

SUPPLEMENTAL INFORMATION

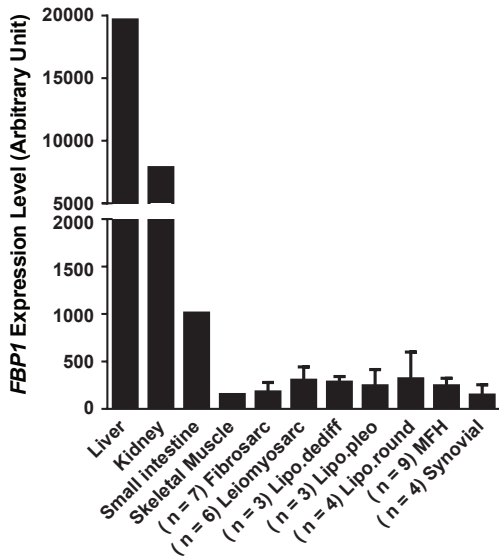
Fructose-1,6-bisphosphatase 2 inhibits sarcoma progression by restraining mitochondrial biogenesis

Peiwei Huangyang, Fuming Li, Pearl Lee, Itzhak Nissim, Aalim M. Weljie, Anthony Mancuso, Bo Li, Brian Keith, Sam S. Yoon, M. Celeste Simon

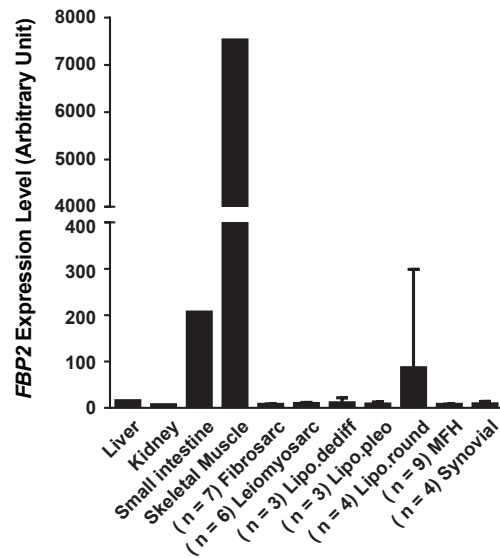
Supplemental information includes seven figures and one table.

Figure S1

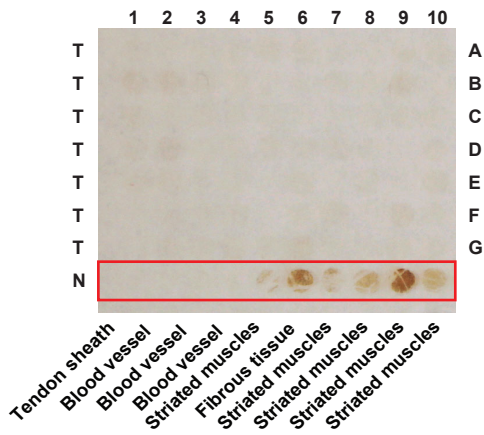
A



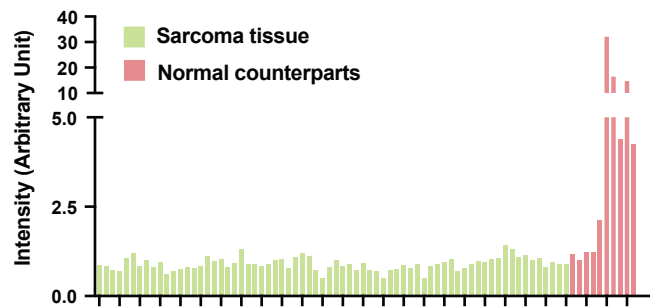
B



C



D



| | | | |
|--------------------------------|--------------------------------------|-----------------------------------|--------------------------------|
| A1: neurofibroma | C1: neurilemmoma | E1: pleomorphic rhabdomyosarcoma | G1: fibrous histiocytoma |
| A2: leiomyosarcoma | C2: neurilemmoma | E2: hemangioendothelial sarcoma | G2: mucoid type liposarcoma |
| A3: fibroma | C3: neurilemmoma | E3: liposarcoma | G3: mucoid type liposarcoma |
| A4: alveolar soft part sarcoma | C4: neurilemmoma | E4: mucoid type liposarcoma | G4: alveolar soft part sarcoma |
| A5: fibrous histiocytoma | C5: alveolar rhabdomyosarcoma | E5: liposarcoma | G5: fibrous histiocytoma |
| A6: glomus tumor | C6: neurilemmoma | E6: liposarcoma | G6: fibrous histiocytoma |
| A7: rhabdomyosarcoma | C7: neurilemmoma | E7: lipoma-like liposarcoma | G7: fibrous histiocytoma |
| A8: leiomyosarcoma | C8: fibrous histiocytoma | E8: mucoid type liposarcoma | G8: fibrous histiocytoma |
| A9: leiomyosarcoma | C9: neurilemmoma | E9: lipoma-like liposarcoma | G9: fibrous histiocytoma |
| A10: leiomyosarcoma | C10: neurilemmoma | E10: fibrous histiocytoma | G10: neurilemmoma |
| B1: leiomyosarcoma | D1: pleomorphic leiomyosarcoma | F1: fibrous histiocytoma | |
| B2: leiomyosarcoma | D2: synovial sarcoma | F2: pleomorphic liposarcoma | |
| B3: neurofibroma | D3: synovioma | F3: pleomorphic liposarcoma | |
| B4: osteosarcoma | D4: synovial sarcoma | F4: mucoid type liposarcoma | |
| B5: leiomyosarcoma | D5: synovial sarcoma | F5: lipoma-like liposarcoma | |
| B6: leiomyosarcoma | D6: tenosynovial giant cell tumor | F6: mucoid type liposarcoma | |
| B7: neurilemmoma | D7: embryonic rhabdomyosarcoma | F7: round cells liposarcoma | |
| B8: neurilemmoma | D8: round cells liposarcoma | F8: mucoid type liposarcoma | |
| B9: neurilemmoma | D9: epithelioid hemangioendothelioma | F9: alveolar rhabdomyosarcoma | |
| B10: neurofibroma | D10: hemangiopericytoma | F10: alveolar soft tissue sarcoma | |

Figure S1, Related to Figure 1. *FBP2* expression is downregulated in sarcoma.

