# **Supplementary Information**

## Type I fatty acid synthase (FAS) trapped in the octanoylbound state

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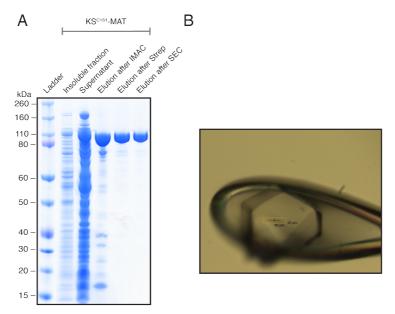
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## **Supplementary Tables**

Table S1: Dimerization interface of the KS domain

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**Figure S1: Purification and crystallization of the KS-MAT didomain.** (A) SDS-PAGE (NuPAGE 4-12 % Bis-Tris) of the purification strategy of the KS-MAT didomain. A tandem purification using Ni-chelating and Strep-Tactin affinity chromatography was followed by size exclusion. (B) Photograph of the octanoyl-CoA soaked crystal within a nylon loop at the synchrotron.

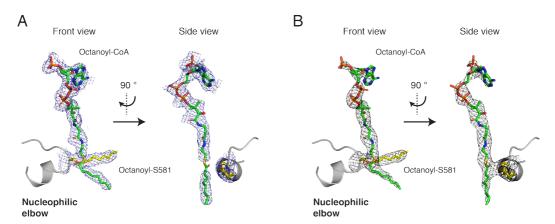


Figure S2: Validation of ligand placement in the MAT domain. Octanoyl-CoA and the covalent bound octanoyl-S581 were placed based on unbiased electron density maps. The FEM map (A) is shown in blue and the Polder map (B) in black. Contour levels are at 3  $\sigma$ .

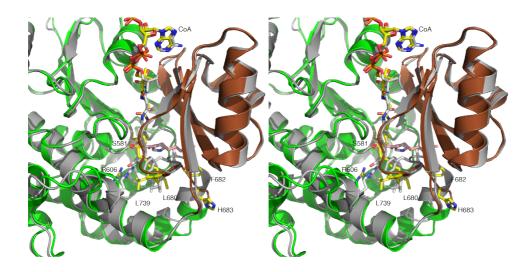
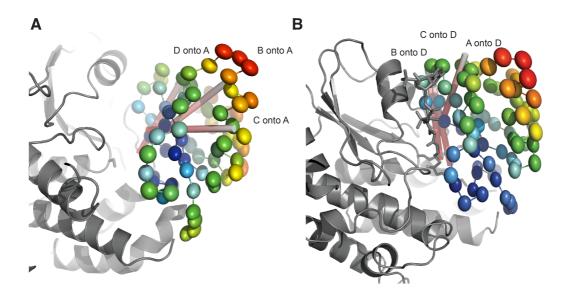


Figure S3: Stereo cartoon diagram to illustrate conformational changes in the MAT domain. Substrate bound chains (D) of the octanoyl-bound and malonyl-bound (grey) structures superimposed based on the  $\alpha/\beta$ -hydrolase fold. Residues S581, R606, L680, F682, H683 and L739 and bound moieties are shown in stick representation. Green and brown colouring indicate the regions of  $\alpha/\beta$ -hydrolase fold and the ferredoxin-like fold respectively for the octanoyl-bound model.



**Figure S4:** Anisotropic movement of C-alpha atoms in the ferredoxin-like subdomain. The anisotropic movement, as derived from the TLS tensors, is depicted by thermal ellipsoids colored by B-values (blue – low values; and red – high values) for the ferredoxin-like fold (616-684) region of chain A (A) and chain D (B). Rotation axes derived from superposition of ferredoxin-like fold (616-684) of chains B (A), C (B), and D (C) onto A (D) are shown in redwhite cylindrical bars. For orientation, the rest of the KS-MAT chain is shown in grey.

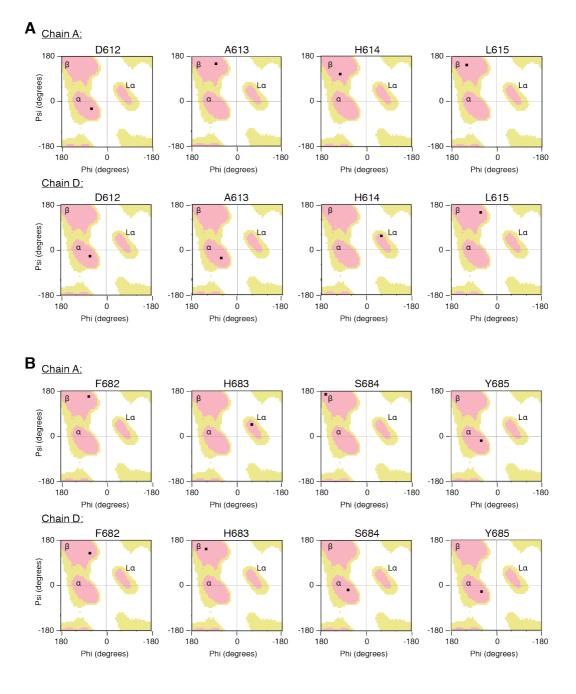


Figure S5: Ramachandran plots for crucial residues in both subdomain linkers. Significant changes in dihedral angles were seen for residues in linker 1 (A) 612-615 and linker 2 (B) 682-685 leading to changes to different allowed regions. Plots were created in coot. Used abbreviations indicate  $\alpha$  – right-handed helical region, L $\alpha$  – left-handed helical region and  $\beta$  –  $\beta$ -sheet region.

