Chronic IL-1β-induced inflammation regulates epithelial-to-mesenchymal transition memory phenotypes via epigenetic modification in non-small cell lung cancer

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GAPDH

CK18

GAPDH

В

Figure S1 related to Figure 1. Chronic cytokine exposure leads to EMT memory in NSCLC. A, Phase-contrast microscopy showed decreased cell-to-cell contacts and increased cell protrusions (indicated by arrow head) following 48-hour IL-1 β exposure in A549 cells. **B**, Gene expression of indicated mesenchymal markers by RT-PCR. **C**, High dose of IL-1 β exposure did not lead to EMT memory following 9-day IL-1 β exposure in A549 cells. **D**, A screening of EMT in indicated NSCLC cell lines following 48-hour IL-1 β treatment. **E**, EMT markers in A427 and H460 cell lines following 48-hour IL-1 β exposure. E-cadherin was not detected in these two cell lines. **F**, Prolonged EMT phenotype was observed in A427 and H460 following chronic IL-1 β exposure. L, low dose (1 ng/ml); H, high dose (10 ng/ml). "p", previously treated with IL-1 β . All results were reported as mean ± SEM. ** *P*<0.002, **** *P*<0.0001.



Figure S2 related to Figure 2. EMT-associated phenotypes induced by chronic IL-1β exposure are also memorized. A, Chemotherapy-induced apoptosis following the acute IL-1β exposure in H460 and A427 cells. **B**, Surface PD-L1 expression following the acute IL-1β exposure in H460 and A427 cells. **C**, A heatmap of differentially expressed genes at indicated time points following IL-1β exposure, which were divided into four clusters. C1 and C4, genes sensitive to IL-1β exposure; C2 and C3, genes memorizing their expression patterns following IL-1β withdrawal. **D**, Pathway analysis in A549 cells showing that pathways, such as the NF-κB and AP-1 pathways, and phenotypes, such as cell differentiation and migration, are IL-1β sensitive. Cis, cisplatin; Etop, etoposide; Doxr, doxorubicin. All results were reported as mean \pm SEM. **** *P* <0.0001, n.s, not significant.



Ε





G



Figure S3 related to Figure 3. Accumulation of SLUG is indispensable for the establishment of the memory phenotypes. **A**, Relative expression of EMT-associated transcription repressors by RT-qPCR following the acute IL-1 β exposure. **B**, Knockdown of SLUG or ZEB2 is sufficient to prevent E-cadherin downregulation following the acute IL-1 β exposure. **C**, Morphological changes of the untreated, LS, and HS cells by bright-field microscopy. **D**, Morphological changes with various levels of SLUG at day 21 of SLUG overexpression (21d) and day 7 after the overexpression was terminated (-7d). **E-G**, Evaluation of EMT markers, apoptosis resistance, and surface PD-L1 expression with various levels of SLUG overexpression at indicated time points. "p", previously treated with IL-1 β . "oe", overexpression. Cis, cisplatin; Etop, etoposide; Doxr, doxorubicin. All results were reported as mean \pm SEM. * *P*<0.003, ** *P*<0.002, **** *P*<0.0002, **** *P*<0.0001, n.s, not significant.





Ε





F

30m

IL-1β

p-c-JUN

c-JUN

p-Fra-1

Fra-1

GAPDH

IL-1β p-ERK1/2 ERK1/2 SLUG GAPDH





Figure S4

Figure S4 related to Figure 4. Pathways mediating acute EMT are not required for the maintenance of chronic EMT or EMT memory. A, Signaling pathways activated by IL-1 β within 48 hours in A549 cells. B, A dose-dependent response of E-cadherin expression to the ERK or JNK inhibitor in acute EMT. C, Expression levels of the transcription factor AP-1 components by RT-qPCR at 48 hours following the IL-1 β treatment. D, Phosphorylation of Fra-1 and c-JUN upon 30 minutes of IL-1 β exposure, examined by immunoblotting. E, Relative expression of Fra-1, SLUG and ZEB2 with the JNK inhibition followed by the acute IL-1 β exposure by RT-qPCR. F, Persistent activation of the ERK pathway in chronic EMT and EMT memory. G, EMT markers following 72-hour inhibition. U0126, ERK pathway inhibitor; JNKi II, JNK inhibitor; "U+J", U0126 and JNKi II combination treatment. "-d", days after IL-1 β withdrawal; "p", previously treated with IL-1 β . "n.d.", non-detectable. All results were reported as mean ± SEM. ** *P*<0.002, **** *P*<0.0001, N.D., not detectable.

Α

В







HDACs



DNMTs



С

Relative Expression



Figure S5 related to Figure 5. Dynamic epigenetic modifications of the *CDH1* **promoter in IL-1β-induced EMT. A**, Increased DNA methylation in the *CDH1* promoter upon the chronic TNF- α (10 ng/ml) or TGF- β (5 ng/ml) exposure. IL-1 β served as a positive control. **B**, RT-qPCR revealed the expression levels of various histone methyltransferases (HMTs), lysine demethylases (KDMs), histone deacetylases (HDACs), and DNA methyltransferases (DNMTs) following the IL-1 β treatment at indicated time points. **C**, HDAC9 knockdown did not affect E-cadherin expression in acute EMT. T α , TNF- α ; T β , TGF- β . M, methylated; U, unmethylated.











В

Figure S6 related to Figure 6. Blockade of epigenetic modifications restores E-cadherin expression and cellular sensitivity to chemotherapy in EMT memory. A, Epigenetic inhibitors were added following 4 days of IL-1 β exposure. TSA and EPZ reversed the downregulation of E-cadherin in acute EMT. Quantification of E-cadherin expression was normalized to samples without IL-1 β exposure. **B**, E-cadherin expression was evaluated following EZH2 knockdown for 72 hours in acute, chronic EMT, and EMT memory. **C**. E-cadherin expression in chronic EMT following SETDB1 or SUV39H1 knockdown for 72 hours. AZA, 5'-azacytidine-2'-deoxycytidine (decitabine), DNA methyltransferase inhibitor; TSA, pan HDAC inhibitor; EPZ, EPZ-6438, EZH2 inhibitor; BIX, BIX01294, G9a inhibitor; OG-L, OG-L002, LSD1 inhibitor. "p", previously treated with IL-1 β .

Table S1 related to Figure 2. EMT-associated phenotypes induced by chronic IL-1β exposure are also memorized. Differentially expressed gene upon IL-1β exposure.

Table S2 related to Figure 2. EMT-associated phenotypes induced by chronic IL-1 β exposure are also memorized. Pathway analysis based on the Molecular Signature Database.