(A and B) *FBP1* (A) and *FBP2* (B) mRNA expression in multiple normal tissues and a variety of human sarcoma samples. Fibrosarc, fibrosarcoma; Lipo.dediff, dedifferentiated liposarcoma; Lipo.pleo, pleomorphic liposarcoma; Lipo.round, round cell liposarcoma; MFH, malignant fibrous histiocytoma.

(C) Immunohistochemistry staining of a representative sarcoma microarray with *FBP2* antibody. T, sarcoma tumor tissues; N, normal counterparts. Red box indicates normal counterparts. The sarcoma type of each sample is listed below.

(D) Quantification of IHC staining of 70 sarcomas and 10 normal tissues with *FBP2* antibody from (C).

Figure S2

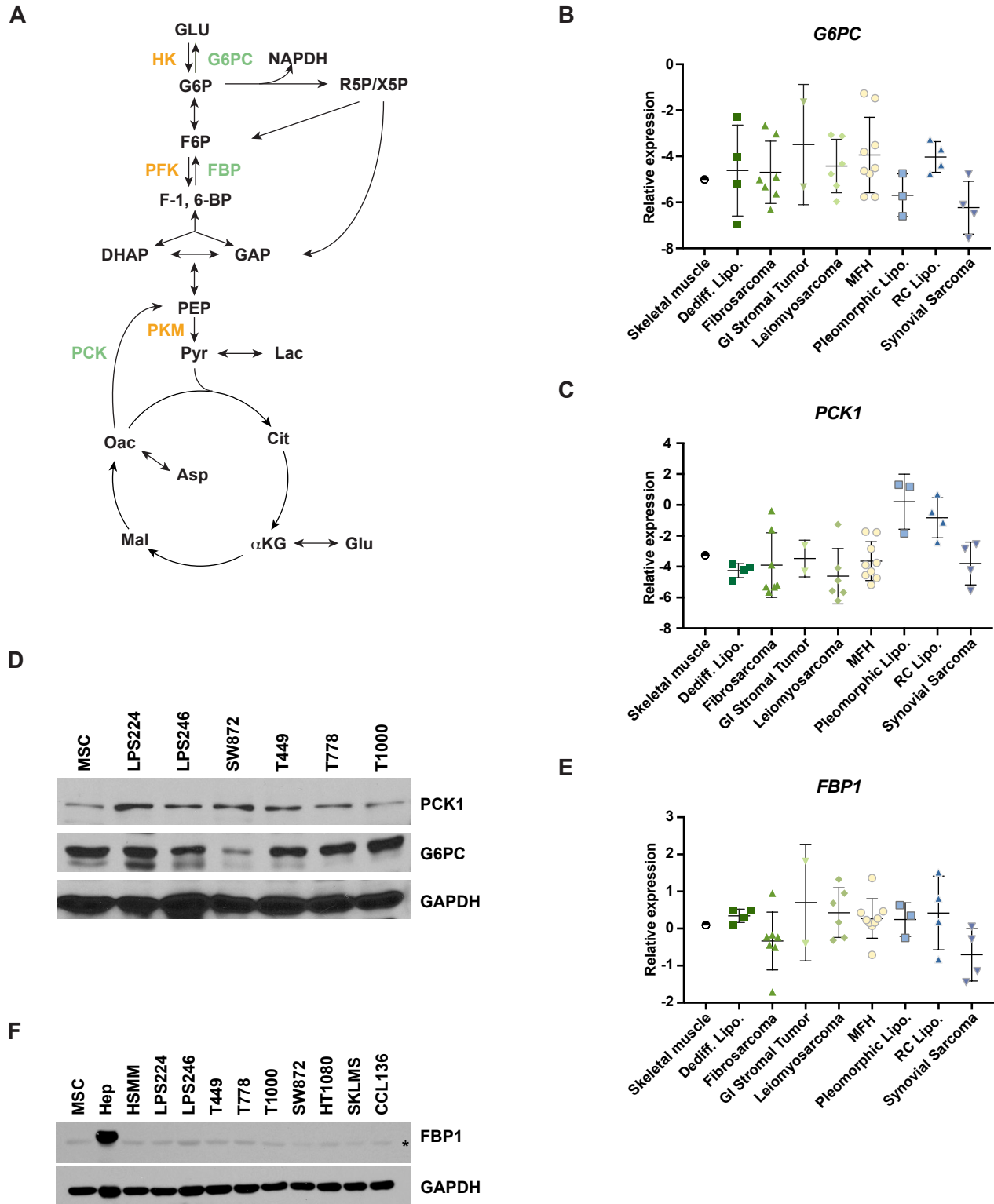


Figure S2, Related to Figure 1. The expression of other gluconeogenic enzymes exhibit no consistent change in sarcoma.

