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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, seeAuthors & Referees and theEditorial Policy Checklist.

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ГОІ	all statistical allalyses, commit that the following items are present in the figure regend, table regend, main text, or intenious section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
×	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection BD FACSDiva v8.0.1; MaxQuant v1.5.3.30; Agilent 1290; G6460 triple Quad MS; Illumina HiSeq 2000 RNA

Data analysis Graphpad Prism V6; STATA v13; Flow-jo software v10.5.3;

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

https://xenabrowser.net (TCGA melanoma cohort) (Figure 7); www.oncomine.org (oncomine database) (Figure 1);

The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE partner repository with the dataset identifier PXD016535 (Figure 6).

Field-specific reporting

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<u>Lite scien</u>	ices st	udy design	
All studies must disc	close on these	e points even when the disclosure is negative.	
Sample size	No specific statistical test was used to predetermine the sample size.		
Data exclusions	s N/A		
Replication	All experiment	ts were representative of at least two independent experiments.	
Randomization	No method of	f randomization was used.	
Blinding	In vivo and in v	vitro experiments were monitored in a non-blinded fashion.	
We require information system or method listed Materials & exp n/a Involved in the	on from authors ed is relevant to perimental s e study cell lines pgy d other organisi earch participar	n/a Involved in the study ChIP-seq Flow cytometry MRI-based neuroimaging MRI-based neuroimaging	
Antibodies		Ill information are included in the methods	
	Antibodies used All information are included in the methods.		
Validation Primary antibodies were validated by the manufacturer.		rimary antibodies were validated by the manufacturer.	
Eukaryotic ce	ell lines		
Policy information a	about <u>cell line</u>	<u>s</u>	
Cell line source(s)		All information are included in the methods.	
Authentication		All information are included in the methods.	
Mycoplasma cont	amination	Cell lines were routinely tested for the absence of mycoplasma contamination by PCR.	
Commonly miside (See <u>ICLAC</u> register)		None	
Animals and	other or	ganisms	
Policy information a	about <u>studies</u>	involving animals; ARRIVE guidelines recommended for reporting animal research	
Laboratory animal	All information are included in the methods.		
Wild animals	<u></u>	None	
Field-collected sar	mples	Tumours, lymph nodes, lungs	
Ethics oversight	All experimental protocols were approved by the local ethic committee (Midi-Pyrénées, France) and are in compliance with French and European regulations on care and protection of laboratory animals.		

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about <u>clinical studies</u>

 $All\ manuscripts\ should\ comply\ with\ the\ ICMJE \underline{guidelines\ for\ publication\ of\ clinical\ research}\ and\ a\ completed \underline{CONSORT\ checklist}\ must\ be\ included\ with\ all\ submissions.$

Clinical trial registration	None
Study protocol	N/A; tumour samples were collected retrospectively.
Data collection	Tumour samples were collected retrospectively.
Outcomes	N/A

Flow Cytometry

Plots

Confirm that:

- The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).
- The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).
- 🗶 All plots are contour plots with outliers or pseudocolor plots.
- 🗶 A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation	All information are included in the methods.
Instrument	BD LSR Fortessa X-20
Software	BD FACSDiva v8.0.1
Cell population abundance	N/A
Gating strategy	See supplementary Figure 2.

x Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.