



Supplementary Information for

Large-scale contractions of Friedreich's ataxia (GAA)<sub>n</sub> repeats occur during DNA replication owing to their triplex-forming potential in yeast

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## Supplementary Information Text

### Methods

**Fluctuation tests.** To compare the rates of contraction in the conditions of high and low transcription, the cells were first grown overnight in a liquid complete media with raffinose as a sugar source. Cells were then diluted and plated on YNB media, which contained glucose or galactose as a sugar source and was additionally supplemented with uracil and adenine. Colonies from the media with galactose were grown for 60 hours instead of 40 before plating the fluctuation test. The rest of the fluctuation test was identical to the protocol described above.

**Cloning of the cassettes.** The *P<sub>Gal1</sub>-UR-(GAA)<sub>124</sub>-A3-TRP1* cassette was constructed as previously described (1). To create cassettes with other repeats, we ordered DNA fragments from GenScript which were identical to a piece of the *P<sub>Gal1</sub>-UR-(GAA)<sub>124</sub>-A3-TRP1* cassette, except had other repeats in place of (GAA)<sub>124</sub> repeats. The fragments were cloned into the *P<sub>Gal1</sub>-UR-(GAA)<sub>124</sub>-A3-TRP1* cassette using *SgrAI* and *NcoI* or *SgrDI* and *BglIII* restriction enzymes. To get the inverted cassettes, we amplified the direct cassettes with primers carrying *SgrDI* and *BglIII* recognition sites in their 5'-tails and cloned back to the vector using the same enzymes, as previously described (1). All constructs were Sanger sequenced and integrated into yeast as linear fragments. Their integration was confirmed using PCR.

**Yeast strain construction.** All of the strains used in this study are derivatives of SMY710 (CH1585 *MATa leu2-Δ1 trp1-Δ63 ura3-52 his3-200 ade2Δ::KanMX4 HAP1-wt*) (2), except for Pol  $\alpha$  mutants and their corresponding WT strain which are derivatives of CH1585 (*MATa leu2-Δ1 trp1-Δ63 ura3-52 his3-200*) (3). To construct gene knockouts, we used gene replacement with *hphMX6*, or *HIS3* as selectable markers. All knockouts were confirmed using PCR with internal primers and either external or junction primers. *Rad27* missense mutants were created via two-

step transformation protocol as previously described (4). All other missense mutants were made using a simplified CRISPR-Cas9 approach (5). To overexpress the yeast RPA, a plasmid containing the *RFA1*, *RFA2* and *RFA3* genes with their flanking sequences, the *HIS3* marker and 2-micron origin of replication cloned into the pUC57 vector was ordered from *Gene Universal*. Plasmid was transformed into yeast and the cells were grown on media lacking histidine to facilitate plasmid retention. A complete list of all yeast strains used in this study can be found in the supplemental tables.

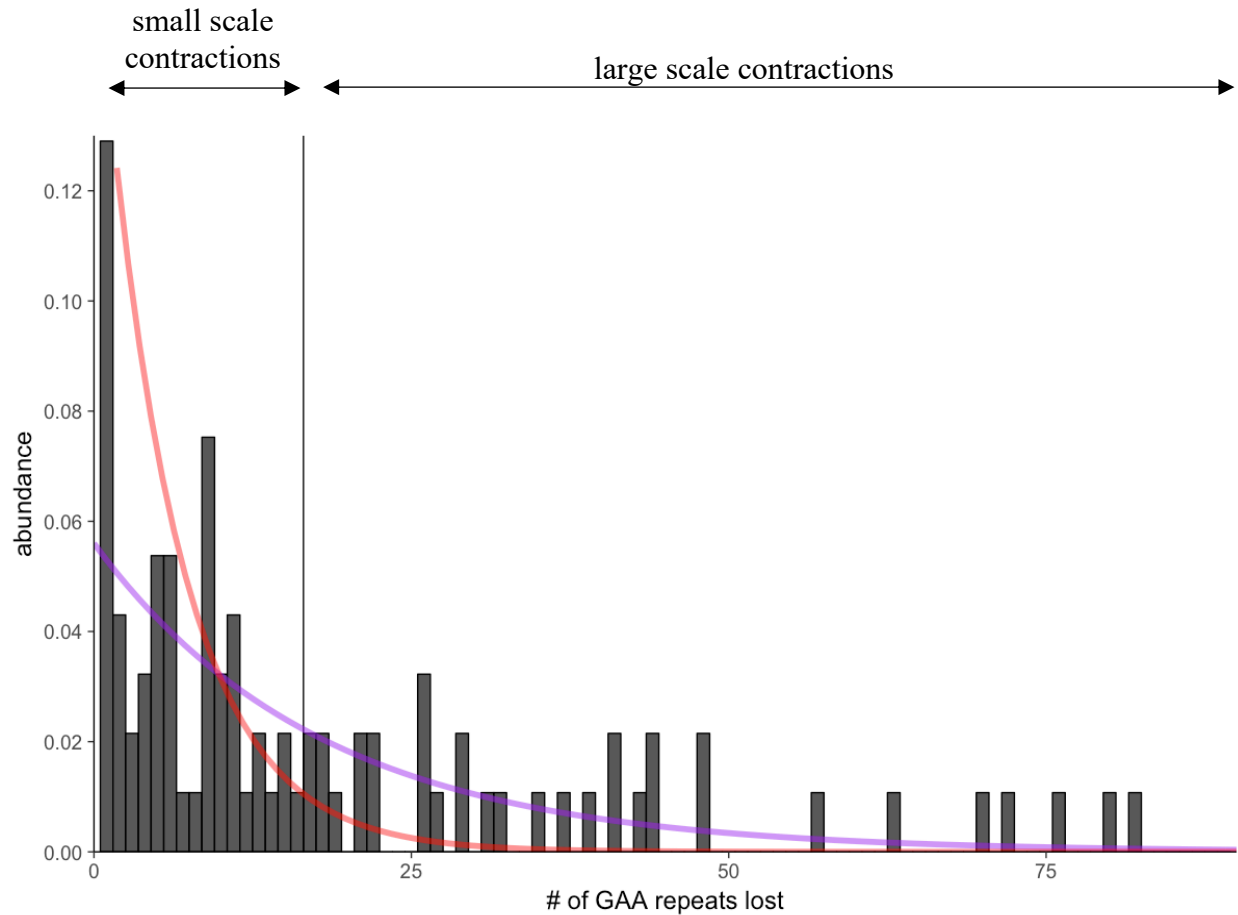
**Spot tests.** Yeast cells were plated on complete media supplemented with uracil and adenine (YPDUA) with no or 0.5 mM H<sub>2</sub>O<sub>2</sub>. After they formed colonies, the colonies were dissolved in water and 5-fold dilutions were made. 4μl of each dilution was spotted on YPDUA and Gal<sup>+</sup>Ura<sup>-</sup> media.