(A) Illustration of central carbon metabolism, including glycolysis, gluconeogenesis, pentose phosphate pathway, and the TCA cycle. Enzymes controlling glycolysis (HK, hexokinase; PFK, phosphofructokinase; PKM, pyruvate kinase type M) are highlighted in orange, while enzymes controlling gluconeogenesis (G6PC, glucose-6-phosphatase catalytic subunit; FBP, fructose-1,6-bisphosphatase; PCK, phosphoenolpyruvate carboxykinase) are highlighted in green. G6P, glucose-6-phosphate; F6P, fructose 6-phosphate; F-1,6-BP, fructose 1,6-bisphosphate; DHAP, dihydroxyacetone phosphate; GAP, glyceraldehyde 3-phosphate; R5P, ribose 5-phosphate; X5P, xylulose 5-phosphate; PEP, phosphoenolpyruvate; Pyr, pyruvate; Lac, lactate; Cit, citrate; α KG, α -ketoglutarate; Glu, glutamate; Mal, malate; Oac, oxaloacetate; Asp, aspartate.

(B and C) *G6PC* (B) and *PCK1* (C) mRNA expression based on OncoPrint analysis of the Detwiller *et al.* sarcoma patient samples data set (Detwiller *et al.*, 2005). Values are normalized to median-centered intensity and shown on a log₂ scale. Dediff. lipo., dedifferentiated liposarcoma; MFH, Malignant Fibrous Histiocytoma; Pleomorphic lipo., pleomorphic liposarcoma; RC lipo., round cell liposarcoma.

(D) Immunoblot analysis of PCK1 and G6PC protein levels in various human sarcoma cell lines. MSC, mesenchymal stem cells. MSCs served as normal control, while GAPDH served as a loading control.

(E) *FBP1* mRNA expression based on OncoPrint analysis of the Detwiller *et al.* sarcoma patient samples data set (Detwiller *et al.*, 2005).

(F) Immunoblot analysis of FBP1 protein levels in various human sarcoma cell lines. MSC, mesenchymal stem cell; hep, human primary hepatocyte. MSC served as normal control and Hep served as positive control for FBP1 expression. GAPDH served as a loading control. * indicates non-specific band.

Figure S3

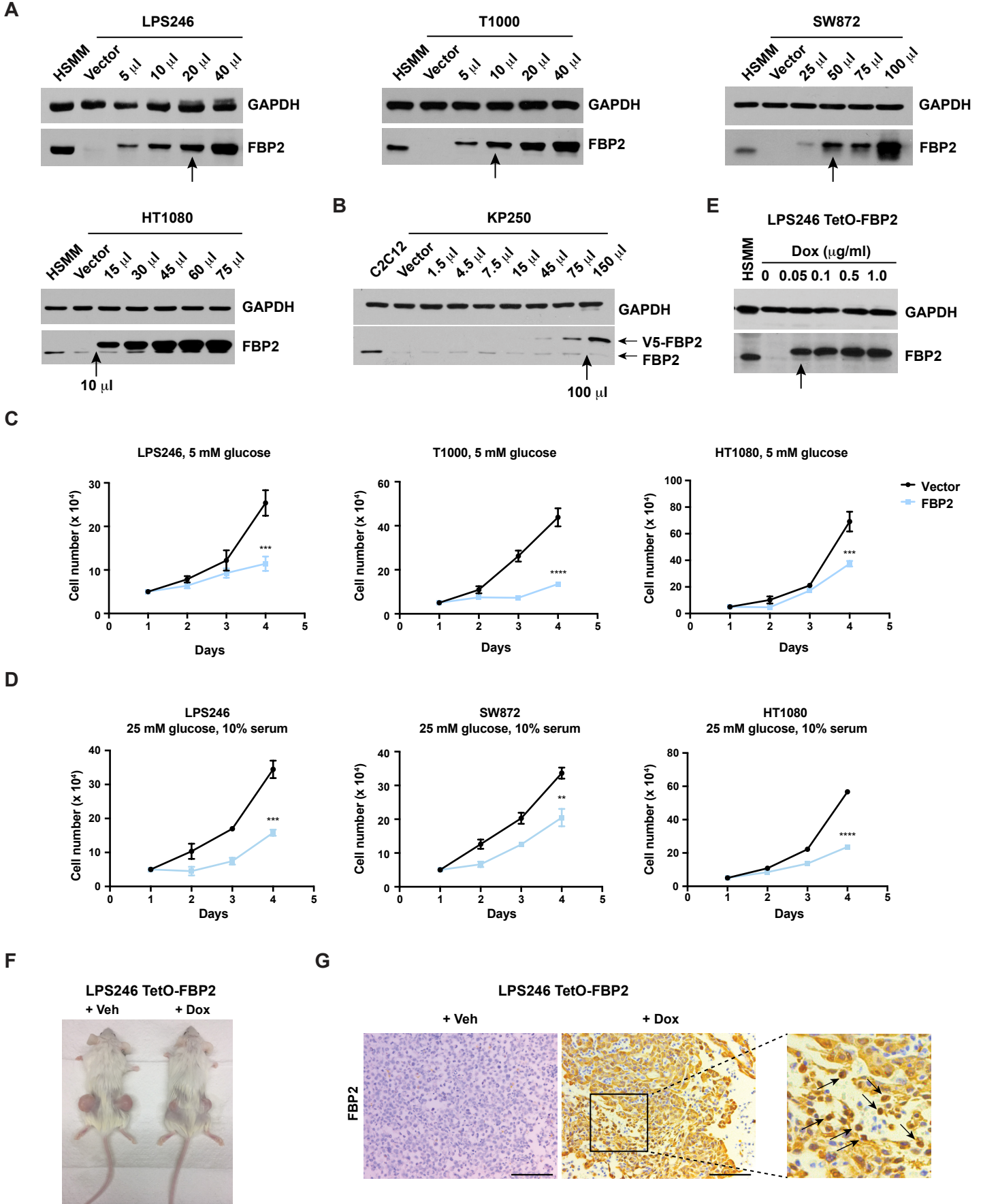


Figure S3, Related to Figure 2. FBP2 re-expression inhibits tumor growth.

