

Figure 2—Source data 2. Thirty-four residue pairs shown conserved rearrangements of residue contacts upon activation.

This PDF file includes:

Auxiliary Supplementary Figs. S1 to S8

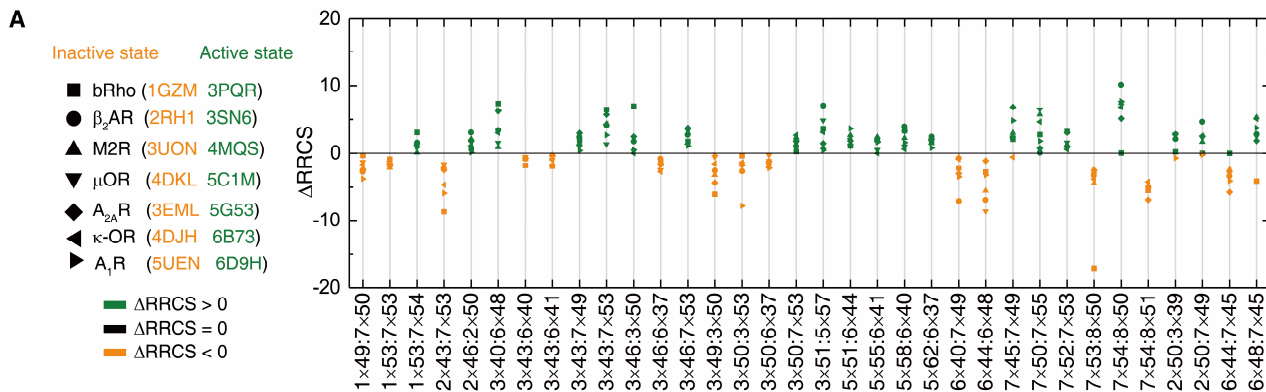


Fig. S1. Residue pairs with conserved rearrangements of residue contacts upon activation.

(A) For 7 receptors with both inactive- and active-state structures available (Rho, β_2 AR, M2R, μ OR, A_2 AR, κ -OR and recently determined A_1 R), 34 intra-receptor residue pairs showed almost identical tendency of Δ RRCS upon activation, *i.e.*, their Δ RRCS are almost entirely positive or negative. Considering there is no sodium pocket for rhodopsin, four related residue pairs around sodium pocket ($2 \times 46:2 \times 50$, $2 \times 50:3 \times 39$, $2 \times 50:7 \times 49$ and $6 \times 44:7 \times 45$) were neglected for bRho. (B) Four residue pairs (see Supplementary Figs. 2-8 for the rest 30 residue pairs) have significant different RRCS values between inactive- and active- states, *i.e.*, they evolve similar conformational changes upon receptor activation at the RRCS-level (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active-state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