**Distributions of GAA repeat sizes.** To build a distribution of repeat sizes in WT colonies without applied selection, the strains were plated to single colonies and PCR of individual colonies was performed. *Image Lab* software (*Bio-Rad*) was used to estimate the size of the repeat tracts in each sample.

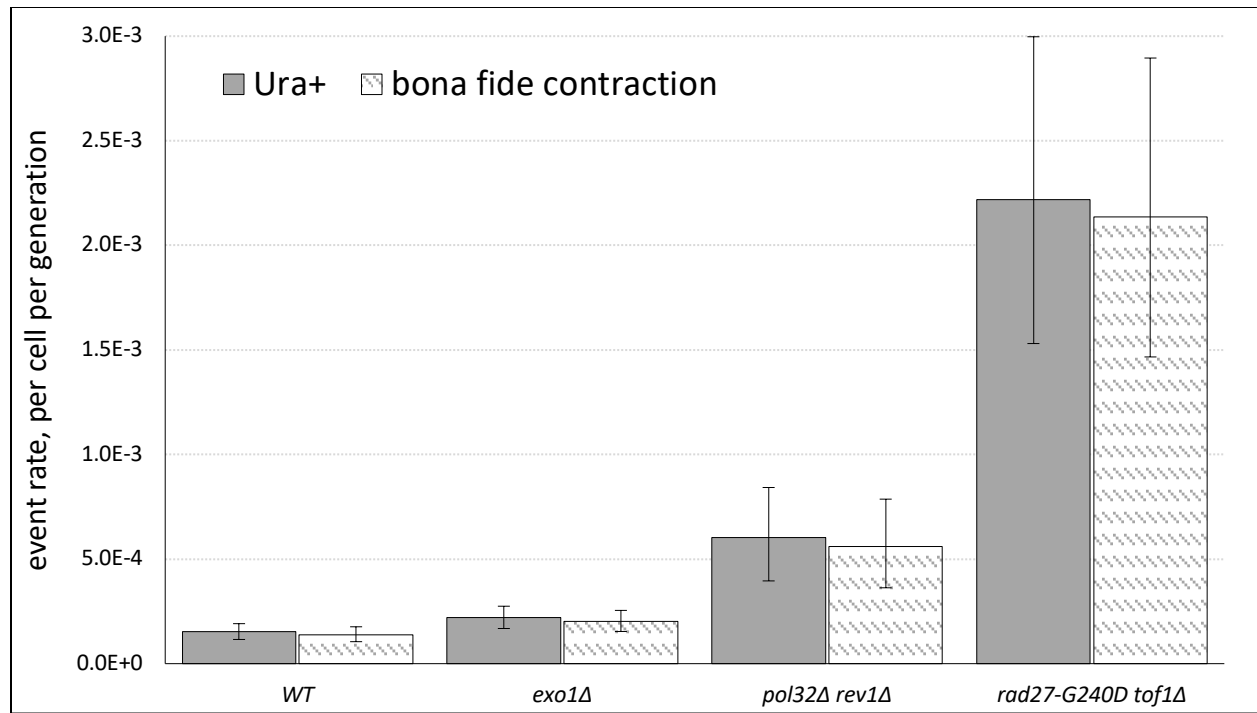
To build a distribution of deleted repeats after selection, 8 to 16 colonies were randomly picked from each of the selective media. Colony PCR was performed with A2 and B2 primers. *Total Lab Count* or *Image Lab* software was used to estimate the size of the repeat tracts in each sample. If two or more colonies from the same selective plate showed an identical size of repeat deletion, that value was only accounted once.