(A and B) Immunoblot showing FBP2 expression levels over lentiviral titration in LPS246, T1000, SW872 and HT1080 cells compared to HSMM (A), and in KP250 cells compared to C2C12 (B). GAPDH served as a loading control. Arrows indicate virus volumes used for all experiments.

(C and D) Growth of LPS246, T1000, and HT1080 cells in low glucose medium (10% FBS and 5 mM glucose) (C) and replete medium (10% FBS and 25 mM glucose) (D), with or without ectopic FBP2 expression. $n = 3$, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

(E) Immunoblot showing FBP2 expression level with increased doxycycline concentration in LPS246 cells compared to HSMM. GAPDH served as loading control. The arrow indicates dox concentration used for all experiments.

(F) Representative xenograft tumors in mice fed with vehicle or dox chow.

(G) Representative immunohistochemistry image of FBP2 staining on LPS246 TetO-FBP2 tumor sections.

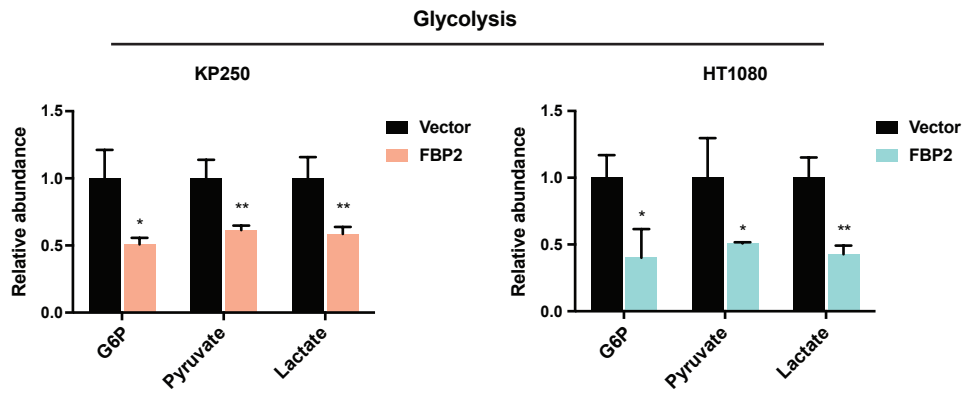
Scale bars: 100 μm . Arrows indicate nuclear staining of FBP2.

Error bars represent SD of three experimental replicates. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

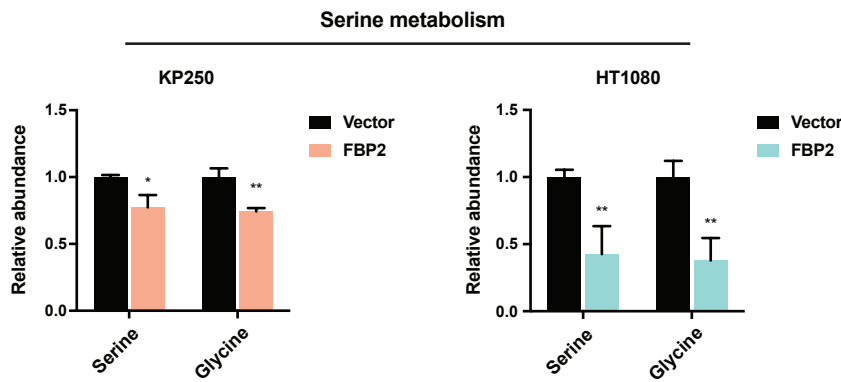
n.s., not significant.

