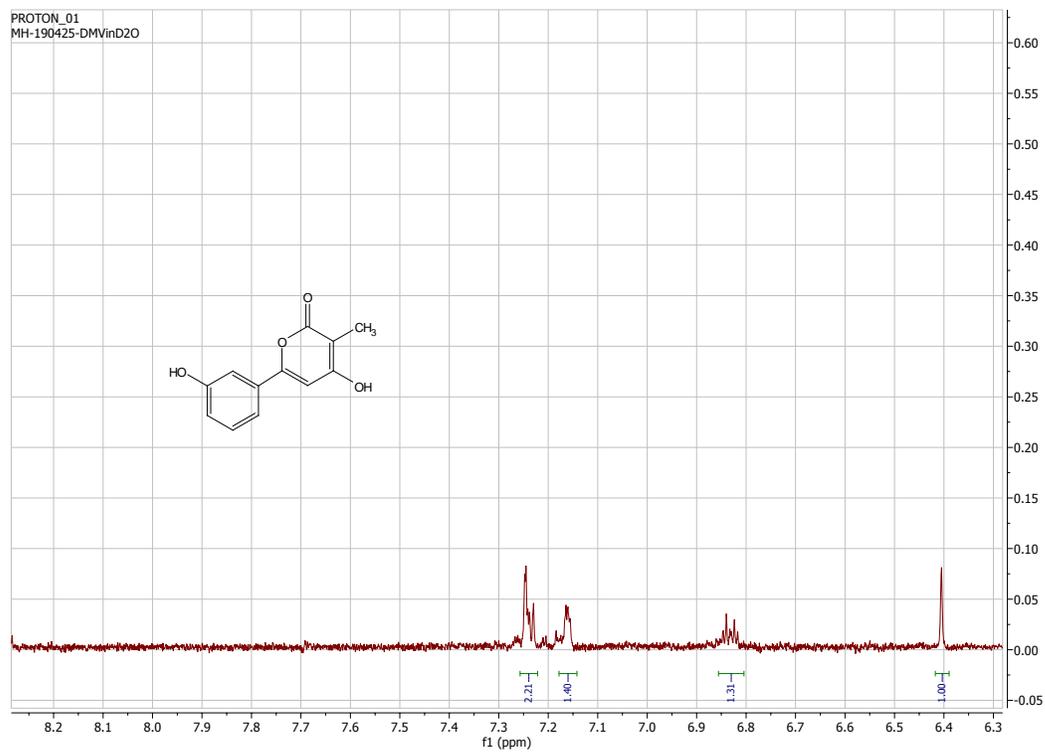
Supplementary Figure 23. ^1H - ^{13}C -HSQC of deshydroxyvenemycin (5) in DMSO- d_6 .
a) Full spectrum. b) Expanded region.



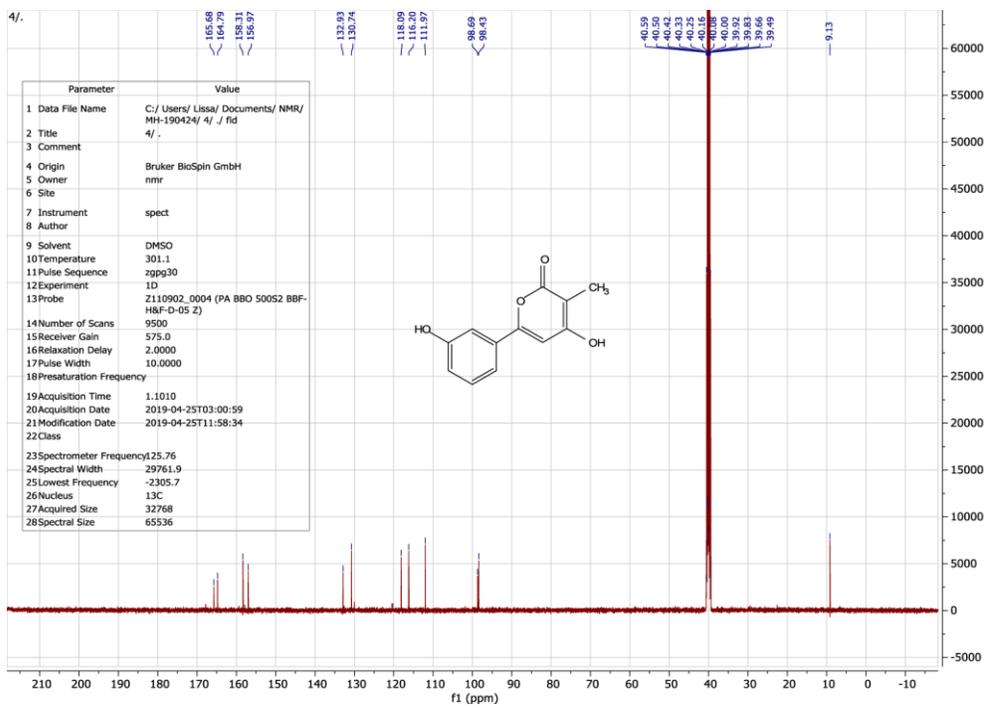
Supplementary Figure 24. Additional characterization of deshydroxyvenemycin (5).
a) High resolution MS. 203.0350 m/z calculated, 203.0310 m/z found. **b)** Absorbance spectrum. **c)** X-ray crystal structure.