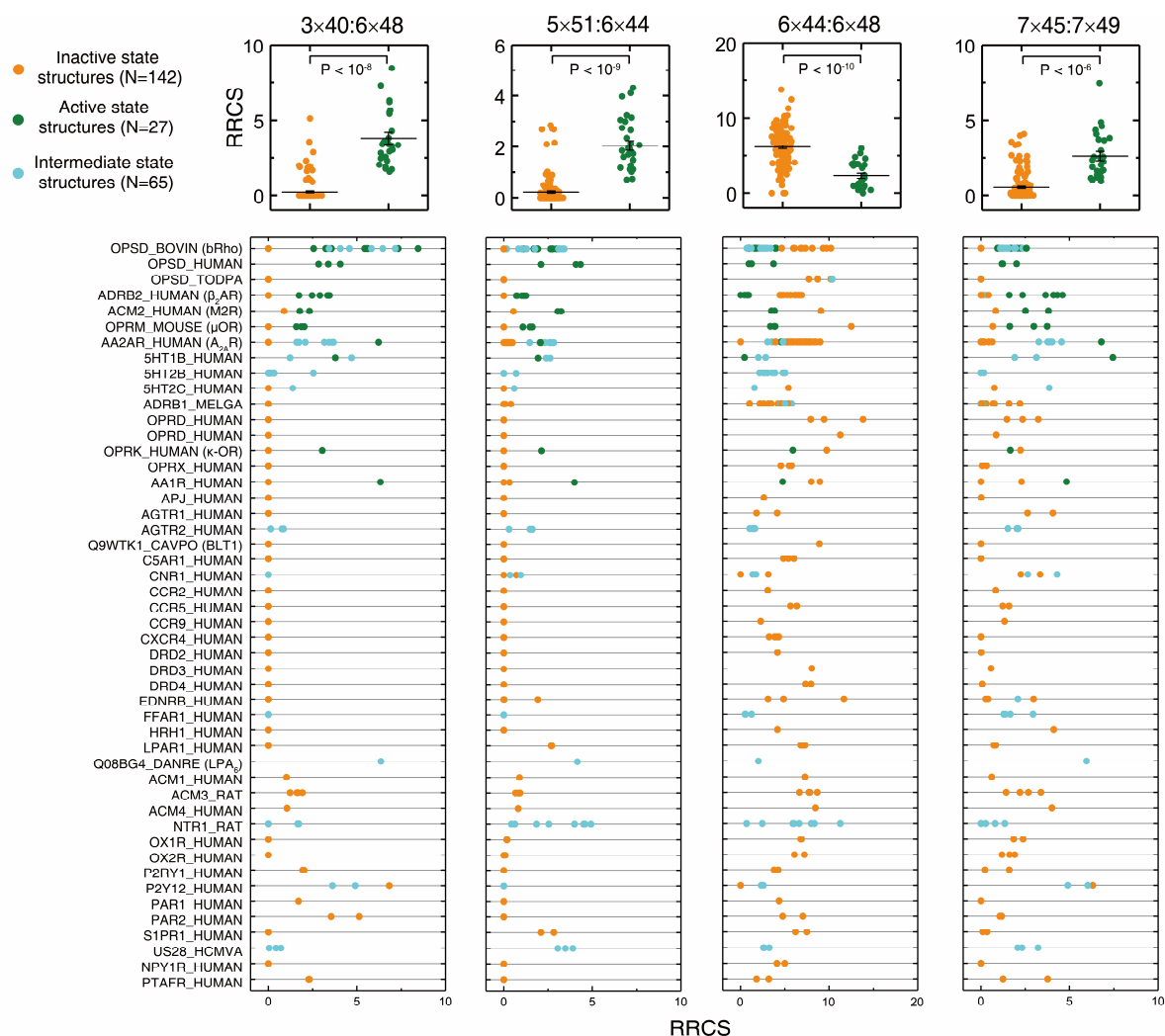


Fig. S2. Conserved signal initiation step involving changes in residue contacts at the bottom of the ligand-binding pocket. These residue pairs have significant different RRCS between inactive- and active- states, i.e., they evolve similar rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

