

**AN OPTIMISED PROTOCOL HARNESSING LASER CAPTURE MICRODISSECTION
ON MATCHED PRIMARY AND METASTATIC COLORECTAL TUMOURS**

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The authors declare no conflict of interest.

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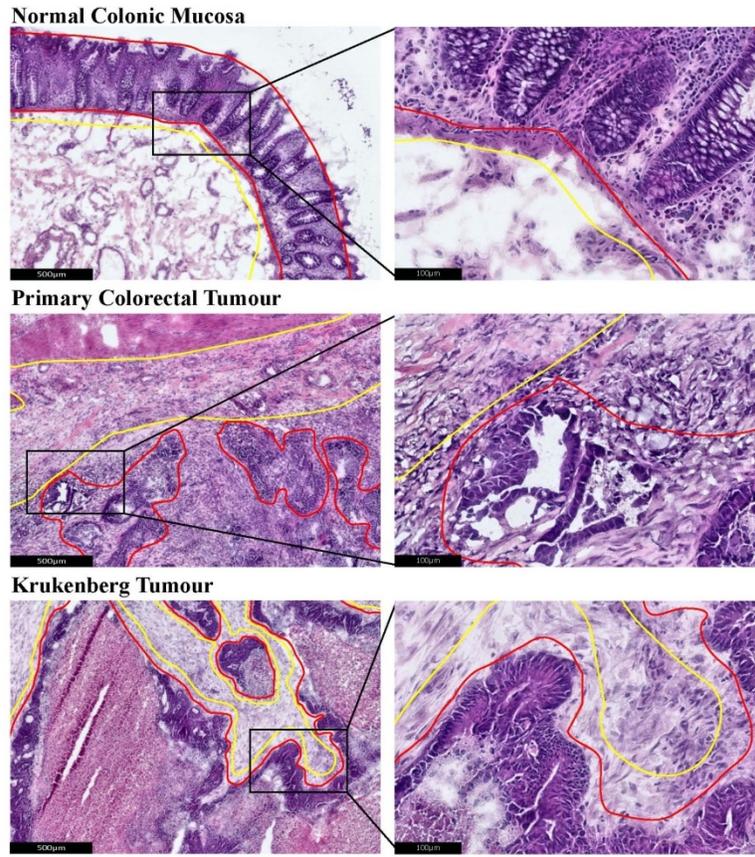
Supplementary Information

Supplementary Table S1: List of microdissected samples used in RNA-Seq analysis						
	Normal colonic mucosa		Primary colorectal tumour		Krukenberg tumour	
	Stroma	Normal Epithelium	Stroma	Normal Epithelium	Stroma	Normal Epithelium
Patient 1	1	1	1	1	1	1
Patient 2	1	1	1	1	2	2
Patient 3	0	0	1	1	1	1
Patient 4	0	0	0	0	1	1