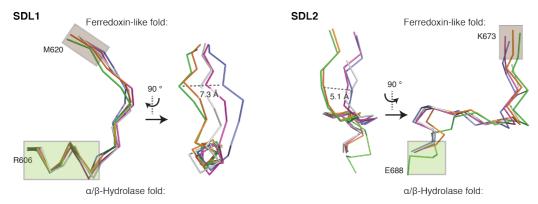
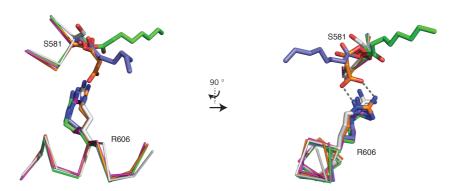
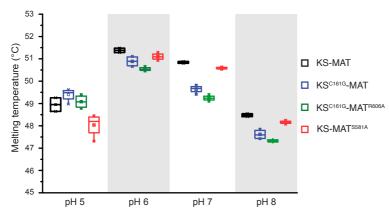


Figure S6: Dynamics of the MAT domain is determined by flexible subdomain linkers. SDL1 (612-617, left panel) and SDL2 (675-684, right panel) are shown as ribbons after  $\alpha/\beta$ -hydrolase fold based alignments. Both linker stretches are shown from two different perspectives rotated by 90°. Chain A (blue), chain C (grey) and chain D (green) from the octanoyl-CoA soaked crystal (PDB code 6rop), malonyl-bound (orange) (PDB code 5my0; chain D) and apo human MAT (purple) (PDB code 3hhd; chain A) were used. Green and brown rectangles indicate secondary structure elements of the  $\alpha/\beta$ -hydrolase- and ferredoxin-like subdomains, respectively.



**Figure S7: Rotational freedom of residue R606.** α/β-Hydrolase based superposition (BB of 488-615) of chain A (blue), chain C (white) and chain D (green) from octanoyl-CoA soaked crystals (PDB code 6rop) with the malonyl-bound structure (orange) (PDB code 5my0; chain D) and of porcine FAS (purple) (PDB code 2vz9; chain A). Important residues S581 and R606 are shown in sticks with covalent modifications of the serine represented also in sticks. For clarity residue stretch 580-583 and 601-610 are depicted as ribbons. R606 in octanoyl-bound active sites adopts the same conformation as R606 in the porcine FAS, whereas R606 in chain C (unbound) possesses the rotameric state of the unbound active sites previously found in human KS-MAT (3hhd).



**Figure S8**: **Stability of select KS-MAT variants.** Melting temperatures were determined by a thermal shift assay as described in the Methods section. The four constructs were tested in phosphate buffers at different pH value. Four replicates are shown as dot plot.

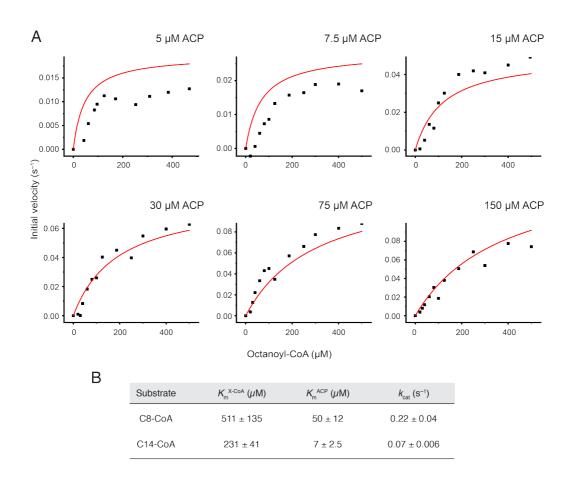


Figure S9: Global Michaelis-Menten fit of KS-mediated transacylation data. (A) Initial velocities were plotted against octanoyl-CoA (C8-CoA) concentrations at six fixed ACP concentrations. Data were fit globally with the Michaelis-Menten equation assuming a pingpong bi-bi mechanism. (B) Absolute kinetic parameter derived from the respective global fits for C8- and C14-CoA, respectively. No parameters constraints were set for the fit function.

Table S1: Dimerization interface of the KS domain

Interface	Residues per side		Solvent-accessible interface area		Solvent energy gain per side	Hydrogen bonds	Salt bridges
	#	%	$\mathring{A}^2$	%	kcal/mol		
KS chain A	73	8.6	2583	8.1	-14.9	38	0
KS chain B	71	8.5	2578	8.0	-14.8		0
KS chain C	72	8.5	2586	7.9	-14.5	34	0
KS chain D	75	8.8	2582	8.1	-15.0		0

Table S2: Primers and Plasmids

Number	Name	Length	Orientation	Sequence (5'3')	Construct Template
AR301	mMAT_S580X_rev	20	reverse	gtgcccaatgatgccgtcag	pAR69/pAR70
AR310	mMAT_S580A_for	38	forward	ggcatcattgggcacGccttgggagaggttgcctgtgg	pAR69/pAR70

Number	Construct	Name
pAR069*	KS-MAT	pAR69_STRI_m(KS_MAT)_H8_pET22b
pAR070*	KS <sup>C161G</sup> -MAT	pAR70_StrepI_m(KS(C161G)_MAT)_H8_pET22b
pAR071*	KS <sup>C161G</sup> -MAT <sup>R606A</sup>	pAR71_StrepI_m(KS(C161G)_ATmut(R(606A))_H8_pET22b
pAR159	KS-MAT <sup>S581A</sup>	pAR159_StrepI_m(KS_MAT(S581A))_H8_pET22b
pAR160	KS <sup>C161G</sup> -MAT <sup>S581A</sup>	pAR160_StrepI_m(KS(C161G)_MAT(S581A))_H8_pET22b
pAR352*	(holo)-ACP	pAR352_StrepII_mACP_H8_RBS_SFP_pET22b