

SUPPLEMENTAL INFORMATION

Figure S1 Optimizing dissociation and primary analysis metrics *Related to Figure 1*

A-B, Plot of qPCR results shown as dCt ([Ct_Gapdh]-[Ct_gene-of-interest], *Gapdh* set to zero) comparing cDNA yields for representative cell type enriched gene markers and stress response genes (*Egr1*, *Fos*) from cold active protease (CAP) dissociation varying temperature (A) and time (B) of dissociation. ‘Undissociated’ and ‘No CAP’ samples were directly extracted adult kidney samples. **C**, Sequencing metrics reported by Cell Ranger for all 12 primary tissue samples (4 kidneys x 3 zones). **D**, Primary analysis dataset graphically represented as violin plots of all 30 clusters (ordered as in Figure 1C) showing per cluster levels for nGenes, nUMI and percent mitochondrial genes. Notably, distal tubule cells (clusters 6,12), have high percent mitochondrial genes while showing typical RNA complexity (nUMI), suggesting high metabolic rates in good quality cells. **E**, tSNE and stacked barplots of the primary dataset (as in Figure 1B-D), illustrating the distribution (left) and composition (right) of the clusters with respect to the four replicates. F: female, M: male, UE: ureteric epithelium, Int: interstitial.

Figure S2 Nephron dataset metrics and sex bias validation for the proximal tubule *Related to Figure 2*

A, N dataset as violin plots of the 19 clusters in Figure 2B, showing per cluster levels for nGenes (top), nUMI (middle) and percent mitochondrial genes (bottom). **B**, tSNE and stacked barplots of N dataset illustrating the distribution (left) and composition (right) of the clusters by replicate. **C**, Expression of sex biased, PT S3+S2 enriched markers *Ascm3* (m) and *Prlr* (f) in adult kidney tissue by RNAscope *in situ* and antibody labelling; inset scale bar 20µm. **D**, Expression of segment restricted, sex biased PT S2 markers are non-overlapping and demarcate the boundary between PT S2 and S3 segments. Scales and probes indicated on each panel. **E-F**, Gene Ontology analyses showing the top five GO Terms returned for genes showing a sex specific enrichment in female or male S3 (E) or throughout all female and male PT segments (F. RC: renal corpuscle, f-PT: female proximal tubule, m-PT: male proximal tubule, tl-LoH, Loop of Henle-thin limbs, DT: distal tubule.

Figure S3 Localization of thin Loop of Henle markers in early vs late forming nephrons *Related to Figure 3*

A-B, Low power views of thin limbs of Loop of Henle (tl-LoH) markers in tdT-labelled adult kidney sections; boxes indicate region of inset below each panel. Boxes also show where high power images in Figure 3E, F were collected relative to medullary depth shown at left; text above field details genetic combinations and tamoxifen (TX) injections; scale bar and probes indicated on each panel; arrowheads: a1, *Fst*⁺/*Apq1*⁺/tdT⁺; a2, *Slc39a8*⁺/*Apq1*⁺/tdT⁺; a3, *Sptssb*⁺/*Clnka*⁻/tdT⁺; a4,

Sptssb⁺/*Clcnka*⁺/tdT⁺; a5, *Crfl1*⁺/*Clcnka*⁺/tdT⁺; a6, *Slc14a2*⁺/*Apq1*⁻/tdT⁻; b1-2, *Slc14a2*⁺/*Apq1*⁻/tdT⁺; b3-4 *Slc39a8*⁺/*Apq1*⁻/tdT⁺; b5, *Fst*⁺/*Apq1*⁻/tdT⁻.

Figure S4 Dataset metrics for ureteric epithelial cell groupings *Related to Figure 4*

A, UE dataset as violin plots of the 16 clusters in Figure 4B, showing per cluster levels for nGenes (top), nUMI (middle) and percent mitochondrial genes (bottom). **B**, tSNE and stacked barplots of UE dataset illustrating the distribution (left) and composition (right) of the clusters by replicate; IC, intercalated cells, type A and B; CNT, connecting tubule; CCD, cortical collecting duct; OMCD, IMCD, outer and inner medullary collecting duct, types1-3; dME, deep medullary epithelium.

Figure S5 Nephron lineage derived principal and intercalated cell types at connection to collecting system *Related to Figure 5*

A, General cell markers of intercalated cells (IC, *Atp6v1b1*) and principal cells (PC, *Aqp2*) in nephron (N) and ureteric epithelium (UE) around the junction with collecting system. **B-D**, Cell specific analysis of indicated markers in PC, IC and other epithelial cell types around the junction with collecting system. **E,F**, tSNE projections of re-clustered cortical epithelial cell types from N and UE datasets displayed as zone (E) or lineage (F) of origin. **G**, Differentially expressed genes for each cluster in Figure 5I. **H**, Feature plots showing expression of key marker genes defining cell types. **I**, Feature plot displaying *Hoxd10* expression across clusters in Figure 5I. **J**, Rare *Ckb*⁺ cells (arrowhead) within the *Slc12a3*⁺ distal convoluted tubule cluster 8 in Figure 5I. **K**, NonA-nonB IC identified by double label for *Rhbg* and *Slc26a4*; arrowheads: white, IC-B; orange, nonA-nonB IC. **L-M**, Immunofluorescent and RNAscope validation (arrowheads) of *Ache* in UE-PCs and *Sh2d4b* in UE-IC-B, respectively; merged panel on left.

