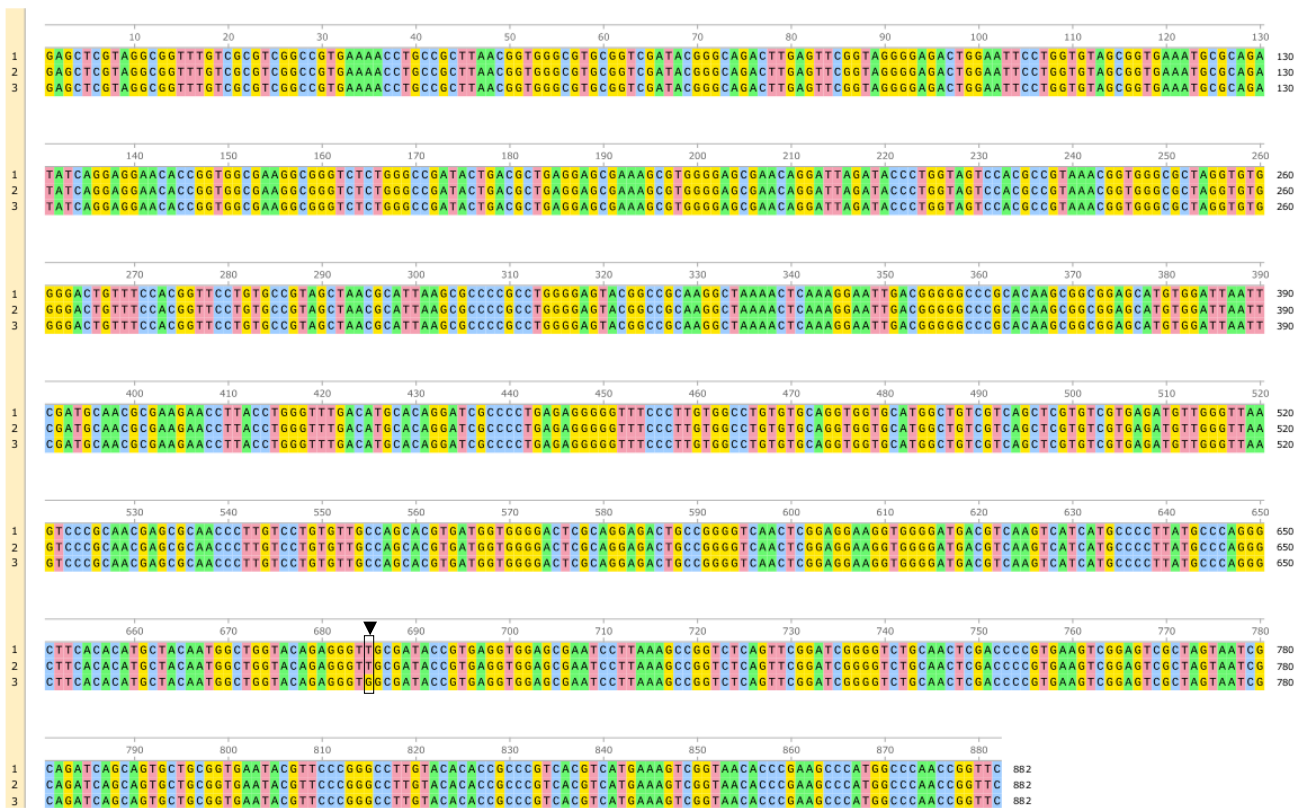
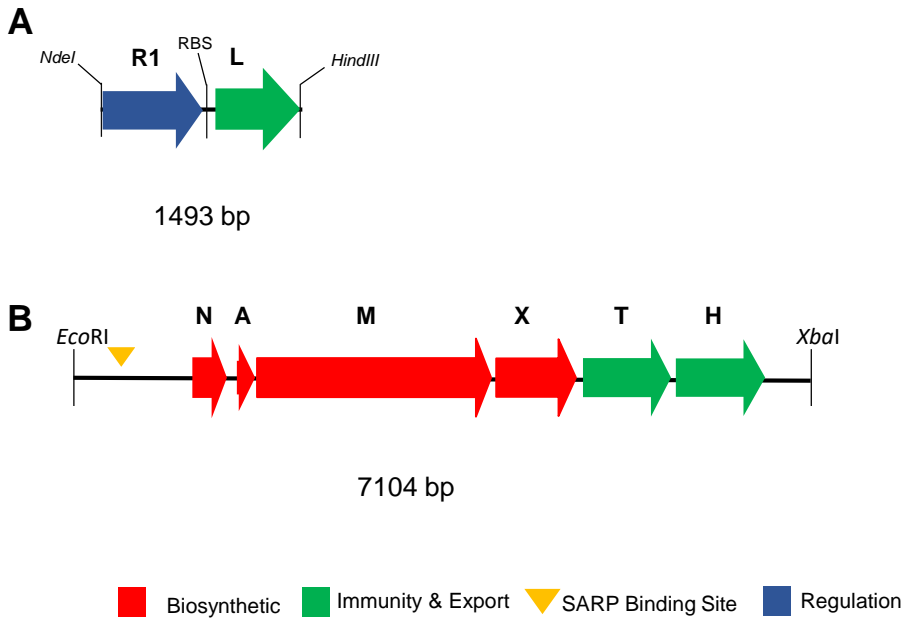


