

Current Biology, Volume 30

Supplemental Information

**Galanin Signaling in the Brain Regulates
Color Pattern Formation in Zebrafish**

Anastasia Eskova, Hans Georg Frohnhöfer, Christiane Nüsslein-Volhard, and Uwe Irion

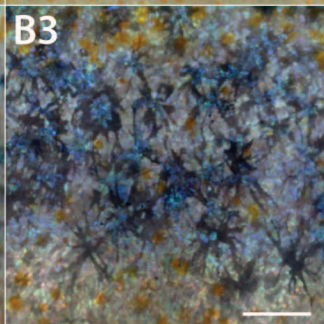
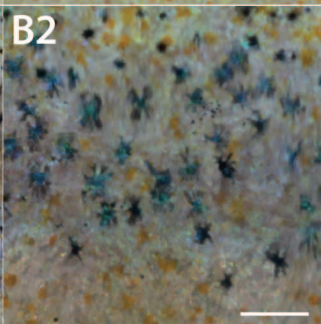
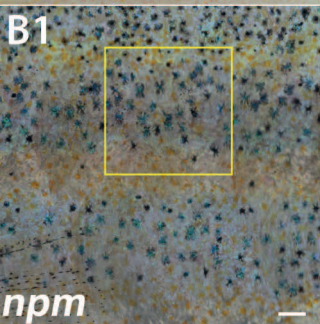
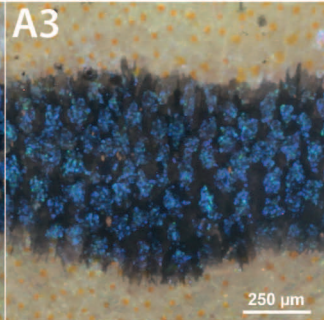
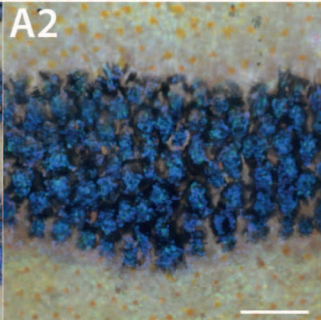
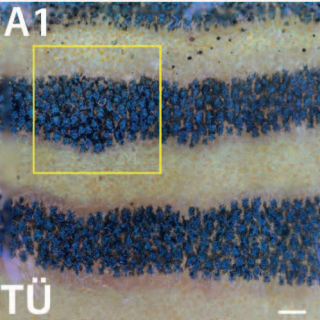
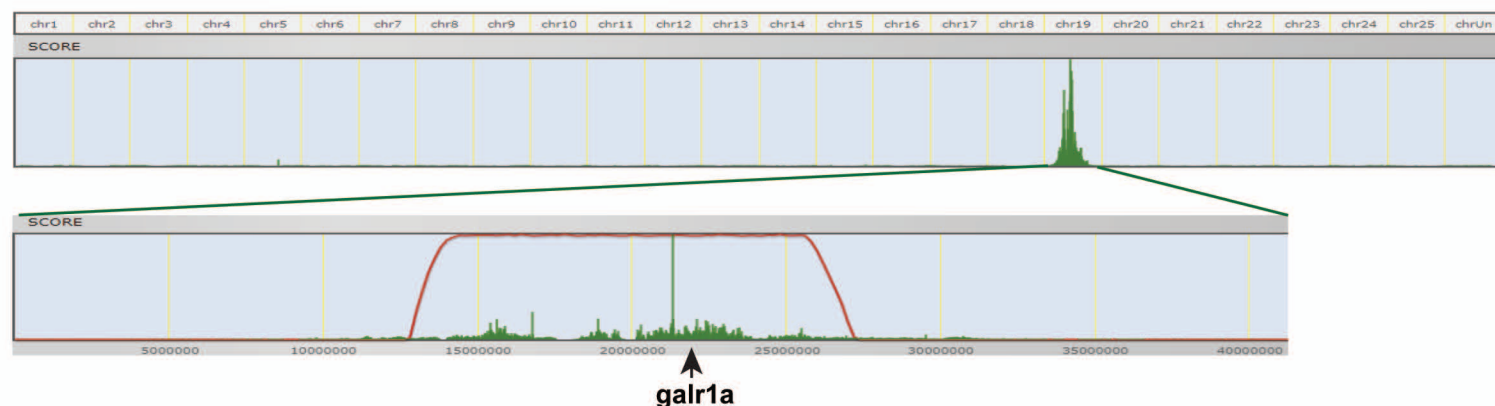
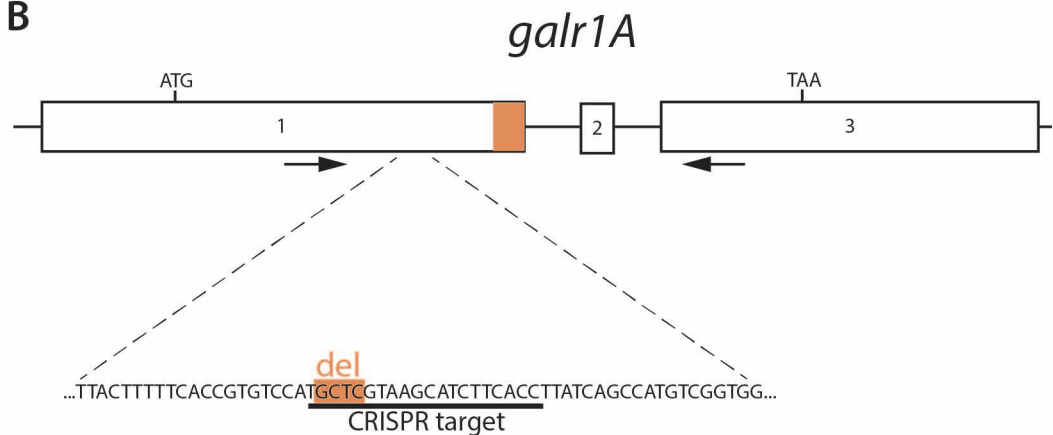
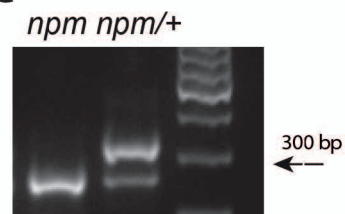