Figure S6 Progressive shift of transcription factors from the ureteric lineage into the distal nephrogenic lineage in early nephrogenesis *Related to Figure 5*

A-D, Human (week 15) and mouse (day of birth) kidney sections immunostained to examine cellular distributions of transcription factors first expressed in the UE and subsequently in distal cell types of the developing nephron. Nephron development initiates with the epithelial renal vesicle (RV), proceeds through a Comma-shaped body (CSB) to an S-shaped body (SSB). A patent luminal interconnection is generated between the nephron and collecting system by the SSB stage; dashes follow proximal-distal axis of nephron precursors; arrowheads indicate transcription factors detected in distal nephron. Scale bar 10µm.

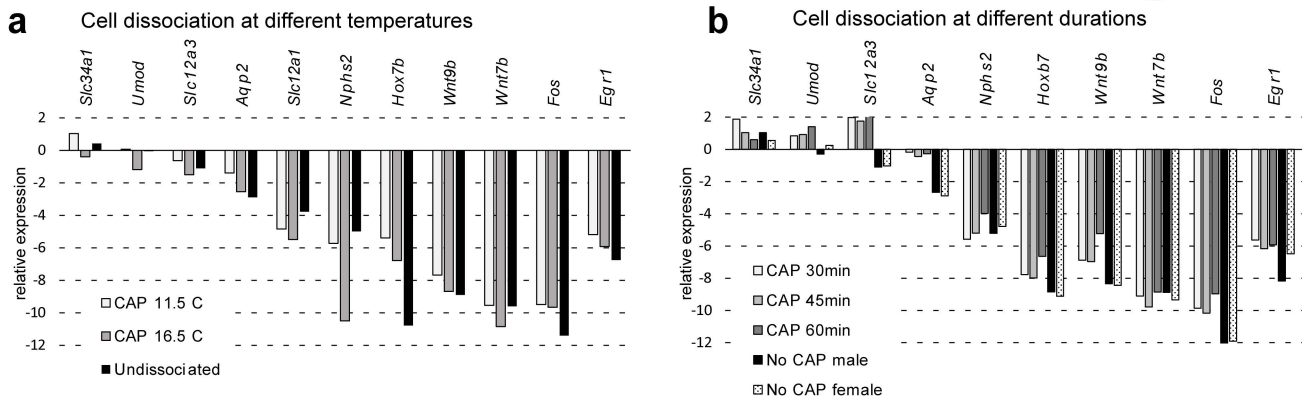
Figure S7 Enriched expression of transcription factors in clusters of nephron and ureteric epithelium datasets *Related to Figure 2 and Figure 4*

A-B, Dot plots showing differential expression of transcription factors along the proximal-distal axis of the mature nephron (A) and ureteric epithelium of the collecting system (B), with a schematic (below) indicating the region sampled: RC, renal corpuscle; f-PT, female proximal tubule; m-PT, male proximal tubule; tl-LoH, thin limbs of Loop of Henle; DT, distal tubule; IC, intercalated cell, type a and b; CNT, connecting tubule; CCD, cortical collecting duct; OMCD, outer medullary collecting duct; IMCD, inner medullary collecting duct; dME, deep medullary epithelium.

Figure S8 Kidney Cell Explorer batch search outputs heatmaps spanning all metacells
Related to Figure 6

A-C, Heat maps of gene expression for multi-gene searches in metacells of nephron and collecting duct showing examples for batch searches with genes linked to hormone regulation (A), nephrotic syndrome (B) and specific physiological activities (C).

Figure S1



c Parameters for replicates and single cell RNA sequencing

Replicates	Samples	Cell Number	Genes per Cell	UMIs per Cell	Number of Reads	Mean Reads per Cell	Total Genes Detected
F1	F1_Z1	3,730	1,199	2,786	227,832,770	61,081	17,584
	F1_Z2	3,188	1,178	2,608	227,975,422	71,510	17,315
	F1_Z3	1,805	1,361	3,021	259,414,621	143,720	15,978
F2	F2_Z1	3,216	1,407	3,571	231,671,396	72,037	17,629
	F2_Z2	3,788	1,380	3,295	236,289,620	62,378	18,078
	F2_Z3	3,295	1,738	4,339	247,417,471	75,088	17,503
M1	M1_Z1	3,393	1,494	3,873	235,710,784	69,469	17,589
	M1_Z2	3,526	1,288	2,896	239,168,214	67,829	17,651
	M1_Z3	3,580	1,554	3,813	238,383,845	66,587	17,420
M2	M2_Z1	4,095	1,439	3,393	236,710,207	57,804	18,077
	M2_Z2	3,646	1,258	2,946	230,655,689	63,262	17,828
	M2_Z3	3,450	1,445	3,379	255,389,355	74,025	17,508