Figure S4

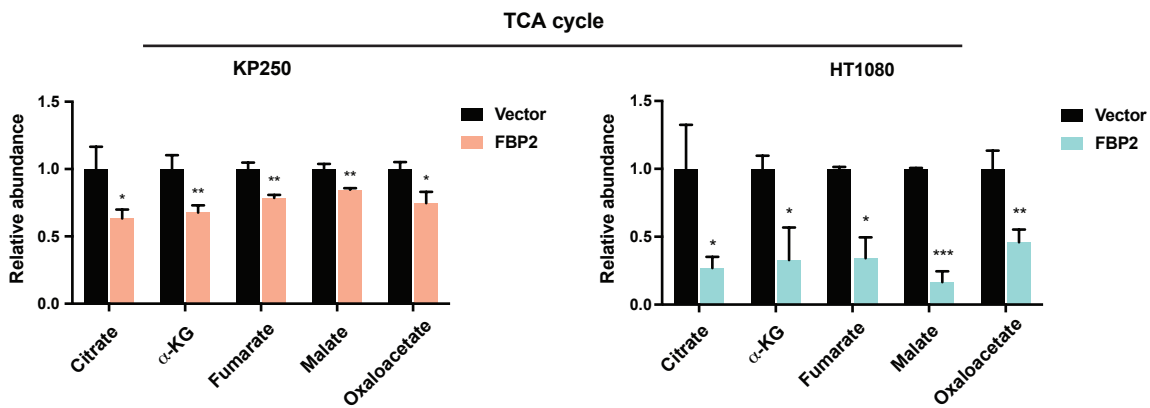
A



B



C



D

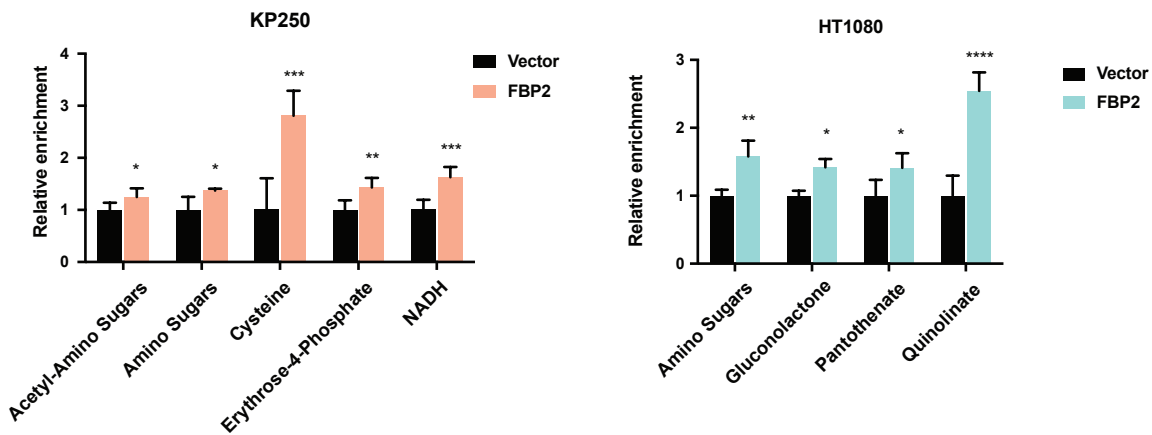


Figure S4, Related to Figure 3. Ectopic FBP2 expression opposes glycolysis and the TCA cycle.

(A-C) Pan-metabolomic analysis of steady-state metabolites in KP250 cells (left) and HT1080 cells (right) expressing vector control or FBP2. Relative abundance of metabolites in glycolysis (A), serine metabolism (B) and TCA cycle (C) are shown.

(D) Representative examples of metabolites whose relative abundance is higher in KP250 cells (left) or HT1080 cells (right) upon FBP2 restoration, compared to cells expressing control vector.

Error bars represent SD of three experimental replicates. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

n.s., not significant.

Figure S5

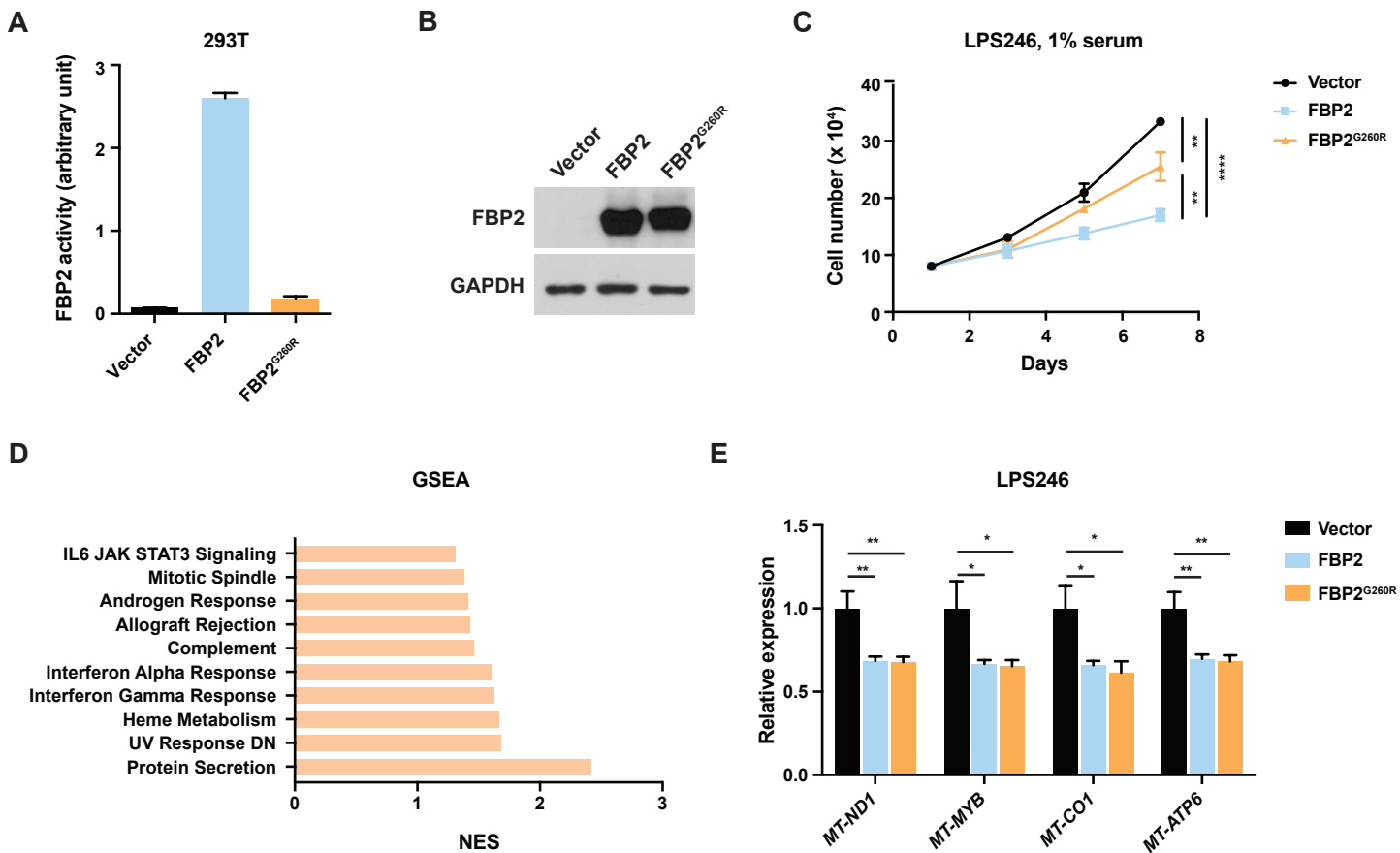


Figure S5, Related to Figure 4. FBP2 inhibits mitochondrial gene expression in a catalytic activity-independent manner.

