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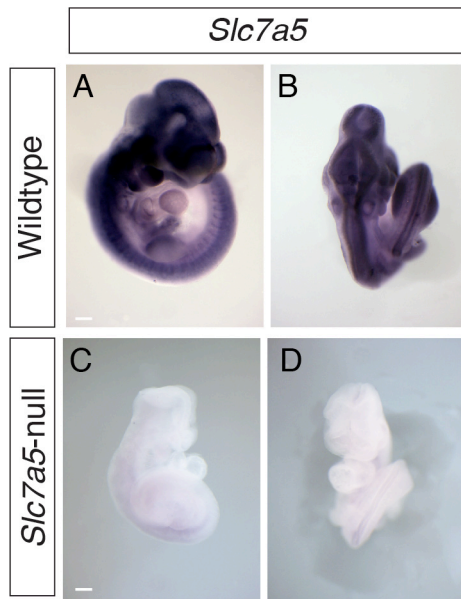
Appendix Figure S7 *Chac1* and *Trib3* transcripts are just detectable in E8.5 wildtype embryos

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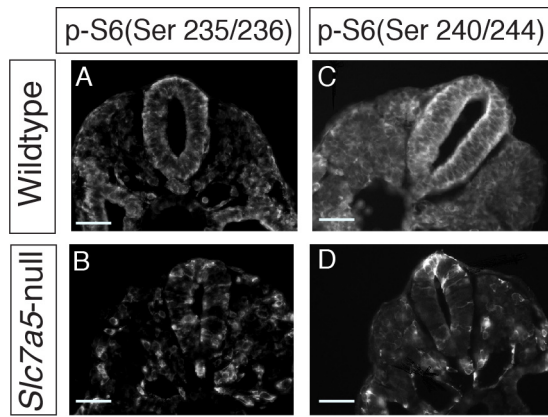
Appendix Table S1 Sequence of the primers used to generate in situ hybridization probes specific for the murine *Slc7a5*, *Slc3a2*, *Chac1* and *Trib3*

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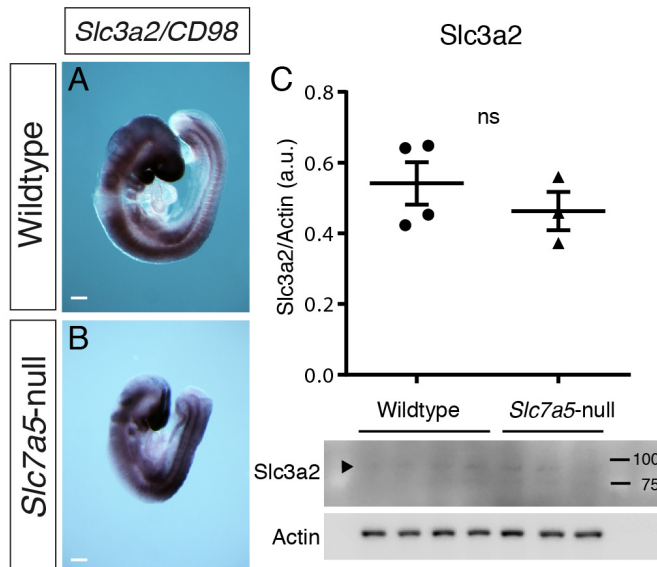
**Appendix Figure S1 Validation of specificity of *Slc7a5* mRNA probe**

In situ hybridisation for *Slc7a5* mRNA in (A, B) wildtype and (C, D) *Slc7a5*-null embryos. Two whole litters including n=3 mutant embryos were assessed for *Slc7a5* expression. All embryos in each litter were processed in parallel and the BCIP/NBT reaction stopped at the same time for all embryos: while strong expression of *Slc7a5* was detected in littermates, all *Slc7a5*-null embryos lacked transcripts, consistent with RNA-seq data comparing wildtype and *Slc7a5*-null embryos. Scale bar 200 μ m.