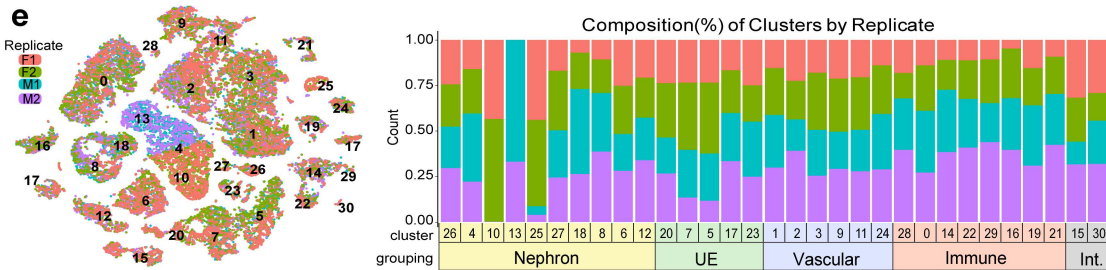
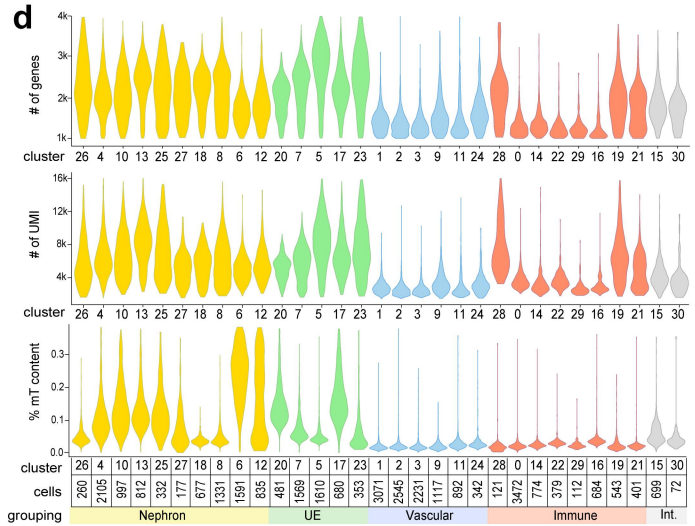