(A) Enzymatic activity of FBP2 in 293T cells expressing vector, wild-type FBP2 and FBP2^{G260R}.

(B) Protein levels of ectopically expressed FBP2 and FBP2^{G260R} in LPS246 cells. GAPDH was used as a loading control.

(C) Growth of vector control, FBP2- or FBP2^{G260R}-expressing LPS246 cells grown in 1% serum medium.

(D) GSEA comparing vehicle-treated (n = 5) and dox-treated (n = 4) LPS246 TetO-FBP2 cells. The 50-gene “Hallmark signatures” set from MsigDB was queried, revealing top 10 gene sets upregulated in dox-treated groups, as shown with the normalized enrichment score (NES).

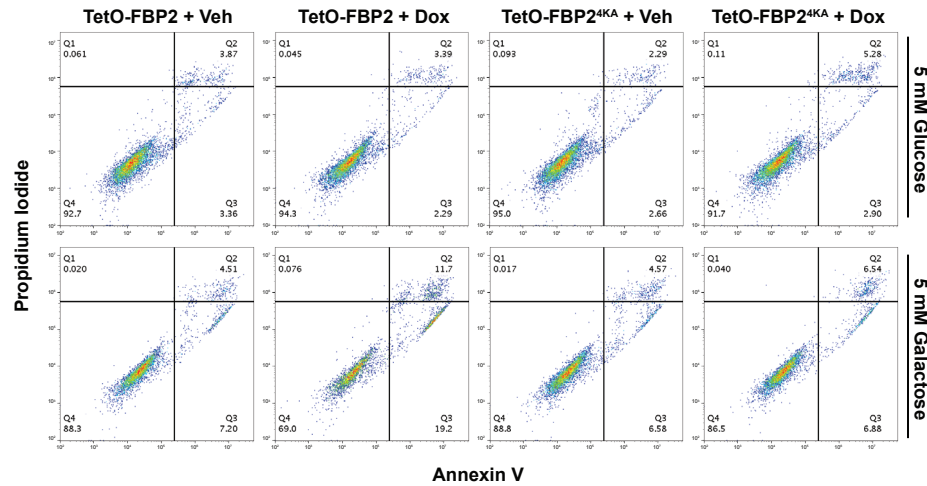
(E) qRT-PCR analysis of *MT-ND1*, *MT-CYB*, *MT-CO1*, and *MT-ATP6* in LPS246 cells constitutively expressing vector, FBP2 or FBP2^{G260R}.

Error bars represent SD of three experimental replicates. *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001.

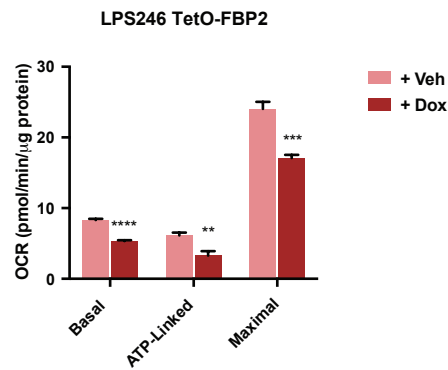
n.s., not significant.