**RT-qPCR.** Three independent colonies from each strain and condition were grown overnight. In the morning, the cells were diluted to an OD<sub>600</sub> of 0.1 and harvested when they reached an OD<sub>600</sub> of 0.5. RNA was isolated using RNeasy Kit (*Qiagen*) following the

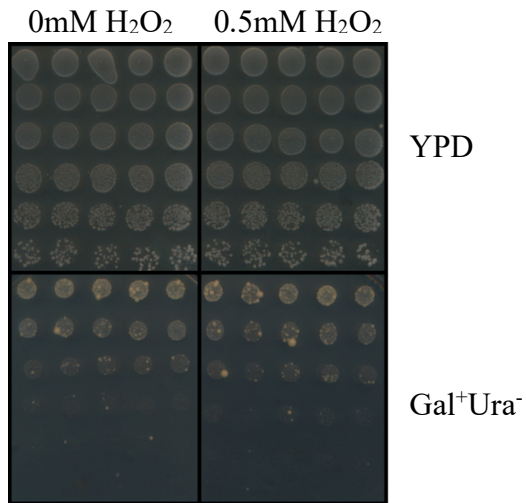
manufacturer's instructions. Extracted RNA was treated with Turbo DNase (*ThermoFisher*) following the manufacturer's instructions. Reverse transcription was performed using SuperScript IV Reverse Transcriptase (*ThermoFisher*) following the manufacturer's instructions. Random hexamers and polyT primers were used to estimate URA3 transcription and RPA overexpression correspondingly. Next, qPCR was performed using SYBR Select Master Mix (*ThermoFisher*) on a QuantStudio 6 Flex Real-Time PCR system (*ThermoFisher*). To compare different samples to one another, mRNA levels were normalized to the amount of *ACT1* mRNA. Standard curves based on 10-fold serial dilution of gDNA were used for the quantification of the results. Two-sample *t*-test was used to estimate the significance.



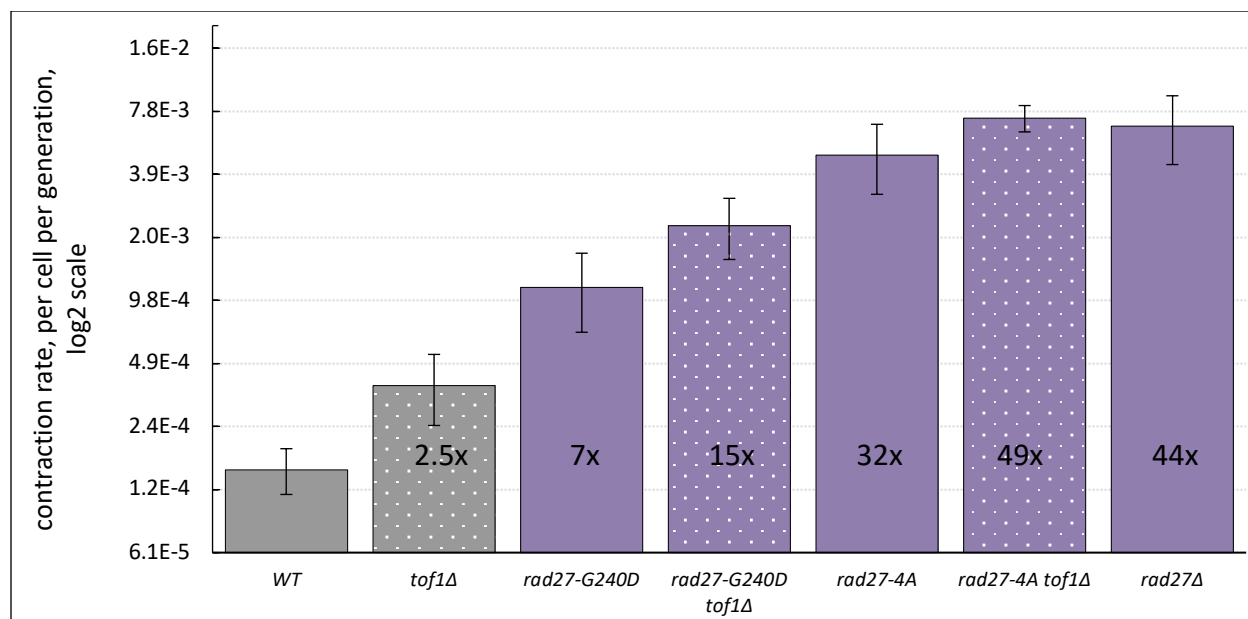
**Fig. S1.** Distribution of contracted repeats in the inverted cassette in the absence of selection. Red line represents exponential distribution fit for the distribution of all contractions (Kolmogorov-Smirnov test,  $P=0.13$ ). Purple line represents exponential distribution fit for the small-scale contractions (Kolmogorov-Smirnov test,  $P=0.24$ ). The vertical bar denotes the border between small and large scale contractions as classified by  $k$ -means clustering analysis.



**Fig. S2.** Comparison of the Ura<sup>+</sup> and *bona fide* contraction rates for various mutants. Error bars represent 95% confidence intervals.