Figure S1. Melanophore shape in *npm* mutants, Related to Figure 1. In wild type (A) melanophores display a compact shape and fill the dark stripe area; melanosomes are strongly dispersed in the cells after short anaesthesia (2 min, A2), and fully dispersed after extended periods of anaesthesia (30 min, A3). In *npm* mutants (B) melanosomes only disperse after prolonged anaesthesia (B3) revealing a much more stellate shape of melanophores, and showing that they contact one another.

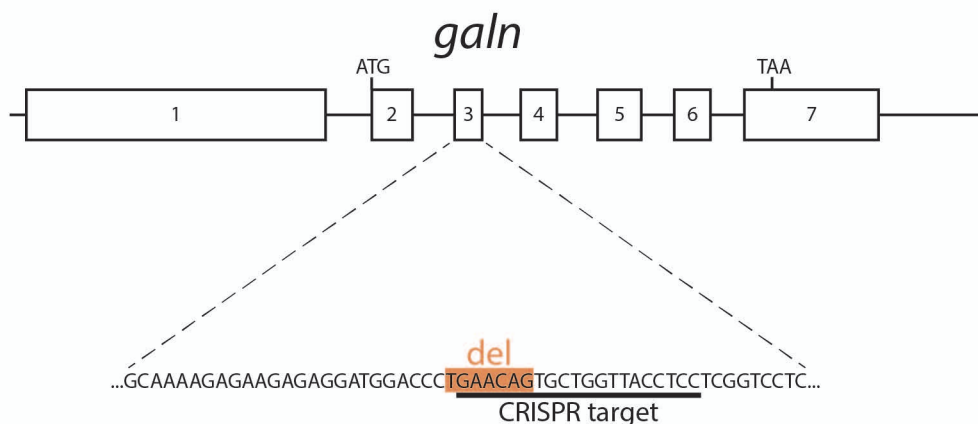
A**B****C****D**

GALR1_H.sap	1	MELAVGNLSEGNASWPEPPAPEGPLFVGIVENFVTLVVFGLIFALGVLGNSLVITVLARSKPGKPRSTTNLFI LNLSIADLAYLLFCIPFQATVYALPT	100
Galr1a_D.rer	1	MGTQNSDLDRPRSNI DLEAPEKN-LFGIGTDNLTLLIFGLIFTLGVLGNSLVITVLAQRKPGQQRSTTNIFILNLSVADLSYLLFCIPFQSTVYMLPT	99
Galr1A in npm	1	MGTQNSDLDRPRSNI DLEAPEKN-LFGIGTDNLTLLIFGLIFTLGVLGNSLVITVLAQRKPGQQRSTTNIFILNLSVADLSYLLFCIPFQSTVYMLPT	99
Galr1A_mut	1	MGTQNSDLDRPRSNI DLEAPEKN-LFGIGTDNLTLLIFGLIFTLGVLGNSLVITVLAQRKPGQQRSTTNIFILNLSVADLSYLLFCIPFQSTVYMLPT	99

