

# Supplementary Materials

## **Robust generation of erythroid and multilineage hematopoietic progenitors from human iPSCs using a scalable monolayer culture system**

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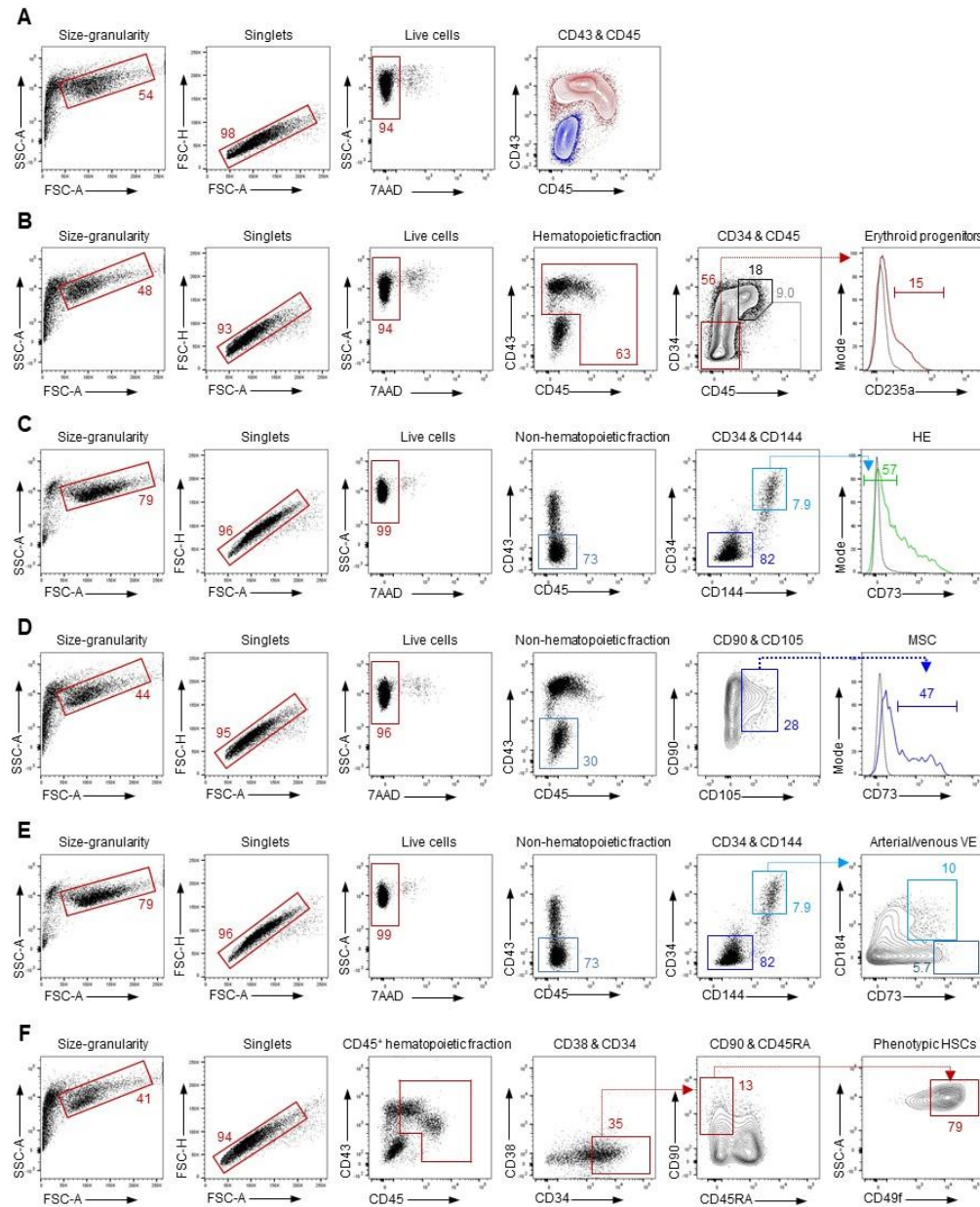