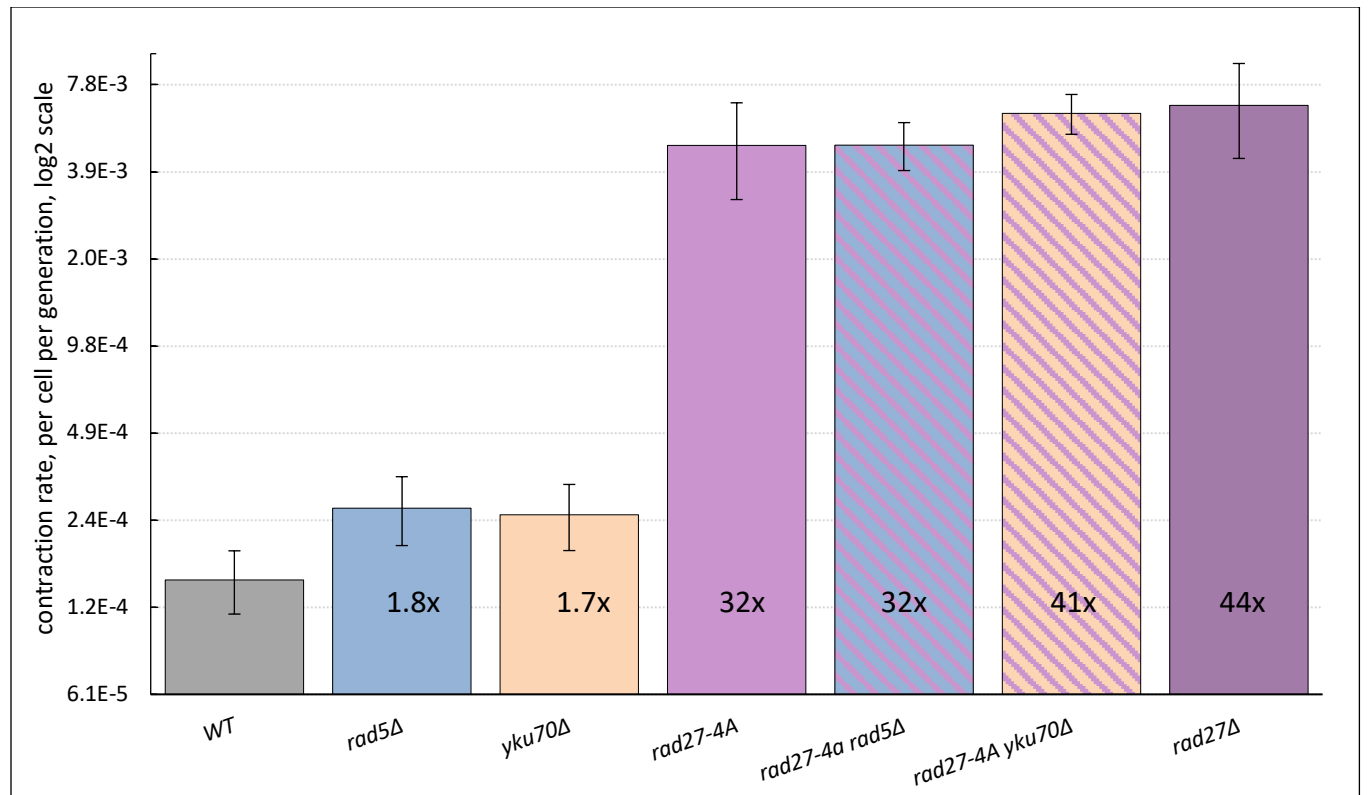


**Fig. S3.** Colonies of WT cells bearing the (GAA)<sub>124</sub> repeat tract in the direct cassette were grown on YPD plates supplemented with adenine, uracil, and either 0 or 0.5mM of H<sub>2</sub>O<sub>2</sub>. Figure shows 5 colonies from each plate with intact repeat tracts spotted onto complete (YPD) or selective media (Gal<sup>+</sup>Ura<sup>-</sup>).