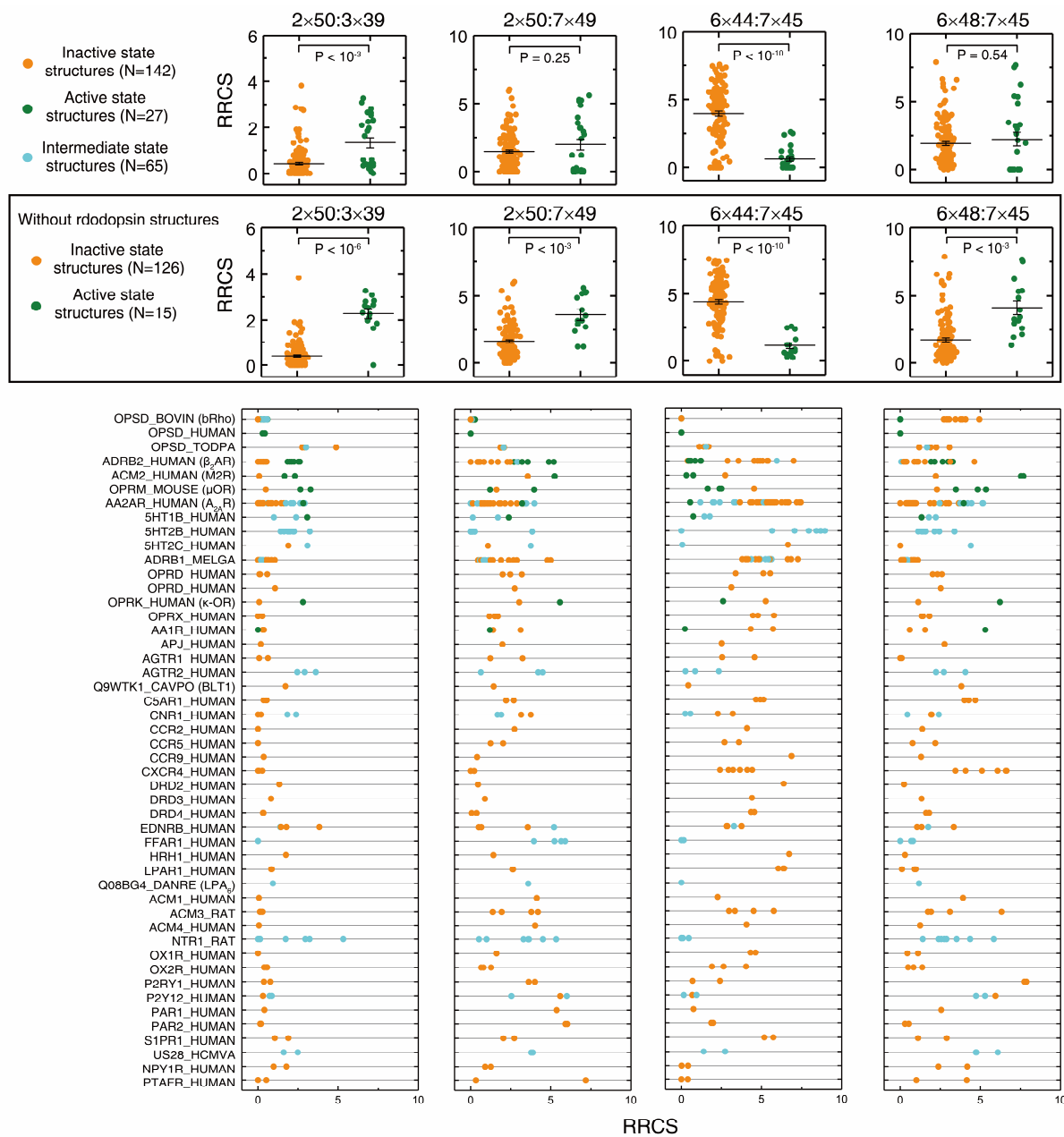


Fig. S3. Residue pairs around sodium pocket share conserved rearrangements of residue contacts upon activation. Considering there is no sodium pocket (residues at 2×50 , 3×39 , 7×45 and 7×49) for rhodopsin, four related residue pairs ($2 \times 46:2 \times 50$, $2 \times 50:3 \times 39$, $2 \times 50:7 \times 49$ and $6 \times 44:7 \times 45$) were omitted for rhodopsin. These residue pairs have significant different RRCS between inactive- and active- state, i.e., they evolve rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean).