GALR1_H.sap	101	WVLGAFICKFIHYFFTVSMLVSI FTLAAMSVDRYVAIVHSRRSSSLRVSRNALLGCVGIWALS IAMASPVAYHQGLFHPRASNTFCWEQWDPDRHKKAY	200
Galr1a_D.rer	100	WILGAFICKFIHYFFTVSMLVSI FTLSAMSVDRYIAIVHCRKSSSIRVARHALIGVLVIWVLSFAMATPVAYYQGIVES-EDNSTFCWEVWPDHRRKIY	198
Galr1A in npm	100	WILGAFICKFIHYFFTVSMLVSI FTLSAMSVDRYIAIVHCRKSSSIRVARHALIGVLVIWVLSFAMATPVAYYQGIVES-EDNSTFCWEVWPDHRRKIY	198
Galr1A_mut	100	WILGAFICKFIHYFFTVSM*	118

GALR1_H.sap	201	VVCTFVFGYLLPLLLFCYAKVLNHLHKKLNMSKKSEASKKTAQTVLVVVVVFGLISWLPHHI IHLWAEFGVFP LTPASF L RITAHCLAYSNSVNE	300
Galr1a_D.rer	199	VVCTFVFGYVLP LLLSFCYAKVLNHLHKKLRNVSKSEASKKTAQTVLVVVVVFCLSWLPHHVHLWVEFGSFPLNQASFVLRVAACHCLAYSNSVNP	298
Galr1A in npm	199	VV-----LNHLHKKLRNVSKSEASKKTAQTVLVVVVVFCLSWLPHHVHLWVEFGSFPLNQASFVLRVAACHCLAYSNSVNE	277

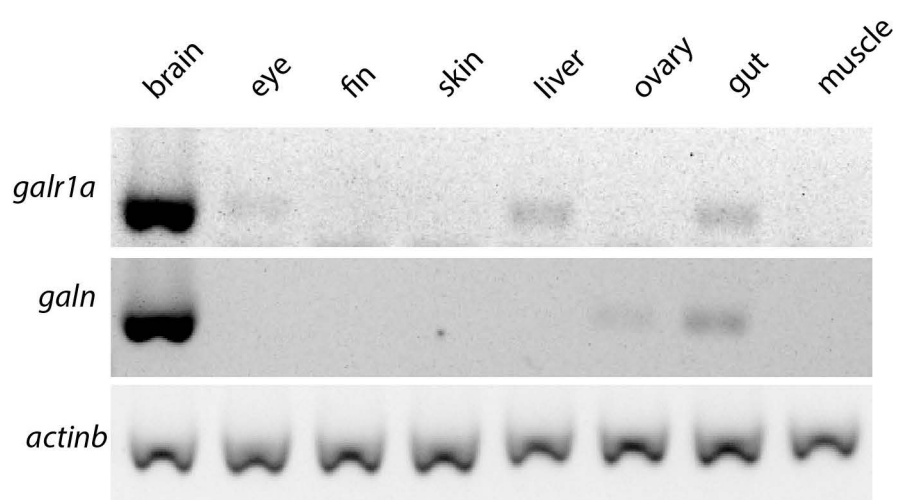
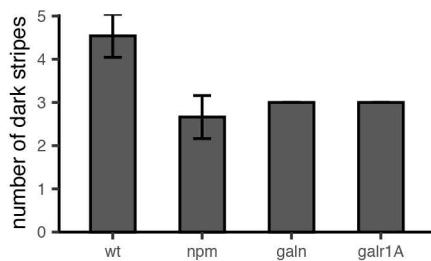
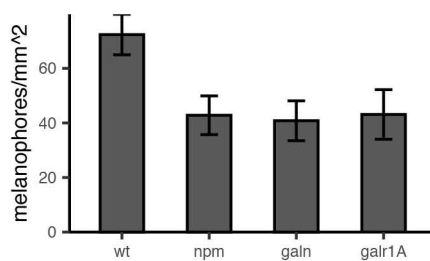
GALR1_H.sap	301	TIYAFLS ENFRKAYKQVFKCHIRKDSHLSDTKESKRIDTPPSTNCTHV	349
Galr1a_D.rer	299	VIYAFLS ENFRQAYKQVFRQVASECPTNEAREMKSKEAAPSTNCTTV	347
Galr1A in npm	278	VIYAFLS ENFRQAYKQVFRQVASECPTNEAREMKSKEAAPSTNCTTV	326