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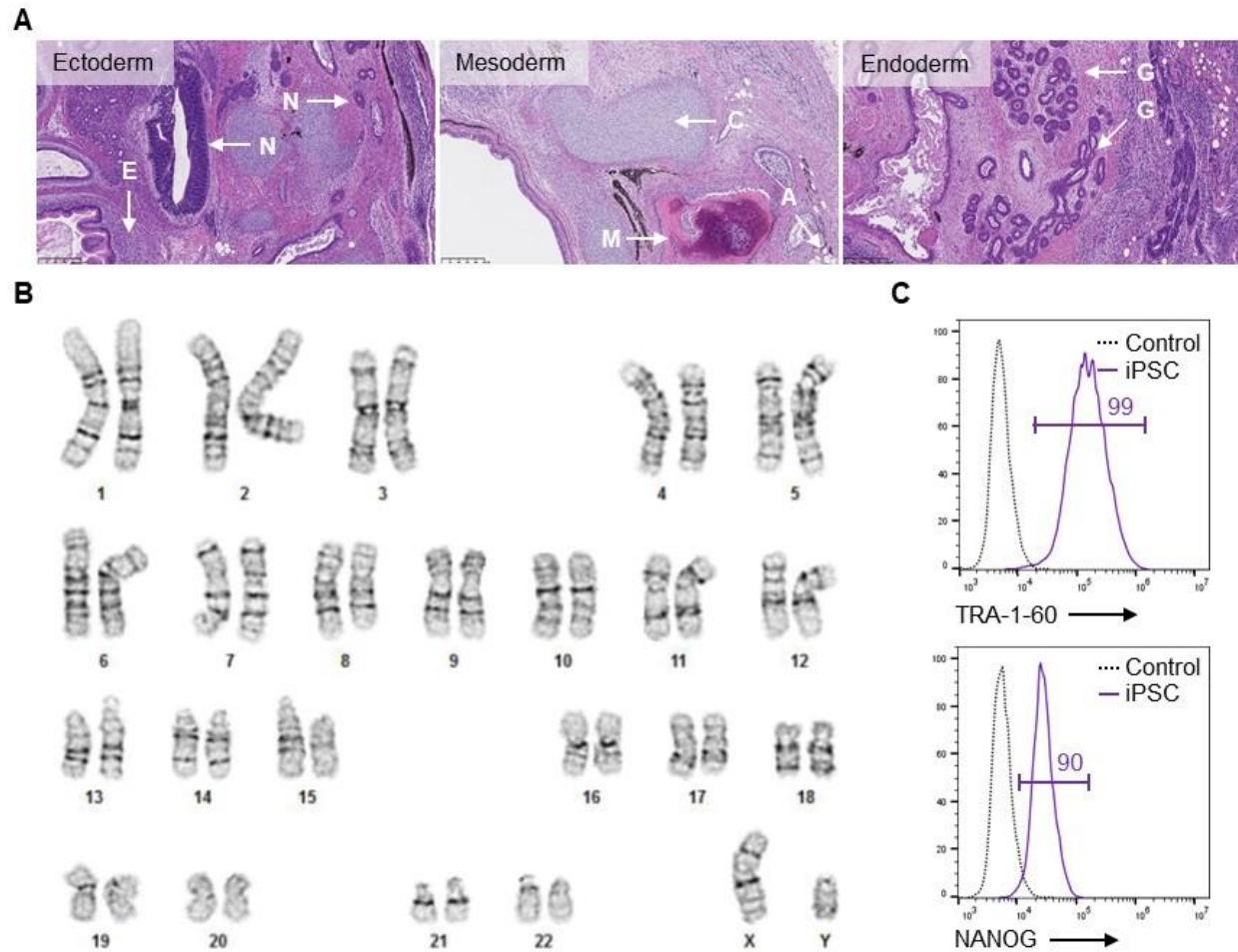
### **This PDF file includes:**