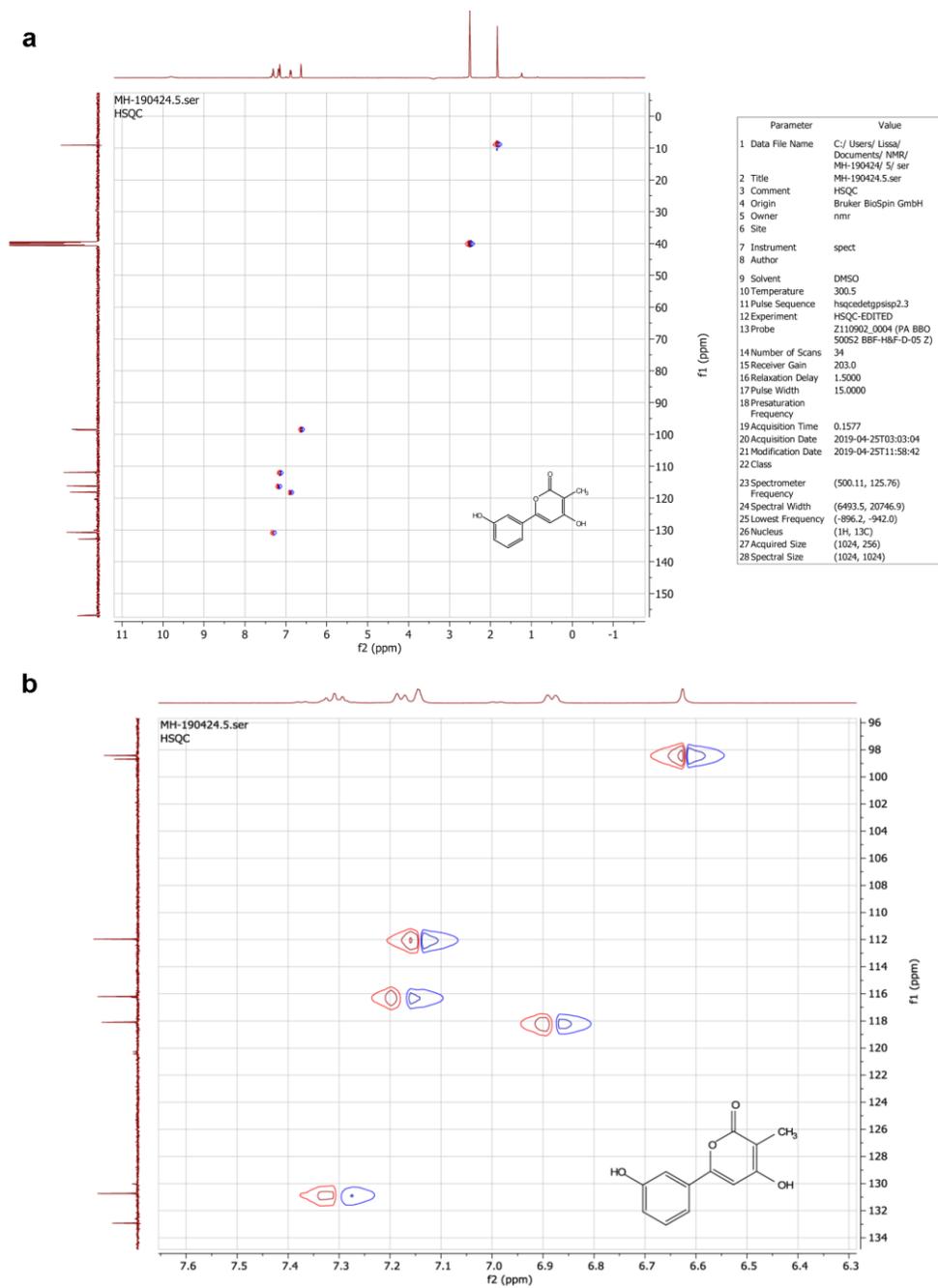
Supplementary Figure 25. ^1H NMR of deshydroxymethylvenemycin (**6**) in DMSO-d_6 .
 a) Full spectrum. b) Expanded region.

