

Reviewer Report

Title: Multifaceted Hi-C benchmarking: what makes a difference in chromosome-scale genome scaffolding?

Version: Original Submission **Date: 7/29/2019**

Reviewer name: Jay Ghurye

Reviewer Comments to Author:

I thoroughly enjoyed reading the manuscript benchmarking HiC data for assembly through different aspects. To my knowledge, this is the first study that comprehensively studies this topic. This is a novel study and I think the topic of the manuscript will receive tremendous interest. However, I have some queries/concerns that I would like authors to address.

- I see in Supplementary Table S2 the percentage of long and short range read pairs. However less than 20 kbp and greater than 20 kbp is not very informative. Can you stratify more? Like percentage of read pairs between 10k -100k, 100k -1Mbp, 1Mbp-10Mbp, and 10 Mbp and above. This would highlight in what range the utility of iconHi-C protocol.

- I understand from Figure 9 the bulk assembly contiguity statistics. However, it doesn't tell much about how correct is the assembly. I would like to see a contact matrix for a couple of assemblies that authors think are the best. Also, a heatmap for iconHi-C assembly constructed using other Hi-C datasets is also interesting to see. Such a comparison would highlight the valuable contact information that's probably missed in iconHi-C or other Hi-C datasets.

- I saw the library QC results for GM12878, however I was not able to see any scaffolding results for it with different Hi-C datasets. Since we have a known reference genome, we can get a solid evidence that which parameter setting and what type of Hi-C library provides the best assembly in terms of both contiguity and accuracy.

- This may be out of the scope of this manuscript. Did authors find out minimum amount Hi-C read pairs required for good scaffolding? Such a discussion or recommendation would guide the amount of sequencing needed for the scaffolding project and would reduce the cost.

- The scope of the manuscript is mainly understanding the effect of different parameters on scaffolding. But, do authors have any intuition about usage of iconHi-C in other 3D genomic application such as detecting TADs, chromatin loops, etc? Some discussion would be helpful.

- Figure 8 and Figure 9 is kind of hard to understand. I would appreciate if the data is displayed in a tabular format.

Methods

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