Figure S6

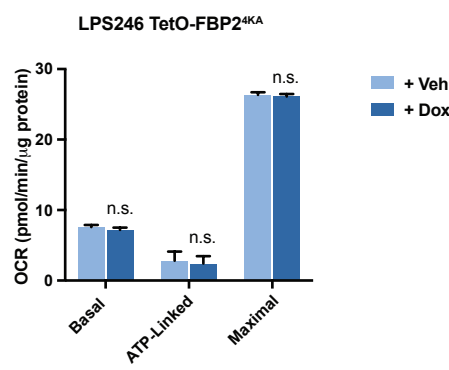
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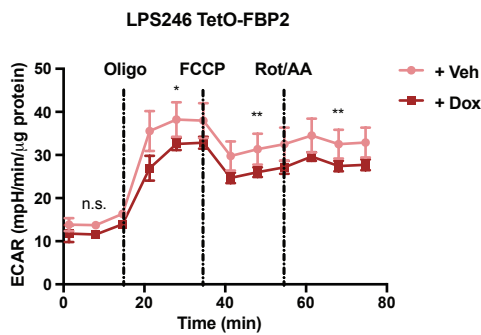
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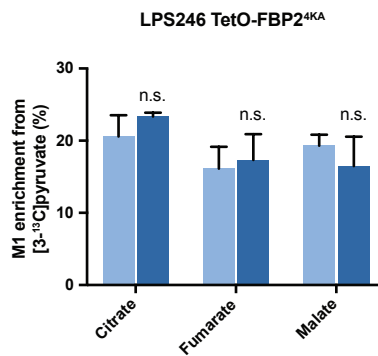
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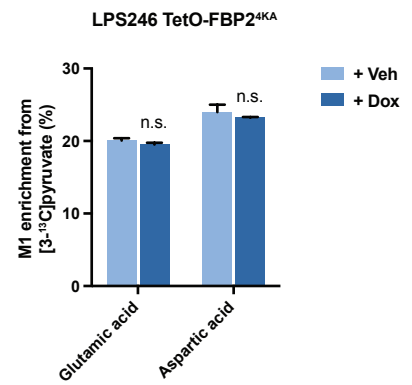
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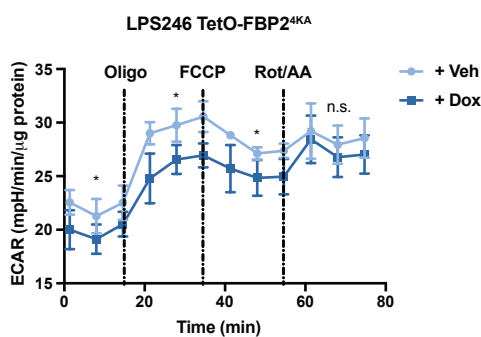
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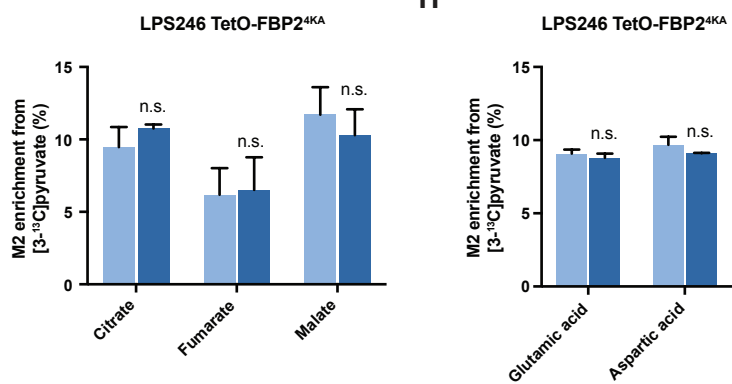
F



G



H



H

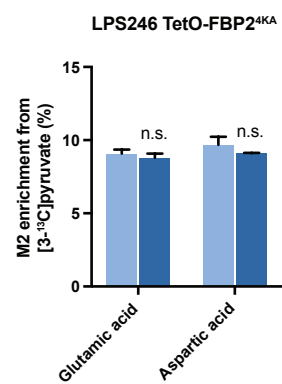


Figure S6, Related to Figure 6. FBP2 re-expression affects mitochondrial respiratory capacity.

(A) Indicated cells were cultured in 5 mM glucose and 5 mM galactose medium. Flow cytometry plots of Annexin V/PI staining of indicated cells.

(B and C) Histogram showing basal, ATP-linked and maximal respiration in LPS246 TetO-FBP2 cells (B) or LPS246 TetO-FBP2^{4KA} cells (C) treated with vehicle or dox. Data are presented as mean \pm SD of three reading cycles of n = 9 wells pooled from three independent experiments. **p < 0.01, ***p < 0.001, ****p < 0.0001.

(D) Relative extracellular acidification rate (ECAR) normalized to protein abundance in LPS246 TetO-FBP2 cells (upper panel) or LPS246 TetO-FBP2^{4KA} cells (lower panel) treated with vehicle or dox. Data are presented as mean \pm SD of three reading cycles of n = 9 wells pooled from three independent experiments.

(E-H) M1 isotopomer distribution of indicated TCA metabolites (E) and amino acids (F), and M2 isotopomer distribution of indicated TCA metabolites (G) and amino acids (H) in LPS246 TetO-FBP2^{4KA} cells with vehicle or dox treatment, labelled with [3-¹³C]pyruvate. Error bars represent SD of three experimental replicates. *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001. n.s., not significant.

