

1 **Additional File 1 for**

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3 **Mobile resistome of human gut and pathogen drives anthropogenic bloom of antibiotic**
4 **resistance**

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12 **This file includes:**

13 **Figures S1.** Correlations of ARG density with microbiological, genetic, physicochemical,
14 geographic, and socio-economic parameters.

15 **Figure S2.** Proportions of losses and gains of ARGs and OTUs between river samples
16 according to geographic distance.

17 **Figure S3.** Relative abundances of mobile genetic elements (MGEs) in the metagenome
18 contigs of Han River.

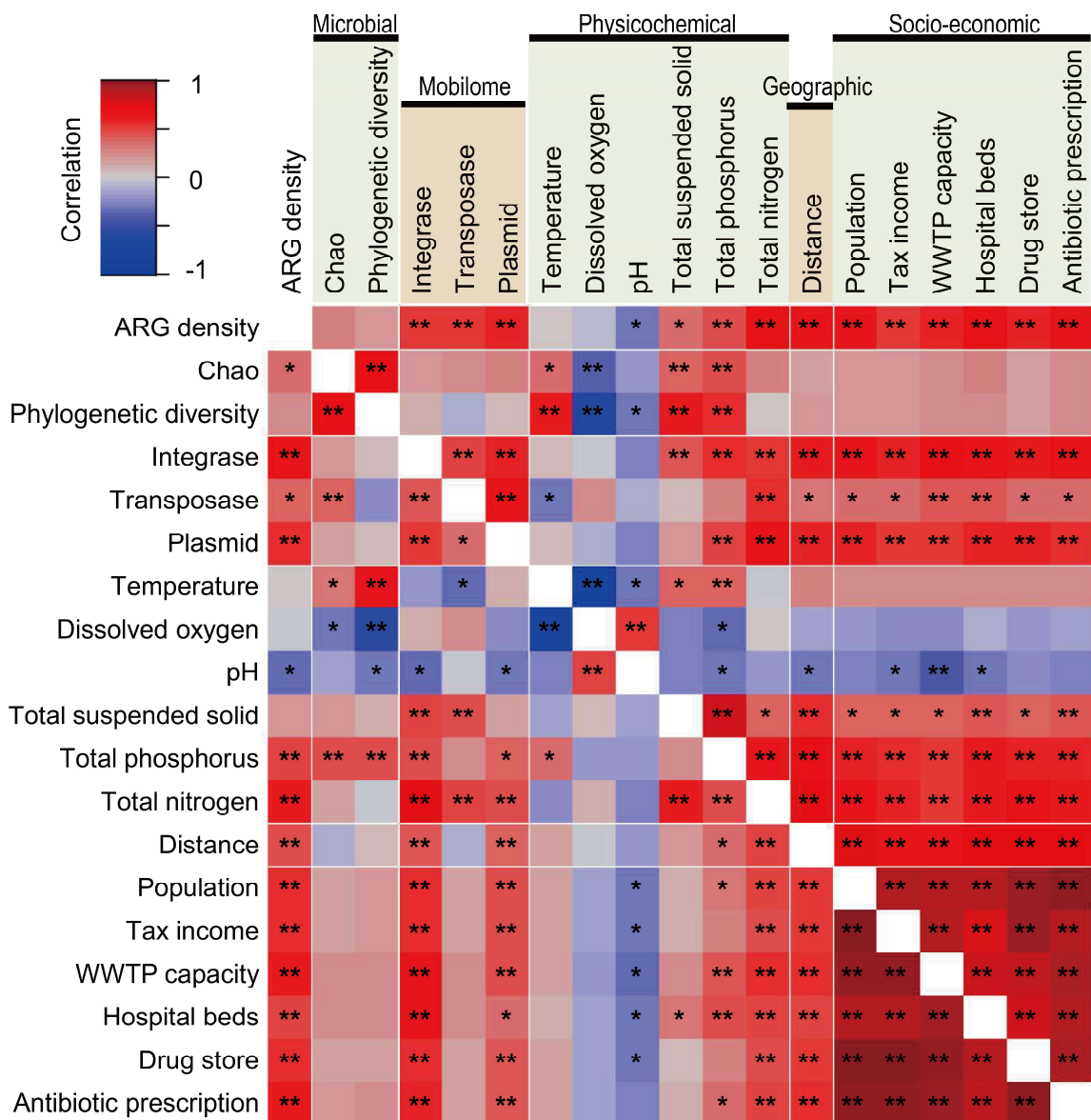
19 **Figure S4.** Co-occurrence of ARGs and MGEs in metagenome contigs.