The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

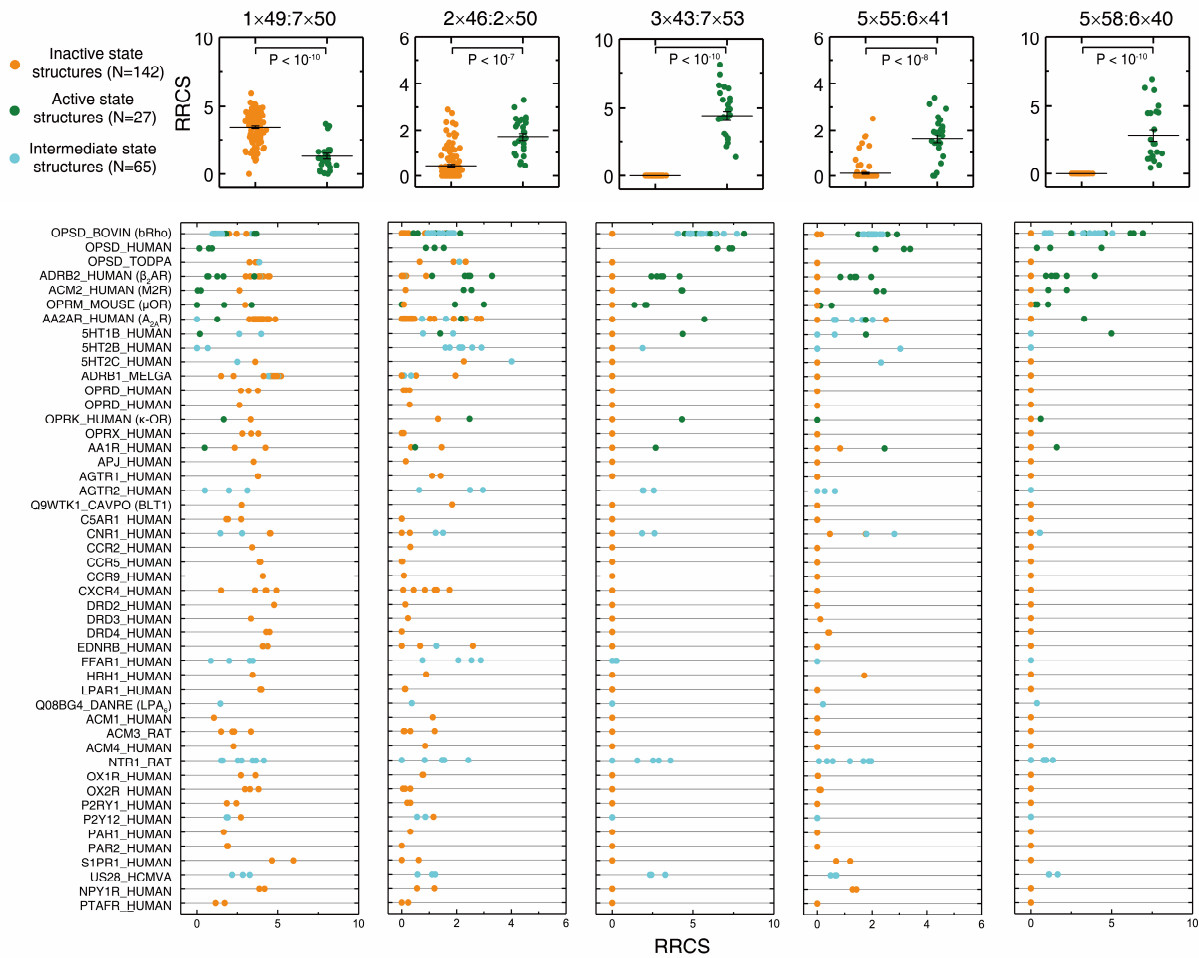


Fig. S4. Residue pairs around hydrophobic lock (3 × 43, 6 × 40 and 6 × 41) in layer 2 share conserved rearrangements of residue contacts upon activation. These residue pairs have significant different RRCS between inactive- and active- states, i.e., they evolve rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