1. KY3  
2. KY7  
3. KY21

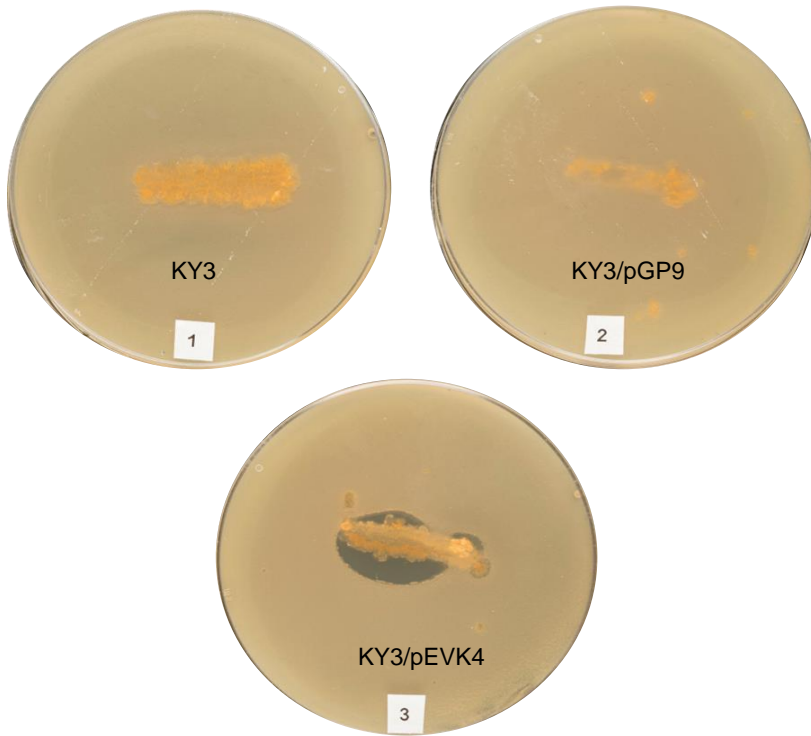


**FIG S1 Alignment of *Saccharopolyspora* sp. KY3, KY7 and KY21 16S rDNA sequences.** The alignment was performed with Clustal Omega (v1.2.4) and the figure was generated by SnapGene Viewer (v4.2.11). The difference between KY21 to strains KY3 and KY7 is indicated with a black arrow and a box at position 685.