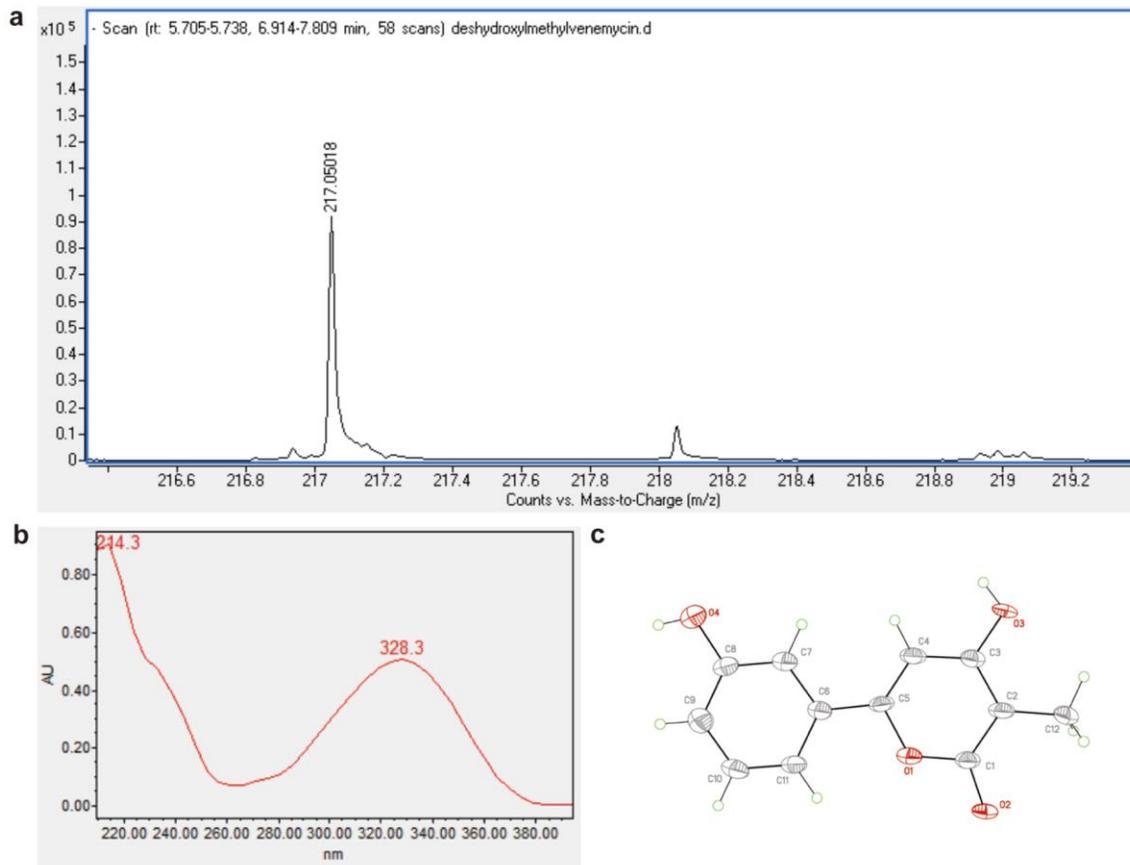


Supplementary Figure 26. ¹H NMR of deshydroxymethylvenemycin (6) in D₂O.

