

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Data analysis

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	It is generally considered that a sample size containing at least 3 biological replicates can provide adequate statistical power in biochemical analysis. We have described the exact sample size for each experiment in our manuscript.
Data exclusions	No data captured was excluded from the subsequent analyses.
Replication	The exact number of replication for all experiments was described in figure legends and our attempts at replication were successful.
Randomization	Samples and animals were allocated randomly.
Blinding	The experimenters were blinded to the animal genotype, grouping information and data analysis.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

n/a	Involvement in the study	n/a	Involvement in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies	<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines	<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology	<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms		
<input checked="" type="checkbox"/>	<input type="checkbox"/> Human research participants		
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data		

Antibodies

Antibodies used	The following antibodies were used for immunoblotting or immunostaining: rabbit anti-S6K (2708), rabbit anti-p-S6K (T389) (9205), rabbit anti-mTOR (2983), rabbit anti-pS6 (S235/236) (4858) and rabbit anti-S6 (2217) were from Cell Signaling Technology, mouse anti-DSTYK (sc-374487), mouse anti-LAMP1 (sc-20011) were from Santa Cruz Biotechnology, rabbit anti-V5 (14440-1-AP), rabbit anti-RAB7A (55469-1-AP), rabbit anti-DSTYK (20102-1-AP), rabbit anti-TFEB (13372-1-AP) and rabbit anti-LAMP3 (12632-1-AP) were from Proteintech, mouse anti-V5 (ab27671) was from Abcam, mouse anti-actin (AC-74, A5316) was from Sigma. Alexa Fluor 488 donkey anti-rabbit IgG (A21206), Alexa Fluor 568 donkey anti-rabbit IgG (A10042) and Alexa Fluor 594 donkey anti-mouse IgG (A21203) were from Invitrogen.
Validation	All antibodies are from commercial sources and their validation data are available on the manufacturer's website. The appropriate dilution for all the antibodies was determined through preliminary experiments.

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	The COS-7 and HeLa cell lines were purchased from the Stem Cell Bank, Chinese Academy of Sciences (Shanghai, China).
Authentication	The COS-7 and HeLa cell lines were verified by short tandem-repeat DNA profiling before the study.
Mycoplasma contamination	All the cell lines had been tested and confirmed negative for mycoplasma contamination.
Commonly misidentified lines (See ICLAC register)	No commonly misidentified cells were used in this study.

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals	The detailed information about zebrafish strains and generation of transgenic lines used in this study has been provided in our METHODS section. The development stage in zebrafish experiments are included in the manuscript. All zebrafish were housed in semi-closed recirculation housing systems (ESEN, China) and were kept at a constant temperature (27–28°C) on a 14:10 hour
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light:dark photoperiod. All in vivo experiments and protocols were approved by Institutional Animal Care and Use Committee of the Research Institute of Surgery, Daping Hospital.

Wild animals

No wild animals involved in this study.

Field-collected samples

The study did not involve samples collected from the field.

Ethics oversight

All in vivo experiments and protocols were approved by Institutional Animal Care and Use Committee of the Research Institute of Surgery, Daping Hospital IACUC protocol SCXK- (Army) 2007-017.

Note that full information on the approval of the study protocol must also be provided in the manuscript.