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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see <u>Authors & Referees</u> and the <u>Editorial Policy Checklist</u>.

Statistics	
For all statistical analy	ses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a Confirmed	
The exact sar	nple size (n) for each experimental group/condition, given as a discrete number and unit of measurement
A statement	on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
The statistica Only common	l test(s) used AND whether they are one- or two-sided tests should be described solely by name; describe more complex techniques in the Methods section.
🗶 🔲 A description	of all covariates tested
A description	of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
A full descrip AND variation	tion of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) in (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
For null hypo Give P values a	thesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted is exact values whenever suitable.
For Bayesian	analysis, information on the choice of priors and Markov chain Monte Carlo settings
For hierarchie	cal and complex designs, identification of the appropriate level for tests and full reporting of outcomes
Estimates of	effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.
Software and	code
Policy information abo	out <u>availability of computer code</u>
Data collection	N/A
Data analysis	Custom Matlab (R2015a) code was used to analyse the periodicity, intensity of puncta in Figure S5.
	tom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.
Data	
Accession codes, urA list of figures that	out <u>availability of data</u> include a <u>data availability statement</u> . This statement should provide the following information, where applicable: nique identifiers, or web links for publicly available datasets have associated raw data y restrictions on data availability
_	and analysed for Figures 1, 3, 6 and Supplementary Figures 2 and 3 are available in the Source Data file. Datasets generated and analysed and Supplementary Figures 5 and 6 are available from the corresponding author on reasonable request.
Field-spec	ific reporting
Please select the one b	pelow that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.
X Life sciences	Behavioural & social sciences

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

The sample size used here was based on our previous published results, which indicate a sufficient power to detect differences between wild-type and other mutants. The sample size reflects the nature of the assay and it is also used in most leading studies on neuronal development and maintenance in C. elegans.

Data exclusions

The only animals that were excluded from the analysis were those animals that died following axotomy, because axonal degeneration could not be confirmed in these animals (Figure S2A). This criterion was pre-established as a condition for the scoring of axonal degeneration. In rare cases animals also perished following RNAi knockdown of unc-70 (Figure S3F). These animals were excluded from scoring axonal breaks. This criterion was pre-established as a condition for the scoring of axonal breaks.

Replication

The expression of any transgene from semi-stable extrachromosomal array was replicated in multiple independent lines. Where presented in the main figures a representative strain is depicated and the raw data for all replicates are presented in Supplementary Table 2.

Randomization

No specific randomization was used. The animal groups were determined by their genotype (wild-type, mutant, transgenic, double mutant, triple mutant), with each population considered isogenic. A number of animals from each population was taken from the petri dish with no bias (the first one in the field of view) other than age (L4 stage) and sex (hermaphrodites).

Blinding

C. elegans wild-type and a large set of mutant animals were used in this study. The genotypes of the animals was known to the investigator and was often associated with an obvious visible phenotype making blinding impractical.

Reporting for specific materials, systems and methods

Mathada

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Methous	
n/a Involved in the study	
ChIP-seq	
🗷 🔲 Flow cytometry	
MRI-based neuroimaging	
·	

Animals and other organisms

Materials & experimental systems

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals	Caenorhabditis elegans
Wild animals	The study did not involve wild animals.
Field-collected samples	The study did not include field-collected samples.
Ethics oversight	No ethical approval was required.

Note that full information on the approval of the study protocol must also be provided in the manuscript.