Figure S2

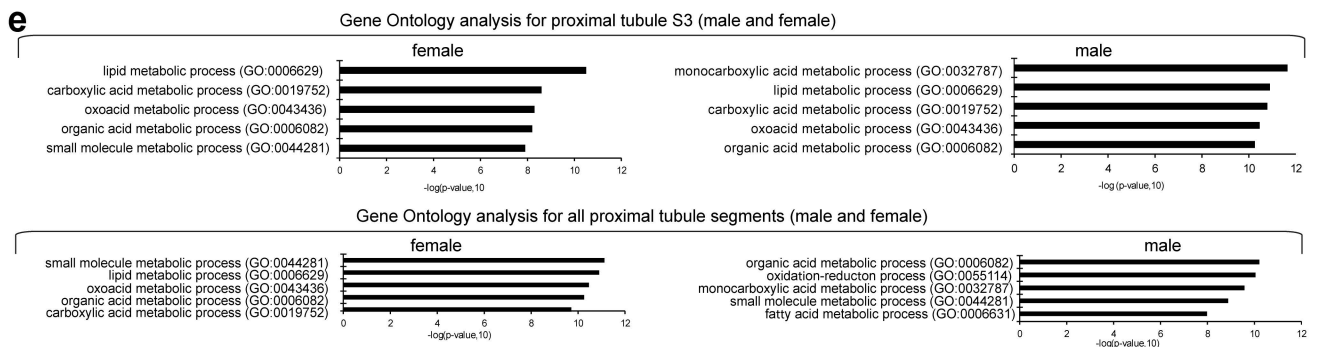
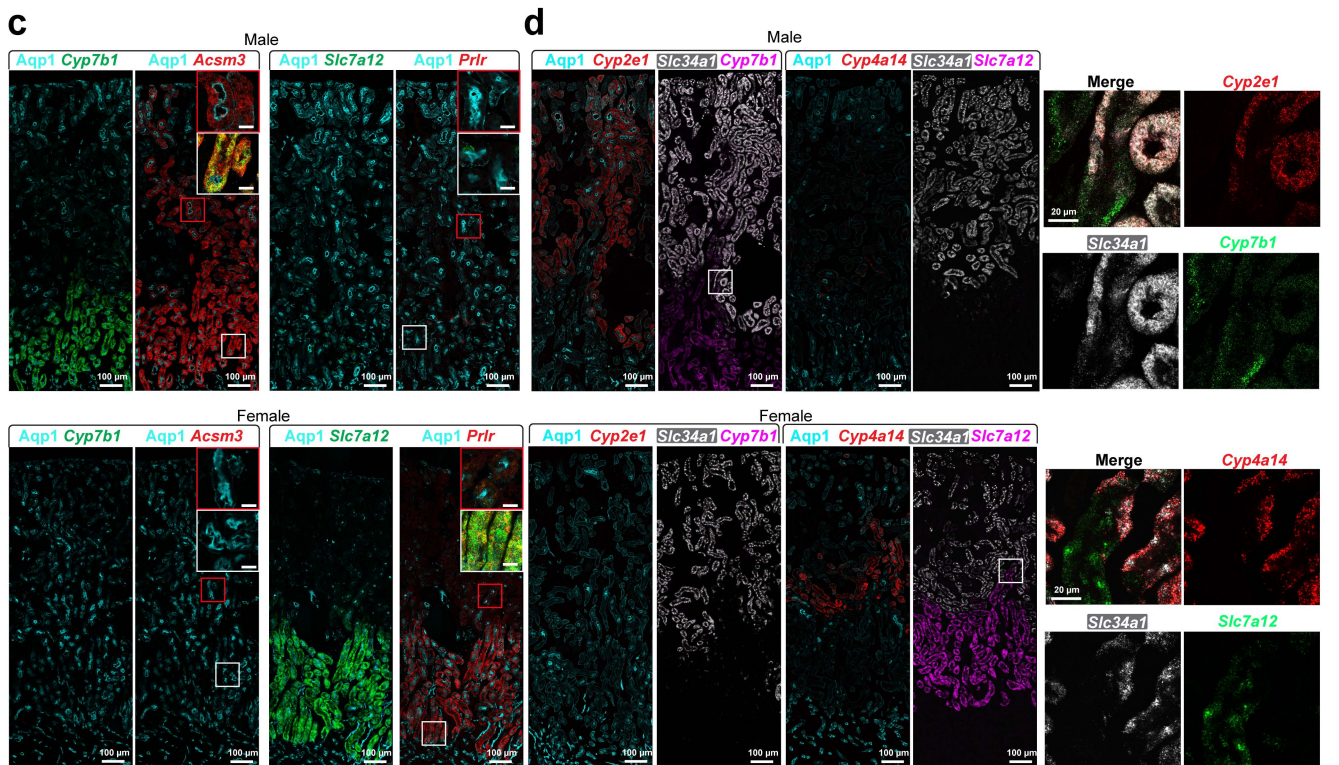