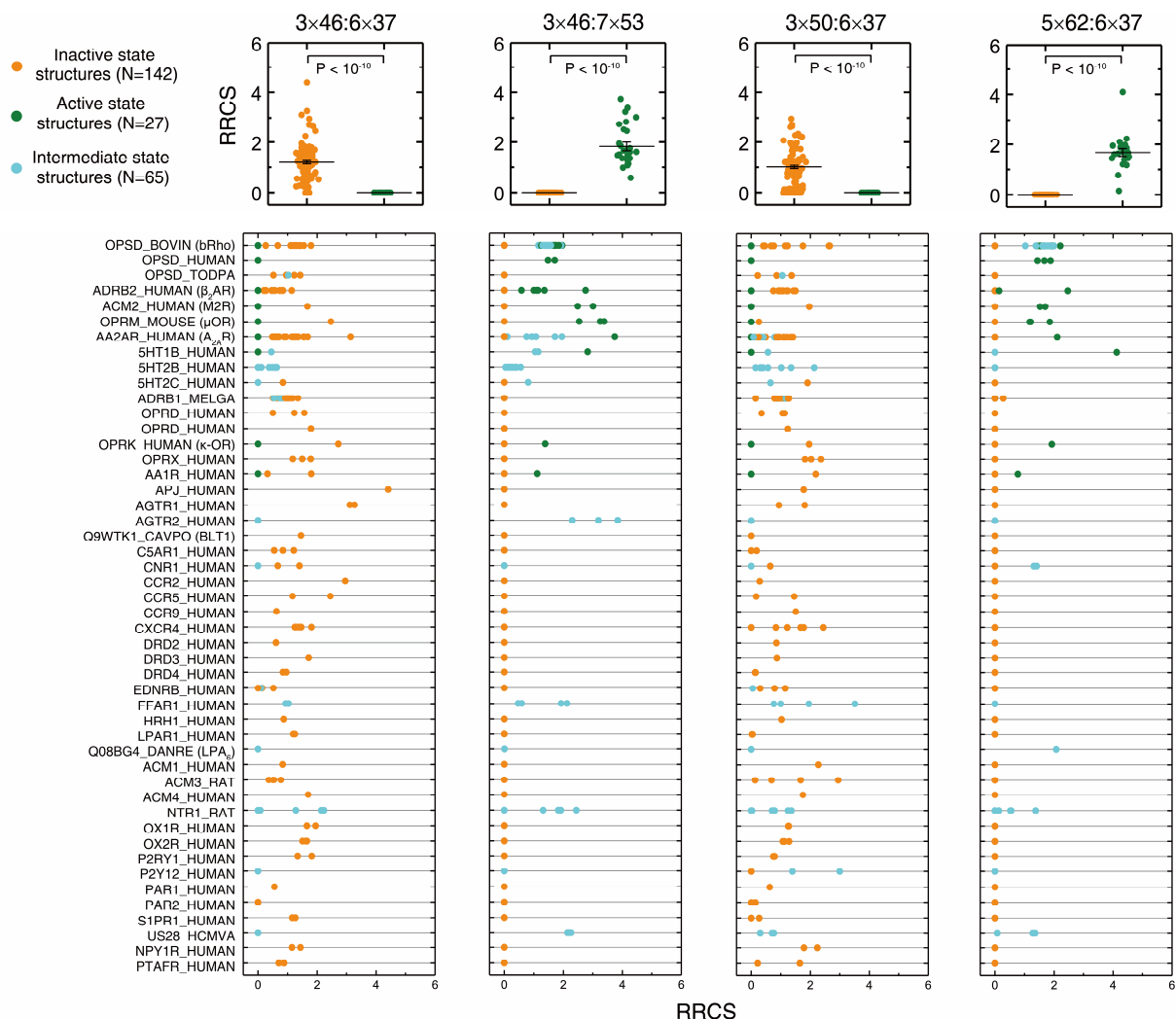


Fig. S5. Microswitch residues (3×46 , 6×37) share conserved rearrangements of residue contacts upon activation. These residue pairs have significant different RRCS between inactive- and active- state, i.e., they evolve similar rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