Figure S7

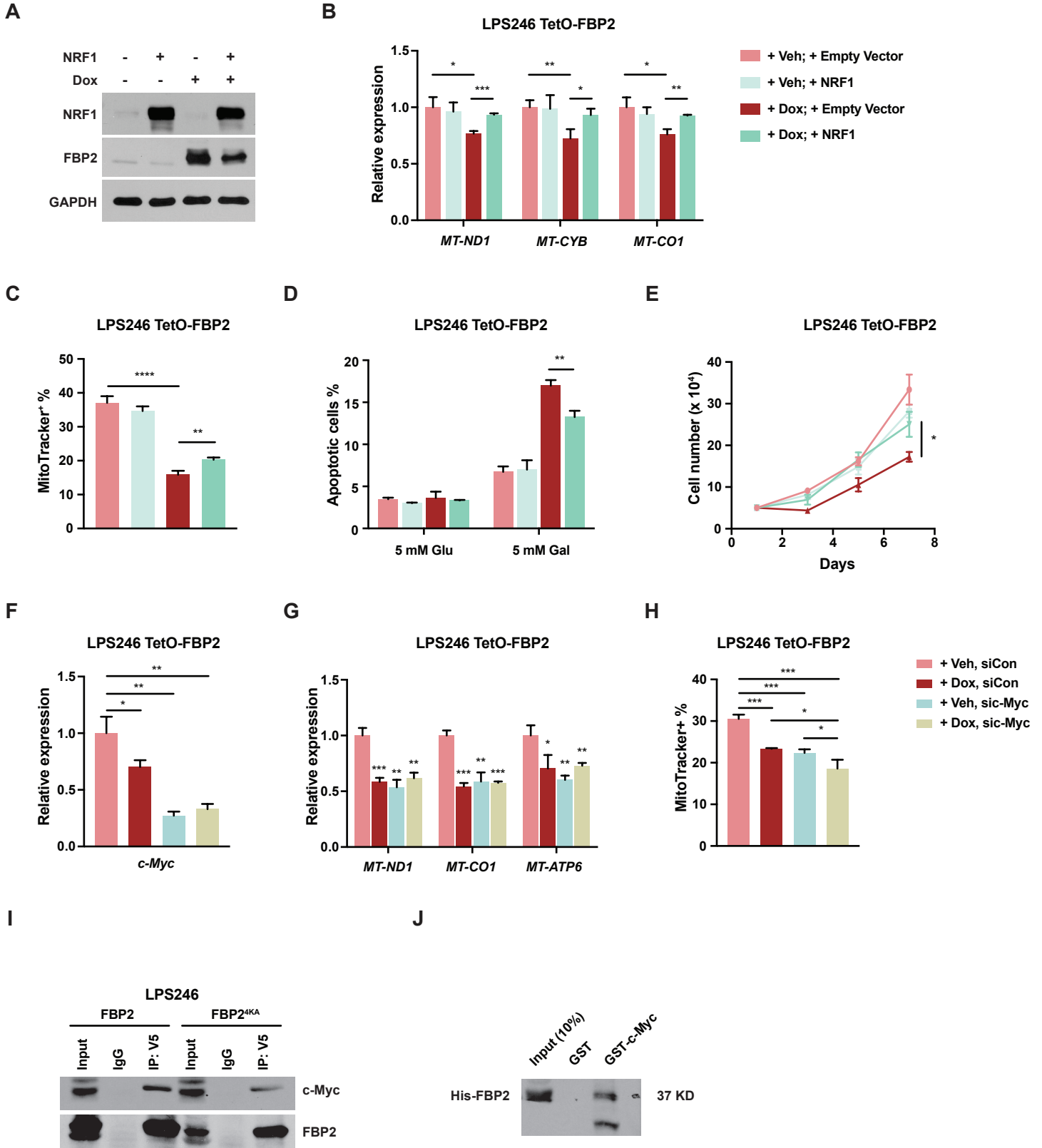


Figure S7, Related to Figure 7. NRF1 partially rescues FBP2-mediated inhibition of mitochondrial biogenesis; FBP2's effects on mitochondrial biogenesis are highly dependent on c-Myc.

(A) Immunoblot analysis for ectopic expression of vector control or NRF1 in LPS246 TetO-FBP2 cells.

(B) qRT-PCR analysis of *MT-ND1*, *MT-CYB*, and *MT-CO1* in indicated cells.

(C) LPS246 TetO-FBP2 cells with or without NRF1 expression were stained with MitoTracker Green FM probe. Fluorescence intensity corresponding to mitochondrial mass is shown in histogram.

(D) Indicated cells were cultured in 5 mM glucose and 5 mM galactose medium. Apoptotic cells were measured through Annexin V/PI staining followed by flow cytometry.

(E) Growth of indicated cells in low serum medium (1% FBS).

(F and G) qRT-PCR analysis of *c-Myc* (F) and *MT-ND1*, *MT-CO1* and *MT-ATP6* (G) in LPS246 TetO-FBP2 cells under four conditions (+ Veh, siControl; + Dox, siControl; + Veh, sic-Myc; + Dox, sic-Myc).

(H) LPS246 TetO-FBP2 cells treated with four conditions stained with MitoTracker Green FM probe. Histogram shows the quantification of fluorescence intensity corresponding to mitochondrial mass.

(I) V5 tagged FBP2 or FBP2^{4KA}-expressing LPS246 cell lysates were immunoprecipitated with IgG, or V5 antibody and blotted for endogenous c-Myc. IP, immunoprecipitate.

(J) GST pull-down analysis between recombinant His-FBP2 and recombinant GST or GST-tagged c-Myc and blotted using FBP2 antibody.

Error bars represent SD of three experimental replicates. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$. n.s., not significant.

Table S1. Related to STAR METHODS section. Primers for qRT-PCR.

| | | |
|-------------------------------|--------------------------|--|
| 18S | Life Technologies | HS03928985_G1 |
| MT-ND1 | Life Technologies | HS02596873_S1 |
| MT-MYB | Life Technologies | HS02596867_S1 |
| MT-CO1 | Life Technologies | HS02596864_G1 |
| MT-ATP6 | Life Technologies | HS02596862_G1 |
| TFAM | Life Technologies | Hs00273372_s1 |
| NRF1 | Life Technologies | Hs00602161_m1 |
| c-Myc | Life Technologies | Hs00153408_m1 |
| CCDN2 | Life Technologies | Hs00153380_m1 |
| eIF2A | Life Technologies | Hs00230684_m1 |
| NPM1 | Life Technologies | Hs02339479_g1 |
| PSAT1 | Life Technologies | Hs00795278_mH |
| TFAM_Neg | (Li et al., 2005) | Forward: GATGAATGAGGCAGAATCAGC Reverse: CTTTCACCCATCTTGAAGTCC |
| TFAM_1 | (Li et al., 2005) | Forward: GGCATACTTGTCACACGTTCTCA Reverse: CAGTCTGGCCTCCAGCTTG |
| TFAM_2 | (Li et al., 2005) | Forward: TTAGGTTTGCGAATCCCCG Reverse: CGCCCGTGACGGTCA |
| β -globin (genomic DNA) | (Dickinson et al., 2013) | Forward: CAACTTCATCCACGTTCCACC Reverse: GAAGAGCCAAGGACAGGTAC |