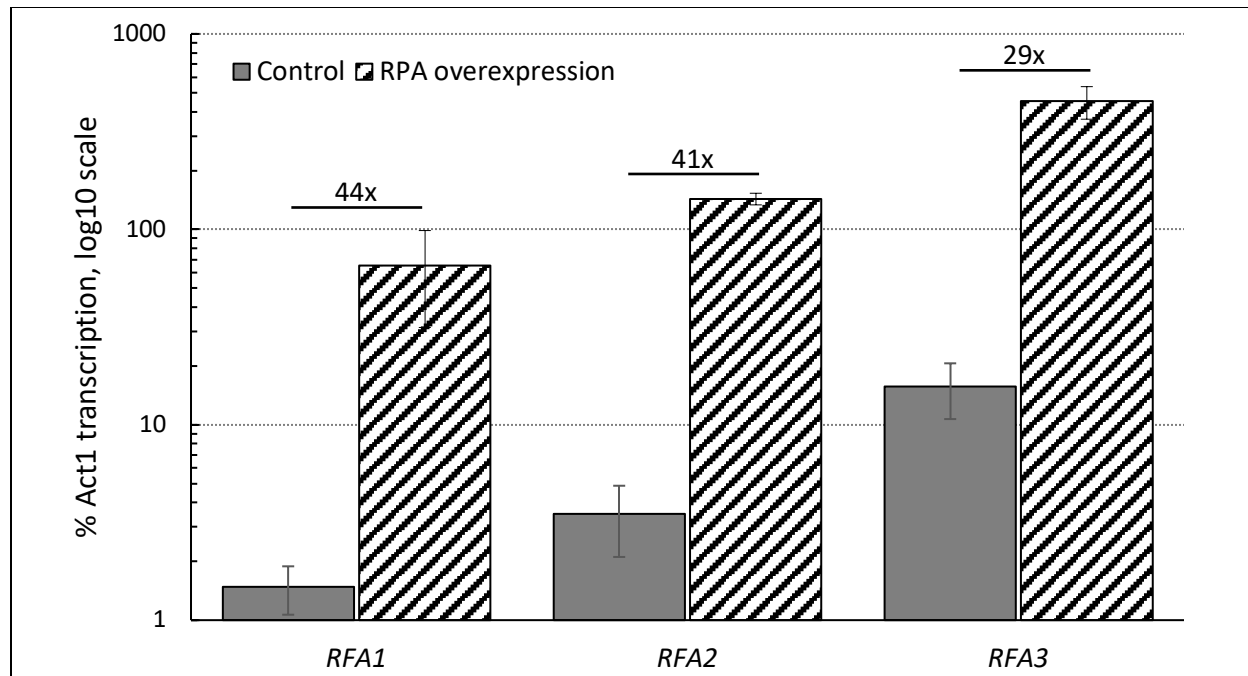


**Fig. S4.** Effect of knocking out *TOF1* on GAA repeat contraction rates in two *rad27* mutants. Error bars represent 95% confidence intervals. Numbers on bars show fold change relative to WT.

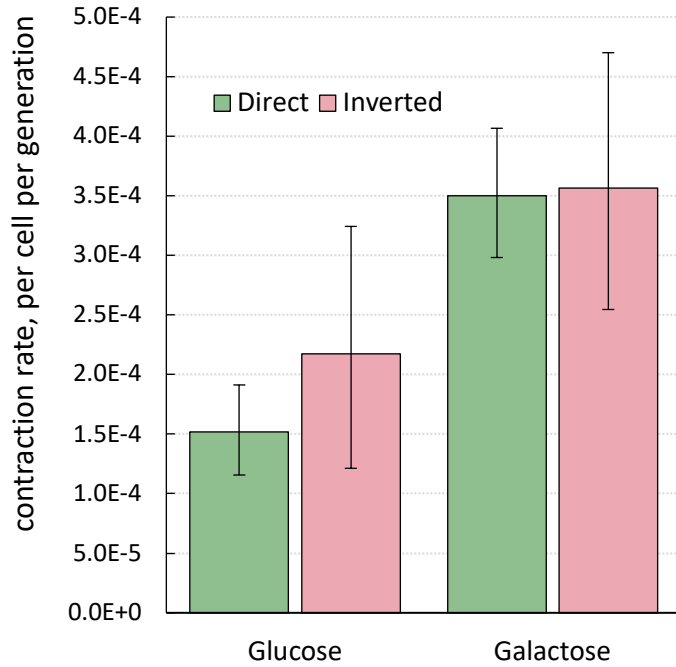




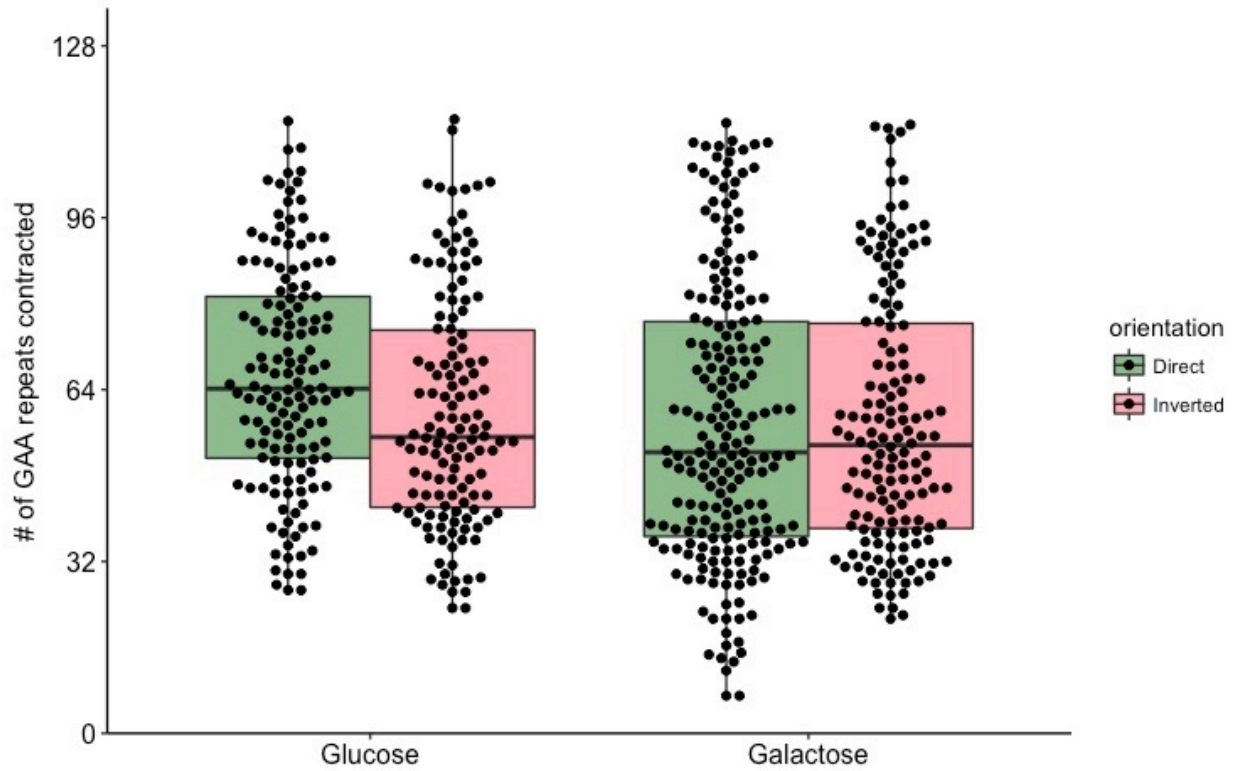
**Fig S5.** Effect of knocking out *RAD5* or *YKU70* on GAA repeat contraction rates in the *rad27-4A* mutant. Error bars represent 95% confidence intervals. Numbers on bars show fold change relative to WT.