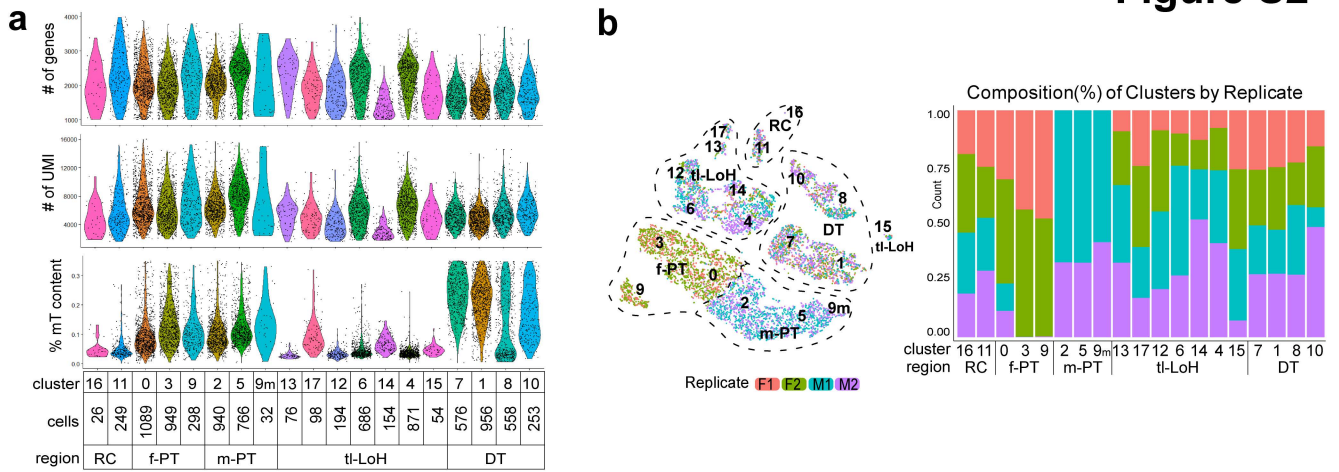
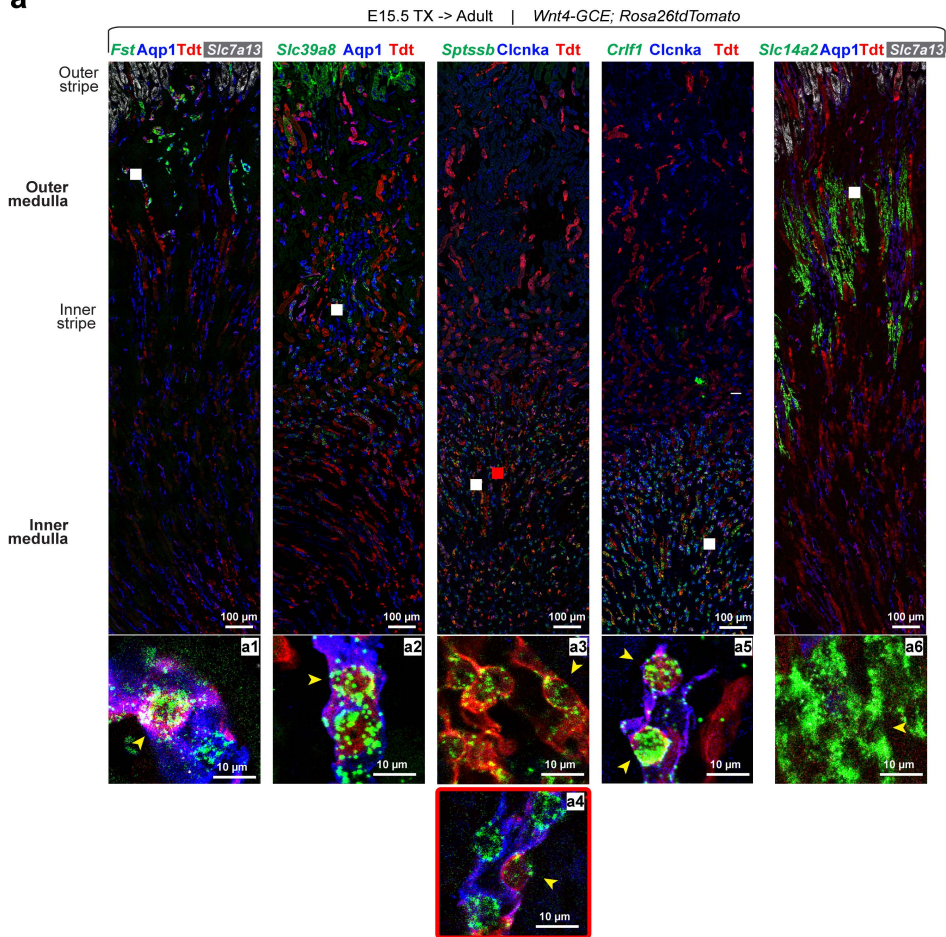