Figures S1-S6

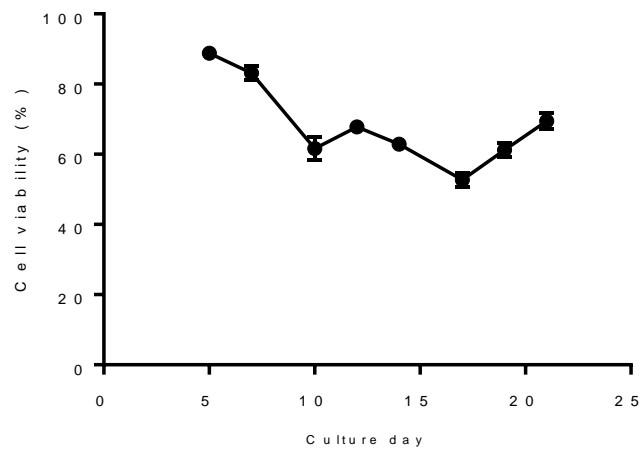
Tables S1-S3



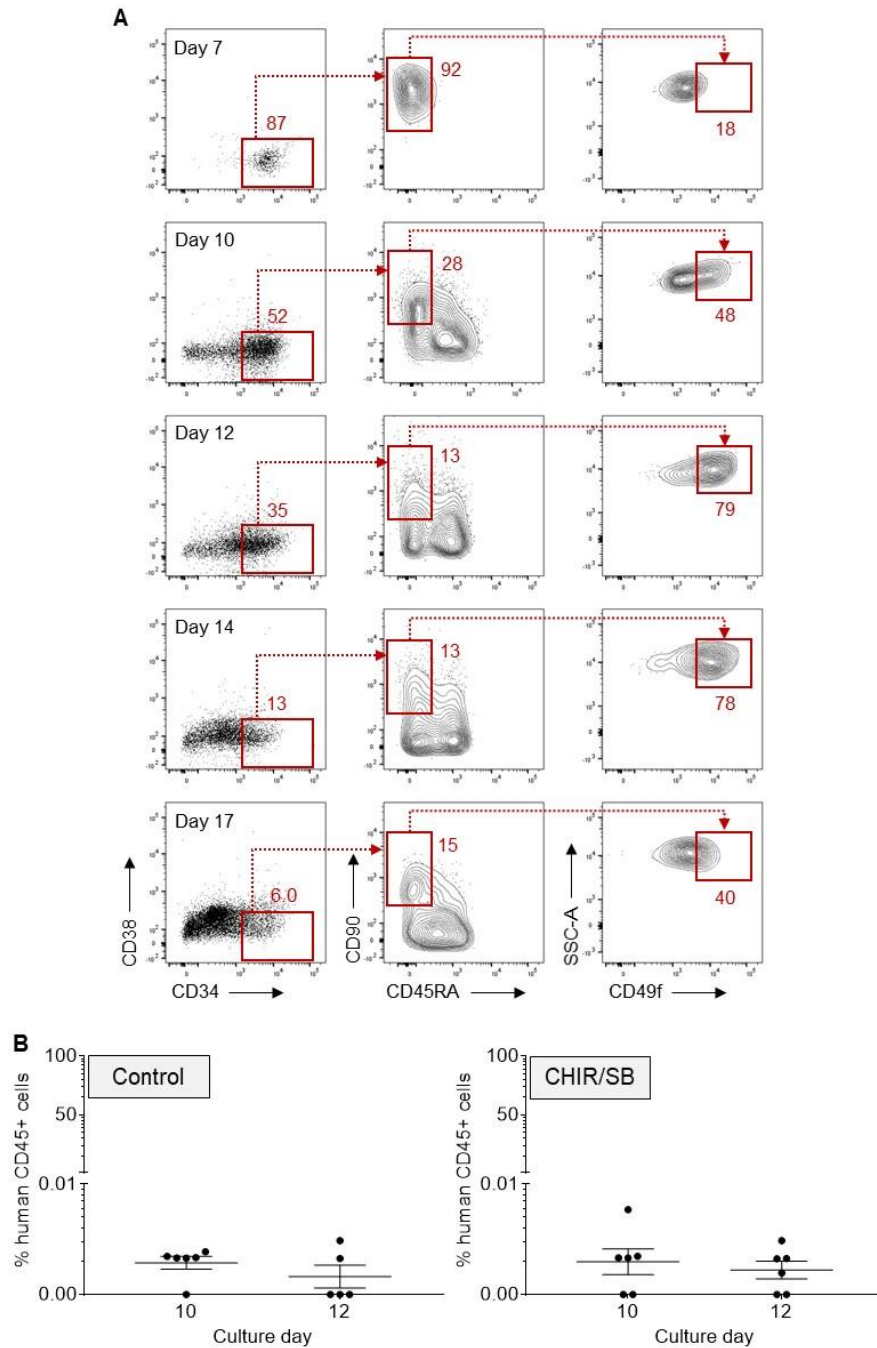
**Figure S1 | Complete gating strategies used for flow cytometry characterization of the cellular constituents arising during human iPSC differentiation with STEMdiff™ hematopoietic kit. (A)** Representative gating strategy for hematopoietic and non-hematopoietic fractions at day 12 of differentiation; associated with Fig. 1C. **(B)** Representative gating strategy for hematopoietic populations at day 10 of differentiation; associated with Fig. 2A. **(C)** Representative gating strategy for populations of mesenchymal cells, vascular endothelium (VE) and hemogenic endothelium (HE) at day 5 of differentiation; associated with Fig. 3A. **(D)** Representative gating strategy for populations of mesenchymal stromal cells (MSC) at day 10 of differentiation; associated with Fig. 3C. **(E)** Representative gating strategy for populations of arterial and venous VE at day 5 of differentiation; associated with Fig. 3F. **(F)** Representative gating strategy for populations of immunophenotypic HSCs at day 12 of differentiation; associated with Fig. S4A.