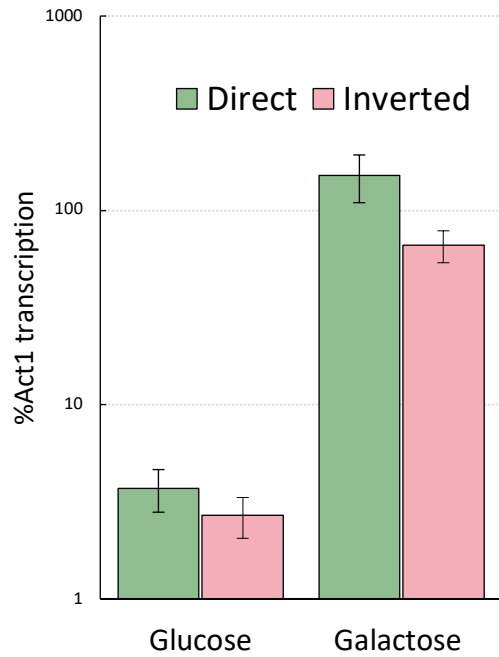
**Fig S6.** Levels of RPA overexpression. RPA overexpression was achieved by transforming yeast cells with a multicopy plasmid bearing three yeast RPA subunits. Total RNA was isolated from cells with (RPA overexpression) or without (control) the plasmid and reverse transcribed using a polyT primer. The amount of RPA cDNA was estimated using qPCR. The standard curve method was used to quantify transcription levels. Means of three biological replicates are shown and the error bars represent standard deviations. Fold changes in gene expression are shown above each bar.



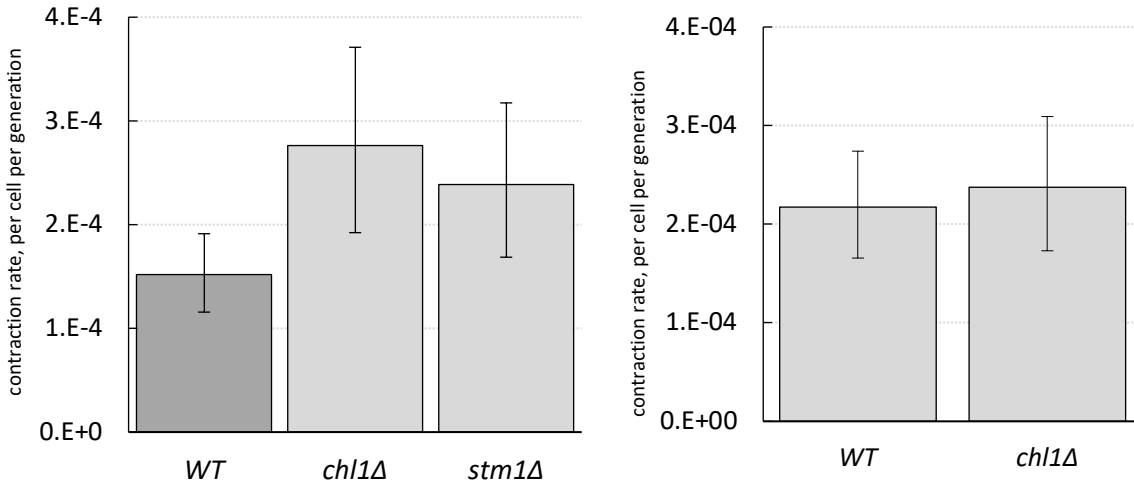
**Fig. S7.** The effect of low (glucose) or high (galactose) transcription state on GAA repeat contraction rate in strains with direct versus inverted cassettes. Bars represent 95%-confidence intervals.