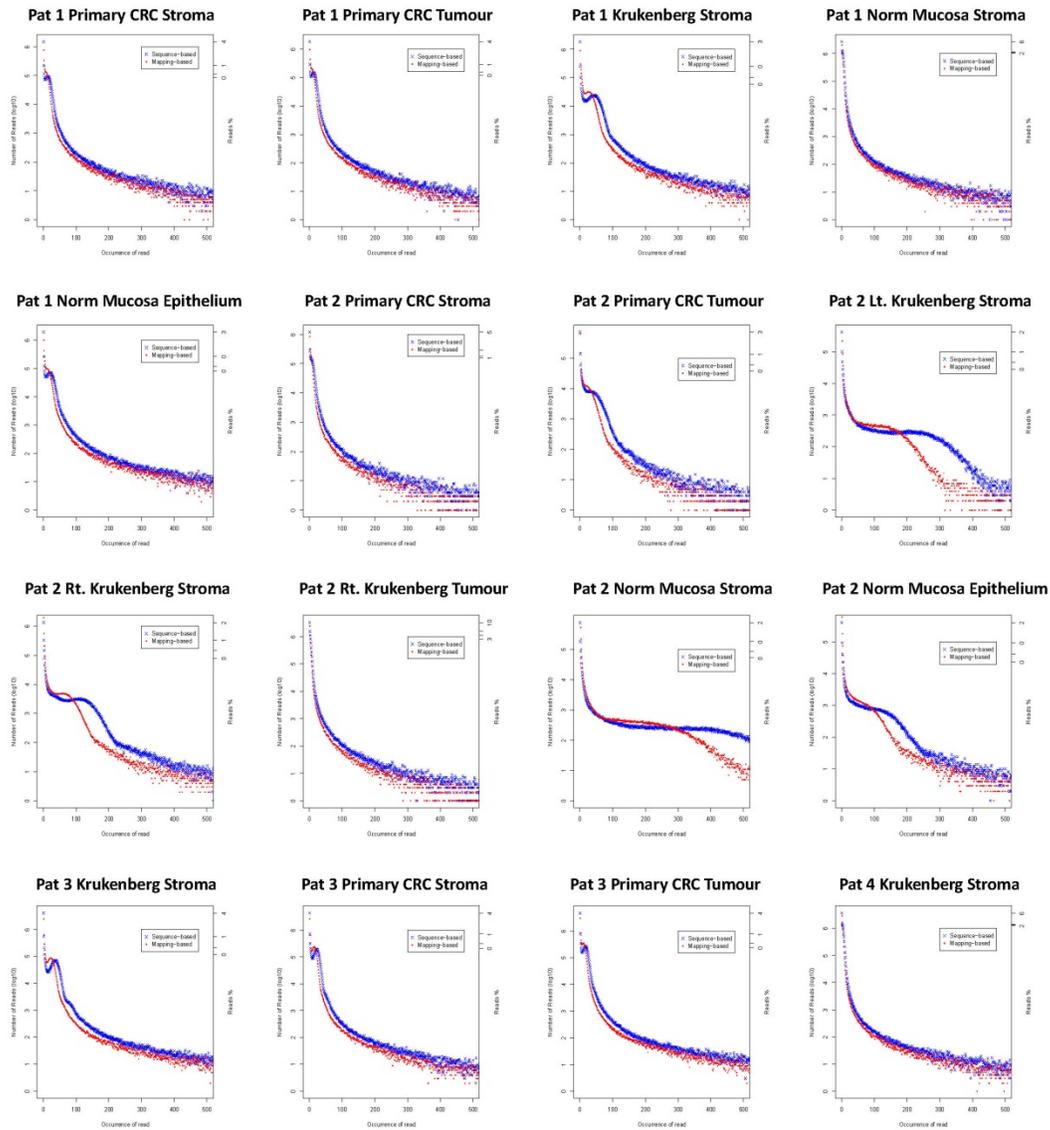
Supplementary Table S2: Gene expression patterns of selected targets	
Gene	Expression pattern
SPINK1	Higher expression in tumour epithelium compared to normal epithelium
ERBB2	Higher expression in metastatic tumour epithelium compared to normal epithelium and primary tumour epithelium
COL1A1	Higher expression in tumour-associated stroma compared to normal stroma
SPARC, TIMP1, IGFBP7	Higher expression in metastatic stroma compared to normal stroma and primary tumour-associated stroma
ENO1	Higher expression in metastatic epithelium and metastatic stroma compared to the respective tissue compartments in normal mucosa and primary tumour
VEGFA, S100A11	Higher expression in tumour epithelium and stroma compared to respective tissue compartments in normal mucosa
PLA2G2A	Lower expression in tumour epithelium compared to normal epithelium
CEACAM1	Lower expression in tumour compared to normal mucosa