Figure S3

a



b

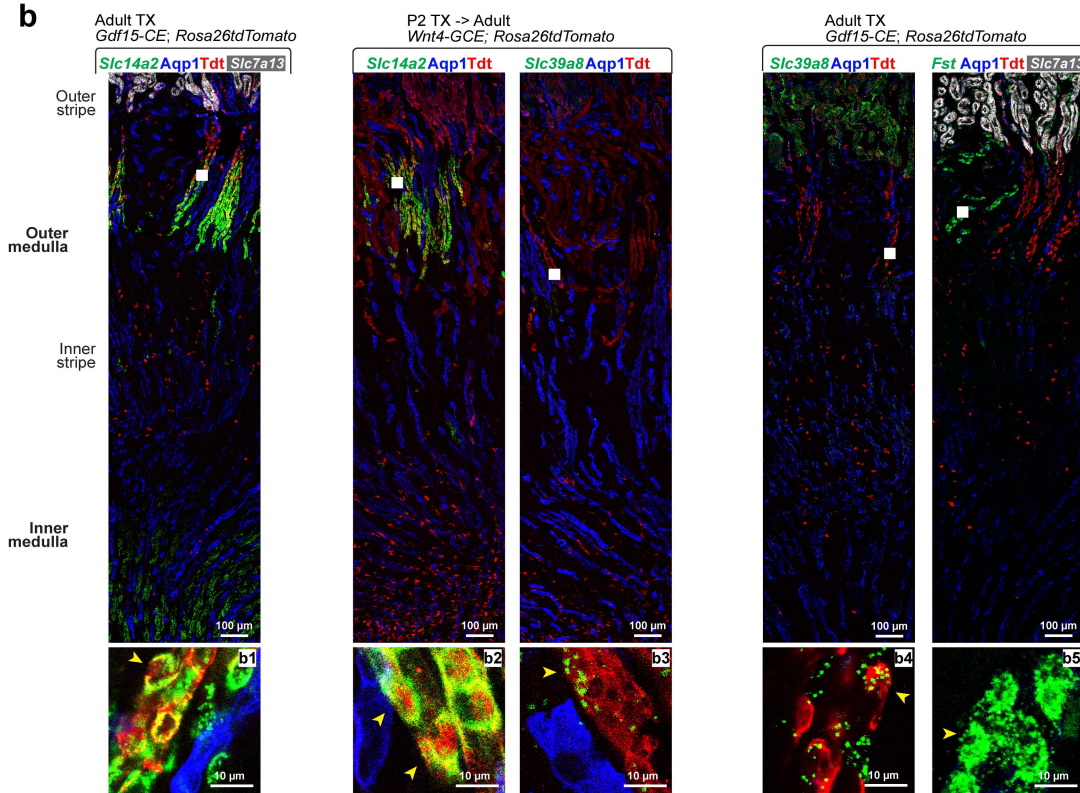
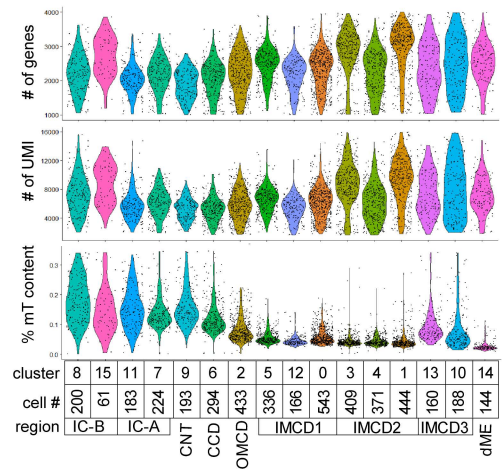