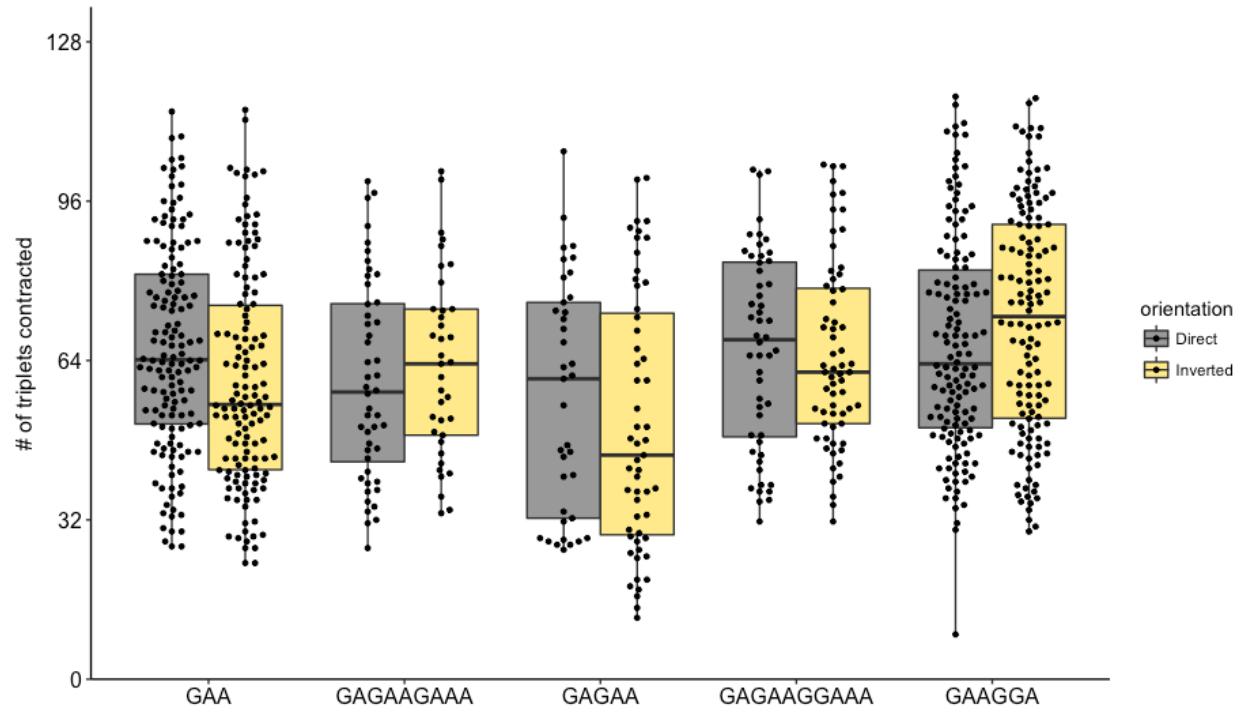
**Fig. S8.** Distribution of contraction sizes for direct versus inverted cassettes in colonies with repressed (glucose) and induced (galactose) transcription through the reporter. Each box represents median and interquartile range; whiskers show 1.5 interquartile range.



**Fig. S9.** *URA3* transcription level in cell cultures with direct versus inverted orientation of the cassette grown under transcription repression (glucose) or induction (galactose) measured by RT-qPCR. The standard curve method was used to quantify transcription levels. Mean values of three biological replicates are shown. Error bars represent standard deviations.



**Fig. S10** The effect of mutations in genes that were reported to interact with DNA triplex on GAA repeat contraction rates in the direct (A) and inverted (B) cassettes. Error bars represent 95% confidence intervals.



**Fig. S11.** Distribution of contraction sizes for direct and inverted cassettes depending on repeat composition. Each box represents median and interquartile range; whiskers show 1.5 interquartile range.

**Table S1.** Fraction of colonies with *bona fide* contractions among Ura<sup>+</sup> colonies for various mutants. Colonies of various strains were recovered from selective media after a fluctuation test and subjected to PCR analysis of the repeat tract length.

	# of total colonies analyzed	# of <i>bona fide</i> contractions	% of <i>bona fide</i> contractions
WT	183	167	91
<i>rad27-G240D</i>	119	117	98
<i>rad27-4A</i>	128	124	97
<i>msh3Δ</i>	159	145	91
<i>exo1Δ</i>	191	178	93
<i>rad27-G240D tof1Δ</i>	191	183	96
<i>pol32Δ rev1Δ</i>	126	116	92
<i>rad27Δ</i>	144	140	97
<i>rad27-G67S</i>	190	181	95