E**F**

prepro-GALN H.sap	1	MARGSALLLASLLAAALSASAGLWSPAKEYRGWTLNSAGYLLGPHAVGNHRFSFDKNGLTSKREL RPED	70
prepro-Galn D.rer	1	MHRCVGVCSLVIVCAFLTETLGMVIAAKEKRGWTLNSAGYLLGPH AIDSHRSLSDKHGLAGKREMP LDE	70
Galn_mut	1	MHRCVGVCSLVIVCAFLTETLGMVIAAKEKRGWTLNSAGYLLGPH AIDSHRSLSDKHGLAGKREMP LDE	70

prepro-GALN H.sap	71	DMKPGSFDRSIPENNIMRTIIEFLSFLHLKEAGALDRLLDLPAAASSEDIERS	123
prepro-Galn D.rer	71	DFKTGALR--IADEDVVHTIIDFLSYLKLKEIGALD---SLPSSLTSEISQP	118
Galn_mut	71	I VTGASATNTDWQEREKCL	89

Figure S2. Mapping of *npm* and details of *galr1A* and *galn* mutations, Related to Figure 4. *npm* maps to chromosome 19 (A) as shown by the output of the SNPTrack software. In (B) a schematic representation of the *galr1A* gene is shown, exons are boxes, introns are not drawn to scale. The region of exon1 that is lost from the transcript by aberrant splicing in *npm* mutants is shown in orange. The CRISPR target site and the induced 4 bp deletion are indicated. (C) RT-PCR results showing a shorter transcript in *npm* mutants, primers used are indicated as arrows in (B). (D) protein sequence alignment of Galr1A from human and zebrafish wild-type, *npm* and *galr1A^{ko}* mutants. Identical amino acid positions are indicated by asterisks, similarities by dots, the seven transmembrane regions are shaded in yellow. In *npm* 21 amino acids are missing. The k.o. allele has a premature stop. In (E) a schematic representation of the *galn* gene is shown, exons are boxes, introns are not drawn to scale. The CRISPR target site in exon 3 and the induced 7 bp deletion are displayed. (F) shows the protein sequence alignment of the Galn precursors (prepro-Galn) from human and zebrafish and the generated k.o. allele. Identical amino acid positions are indicated by asterisks, similarities by dots, the mature peptide is highlighted in yellow. The grey box covers the altered amino acids present in the k.o. allele.

A**B****C****D**

```

prepro-Gal D.re 1 MHRCVGGVCSLIVC-AFLTETLGMVIAAKEKRGWTLNSAGYLLGPHAIDSHRSLSDKHGLAGKREMP-L 68
si-rp71-1c10.8 1 MQSSCALLCISLCVFTAHLSSIHGMTLMNPEKKGWTLSAGYLLGPYA---HRSLNVRHRASGKRDTGNE 67

prepro-Gal D.re 69 DEDFKTGALRIADEDVVHTIIDFLSYLKLKEIGALDSLP-SSLTSEEISQP 118
si-rp71-1c10.8 68 NSSFPTSSY---NDSYLLSILGHLAYLRLKEKGMTEDFSGSFINSGNVKQ- 114