Supplementary Table S3: List of primers

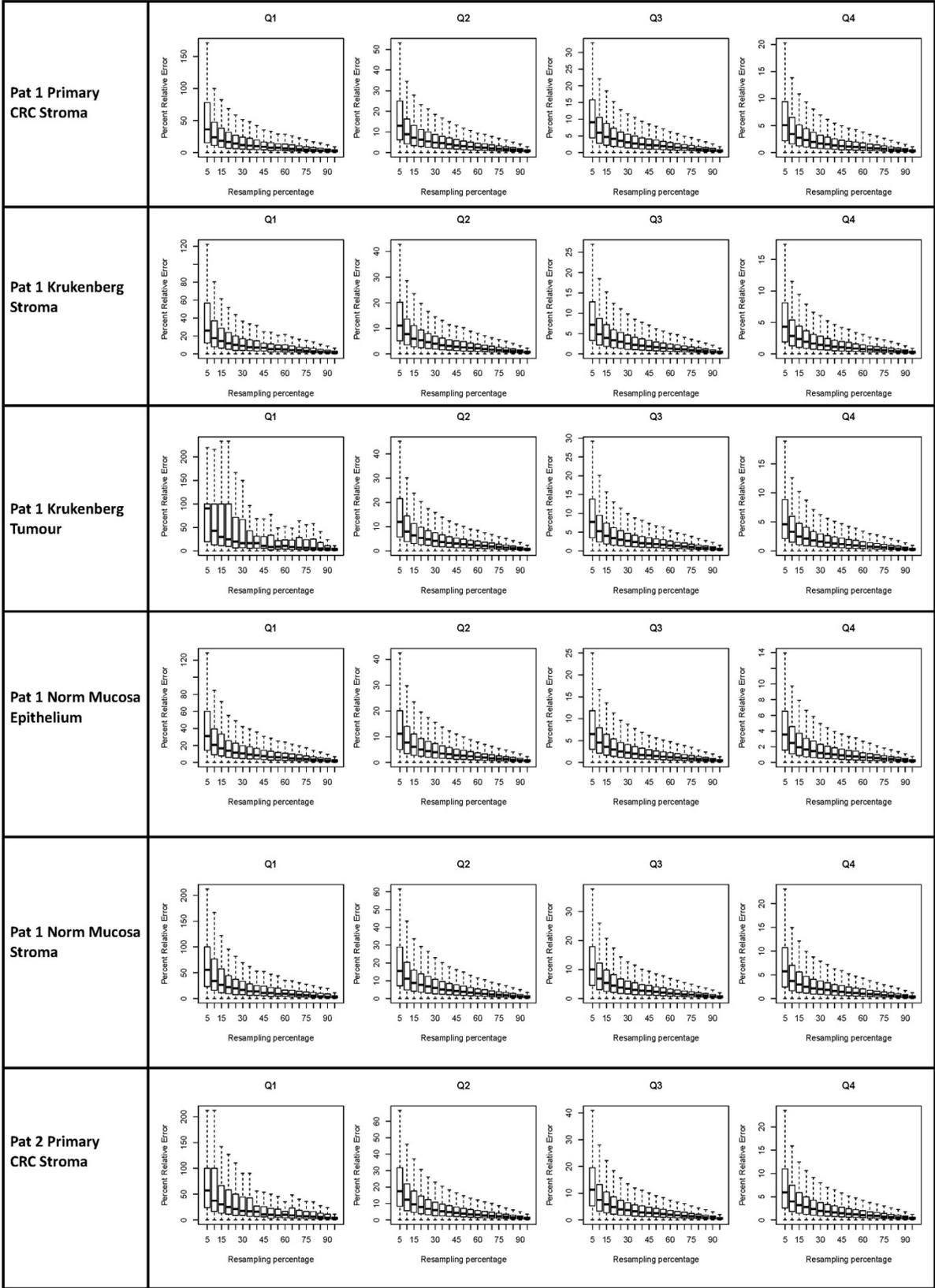
Gene ID	Primer Sequence 5' to 3'
GAPDH F	CAACAGCCTCAAGATCATCAG
GAPDH R	ATGGACTGTGGTCATGAGTC
β ACTIN F	TGTTTGAGACCTTCAACACC
β ACTIN R	AGGTAGTCAGTCAGGTCCCGGCC
ErbB2 F	ACTGGCCCTCATCCACCATA
ErbB2 R	GGTTGGCAGTGTGGAGCAG
SPINK1 F	TGTCTGTGGGACTGATGGAA
SPINK1 R	AGGCCCAAGATTTTTGAATGA
CEACAM1 F	AAGCCCCAAATCAAAGCCAG
CEACAM1 R	CAGCATCCTCCCTCTTGACA
SPARC F	AAACTTTTGGGAGCACGGAC
SPARC R	ACCGATTCACCAACTCCACT
TIMP-1 F	CCTTCTGCAATTCGACCTC
TIMP-1 R	GTATCCGCAGACACTCTCCA
IGFBP7 F	TGGGTGCTGGTATCTCCTCT
IGFBP7 R	TATAGCTCGGCACCTTCACC
COLIA1 F	ATGTGCCACTCTGACTGGAA
COLIA1 R	CTTGTCCTTGGGGTTCTTGC
ENO1 F	AAAGCTGGTGCCGTTGAGAAG
ENO1 R	AGCATGAGAACCGCCATTGAT
S100A11 F	TCTCCAGCCCTACAGAGACT
S100A11 R	TTCATCATGCGGTCAAGGAC