20 **Figure S5.** Horizontal gene transfer networks of ARGs inferred from pathogen genomes and
21 river metagenomes.

22 **Figure S6.** Comparison of ARG compositions between river and human gut metagenomes.

23 **Table S1.** Sample metadata.

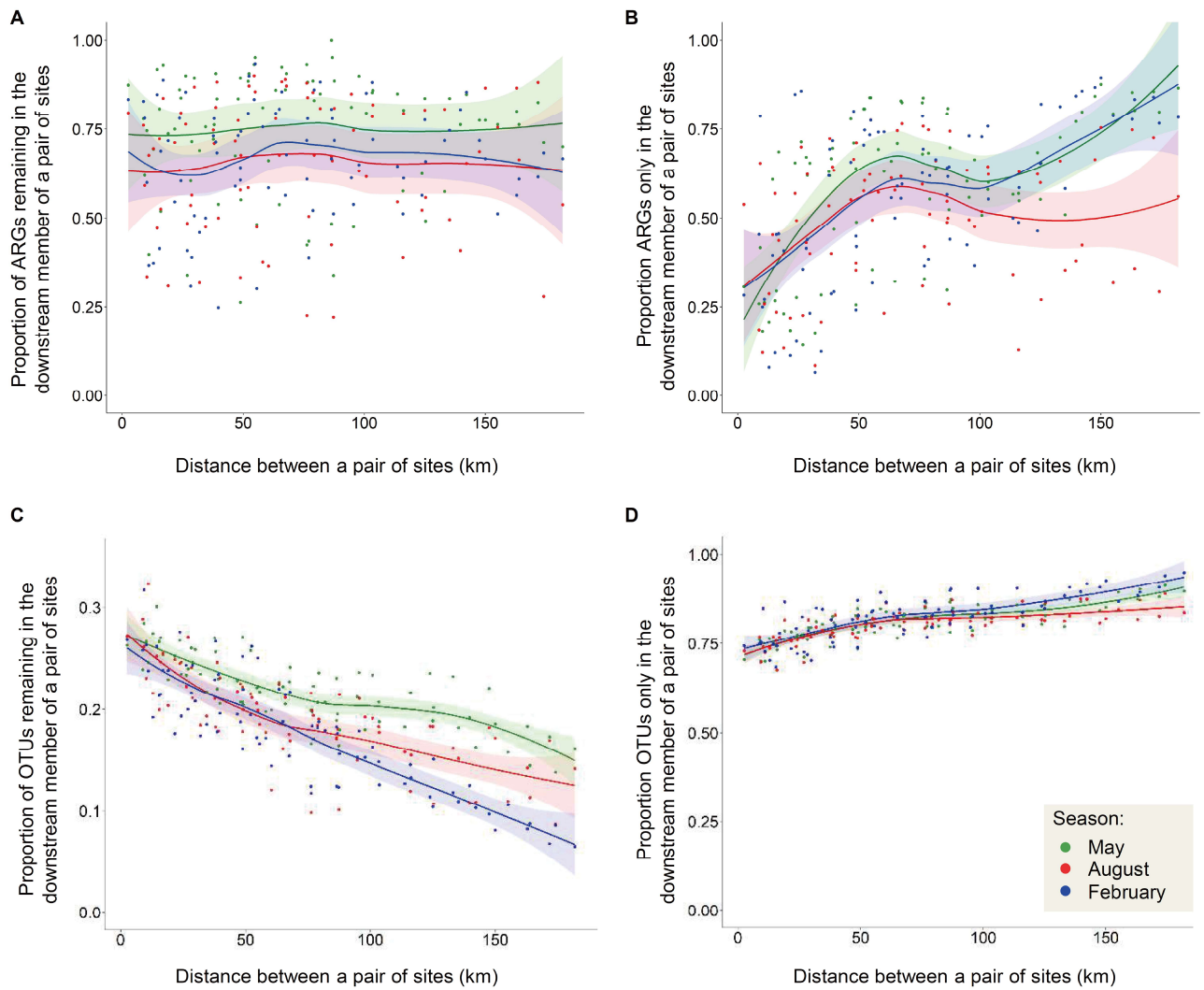
24 **Table S2.** Representative fecal OTUs selected from the OTU table of the Earth Microbiome
25 Project.