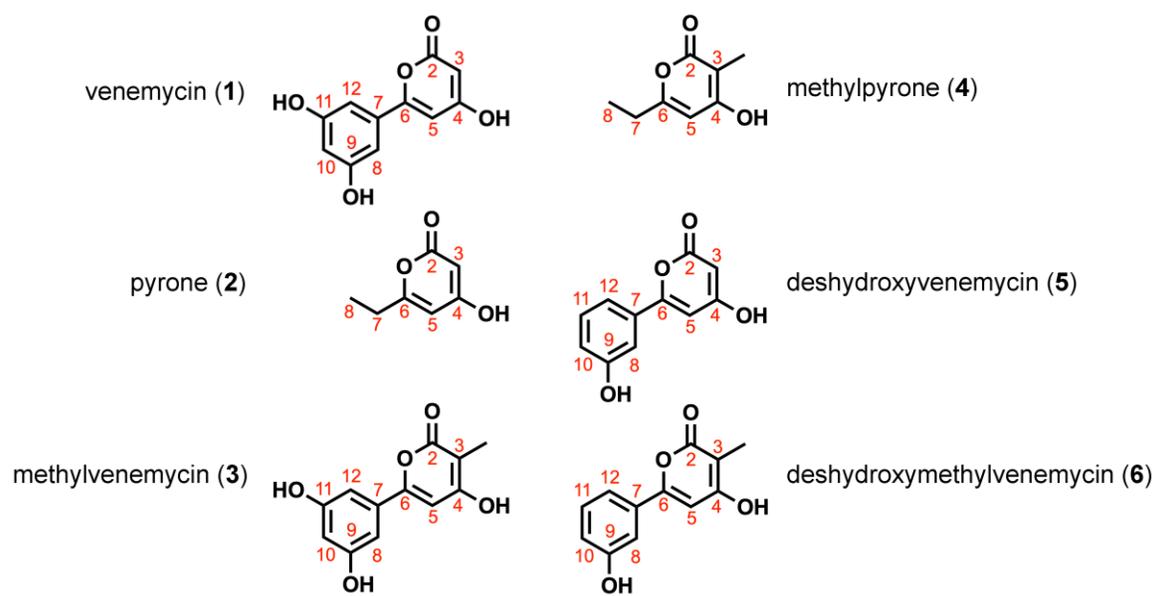


Supplementary Figure 27. ^{13}C NMR of deshydroxymethylvenemycin (6) in DMSO-d_6 .





Supplementary Figure 29. Additional characterization of deshydroxymethylvenemycin (6). **a**) High resolution MS. 217.0506 m/z calculated, 217.0502 m/z found. **b**) Absorbance spectrum. **c**) X-ray crystal structure.



Supplementary Figure 30. Numbering system for polyketide products.



Supplementary Figure 31. TLC of compounds produced by combinatorial biosynthesis. Left to right: venemycin (**1**), methylvenemycin (**3**), deshydroxyvenemycin (**5**), deshydroxymethylvenemycin (**6**), and pyrone (**2**). Run in EtOAc:hexanes:HCOOH, 66:33:1, viewed under 254 nm light.

Supplementary Table 1. Primers used in this study.

VemG F	tgccgcgcggcagccatgGTGACGCAAGTCGATGACATCC GGCCCCTA
VemG R	tggtggtggtggtgctcgagTCAGGGTCCCTTCAGTTCGTTGT CGATGA
VemH F	aagaaggagatatacatatgACGGGCACCGAGGAGAAGCTGG T
VemH R	tggtggtggtggtgctcgagCAGCAGTGACCGTGACCACTGCT CGA
PikAI updated F	tggtgccgcgcggcagccatATGTCTTCAGCCGGAATTACCAG GACCGGT
PikAI updated R	ggcggcagggcgggcccgcGACAACCACCGGGCCTCTTC GAGCA
PikAI traditional F	tggtgccgcgcggcagccatATGTCTTCAGCCGGAATTACCAG GACCGGT
PikAI traditional R	cggcggccggtgctcgacGACCCGGTCGTCGTCATCGGCA TGAGC
VemG AT-ACP- ^c DD F	gcgcgcgcgcctgcgcgc
VemG AT-ACP- ^c DD R	tggtggtggtggtgctcgagTCAGGGTCCCTTCAGTTCGTTGT CGATGA
VemG KS-AT-ACP- ^c DD F	gaccggtgctgctcagcCATGAGCT
VemG KS-AT-ACP- ^c DD R	tggtggtggtggtgctcgagTCAGGGTCCCTTCAGTTCGTTGT CGATGA
PikAIV updated F	gccccgagcgcggctcgggGTCGAGCCGCCGGCCGGTGGC GGCGT
PikAIV updated R	tggtggtggtggtgctcgagCTTGCCCGCCCCCTCGATGCCCT CGAT
PikAIV traditional F	ttcaagcggatccctcagcATTCTTTTCAACATCA
PikAIV traditional R	tggtggtggtggtgctcgagCTTGCCCGCCCCCTCGATGCCCT CGAT
VemH ^N DD-KS F	aagaaggagatatacatatgACGGGCACCGAGGAGAAGCTGGT
VemH ^N DD-KS R	gggctcctcgggtggggcc
VemH ^N DD F	aagaaggagatatacatatgACGGGCACCGAGGAGAAGCTG GT
VemH ^N DD R	cggctcgcgcagcgcgcCCCGCAACT
pET28b-His10 F	agccatcatcatcatcaTCATCATCACAGCAG
pET28b-His10 R	tgatgatgatgatggctGCTGCCCATGGTATATCTCC

Note: Lowercase indicates SLiCE overlapping sequence.

Supplementary References

- 1 Sheldrick, G. M. SHELXT. A program for crystal structure solution. *Acta Cryst* **A71**, 3-8 (2015).
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