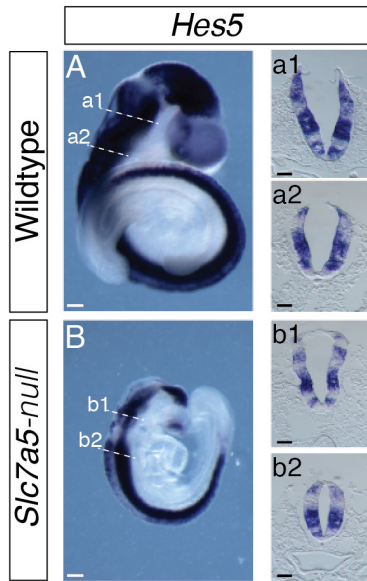
Appendix Figure S2 pS6 detection comparing antibodies against p-Ser 235/236 and pSer 240-244

The patterns of pS6 detection with an antibody recognising p-Ser 235/236 in (A) wildtype and (B) *Slc7a5*-null embryo neural tube (n=5, 18 sections and n=8, 24 sections, respectively, as in Figure 4) were found to be similar to those detected with an antibody recognising pSer 240-244 (a site specifically phosphorylated by mTORC1) in (C) wildtype and *Slc7a5*-null (D) embryos (n=1 embryo, 11 sections; n=2 embryos, 9 and 11 sections respectively). Note both antibodies reveal an aberrant /reduced pattern of pS6 in mutant embryos. Scale bar 50µm.



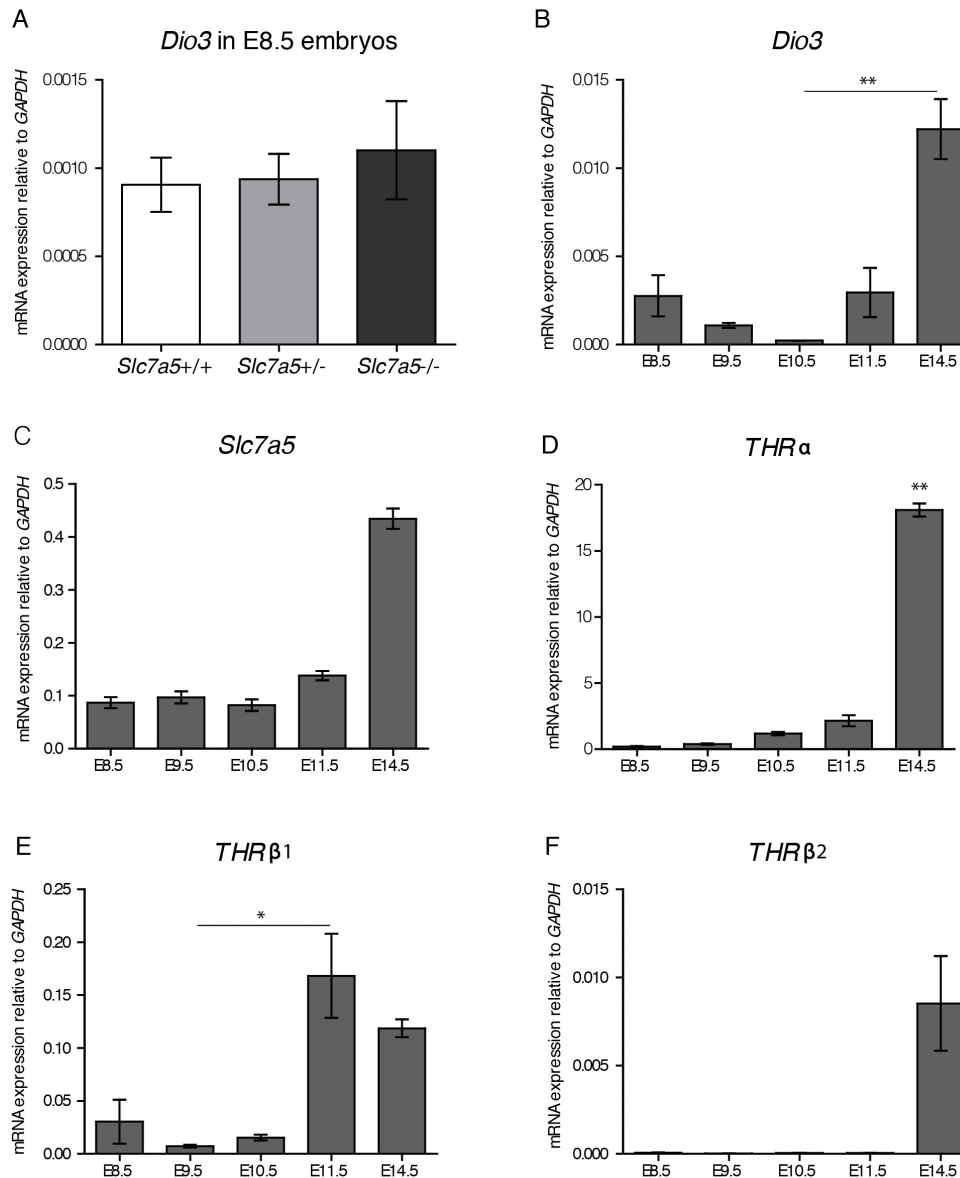
Appendix Figure S3 Transcript and protein levels of Slc7a5 partner Slc3a2/CD98 are not reduced in Slc7a5-null embryos