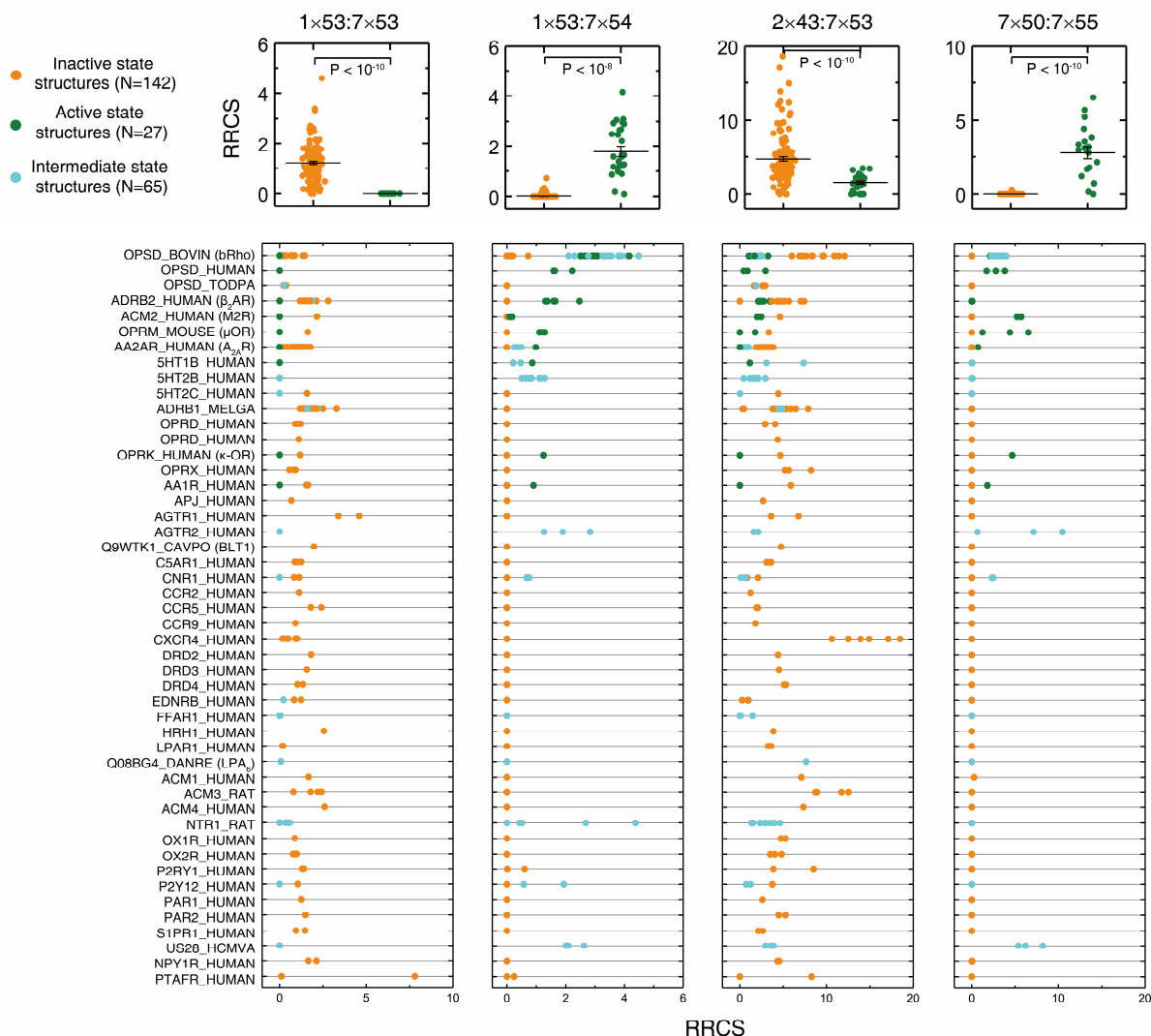


Fig. S6. Translocation of Y^{7x53} during receptor activation. These residue pairs around Y^{7x53} have significant different RRCS between inactive- and active- states, i.e., they evolve similar rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