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27 **Figure S1.** Correlations of ARG density with microbiological, genetic, physicochemical,
 28 geographic, and socio-economic parameters. Pairwise Pearson's and Spearman's correlations
 29 were assessed for various sample parameters. Values measured from 45 samples from three
 30 different seasons were used as input for correlation analyses. Pearson's correlation and
 31 Spearman's rho values are shown as color gradients in lower and upper triangles, respectively.
 32 Significance is indicated by * ($P < 0.05$) and ** ($P < 0.01$). Distance from H7 was scaled from
 33 0 (S1) to 1 (H7). WWTP, wastewater treatment plant.

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35

36 **Figure S2.** Proportions of losses and gains of ARGs and OTUs between river samples

37 according to geographic distance. **a** Distance-decay curve of the proportion of ARGs (clustered

38 at 95% identity) remaining in the downstream member of a pair of sites for comparison. **b**

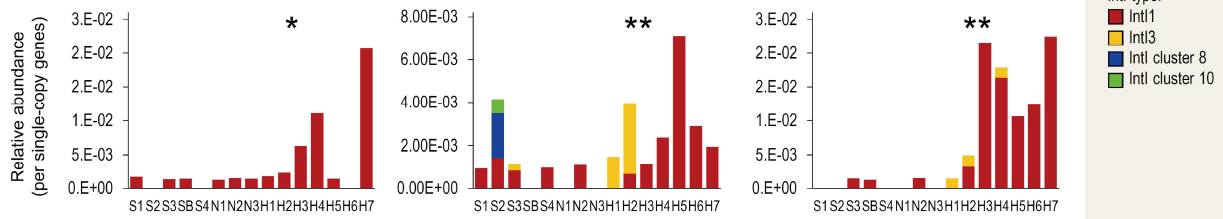
39 Proportion of ARGs only in the downstream member of a pair of sites for comparison. **c**

40 Distance-decay curve of the proportion of OTUs remaining in the downstream member of a

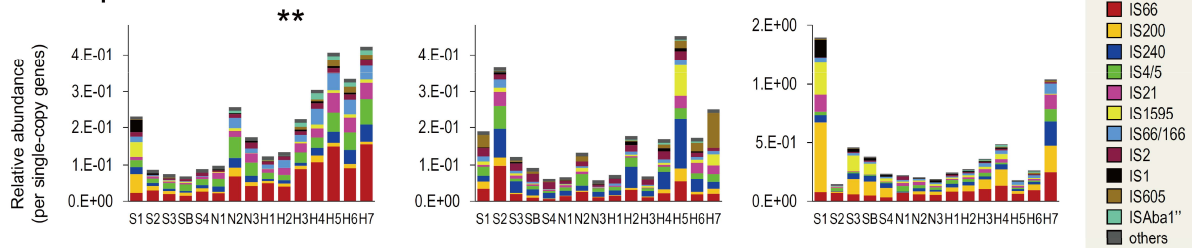
41 pair of sites for comparison. **d** Proportion of OTUs only in the downstream member of a pair

42 of sites for comparison.

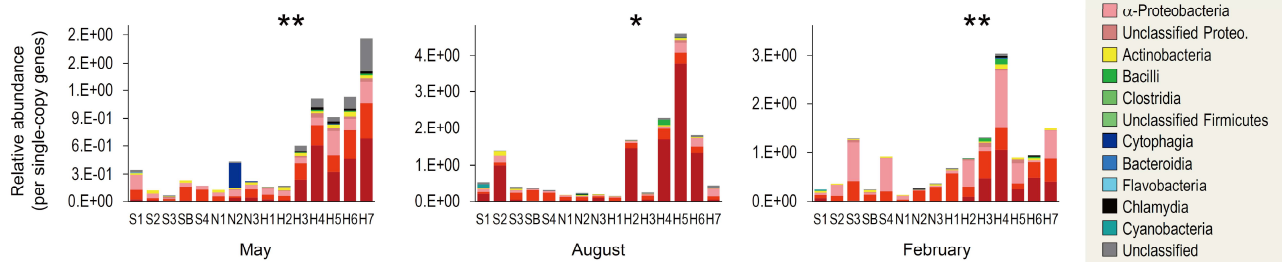
A Integrases



B Transposases



C Plasmids



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44 **Figure S3.** Relative abundances of mobile genetic elements (MGEs) in the metagenome