mRNA in situ hybridization was undertaken to determine whether expression of Slc7a5 partner *Slc3a2/CD98* is reduced in *Slc7a5*-null embryos. *Slc3a2* was cloned (sequence from NM_001161413.1 and probe generated for region 920-1922 - 3' end of coding sequence). *Slc3a2* transcripts were assessed in *Slc7a5*-null (n=3) and heterozygote (n=2) and wildtype (n=2) embryos at E9.5, when the phenotype is apparent. This revealed strong *Slc3a2* expression in all embryos, including (A) wildtype and (B) *Slc7a5*-null embryos, intriguingly the *Slc3a2* pattern of expression is similar to that of *Slc7a5*, Scale bars 200 μ m. (C) Western blotting was used to evaluate levels of Slc3a2 protein in wildtype (n=4) and *Slc7a5*-null (n=3) embryos. Each lane contains lysate from one embryo and expression in each embryo is represented by a dot or triangle, actin is the loading control and error bars indicate SEM.



Appendix Figure S4 Notch signalling is unaffected in *Slc7a5*-null embryos

Expression of notch signalling target *Hes5* was assessed by mRNA in situ hybridization in wildtype littermate (A) and *Slc7a5*-null (B) E9.5 embryos (n=2 each). Panels (a1 – b2) show transverse sections of the dorsal neural tube at the level of the hindbrain (a1, b1) or the spinal cord (a2, b2). Scale bars 200 μm , except for sections 50 μm .