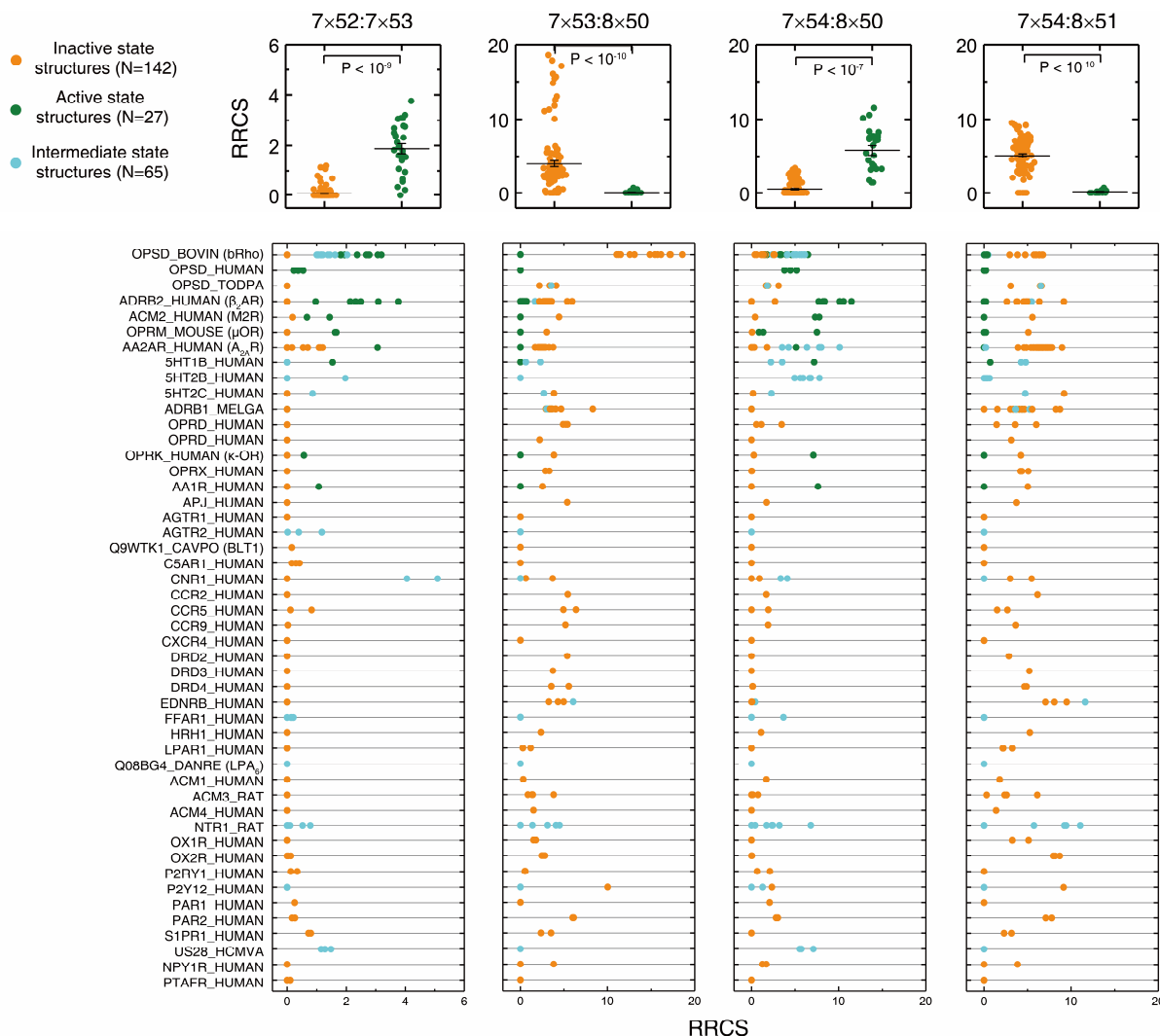


Fig. S7. Residue pairs around $Y^{7 \times 53}$ share conserved rearrangements of residue contacts upon activation. These residue pairs have significant different RRCS between inactive- and active- state, i.e., they evolve similar rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.