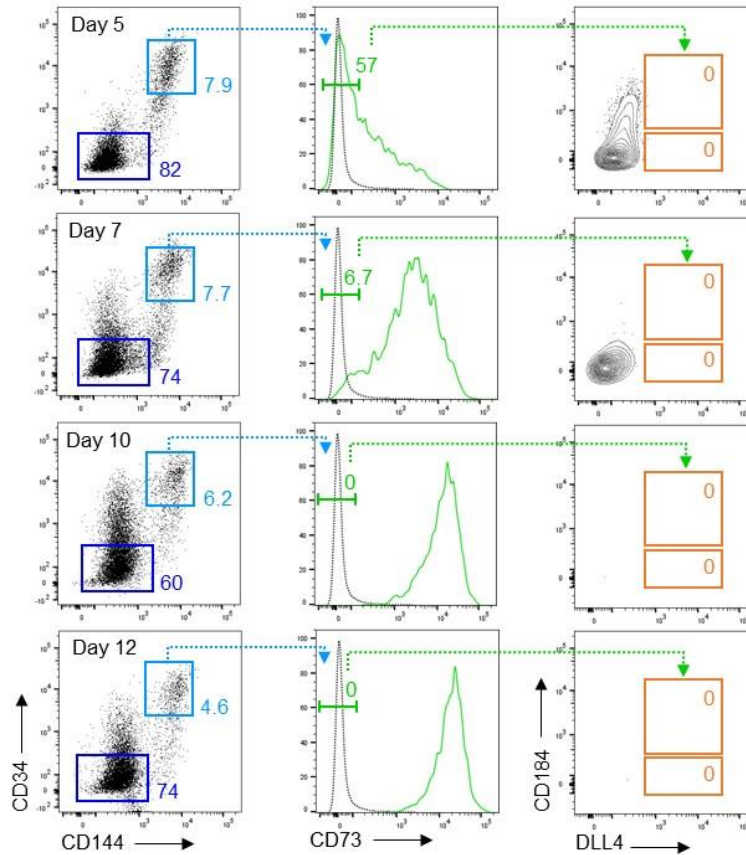
**Figure S2 | Characterization of MCND-TENS2 iPSC line.** (A) Hematoxylin and eosin stain of a teratoma from MCND-TENS2 iPSCs, displaying structures representative of ectoderm, mesoderm and endoderm. E, epidermal tissue; N, neural tissue; M, striated muscle; C, cartilage; A, adipose tissue; G, gut epithelial tissue. Scale bar = 250  $\mu$ m. (B) Cytogenetic analysis showing a normal karyotype (46, XY). (C) Flow cytometry analysis of iPSCs showing expression of pluripotency protein markers, TRA-1-60 and NANOG, compared to isotype controls.