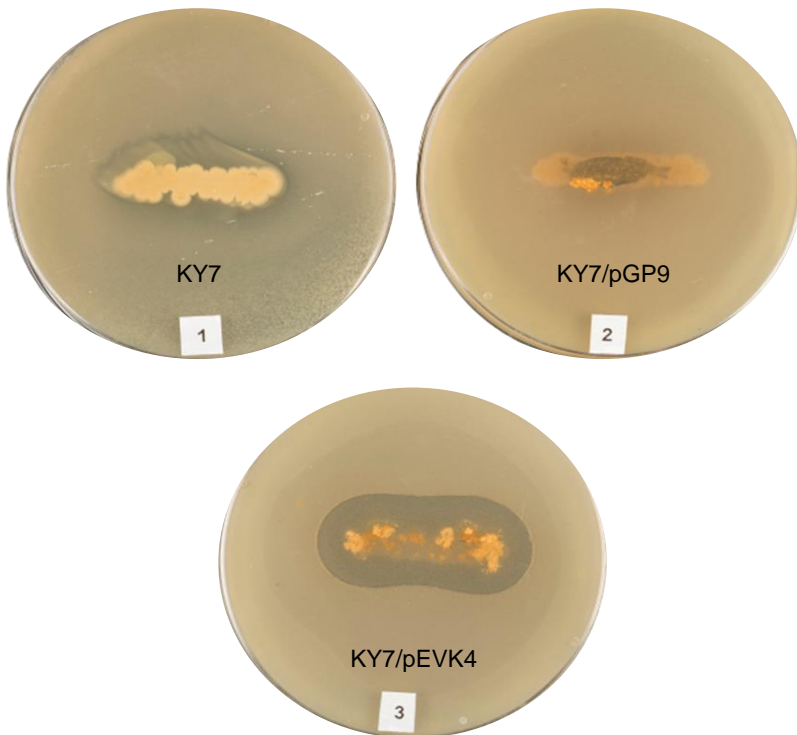


**FIG S2 Schematic of synthetic artificial operons. (A)** The operon consisting of *kyaR1*, encoding a *Streptomyces* antibiotic regulatory protein (SARP), and *kyaL*, encoding a PE-methyl transferase that provides resistance – the homologues of *cinR1* and *cinorf10* respectively. **(B)** The operon carrying genes *kyaN* to *kyaH* as an *EcoRI/XbaI* fragment. These genes are expected to be essential for kyamycin biosynthesis.