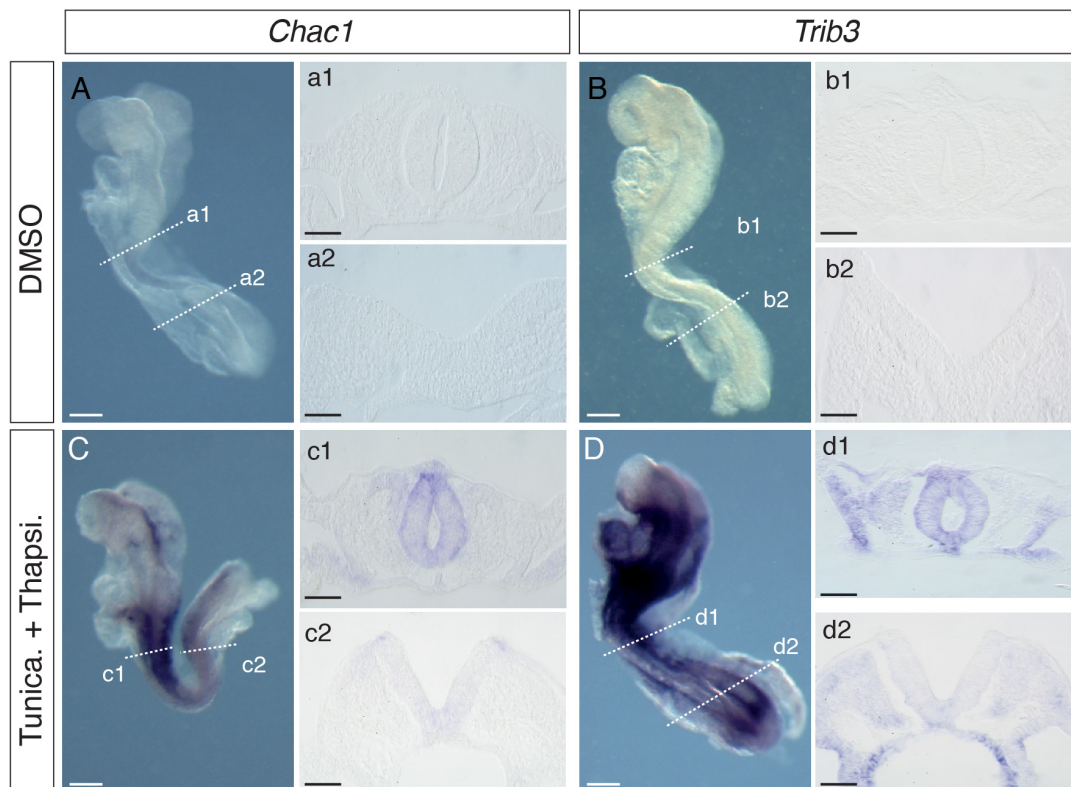


Appendix Figure S5 Expression of genes involved in the thyroid hormone response were unchanged in *Slc7a5*-null embryos and thyroid hormone receptor mRNA levels rise after E11.5 in wildtype embryos

The expression of enzyme type III iodothyronine deiodinase (*Dio3*) is T3 responsive (Bianco and Kim 2006) but was not significantly changed in RNAseq data. (A) This was confirmed using qPCR on wildtype and *Slc7a5*-deficient E8.5 embryos (n=5).

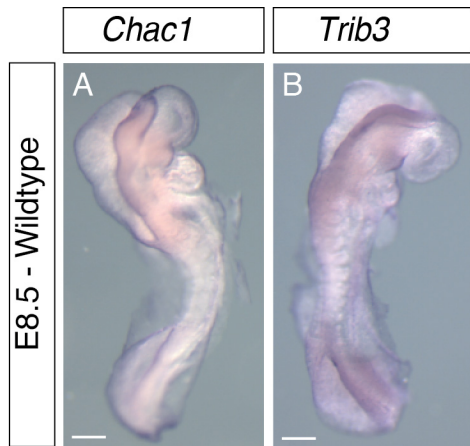
(B-F) qPCR analysis was performed on wildtype embryos at various developmental stages (n=3, except for E9.5 (n=4) using primers specific for (B) *Dio3*, (C) *Slc7a5* or (D, E, F) thyroid hormone receptors *THR*.

Data information: Error bars indicate SEM. (B-F) A one-way Anova test was performed for statistical analysis *p<0.05, **p<0.01. Actual p values for all comparisons are in original source data.



Appendix Figure S6 Exposure to known ER stressors Tunicamycin and Thapsigargin induces localised expression of *Trib3* and *Chac1*

Tunicamycin and Thapsigargin act respectively by inhibition of protein glycosylation or reduction of activity of calcium-dependent ER chaperones (Kuo & Lampen, 1974, Thastrup, Cullen et al., 1990). Exposure to either of these drugs leads to accumulation of unfolded proteins in the ER, which provokes ER stress (Osowski & Urano 2011). Whole E8.5 embryos were suspended in hanging drop culture for 6 hours in the presence of control carrier DMSO or Tunicamycin and Thapsigargin. All embryos were harvested and processed in parallel for mRNA in situ hybridization (n=3 embryos in each condition and for each gene assessed). *Chac1* and *Trib3* transcripts were undetectable in control DMSO-only treated embryos (Figures A,B), but strongly upregulated following Tunicamycin and Thapsigargin treatment, particularly in the forming neural tube (Figures C-c2, D-d2). These data directly link increased expression of *Chac1* and *Trib3* to an ER stress response in the developing mouse embryo and show that at this early stage the developing neural tube is particularly vulnerable to such stress. Scale bars 200 μm , except for sections 100 μm .