45 contigs of Han River. **a** The relative abundances of integrase genes (*intI*) in the

46 metagenome contigs. **b** The relative abundances of transposase genes in the metagenome

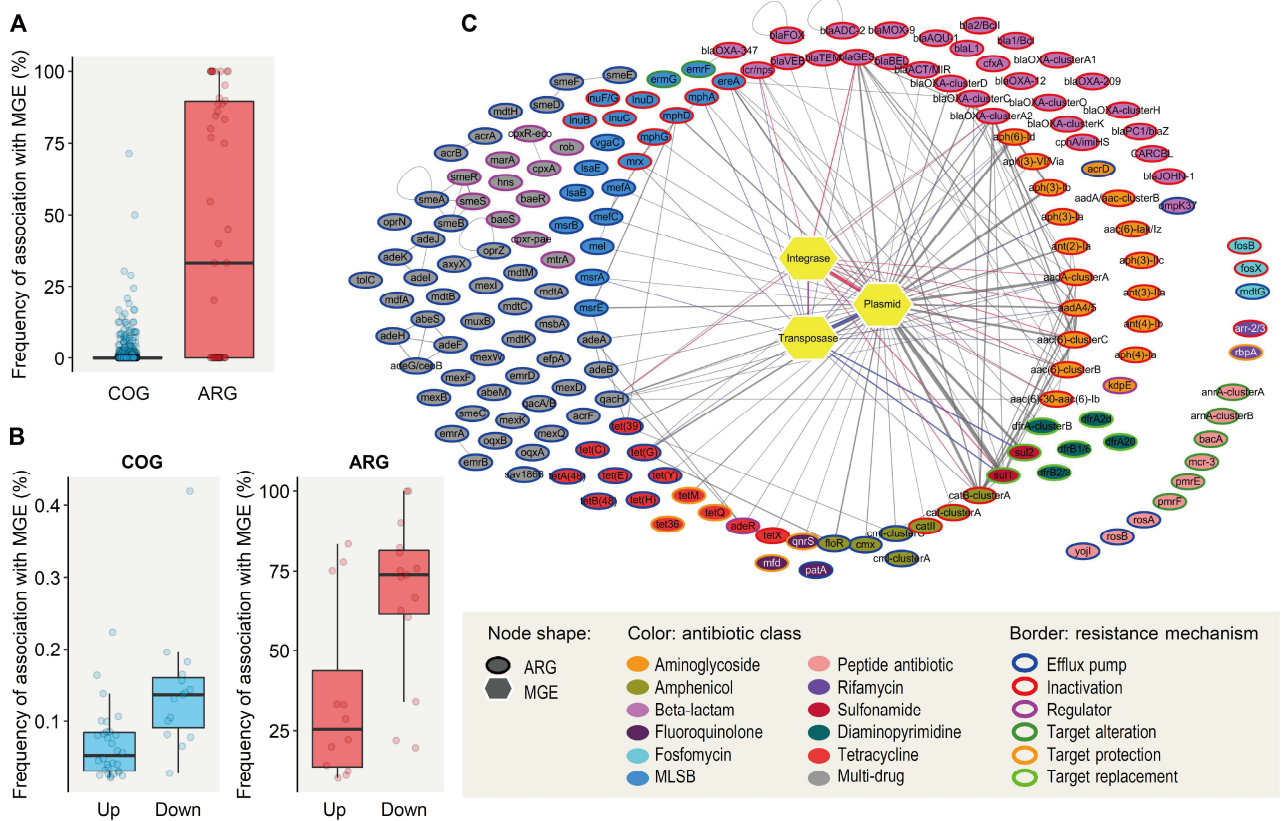
47 contigs. **c** The relative abundances of plasmids determined by the abundances of all genes in

48 plasmid-like contigs. For all three MGEs, relative abundance was normalized by the average

49 abundance of universal single-copy genes. The significance level of the difference between

50 upstream and downstream samples is shown above each plot (* $P < 0.05$, ** $P < 0.01$, t test).

