## Title: Genome assembly provides insights into the genome evolution and flowering regulation of orchardgrass

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**Supplementary Fig. 1 The orchardgrass genome landscape.** Track a represents the seven chromosomes in Mb scale. Track b represents chromosomal distribution of 603 orthologs of *A. thaliana* flowering genes. Track c represents chromosomal distribution of gene models that were annotated, where gene density ranged from 371 bp/Mb to 380,434 bp/Mb. Track d represents the chromosomal distribution of repeat sequences where density of the repeat sequences was 74,261 bp/Mb to 903,485 bp/Mb. Track e represents chromosomal distribution of TE density, and TE density axis range was 74,261 bp/Mb – 895,797 bp/Mb. Track f represents the GC content along the assembled genome, which ranged between 41.0989%/Mb to 48.0363%/Mb. Track g represents the pooled gene expression level of five tissues (root, stem, leaf, flower, and spike). Track h represents chromosomal distribution of SNPs identified from 54 wild germplasm accessions, 11 cultivars and 11 unknown orchardgrass collection samples. The range of SNP density was 3,034/Mb – 111,599/Mb.



Supplementary Fig. 2 The workflow of orchardgrass genome assembling



Supplementary Fig. 3 K-mer frequency distributions in orchardgrass. axis means sequence depth (X), and y axis means frequency of K-mer.



**Supplementary Fig. 4 Scaffold Hi-C contact map data analysis.** the x and y axes indicate the mapping positions of the first and second read in the read pair respectively, grouped into bins. The color of each square gives the number of read pairs within that bin. White vertical and black horizontal lines have been added to show the borders between scaffolds. Scaffolds less than 1 Mb are excluded.



**Supplementary Fig. 5** The chromosome number of the diplod orchardgrass (genotype 2006-1). a) seven pairs of sister chromosomes (Prometaphase, mitosis) b) seven pairs of sister chromosomes (Metaphase, mitosis).



**Supplementary Fig. 6 Consistency between Hi-C and BioNano.** Blue dots mean negative direction, and red dots mean positive direction.



**dSupplementary Fig. 7 The density of TEs surrounding genes.** The density was determined to proportion of genes inserted by TEs in one locus to all genes inserted by TEs. TSS means transcript start point, and TTS means transcript end point. Dgl means orchardgrass, Ata means *A. tauschii*, Bdi means *B. distachyon*, and Tur means *T. Urartu*.



Supplementary Fig. 8 The distribution of desertion time for LTRs/Gypsy (a) and LTRs/Copia (b) for

orchardgrass.



**Supplementary Fig. 9** Synteny analysis of seven chromosomes from orchardgrass (Dgl) to twelve chromosomes from *Oryza sativa* (Osa) and seven chromosomes from *Aegilops tauschii* (Ata).



**Supplementary Fig. 10 REM family in orchardgrass.** (a) expression levels of REMs in different tissues. (b) expression levels of REMs in five flowering stages (BV, before vernalization; VE, vernalization; AV, after vernalization; VG, vegetative growth; BH, before heading, and HT, heading stage. Y axis represents the relative expression and x axis represents the different stages. BX indicates orchardgrass cultivar BAOXING, and DON indicates orchardgrass cultivar DONATA). (c) expansion time of REMs. (d) the density of transposons around REMs and all genes in orchardgrass. The density was determined to proportion of genes inserted by TEs in one locus to all genes inserted by TEs. TSS means transcript start point, and TTS means transcript end point.



Supplementary Fig. 11 Phylogenetic tree of 76 orchardgrass accessions. Colors of inner circle indicate the materials come from different regions: Blue, East of Mediterranean; Yellow, Central Asia; Pink, East Asia; Green, West of Mediterranean; Orange, Northern Europe. Colors of outer circle indicate the wild (red), cultivar (light blue) and uncertain (dark blue) orchardgrass accessions.



**Supplementary Fig. 12** Structure analysis with different number of K value (k=2,3,4,5,6) of 76 orchardgrass.



Supplementary Fig. 13 PCA plot of the first two components (PC1 and PC2) of 43 autotetraploid orchardgrass. blue triangles indicate wild accessions, and green triangles indicate cultivated accessions in orchardgrass.



Supplementary Fig. 14 Phylogenetic tree of 43 autotetraploid orchardgrass. Colors of inner circle indicate the materials come from different regions: Blue, East of Mediterranean; Yellow, Central Asia; Pink, East Asia; Green, West of Mediterranean; Red, Northern Europe. Colors of outer circle indicate the wild (dark green) and cultivar (light green) orchardgrass accessions.



**Supplementary Fig. 15** Analysis of important flowering related orthologues in orchardgrass. (a) Expression levels of several key orthologues associating with flowering time (BV, before vernalization; VE, vernalization; AV, after vernalization; VG, vegetative growth; BH, before heading, and HT, heading stage). (b) Phylogenetic tree of FT orthologues in orchardgrass, rice, *B. distachyon, H. vulgare, A. tauschii*, and *T. urartu*. Red line indicates five FT orthologues experience expansion during orchardgrass evolution.



Supplementary Fig. 16 Nucleotide diversity ( $\pi$ ) estimated in wild (red) and cultivated (green) orchardgrass (a) and the *FST* value (b), and patterns of LD in cultivated (c) and wild (d) in the 4.426 Mb region of orchardgrass chromosome 06.



**Supplementary Fig. 17** Comparison of *AGL61* expression during the five development stages in Orchardgrass. Error bars indicate  $\pm$ s.d.; n = 3 biological replicates.



Module-Sample relationship

**Supplementary Fig. 18** Module-Sample relationship. Heatmap shows the correlation between modules and samples, and the deeper red color represents the higher correlation.



**Supplementary Fig. 19** Expression pattern of genes in green, pink and purple module. (a) and (b) indicated the genes of green module in early and late flowering phenotype, (c) and (d) indicated the genes of pink module in early and late flowering phenotype, and (e) and (f) indicated the genes of purple module in early and late flowering phenotype.