Appendix Figure S7 *Chac1* and *Trib3* transcripts are just detectable in E8.5 wildtype embryos

(A) *Chac1* and (B) *Trib3* expression pattern in wildtype E8.5 embryos after >6 days in situ hybridisation reaction (n=4 for each gene). Scale bars 200 μ m.

Appendix Table S1 Sequence of the primers used to generate in situ hybridization probes specific for the murine *Slc7a5*, *Slc3a2*, *Chac1* and *Trib3*

Gene name	FASTA sequence	Forward primer	Reverse primer	product length
<i>Slc7a5/Lat1</i>	NM_011404.3	GCAATATCACGCTGCTCAAC	CCAAGTGGTAGTCCCGAAG	831
<i>Chac1</i>	NM_026929.4	CAAGCCCTGTGGATTTTCGG	GCGGGGCATAAGACACAAAG	716
<i>Trib3</i>	NM_175093.2	CTTTAGCAGCGGAAGAGGCT	CTGAAGACAAAGCGACGCAG	714
<i>Slc3a2</i>	NM_001161413.1	GGGCTCCCAGGAAGATTTTA	TCAGGTTTTCCAGCTTCAGG	1002

Appendix Table S2 Sequence of the primers used for qPCR

Gene name	FASTA sequence	Forward primer	Reverse primer	product length
<i>Aldh1L2</i>	NM_153543.2	TAACACCTACAACAAGACAGAC	GTATTCATTCAGGGCCTCCTCA	96
<i>ATF4</i>	NM_009716.3	AGCAAAACAAGACAGCAGCC	ACTCTCTTCTTCCCCCTTGC	193
<i>β-Actin</i>	NM_007393.3	ATGCTCCCCGGGCTGTAT	CATAGGAGTCCTTCTGACCCATTC	87
<i>CD98</i>	NM_008577.4	GAGGACAGGCTTTTGATTGC	ATTCAGTACGCTCCCCAGTG	136
<i>Chac1</i>	NM_026929.4	AGTGTGGAAGCCGGACTTTG	CACTCGGCCAGGCATCTTGT	121
<i>CHOP</i>	NM_007837.4	CCACCACACCTGAAAGCAGAA	GGTGCCCCAATTCATCT	150
<i>Dio3</i>	NM_172119.2	TCAGACGACAACCGTCTGTG	AAAATTGAGCACCAACGGGC	189
<i>Fanca</i>	NM_016925.3	CGGGCAGAGTCAAAAAGCAA	AGCAGAGCGGATGAAGGAAG	212
<i>GAPDH</i>	NM_001289726.1	TGTGTCCGTCGTGGATCTGA	CCTGCTCACCACCTTCTTGA	77
<i>Klhdc4</i>	NM_145605.2	CGGAAGGAGGAGGAAGACCT	TCACCTCCGAAAAGGATCAGC	164
<i>Lefty2</i>	NM_177099.3	GGACCTGGAGCGCACAC	GGGTCACAATTGCCTTGAGC	133
<i>Nodal</i>	NM_013611.4	CCATGCCTACATCCAGAGCCTGC	TGGTGTTCAGGAGGACCCTGCC	132
<i>Pck2</i>	NM_028994.2	CGGCTGGAGTTCGAGACTTT	GGGCCAGCCAGCAGTTCTTA	165
<i>Slc7a5</i>	NM_011404.3	CTGGTCTTCGCCACCTACTT	GCCTTTACGCTGTAGCAGTTC	128
<i>Spire2</i>	NM_172287.2	TGGAGCCCGAGCCTACAAC	CTCTTCTGGACCCACGTAGC	227
<i>THRα</i>	NM_178060.3	CCTGGACAAAGACGAGCAGT	GCACTGATTCCGGGTGATCT	184
<i>THRβ1</i>	NM_001113417.1	GGACAAGCACCCATCGTGAAT	CTCTGGTAATTGCTGGTGTGAT	97
<i>THRβ2</i>	NM_009380.3	CCTGTAGTTACCTGGAAACCTG	GGCTTTGTCCCCACACACTA	200
<i>Trib3</i>	NM_175093.2	TCTCCTCCGCAAGGAACCT	TCTCAACCAGGGATGCAAGAG	67