Figure S4

a



b

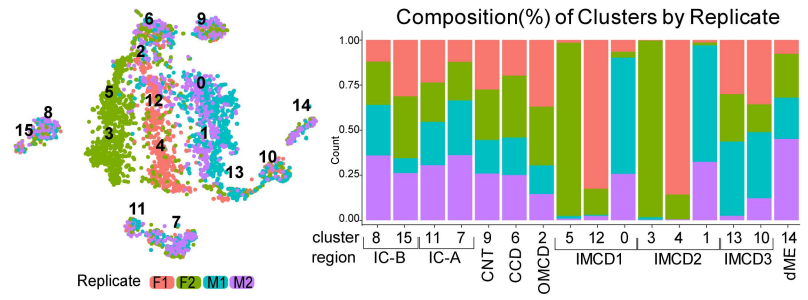


Figure S5

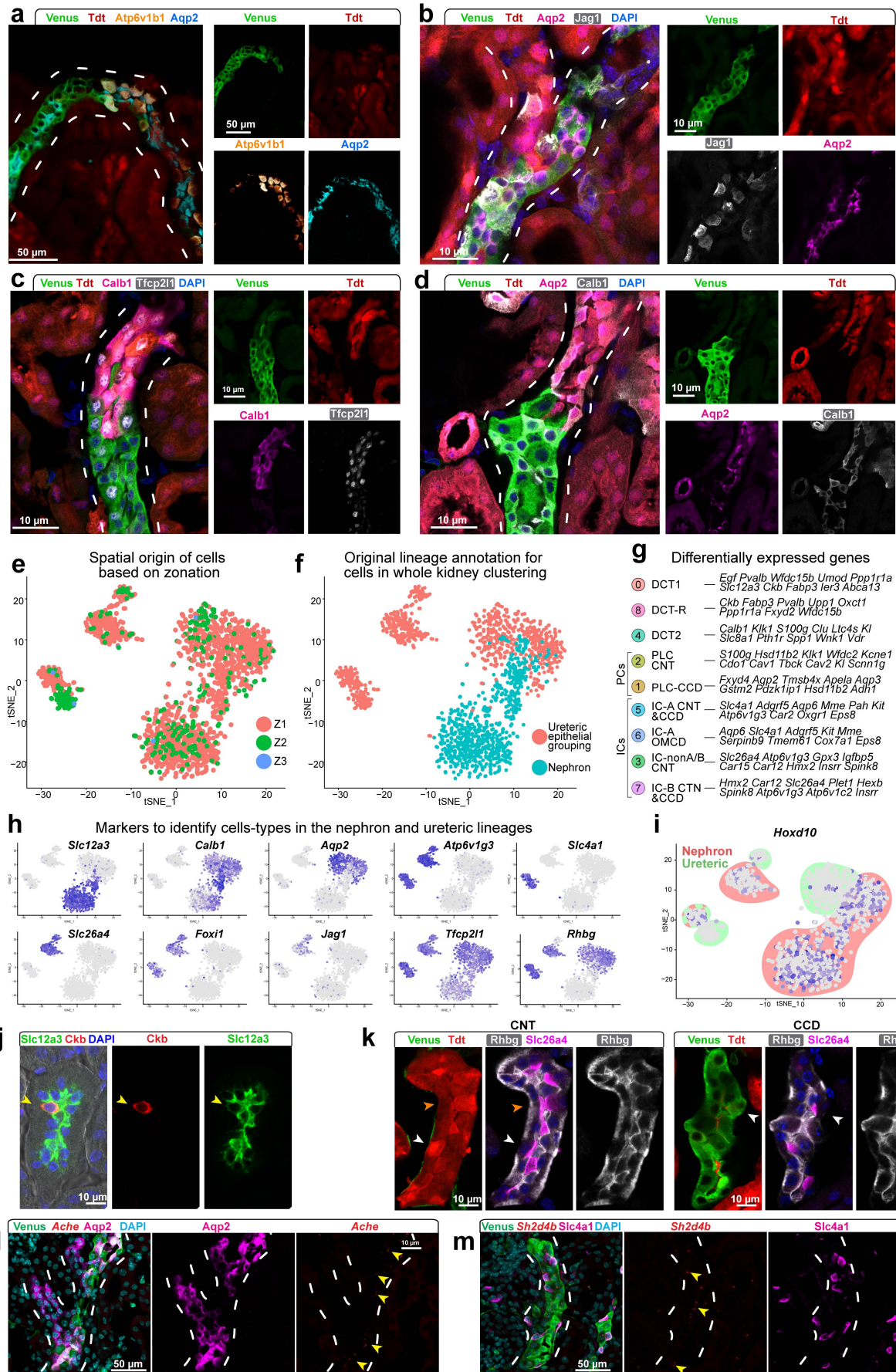


Figure S6

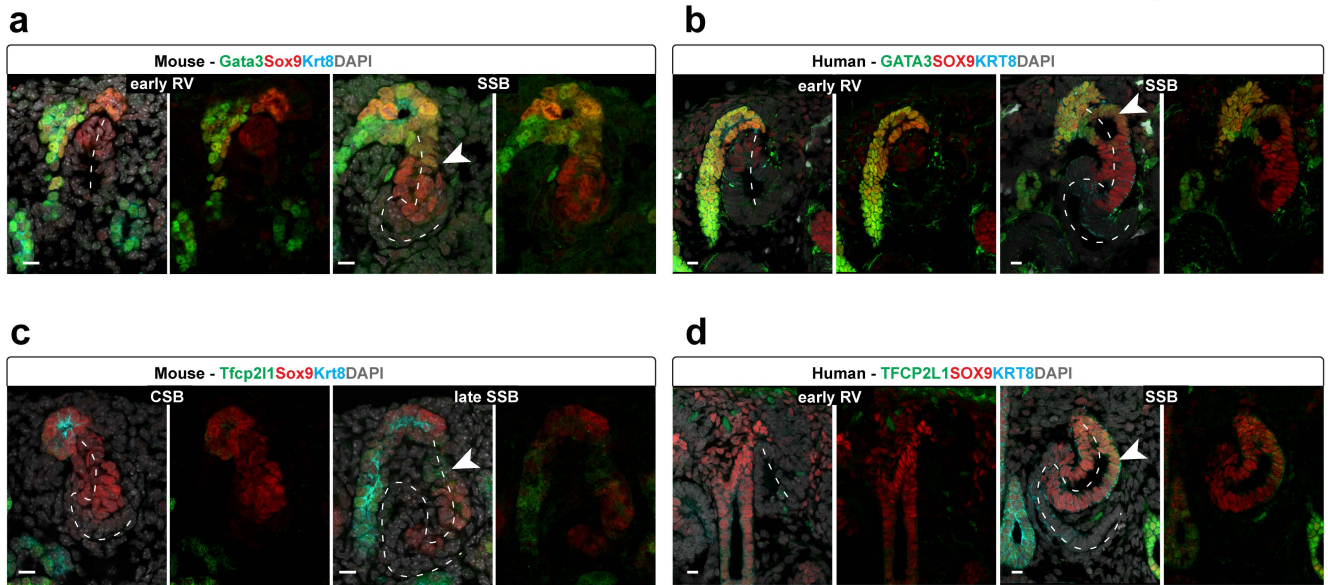


Figure S7

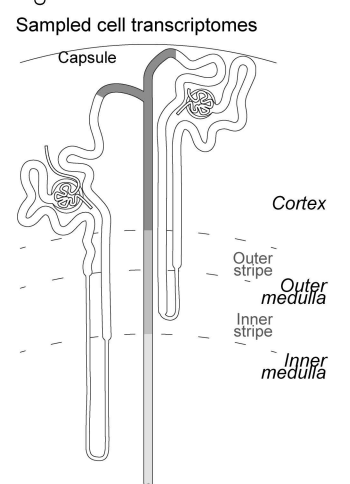
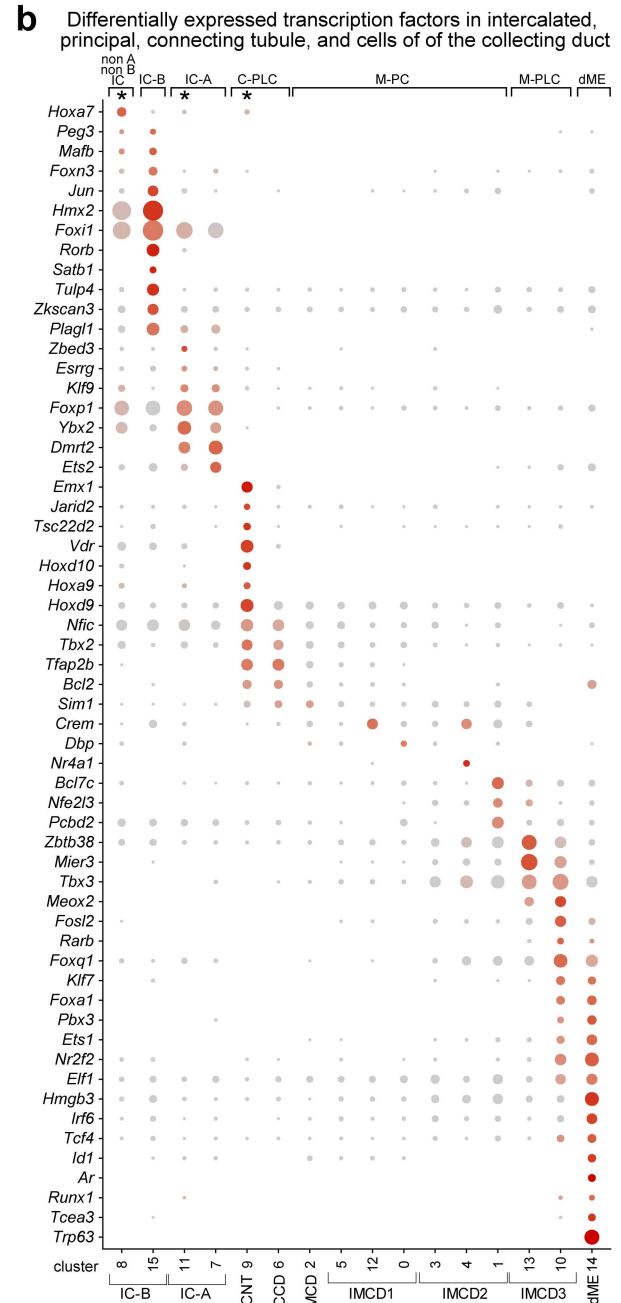