51 For *intI* data, t test was performed for the abundance of *intI1* rather than the total *intI* abundance.



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53 **Figure S4.** Co-occurrence of ARGs and MGEs in metagenome contigs. **a** Boxplot of frequency

54 at which each ARG or COG was detected with MGEs (integrase, transposase, and plasmid) in

55 the same metagenome contig. The frequency was calculated only for gene families (either

56 ARGs or COGs) detected in five or more contigs. **b** Co-occurrence frequencies of ARGs and

57 MGEs (right) and COGs and MGEs (left) in upstream (Up) and downstream (Down) river

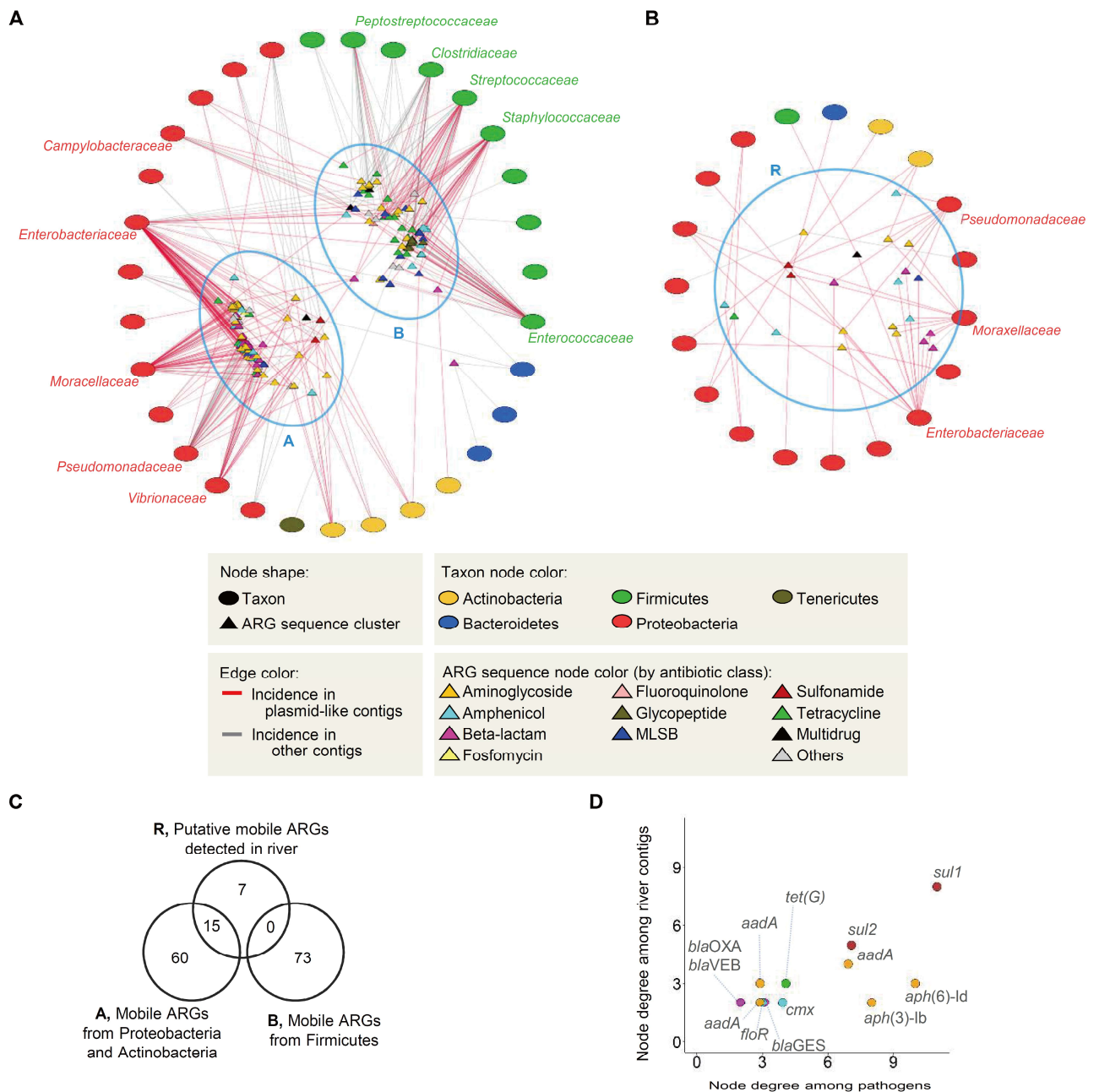
58 samples. **c** Network of co-occurrence of ARGs and MGEs on the same contig. Hexagonal