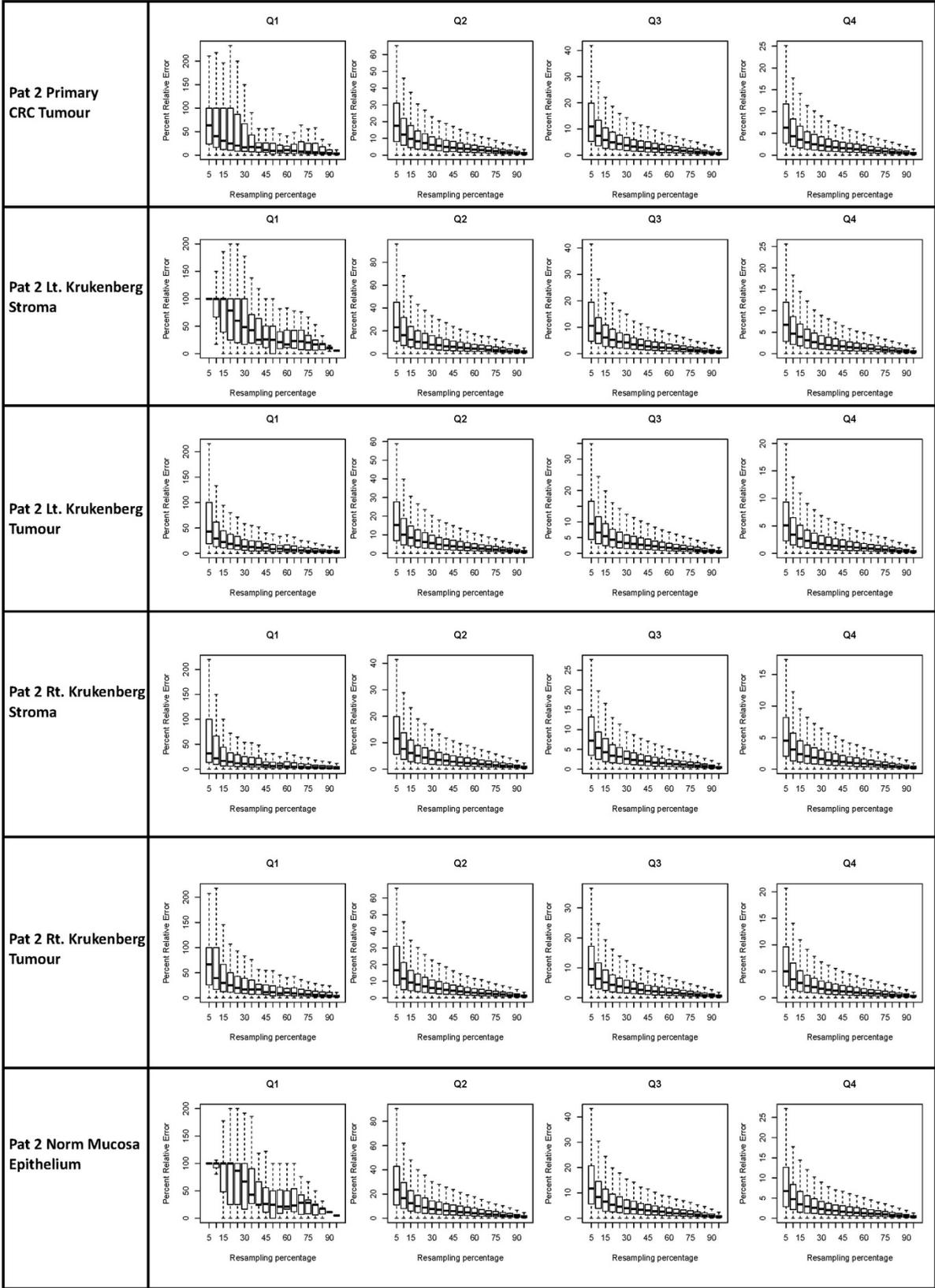


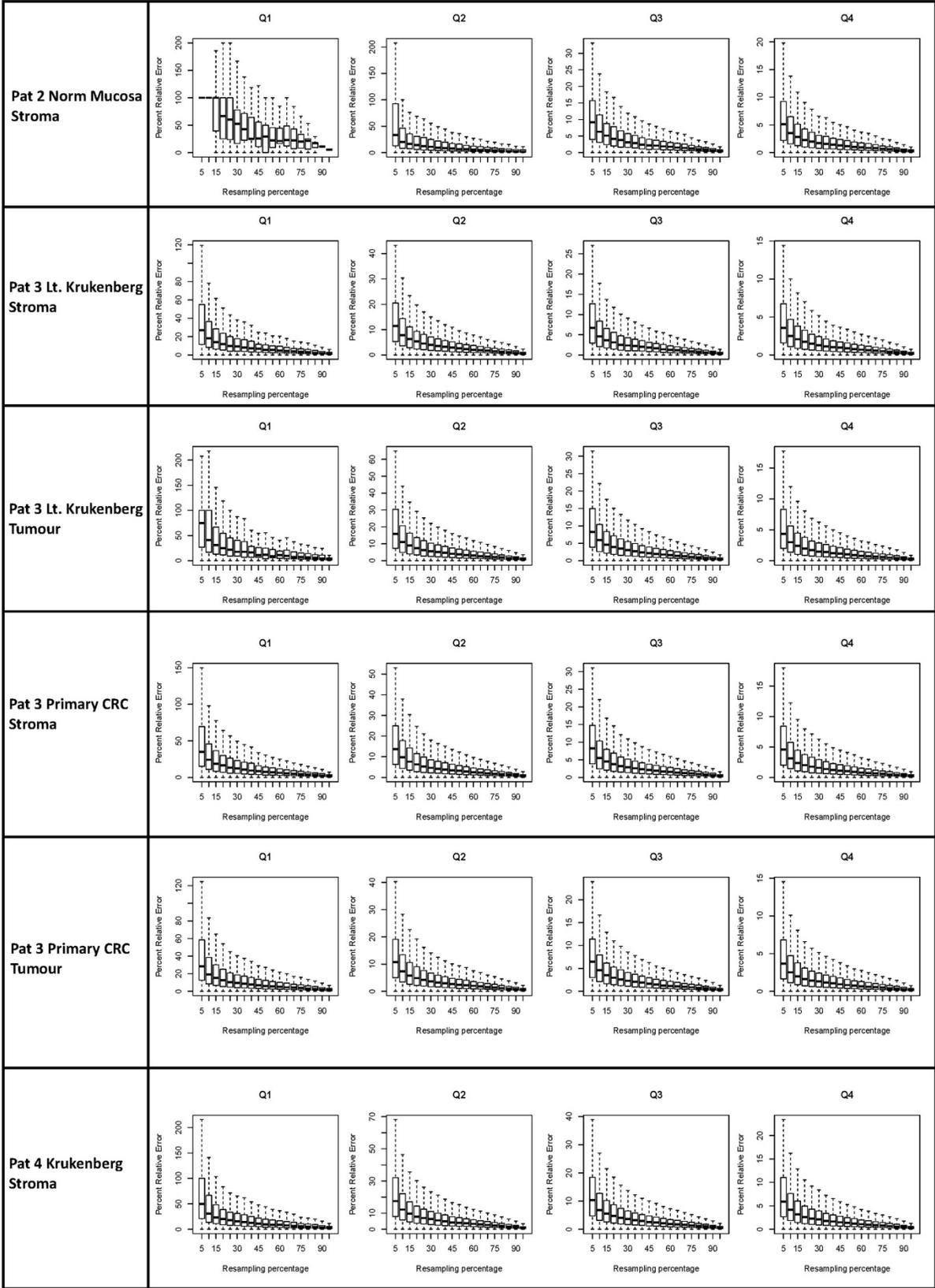
Supplementary Figure S1: H&E stained slides with pathology annotation of stroma and tumour regions for LCM.