```

Figure S3. Expression of *galr1A* and *galn*, Related to Figure 4. RT-PCR results showing the expression of *galr1A* and *galn* in different tissues of adult zebrafish (A). Both genes are highly expressed in the brain. The numbers of dark stripes (B) and the numbers of melanophores (C) in *npm*, *galn* and *galr1A* mutants are shown. (D) shows the alignment of prepro-Galn with the potential Galn-like peptide from zebrafish, si-rp71-1c10.8. Identical amino acid positions are indicated by asterisks, similarities by dots, the mature Galn-peptide is highlighted in yellow.

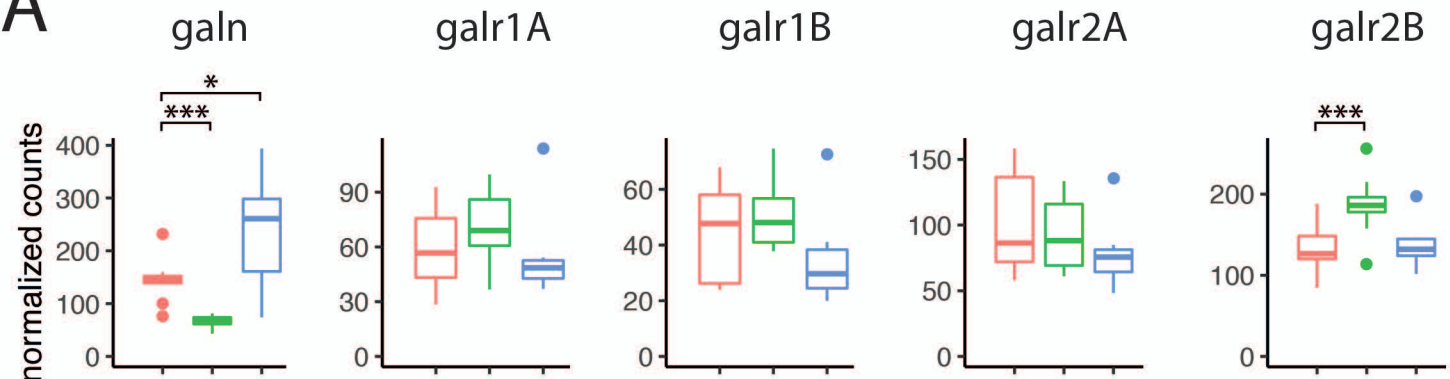
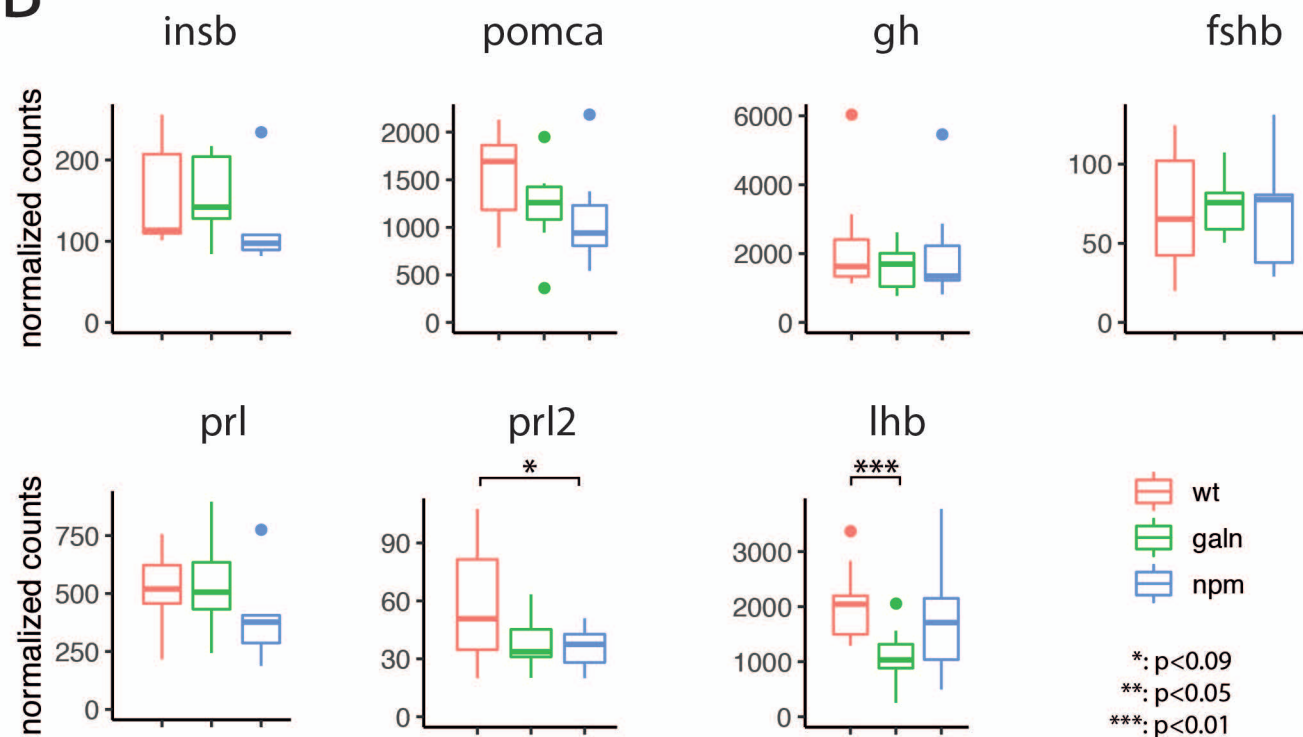
A**B**