59 nodes represent three types of MGEs. Oval nodes represent ARGs. Edges represent the co-

60 occurrence of two nodes (ARGs or MGEs) in the same contig; the edge width is proportional

61 to the number of cases. Edges involving integrase are in red. Edges involving transposase are

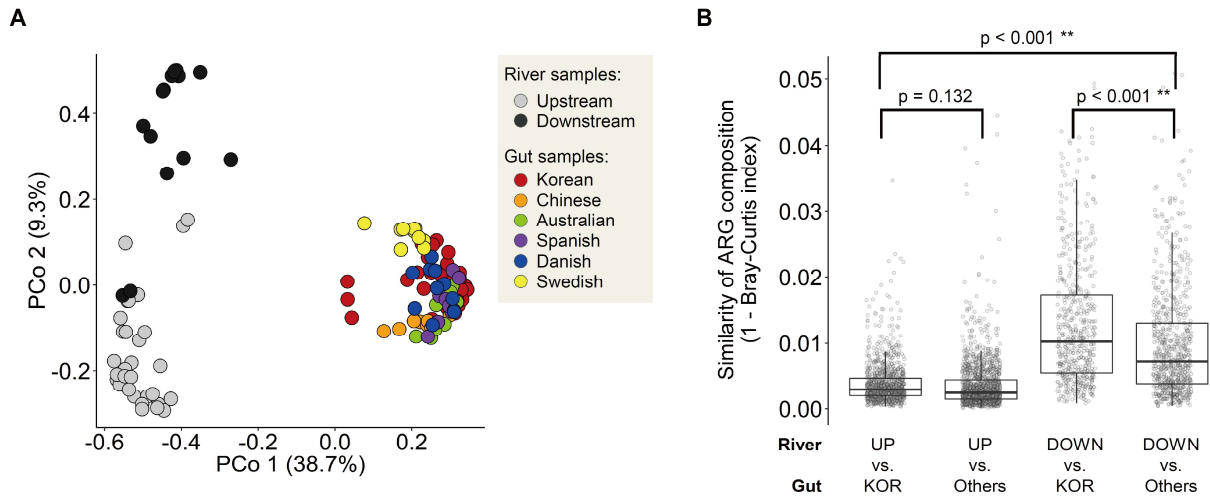
62 in blue. Plasmid edges are in gray.



63

64 **Figure S5.** Horizontal gene transfer networks of ARGs inferred from pathogen genomes and
 65 river metagenomes. **a** Network of bacterial pathogen genomes and ARGs. Taxon nodes
 66 represent pathogen genomes grouped at the family level. ARG nodes represent clusters of ARG
 67 sequences that share $\geq 99\%$ nucleotide identity. Edges indicate detection of ARGs in the
 68 genomes included in the taxon nodes. The occurrence of the same ARG across different taxon
 69 nodes was considered evidence of horizontal transfer of ARGs. Two blue circles encompassing
 70 each set of ARG nodes were designated as “set A (mobile ARGs from Proteobacteria and

71 Actinobacteria)” and “set B (mobile ARGs from Firmicutes)”. **b** Network of river metagenome
72 contigs and ARGs. Taxon nodes represent metagenome contigs assigned at the family level.
73 Edges indicate detection of ARGs in the metagenome contigs. A blue circle surrounding ARG
74 nodes was designated as “set R (mobile ARGs detected in river metagenomes)”. **c** Venn
75 diagram displaying the numbers of ARG nodes shared between the HGT networks of pathogen
76 genomes and river metagenomes. **d** Comparison of node degrees from the HGT networks of
77 pathogen genomes and river metagenomes. Node degree indicates the number of taxon nodes
78 connected to ARGs in each network.



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80 **Figure S6.** Comparison of ARG compositions between river and human gut metagenomes. **a**

81 Principal coordinate analysis of ARG compositions of the Han River and the human gut

82 metagenomes from several countries. **b** Bray–Curtis similarity of ARG compositions between

83 the river and gut metagenomes: upstream river vs. Korean gut, upstream river vs. non-Korean

84 gut, downstream river vs. Korean gut, and downstream river vs. non-Korean gut. Results with

85 P-values less than 0.01 (**) were considered statistically significant.

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87 **Table S1.** Sample metadata.