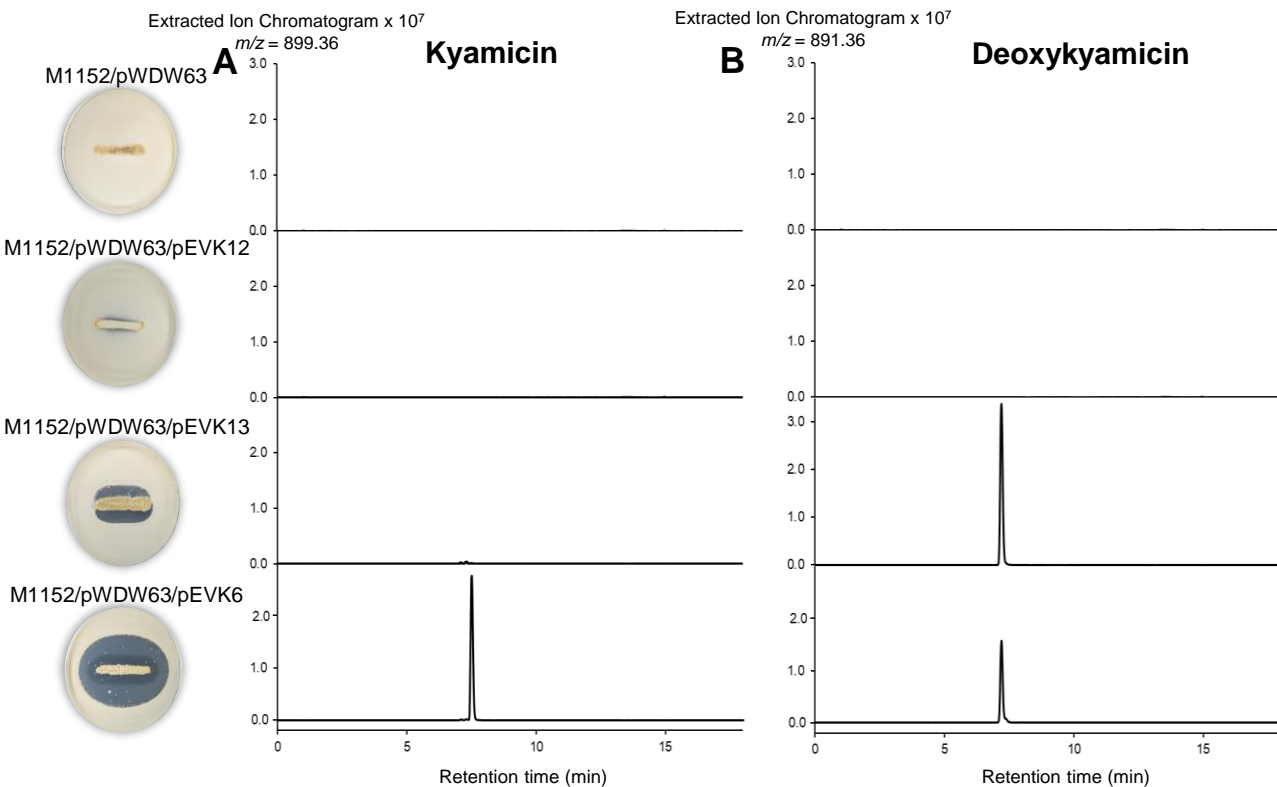
**A**



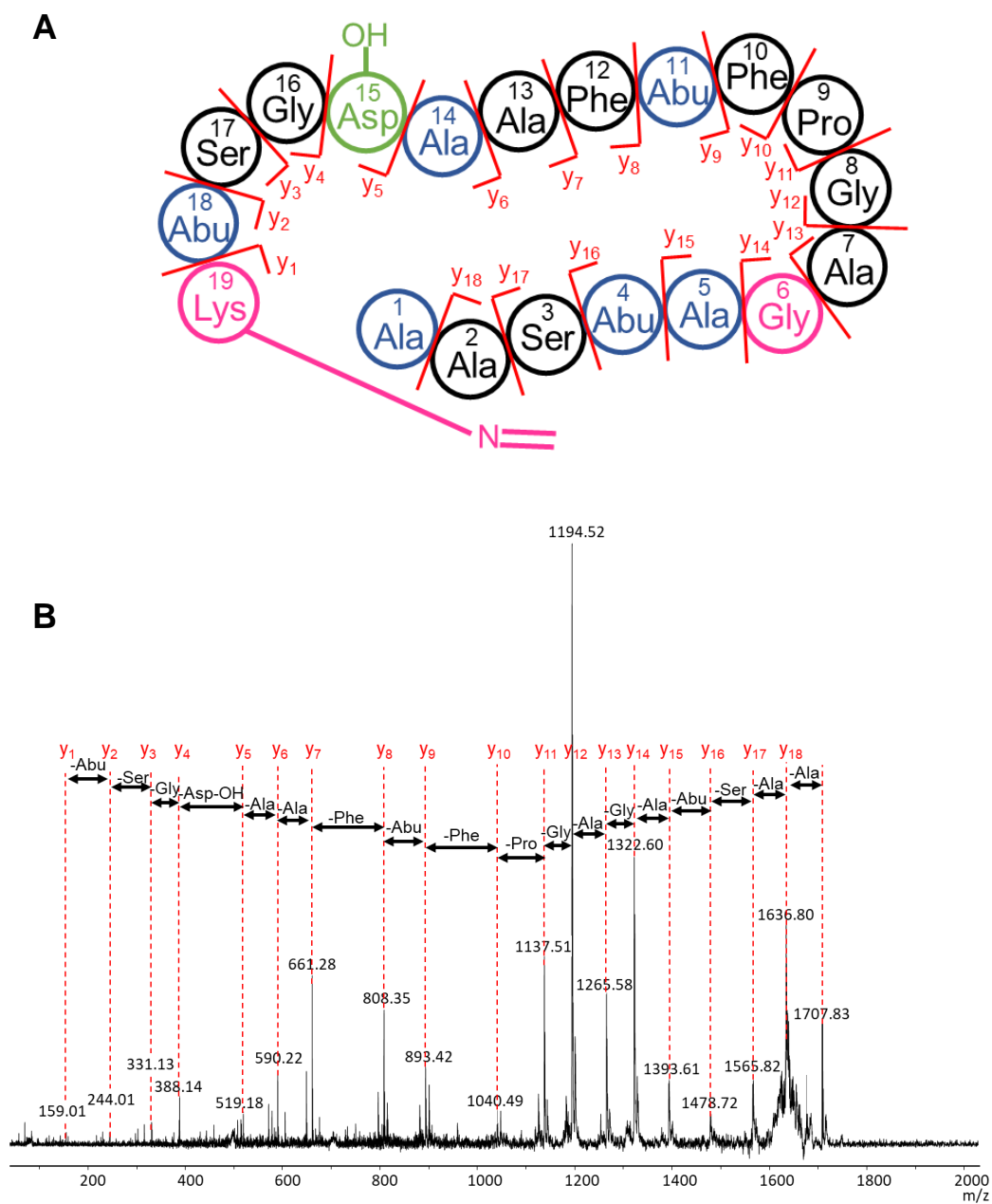
**B**



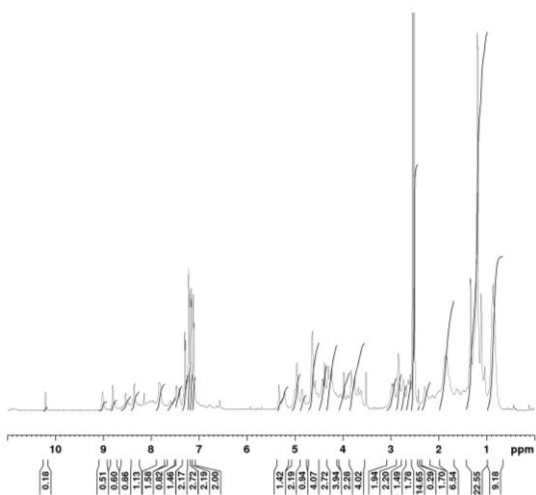
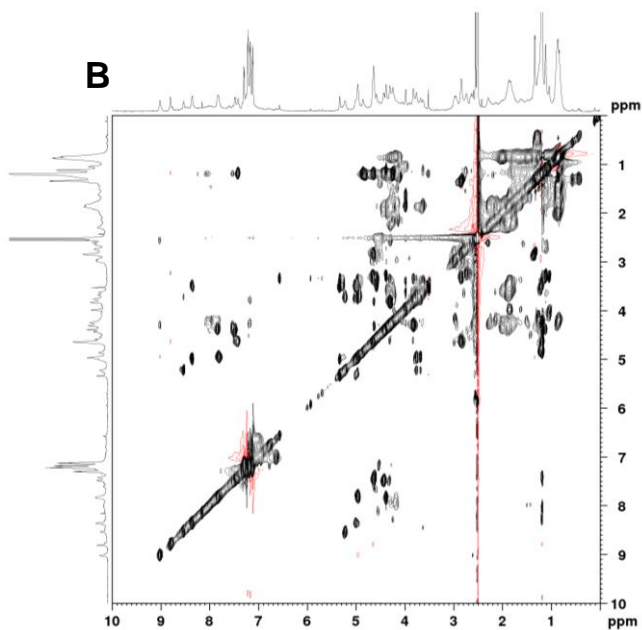
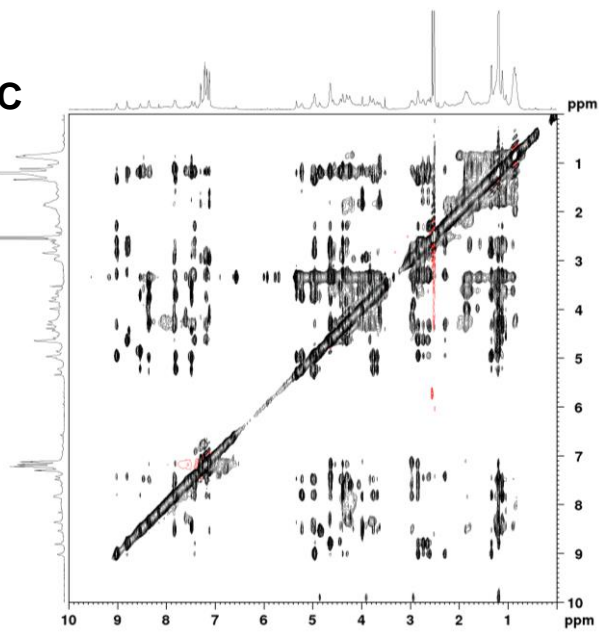
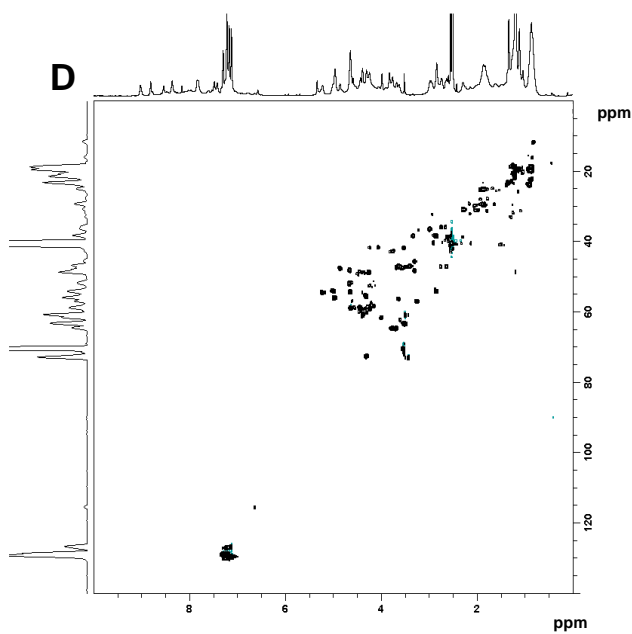
**FIG S3 Activation of kyamicin biosynthesis in KY3 and KY7.** The pEVK4 vector containing *kyaR1* and *kyaL* results in a zone of inhibition, corresponding to the production of kyamicin, in contrast to the pGP9 empty vector control or the wildtype strain. **(A)** Activation of kyamicin production in KY3, and **(B)** in KY7 strains.