Figure S4. Expression levels in *galn^{ko}* and *npm* mutants, Related to Figure 4. (A) *galn* and galanin receptor transcript abundance measured by Nanostring. *galn* levels are low in *galn^{ko}* mutants, presumably due to nonsense-mediated decay; they are possibly upregulated in *npm* mutants. The expression of the different receptor genes is not changed in the mutants with the exception of *galr2B* in *galn^{ko}*. The expression levels of hormones known to be influenced by Galanin signalling in mammals are mostly unchanged in *galn^{ko}* and *npm* mutants (B). Only luteinizing hormone b and (possibly) prolactin2 are affected. The plots depict the mean values (horizontal line), the first and third quartiles (box) and the lowest and highest values no further than 1.5 x IQR (inter-quartile range) from the hinge (whiskers); outliers are plotted as individual points (*: $p < 0.09$; **: $p < 0.05$; ***: $p < 0.01$). $n=9$ for wild-type, $n=8$ for *galn^{ko}*, $n=7$ for *npm*.

Primer_name	5'-sequence-3'	CRISPR Target site
galr1A_CRISPR1	TAGGTGAAGATGCTTACGAGCA	GGTGAAGATGCTTACGAGCA
galr1A_CRISPR2	AAACTGCTCGTAAGCATCTTCA	
galn_CRISPR1	TAGGATGGACCCTGAACAGTGC	GGATGGACCCTGAACAGTGC
galn_CRISPR2	AAACGCACTGTTCAGGGTCCAT	
galr1A_seq_f	TCTGTCCTACCTGCTCTTC	
galr1A_seq_r	AAAGGCAATCCACCAACC	
galn_seq_f	GTGAATACATTTTGTGTAACAGG	
galn_seq_r	ACAAGGTAATAACAAGGATGAAATC	
galr1A_cdna_f	GTCGCCTACTATCAGGGCAT	
galr1A_cdna_r	TTCAGGGGAAAGGAGCCAAAC	
galn_cdna_f	CAGAAACACTCGGGATGGTGA	
galn_cdna_r	CTTCTCGCCCCCTTGAC	

Table S1. List of primers used in this study. Related to STAR Methods.