Site name	GPS position	Sampling time	Physicochemical parameters					
			Temperature (°C)	pH	Dissolved oxygen (mg/L)	Total nitrogen (mg/L)	Total phosphate (mg/L)	Total suspended solids (mg/L)
S1	37°26'0.51" 128°39'27.18"	May 2016	18.5	7.7	9.7	1.9	0.025	4.8
		Aug. 2016	16.5	7.9	6.4	2.3	0.086	29.0
		Feb. 2017	4.4	8.2	13.2	2.9	0.006	1.0
S2	37°0'22.96" 127°59'2.4"	May 2016	11.4	7.7	11.5	2.3	0.008	1.4
		Aug. 2016	18.7	7.5	3.3	2.8	0.016	1.2
		Feb. 2017	4.8	8.3	10.7	1.9	0.006	0.7
S3	37°17'23.51" 127°39'5.92"	May 2016	17.2	8.0	11.1	2.2	0.027	7.7
		Aug. 2016	20.9	8.2	9.1	2.5	0.011	1.4
		Feb. 2017	3.3	8.3	13.8	2.5	0.026	0.9
SB	37°23'46.06" 127°32'40.09"	May 2016	18.2	8.0	11.9	2.4	0.045	13.5
		Aug. 2016	21.9	8.0	8.1	2.3	0.013	2.8
		Feb. 2017	3.6	8.5	12.5	2.8	0.012	1.7
S4	37°29'1.61" 127°29'28.48"	May 2016	20.6	8.5	12.5	2.3	0.034	7.2
		Aug. 2016	22.2	7.8	7.5	2.3	0.013	1.4
		Feb. 2017	2.6	8.2	14.6	2.9	0.010	1.3
N1	38°5'14.95" 127°45'41.24"	May 2016	13.2	7.9	12.1	1.3	0.011	1.9
		Aug. 2016	20.3	7.7	6.5	1.3	0.006	1.7
		Feb. 2017	3.9	7.8	12.0	0.9	0.008	1.0
N2	37°50'5.35" 127°40'41.1"	May 2016	17.1	7.8	13.3	1.4	0.021	5.4
		Aug. 2016	19.7	7.2	7.5	1.7	0.013	4.7
		Feb. 2017	3.7	8.0	12.8	1.6	0.006	1.2
N3	37°40'28.6" 127°23'2.86"	May 2016	16.5	6.9	9.9	1.9	0.013	3.4
		Aug. 2016	24.5	7.2	5.8	1.6	0.008	2.2
		Feb. 2017	3.1	8.0	13.8	1.9	0.007	1.2
H1	37°32'29.75" 127°14'35.2"	May 2016	17.6	8.1	10.1	2.1	0.029	7.2
		Aug. 2016	26.1	7.8	6.0	1.8	0.019	4.1
		Feb. 2017	3.3	8.2	14.5	2.5	0.008	2.9
H2	37°34'31.77" 127°9'38.17"	May 2016	19.1	8.3	11.2	3.2	0.076	4.0
		Aug. 2016	26.2	7.9	7.0	1.9	0.017	2.1
		Feb. 2017	3.5	8.3	15.1	2.6	0.009	2.5
H3	37°32'2.06" 127°2'15.63"	May 2016	21.2	8.1	9.6	2.4	0.038	4.8
		Aug. 2016	25.5	7.6	6.4	2.8	0.038	2.9
		Feb. 2017	3.8	7.4	12.1	4.1	0.021	2.4
H4	37°31'31.66" 127°1'0.82"	May 2016	20.8	7.8	9.4	2.5	0.039	6.4
		Aug. 2016	25.5	7.4	5.3	3.0	0.053	4.0
		Feb. 2017	4.5	7.3	10.6	5.7	0.039	2.5
H5	37°33'0.27" 126°53'46.28"	May 2016	21.3	7.9	9.6	3.3	0.083	6.0
		Aug. 2016	25.6	7.5	5.6	3.5	0.109	11.6
		Feb. 2017	3.2	7.6	12.4	5.8	0.042	2.8
H6	37°35'44" 126°49'1.46"	May 2016	20.3	7.3	9.0	4.7	0.108	24.2
		Aug. 2016	25.3	7.7	7.3	4.2	0.127	46.0
		Feb. 2017	3.6	7.7	13.1	7.5	0.046	40.8
H7	37°39'6.73" 126°43'17.93"	May 2016	23.8	7.6	9.9	4.5	0.352	20.0
		Aug. 2016	24.3	7.7	6.2	4.2	0.190	76.3
		Feb. 2017	3.9	7.5	11.9	8.5	0.097	499.0

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90 **Table S2.** Representative fecal OTUs selected from the OTU table of the Earth Microbiome
 91 Project.