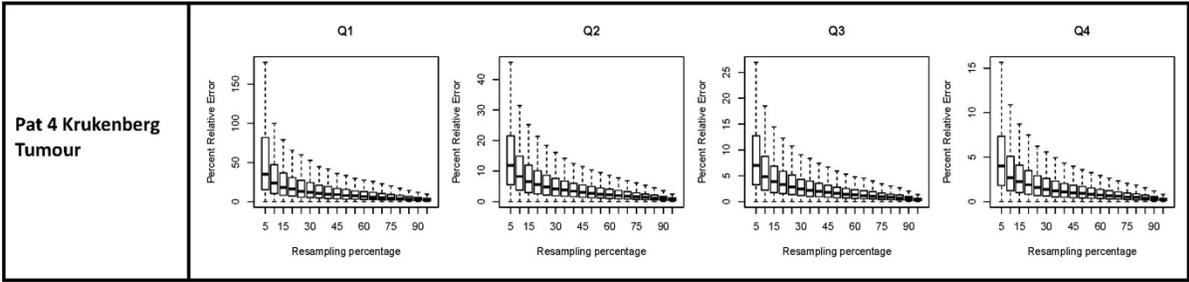


Supplementary Figure S2: (a) The distribution of duplicated reads relative to the total number of sequences for all libraries. The duplication levels relative to the total number of sequences show a distribution that is skewed to the right of the graph. This is expected of deeply sequenced enriched libraries with good library diversity, with the exception of a few samples which may have possible residual ribosomal RNA contamination as observed from the report of overrepresented sequences.