**FIG S4 Dissection of the contribution of *kyaR1* and *kyaL* to kyamicin BGC activation.** Overlay bioassays were carried out with *B. subtilis* EC1524 and agar plugs were taken adjacent to the central streak and analysed by UPLC/MS. The pEVK12 vector containing only *kyaL* does not result in a zone of inhibition. The pEVK13 vector containing only *kyaR1* results in a zone of inhibition, corresponding to the production of deoxykyamicin only. Expression of both pEVK12 and pEVK13 results in a zone of inhibition, corresponding to the production of both kyamicin and deoxykyamicin. Images and LC traces are representative of at least three biological repeats. **(A)** Extracted ion chromatograms are shown where  $m/z = 899.36$  ( $[M + 2H]^{2+}$ ). **(B)** Extracted ion chromatograms are shown where  $m/z = 891.36$  ( $[M + 2H]^{2+}$ ).



**FIG S5 Kyamicin fragmentation.** Following reduction to remove methylanthionine bridges, kyamicin was subject to MALDI-ToF tandem MS, giving the complete y ion ( $\text{NH}_3^+$ ) series. **(A)** Structure of reduced kyamicin and the  $y_1 - y_{18}$  ion series. **(B)** MALDI-ToF tandem MS spectrum with the y ion series indicated with dashed red lines.

**A****B****C****D**

**FIG S6 Kyamicin NMR Spectra. (A)**  $^1\text{H}$  NMR spectrum. **(B)** TOCSY spectrum. **(C)** NOESY spectrum. **(D)** HSQC spectrum.