Gene Name	Accession	Position	Target Sequence
actb2	NM_181601.3	1648-1747	CCTGGGCATATTGTA AAAAGCTGTGTGGAACGTGGCGGTGCCAGACATTTGGTGGGGCCAACCTGTACTACTGACT AATTC AATTCCAATAAAAAGTGACAT
galn	NM_001346239.1	322-421	GGCAGGAAAGAGAGAAAATGCCCTTAGATGAGGATTTCAAGACAGGAGCTCTGAGGATAGCAGATGAGGATGT CGTCCATACCATCATTGACTTTCTTTCCG
galr1a	XM_691123.5	528-627	AAAACAACAGTGATTTGGACAGACCTAGAAGCAACATAGACTTAGAAGCACCTGAAAAAACCTATTTGGCATC GGCACAGACA ACTTAGTCACGCTTCT
gapdh	NM_001115114.1	490-589	CTCACAGTTGTAAGCAATGCCTCCTGCACCACCAACTGCCTGGCTCCTTTGGCAAAGGTATCAATGATAACTTT GTCATCGTTGAAGGTCTTATGAGCA
tg	NM_001329865.1	3086-3185	TCTGTTGGTGTGTTGATGAAGAGGGTCAATACATCGCTGACTCTCTGACGTCTCGTTCTCACTACTCAGATGT GCCAAACTTTATGCCAGAGGCTCCA
trh	NM_001012365.2	691-790	GGTGAACGGACGCTTCAAGTGTTCGGGAAACACACCTGTCTCTCCATGTGCCAGTTTTCGAAGCGACAGTT CAAAGCCTCATTCACTGTGACGCGT
insb	NM_001039064.1	127-226	TTCTCCATCCAGCATCTGTGTGGTTCAAGCCTGGTGGATGCGCTTTACCTAGTGTGTGGCCTAGAGGTTTCTT CTACACCAACAGAGGCCAGGAGAGAC
tshba	NM_181494.2	243-342	TTTATTGTTTCAGAGGGGATGCACCTATCAGGAAGTTGAGTACCGGACAGCCGCTTTCGCCGGATGCCCTTCACA TGCAGATCCTCACTCACCTACCCAG
galr1b	NM_001327843.1	672-771	TAACCGGAGGAATGCGCTTATTGGCGTCTGTGTCATTTGGATGCTTTCTTTATCTTTGCCGTCCCGGTTGCTCAG CACCAAATTTTGACGGATCACCCC
galr2a	XM_021480396.1	629-728	CTCCAGGCCACTATCTACACCATGGACGAGTGGGTTTTTCGGCGGTTTCGTGTGCAAAGCCGTGCACCTTTATTAT TACCTGACCATGTACGCGAGCATCT
galr2b	XM_021474712.1	1344-1443	TGAAAATAAAGTGCACGGCTAATGATGCCTTACAGAAGCCTTATTTAAAAGAAAACACTCAAAGTGAGCCGGAG GATTTCTGCAGCTTTGATCGACATCAC
pomca	NM_181438.3	916-1015	AAGGGGGAGAGGTTGTTATAGGGGGATGTTTTGAATATACTTTCTCCAGCAAACCTCTGGATGAGAGGTTCC TATCATGCATAGAAAACGAAGGTGGGC
gh1	NM_001020492.2	490-589	ATGGACAGCCAAATATGGATGACAACGACTCCCTGCCGTTGCCTTTTGAGGATTTCTACCTGACCGTAGGGGAG ACCAGTCTCAGAGAGAGCTTTCCGCT
lhb	NM_205622.2	435-534	CCCGACTTCTGCATGTCCCAGAGAGAGGATTTCCCGCATACTAGACCTCGGACAACCTCACATCAACCTACACAC ACAGTCGAGCTCAGCATTATTAGAC
fshb	NM_205624.1	261-360	TGTACAAGAGCTACGAGTTAAAGGCTGCTCTGCAGGGGTTGATTCACTCTCGTGTACCCCGTGGCTCTGAGC TGTGAGTGCAACCAGGTTAACTCAGA
prl	NM_181437.3	490-589	CGTACACAAGATGGGCTCGTCTTCTGACAACCTGTCCACTCTCCGTTTAAATGGCAACAACCTTGGTCAGGATAA AACGTCTCGACTTGTCAATTTCCAC
prl2	NM_001162854.1	576-675	CCATGCTCCGATTTCTGACAGCGGAGAAGCCATGAGTGACTACGATCTCTCTACTGCTTTCCGCCGACTCCA ACAAAGTCCAGA ACTATCTAAAAAT

Table S2. List of transcripts and target sequences used for Nanostring expression profiling. Related to STAR Methods.