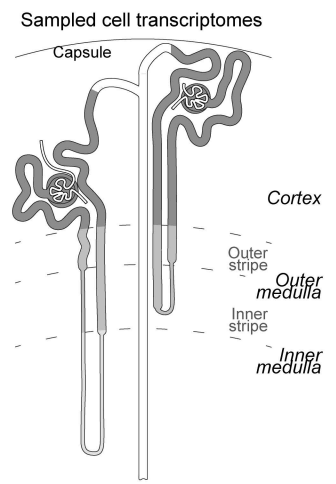
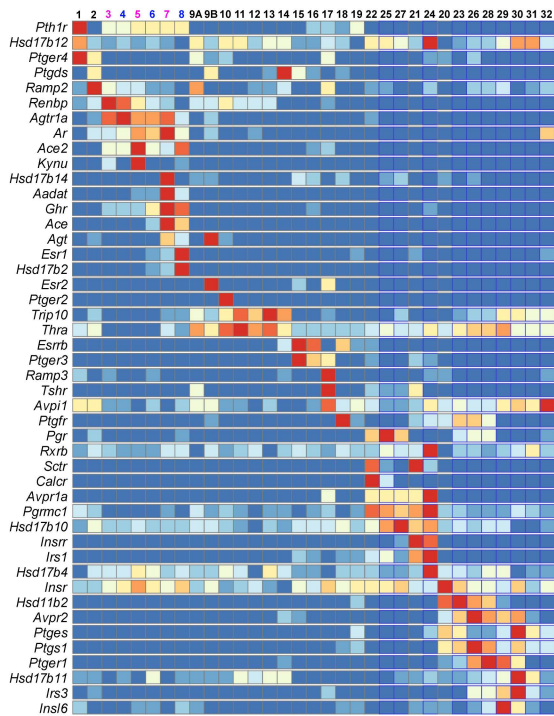
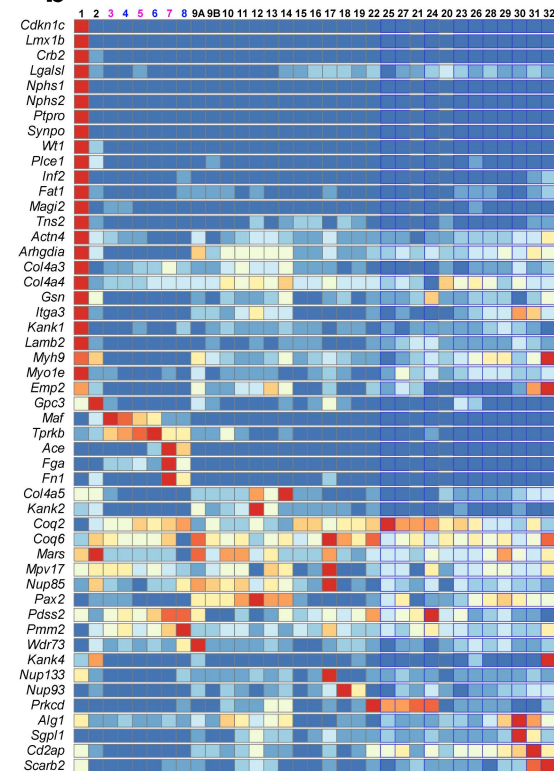


Figure S8

a Hormone related genes grouped by expression profile



b Nephrotic syndrome



c Physiologically active genes grouped by function