**TABLE S1 Recipes for liquid screening media.** Quantities of components are given in g/L. SM = screening media.

	Maltodextrin	Bacto Soy peptone	Bacto Peptone	CaCl <sub>2</sub> [0.02 M]	CaCl <sub>2</sub> *2H <sub>2</sub> O	CaCO <sub>3</sub>	Casamino acids	Corn steep liquor	EDTA	Glucose	Glycerol solution	K <sub>2</sub> HPO <sub>4</sub>	L-Glutamate	L-Proline	Malt extract	Maltose	Meat Extract	MgSO <sub>4</sub> [0.2 M]	MgSO <sub>4</sub> *7H <sub>2</sub> O	Molasses (cane)	MOPS	NaCl	Oatflour	Cotton seed flour	Soluble starch	Soya flour	Tomato paste	Trace salts No.1	Yeast extract	ZnSO <sub>4</sub> *7H <sub>2</sub> O	Distilled water (to 1L)	Tap water (to 1L)	pH	
	g	g	g	mL	g	g	g	g	g	g	g	g	g	g	g	g	g	mL	g	g	g	g	g	g	g	g	g	mL	g	g				
SM3	50.0					2.5				5.0		0.3								3.0						25.0						x		7.0
SM5			20.0			0.4				15.0	10.0						8.0															x		7.2
SM6	20.0							40.0									0.5					2.5										x		7.0
SM7				10.0							20.0	2.0	1.5	15.0			10.0				20.9	0.5						5.0			x		6.5	
SM12			4.0			5.0				50.0							4.0					2.5				10.0						x		7.6
SM14		20.0								10.0							5.0					5.0								0.01	x		7.0	
SM15					0.029		11.5		0.25		23.0	0.5						0.49		20.9	0.5						5.0			x		6.5		
SM18						8.0				15.0									20.0					25.0	40.0						x		7.2	
SM19										2.0													15.0				40.0				x		6.0	
SM20			5.0													20.0	5.0		1.0			3.0							3.0		x		7.2	
SM25			10.0								40.0				21.0																x		6.3	
SM30										2.0													15.0				40.0					x		4.5
SM32			10.0								40.0				21.0																x		4.5	

**TABLE S2 Putative NMR assignments. ND = not determined.**

Amino Acid	Position	$\delta$ H	$\delta$ C	Amino Acid	Position	$\delta$ H	$\delta$ C
1-Cys	NH <sub>2</sub>	ND	-	12-Phe	NH	ND	-
	$\alpha$	ND	ND		$\alpha$	ND	ND
	$\beta$	ND	ND		$\beta$	ND	ND
2-Ala	NH	8.00	-		$\gamma$	-	ND
	$\alpha$	4.24	48.6		$\delta$	7.17	129.3
	$\beta$	1.19	20.9		$\epsilon$	7.21	128.6
3-Ser	NH	8.54	-	$\phi$		127.0	
	$\alpha$	5.22	54.3	13-Ala	NH	7.42	-
	$\beta$	3.76	ND		$\alpha$	4.64	54.1
4-S-Abu	NH	7.83	-	$\beta$	1.19	18.6	
	$\alpha$	4.38	60.8	14-Cys	NH	8.80	-
	$\beta$	3.29	48.0		$\alpha$	3.24	56.9
	$\gamma$	1.11	19.1		$\beta$	2.74	35.7
5-Cys	NH	9.02	-		2.64		
	$\alpha$	4.30	55.4	15-HO-Asp	NH	7.17	-
	$\beta$	2.60	38.5		$\alpha$	4.53	58.6
		2.30			$\beta$	4.30	72.4
			OH		3.82	-	
6-N-Ala	NH	7.81	-	16-Gly	NH	7.48	-
	$\alpha$	4.95	55.8		$\alpha$	4.42	58.7
	$\beta$	3.76	64.4		4.65		
7-Ala	NH	ND	-	17-Ser	NH	8.36	-
	$\alpha$	ND	ND		$\alpha$	5.00	53.8
	$\beta$	1.18	19.4		$\beta$	3.52	70.3
8-Gly	NH	ND	-	OH	5.33	-	
	$\alpha$	ND	ND	18-S-Abu	NH	8.47	-
9-Pro	$\alpha$	3.98	61.4		$\alpha$	3.62	56.2
	$\beta$	1.63	29.2		$\beta$	1.85	24.9
	$\gamma$	1.85		$\gamma$	ND	ND	
	$\delta$	3.62	46.9	19-Lys	NH	ND	-
10-Phe	NH	ND	-		$\alpha$	ND	ND
	$\alpha$	ND	ND		$\beta$	ND	ND
	$\beta$	2.97	36.2		$\gamma$	ND	ND
	$\gamma$	-	ND		$\delta$	ND	ND
	$\delta$	7.29	128.8		$\epsilon$	ND	ND
	$\epsilon$	7.11	129.4		NH	ND	-
	$\phi$		126.7		COOH	-	10.22
11-S-Abu	NH	ND	-				
	$\alpha$	ND	ND				
	$\beta$	ND	ND				
	$\gamma$	ND	ND				