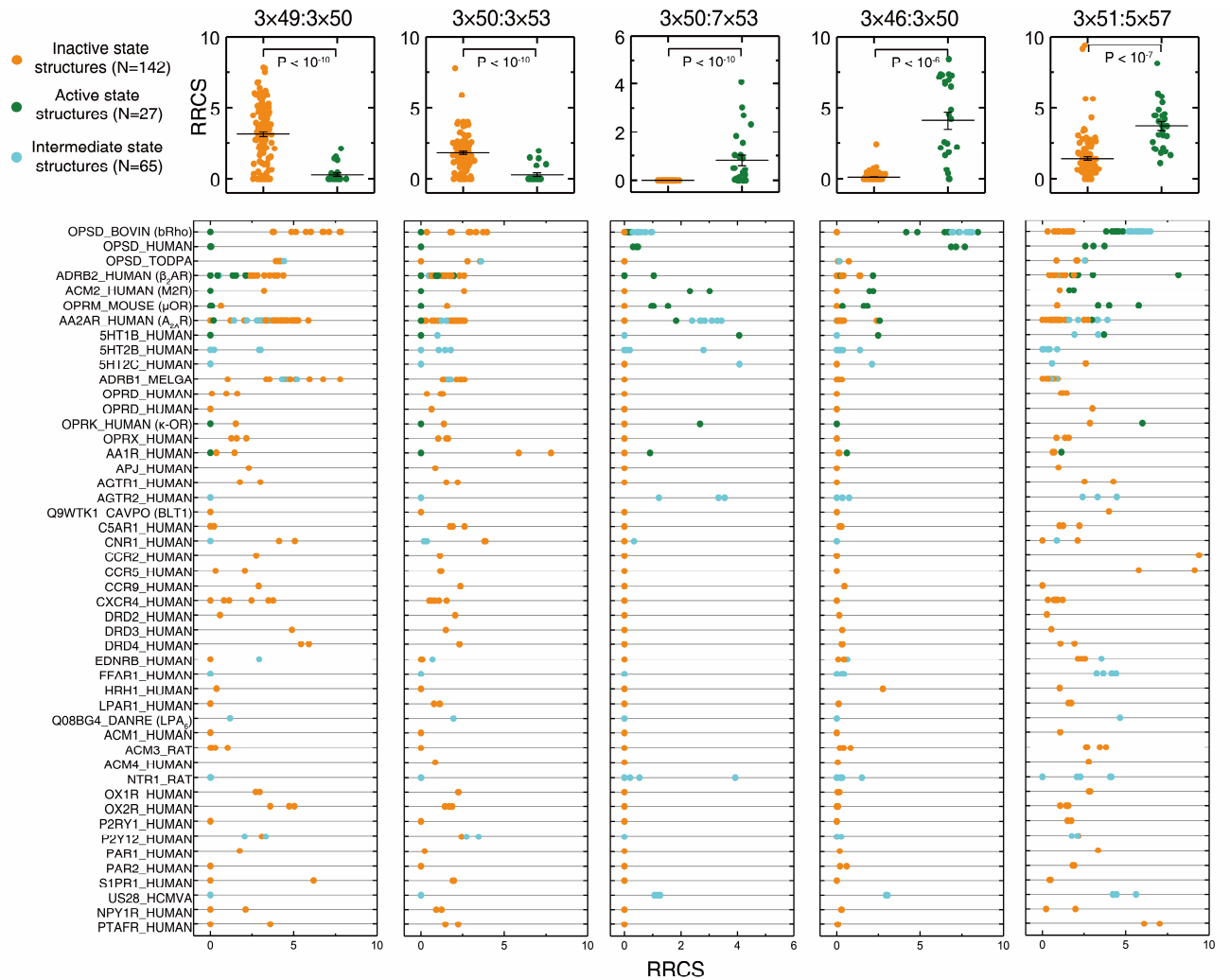


Fig. S8. Residue pairs around G protein-coupling region share conserved rearrangements of residue contacts upon activation. These residue pairs have significant different RRCS between inactive- and active- state, i.e., they evolve rearrangements of residue contacts upon activation (two-sample *t*-test, error bars represent the standard error of the mean). The comparison of RRCS between inactive- and active- state were plotted in receptor-specific manner for 45 class A GPCRs with determined structures, where inactive, active and intermediate states are coloured in orange, cyan and green, respectively.