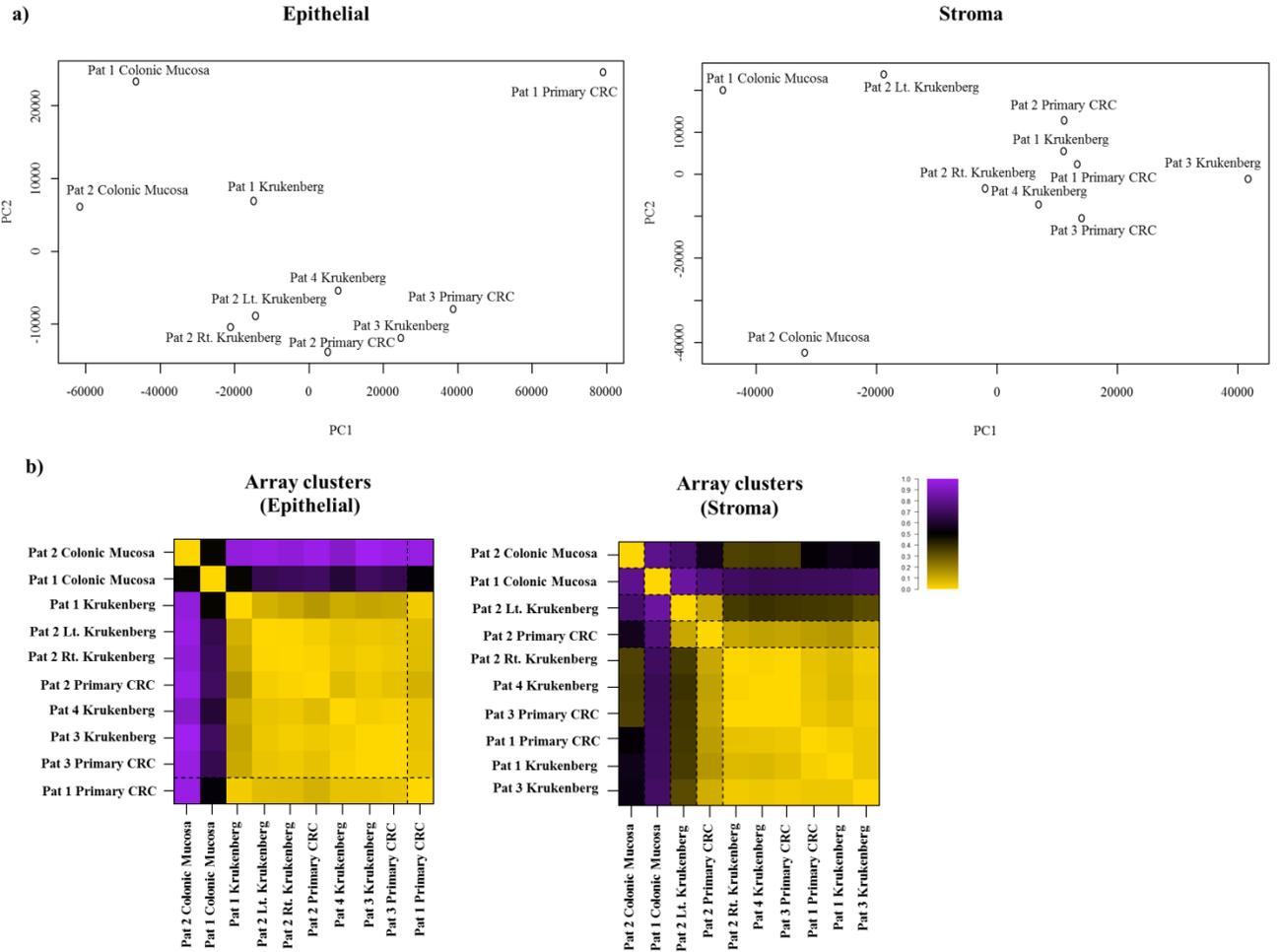




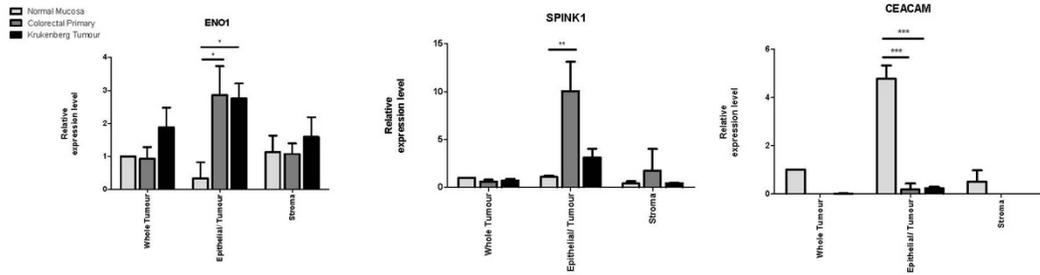




Supplementary Figure S2: (b) Saturation plots of the respective samples illustrate sufficient sequencing depth across all samples.



Supplementary Figure S3: (a) Principal component analysis (PCA) of the epithelium and stroma derived from all the microdissected samples. (b) Distance matrix of epithelium and stroma of all the microdissected samples. *Purple: Different, Yellow: Identical*



Supplementary Figure S4: Validation of RNA-Seq data by qPCR analysis of LCM samples from patient 1. Results illustrating the expression patterns for selected targets. Results are the mean of three biological replicates and standard deviations are shown.