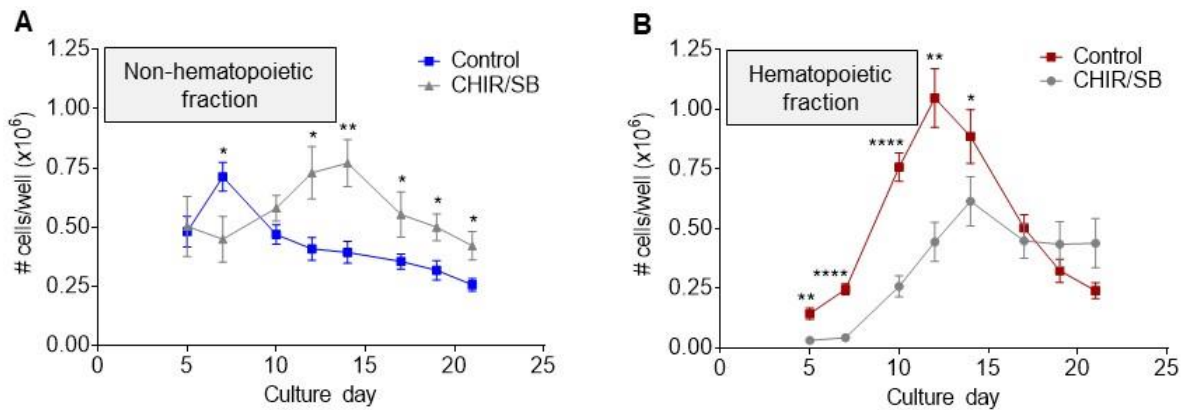
**Figure S3 | Cell viability during human iPSC differentiation with STEMdiff™ hematopoietic kit.** Total cell viability was measured by acridine orange (AO)/propidium iodide (PI) staining for each sample throughout differentiation. Results are shown as mean  $\pm$  SEM (n=8). Associated with Fig. 1E.



**Figure S4 | Production of immunophenotypic HSCs with no long-term engraftment potential during human iPSC differentiation with STEMdiff™ hematopoietic kit.** (A) Representative flow cytometry plots depicting  $CD34^+CD38^-CD45RA^-CD90^+CD49f^+$  cells in gated  $CD43^+CD45^+$  hematopoietic cells at various days of differentiation. The complete gating strategy is shown in Fig. S1F. (B) Human cell engraftment depicted as percentage of human CD45-expressing cells in the bone marrow of recipient NSG mice 16 weeks after transplantation of day 10 or 12 sorted  $CD43^+CD45^+$  hematopoietic cells derived from control (left panel, associated with Fig. 2E) or CHIR/SB-supplemented cultures (right panel, associated with Fig. 5G). Results are shown as mean  $\pm$  SEM; each dot represents an individual mouse (n= 5 to 6 mice per group).



**Figure S5 | Limited hemogenic endothelium (HE) and arterial-type HE formation during human iPSC differentiation with STEMdiff™ hematopoietic kit.** Representative flow cytometry plots of CD34, CD144 (VE-cadherin), CD73, DLL4 and CD184 expression in gated CD43<sup>-</sup>CD45<sup>-</sup> non-hematopoietic cells at various days of differentiation. Gates shown include: CD144<sup>-</sup>CD34<sup>-</sup> mesenchymal cells (dark blue), CD144<sup>+</sup>CD34<sup>hi</sup> vascular endothelium (VE, light blue), CD144<sup>+</sup>CD34<sup>hi</sup>CD73<sup>-</sup> hemogenic endothelium (HE, green), and CD144<sup>+</sup>CD34<sup>hi</sup>CD73<sup>-</sup>DLL4<sup>+</sup>CD184<sup>+/-</sup> arterial-type HE. The complete gating strategy is shown in Fig. S1C.



**Figure S6 | CHIR/SB molecules increase non-hematopoietic cells and decrease hematopoietic production during human iPSC differentiation with STEMdiff™ hematopoietic kit. (A)** Absolute numbers of CD43<sup>+</sup>CD45<sup>+</sup> hematopoietic cells arising from 20-35 iPSC clusters (one well of a 12-well plate) in control and CHIR/SB-supplemented cultures at various days of differentiation (n=6). Associated with Fig. 4A. **(B)** Absolute numbers of CD43<sup>-</sup>CD45<sup>-</sup> non-hematopoietic cells arising from 20-35 iPSC clusters (one well of a 12-well plate) in control and CHIR/SB-supplemented cultures at various days of differentiation (n=6). Associated with Fig. 5A. Results are displayed as mean  $\pm$  SEM. \*P<0.05, \*\*P < 0.01, \*\*\*\*P<0.0001, by two-way unpaired Student's t-tests comparing cell numbers in control vs CHIR/SB groups at each culture day.