Rank	OTU	Median abundance in human distal gut samples (%)	Frequency in human distal gut samples (%)	Frequency in non-human samples (%)	Taxonomy (phylum; class; order; family; genus)	Median abundance of summed OTUs from top to the current rank in human distal gut (%)
1	otu_124280	3.4	94.2	17.1	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Peptostreptococcaceae</i> ; -	3.2
2	otu_88633	3.2	100.0	5.3	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Blautia</i>	9.0
3	otu_190063	2.8	91.9	3.5	Actinobacteria; <i>Actinobacteria</i> ; <i>Bifidobacteriales</i> ; <i>Bifidobacteriaceae</i> ; <i>Bifidobacterium</i>	12.9
4	otu_139096	2.4	98.8	3.8	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Blautia</i>	16.3
5	otu_101273	2.1	93.0	2.4	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Coprococcus</i>	18.4
6	otu_157280	2.0	90.7	5.2	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; -	21.9
7	otu_55941	1.5	96.5	2.8	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Ruminococcaceae</i> ; -	22.9
8	otu_190980	1.4	93.0	3.1	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; -	26.3
9	otu_196163	1.3	98.8	3.1	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Coprococcus</i>	26.7
10	otu_114748	1.0	89.5	2.8	Firmicutes; <i>Erysipelotrichi</i> ; <i>Erysipelotrichales</i> ; <i>Erysipelotrichaceae</i> ; -	28.8
11	otu_106711	0.8	95.3	3.7	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Ruminococcaceae</i> ; -	30.0
12	otu_163574	0.8	90.7	1.4	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Peptostreptococcaceae</i> ; -	31.8
13	otu_190162	0.7	86.0	1.5	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Dorea</i>	32.2
14	otu_86215	0.7	93.0	9.3	Firmicutes; <i>Bacilli</i> ; <i>Lactobacillales</i> ; <i>Streptococcaceae</i> ; <i>Streptococcus</i>	34.4
15	otu_1911	0.6	74.4	1.8	Actinobacteria; <i>Coriobacteriia</i> ; <i>Coriobacteriales</i> ; <i>Coriobacteriaceae</i> ; <i>Collinsella</i>	35.5
16	otu_38125	0.6	90.7	5.2	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Ruminococcaceae</i> ; <i>Faecalibacterium</i>	40.2
17	otu_138262	0.5	96.5	2.3	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; -	42.7
18	otu_156190	0.5	97.7	2.5	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Blautia</i>	43.2
19	otu_85022	0.5	87.2	4.3	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Clostridiaceae</i> ; -	44.5
20	otu_103043	0.4	80.2	1.9	Bacteroidetes; <i>Bacteroidia</i> ; <i>Bacteroidales</i> ; <i>Rikenellaceae</i> ; -	45.3
21	otu_69769	0.3	86.0	2.2	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Ruminococcus</i>	47.7
22	otu_95259	0.3	95.3	1.2	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Dorea</i>	48.2
23	otu_152968	0.3	86.0	1.2	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Blautia</i>	48.3
24	otu_148432	0.2	76.7	1.7	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Clostridiaceae</i> ; -	49.1
25	otu_48195	0.2	81.4	1.4	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Lachnospiraceae</i> ; <i>Coprococcus</i>	49.4
26	otu_2274	0.2	79.1	1.6	Firmicutes; <i>Clostridia</i> ; <i>Clostridiales</i> ; <i>Clostridiaceae</i> ; <i>Clostridium</i>	49.4
27	otu_83279	0.1	69.8	2.4	Bacteroidetes; <i>Bacteroidia</i> ; <i>Bacteroidales</i> ; <i>Bacteroidaceae</i> ; <i>Bacteroides</i>	51.5

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