**Table S1 | Human iPSC lines subjected to STEMdiff™ hematopoietic differentiation**

<b>Cell line</b>	<b>Type</b>	<b>Starting material</b>	<b>Reprogramming method</b>
<b>MCND-TENS1</b>	Normal control	MPB CD34 <sup>+</sup> CD38 <sup>-</sup> cells <sup>(1)</sup>	Sendai virus
<b>MCND-TENS2<sup>(2)</sup></b>	Normal control	MPB CD34 <sup>+</sup> CD38 <sup>-</sup> cells <sup>(1)</sup>	Sendai virus
<b>MCND-TENS3</b>	Normal control	MPB CD34 <sup>+</sup> CD38 <sup>-</sup> cells <sup>(1)</sup>	Sendai virus
<b>MCND-S1</b>	Normal control	MPB CD34 <sup>+</sup> cells <sup>(1)</sup>	Sendai virus
<b>MCND-S3</b>	Normal control	MPB CD34 <sup>+</sup> cells <sup>(1)</sup>	Sendai virus
<b>NL-5</b>	Normal control	Cord blood CD34 <sup>+</sup> cells	Episomal plasmid
<b>ND-2</b>	Normal control	Fibroblasts	Episomal plasmid
<b>NC-1</b>	Normal control	Fibroblasts	Episomal plasmid
<b>NC-4</b>	Normal control	Fibroblasts	Episomal plasmid
<b>NC-5</b>	Normal control	Fibroblasts	Episomal plasmid
<b>113-7</b>	Normal control	Fibroblasts	Episomal plasmid
<b>DBA863-S13</b>	DBA <sup>(3)</sup>	Mononuclear blood cells	Sendai virus
<b>DBA869-S1</b>	DBA <sup>(3)</sup>	Mononuclear blood cells	Sendai virus
<b>DBA872-S1</b>	DBA <sup>(3)</sup>	Mononuclear blood cells	Sendai virus

<sup>(1)</sup> Derived from G-CSF mobilized peripheral blood (MPB)

<sup>(2)</sup> Used in this study

<sup>(3)</sup> Diamond Blackfan Anemia (DBA)



**Table S2 | Antibodies for flow cytometry analysis and FACS**

<b>Antigen</b>	<b>Fluorochrome</b>	<b>Company/Catalogue #</b>	<b>Species</b>	<b>μL/test</b>
<b>CD34</b>	PE-Cy7	BD Pharmingen 560710	Mouse anti-human	10
<b>CD34</b>	PE	BD Pharmingen 555822	Mouse anti-human	20
<b>CD38</b>	APC	BD Pharmingen 555462	Mouse anti-human	20
<b>CD43</b>	BV711	BD OptiBuild™ 743614	Mouse anti-human	5
<b>CD43</b>	FITC	BD Pharmingen 555475	Mouse anti-human	20
<b>CD45</b>	V450	BD Horizon 560367	Mouse anti-human	5
<b>CD45RA</b>	APC-H7	BD Pharmingen 560674	Mouse anti-human	5
<b>CD49f</b>	PE-Cy5	BD Pharmingen 551129	Rat anti-human	20
<b>CD73</b>	PE	BD Pharmingen 550257	Mouse anti-human	20
<b>CD73</b>	FITC	BD Pharmingen 561254	Mouse anti-human	5
<b>CD90</b>	PE-Cy7	BD Pharmingen 561558	Mouse anti-human	5
<b>CD105</b>	AF647	BD Pharmingen 561439	Mouse anti-human	5
<b>CD144</b>	BV605	BD OptiBuild™ 743705	Mouse anti-human	5
<b>CD184</b>	PE-CF594	BD Horizon 562389	Mouse anti-human	5
<b>CD235a</b>	FITC	BioLegend 349104	Mouse anti-human	5
<b>N/A</b>	7-AAD	Thermofisher 00-6993-50	N/A	5

**Table S3 | Taqman™ primers and probes**

<b>Gene</b>	<b>Assay or sequence</b>	<b>Probe</b>
<b>GAPDH</b>	Life Technologies, Hs03929097_g1	VIC
<b>HoxA5</b>	Life Technologies, Hs00430330_m1	FAM
<b>HoxA9</b>	Life Technologies, Hs00365956_m1	FAM
<b>HoxA10</b>	Life Technologies, Hs00172012_m1	FAM