**Table S2.** Primers used in this project

Primer name	Primer sequence	Notes
For cloning		
R-SgrDI	CGGATCCGGAGATCTCGCGTGGGGATGATCCACTAG	
L-BglII	GACCCACGTCGACGCTAGCGGGTAATAACTGATA	
L-SgrDI	TAAGACCGTCGACGCTCGCGTGGGGATGATCCACTAG	For cassette inversion (1)
R-BglII	TGCTTAAGATCTCGCTAGCGGGTAATAACTGATA	For cassette inversion (1)
For Sanger sequencing of the cassettes		
TRP1 PPP R	CTCTGCAAGCCGCAAACCT	
URA3-RT-UnSpl-R	GAGCCCTTGCATGACAATTC	
5' UAS 100 1-R	ATCGAATTTGAGGTCTGCACT	
5'misc-F	GATGGTACGACGGTTTGTAATAGCG	
Alex-URA3-RT-Spl-F	ATCCTAGTCCTGTTGCTGCCAA	
For amplification of the cassettes		
5'misc-F	GATGGTACGACGGTTTGTAATAGCG	
3'misc-R	AAGCACTAACGATTGCGTGATGG	
For integration of the cassettes		
A36a-R	AGGGTCGTTGCCTTCTGGT	
A36b-F	ACGTGTACAGTTCTCTTTACATCATC	
TrpS-F	TCGATTTCTGACTGGGTTGGAAG	
5' UAS 100 1-R	ATCGAATTTGAGGTCTGCACT	
URA3-RT-UnSpl-F	TTGACTGATCTGTAATAACCACGA	
For repeat tract amplification		
A2	CTCGATGTGCAGAACCTGAAGCTTGATCT	
B2	GCTCGAGTGCAGACCTCAAATTCGATGA	
S2	TGCTCGATGTGCAGAACCTGAAGCTTG	
K2	GCTCGAGTGCAGACCTCAAATTCGATG	
For qPCR		
A3-F	CTTCCCTTTGCAAATAGTCCTCTTCC	
A3-R	CAGAATTGTCATGCAAGGGCTCC	
Act1 F	CTCCACCACTGCTGAAAGAG	
Act1 R	GTGATGACTTGACCATCTGG	
RFA1 qPCR F	TCGTGTGACGGATTTTGGTGG	
RFA1 seq R2	GATGAAGTTTGCCTTGCAGGC	
RFA2 qPCR F	TCACGTTTGTTTTGTAGGTGTGG	
RFA2 seq R	CATCGTTACCGGCAGCCAAG	
RFA3 seq F	CGAAACACCAAGAGTTGACCCC	
RFA3 qPCR R	CGTAATCATTTCAACCTCGCTGCC	
For amplification of the pRCC-N plasmid with target gRNA sequence		
DNA2_H547A_pRCCN_F	TATTGCCTAAAGTCATGACAGTTTTAGAGCTAGAAA TAGCAAGTTAAAATAAGG	<i>dna2_H547A</i>
DNA2_H547A_pRCCN_R	TGTCATGACTTTAGGCAATACGATCATTATCTTTCA CTGCGGAG	<i>dna2_H547A</i>
Rev1_pRCC_F	TTGATGAAGCTGTTTGTGTGGTTTTAGAGCTAGAAAT AGCAAGTTAAAATAAGG	<i>rev1-D467A-E468A (CD)</i>
Rev1_pRCC_R	CACACAAACAGCTTCATCAACGATCATTATCTTTCA CTGCGGAG	<i>rev1-D467A-E468A (CD)</i>
RFA1_S351P_pRCCN_F	TCACAATTGTTGACGACTCTGTTTTAGAGCTAGAAAT AGCAAGTTAAAATAAGG	<i>rfa1-S351P</i>
RFA1_S351P_pRCCN_R	AGAGTCGTCAACAATTGTGACGATCATTATCTTTCA CTGCGGAG	<i>rfa1-S351P</i>

RFA1_K45E_pRCC_F	ATAACACCAGGAAATCTGATGTTTTAGAGCTAGAAA TAGCAAGTTAAAATAAGG	<i>rfa1-K45E</i>
RFA1_K45E_pRCC_R	ATCAGATTTCTGGTGTATCGATCATTTATCTTTCA CTGCGGAG	<i>rfa1-K45E</i>
Pol3_D520V_pRCC_F	CATAAGCCTTAAAGGCAGGTGTTTTAGAGCTAGAAA TAGCAAGTTAAAATAAGG	<i>pol3-D520V</i>
Pol3_D520V_pRCC_R	ACCTGCCTTAAAGGCTTATGCGATCATTTATCTTTCA CTGCGGAG	<i>pol3-D520V</i>
For amplification of a template for repair in CRISPR-Cas9 gene editing		
DNA2_H547A_mut_F	AAGACGTTCAATTCTGCAAATGCAATTTCAAGATCC CCGCGGAGAACCAAGTCTTGTTCATGACTTTAGGCAA TATCGTAGCCGAGTTATTGCAAGACTC	<i>dna2_H547A</i>
DNA2_R	CTTCAATATCAATCACATTGGATATAGAAATTGGCT GTGTTCTTCTTGTTCGGAA	<i>dna2_H547A</i>
REV1_mut_F	TACTCTCAAAGATTGAACATATTCAATTTGATTTTA CCTATATCTATTgcagctGCTGTTTGTGTGcgtATAATCCC TGATAA	<i>rev1-D467A-E468A (CD)</i>
REV1_R	TGTGCAACCATTCTGTTCCCTTGA	<i>rev1-D467A-E468A (CD)</i>
RFA1_S351P_mut_F	ATTTGAGCTAACTTCAAGGGCTGGGAAGAAATTCG ATCGTCGTGACATACAATTGTTGACGACCCTGGCTT TTCTATCTCTGTTGGCCTATGGAATC	<i>rfa1-S351P</i>
RFA1_R2	AGGCATATGCCTCAGGAATTTCTGGATTCCGAATCA GGG	<i>rfa1-S351P</i>
RFA1_K45E_F	GTACGATAATCCCACCGGTGGCGTTTATCAAGTTTAT AACACCAGGAAATCTGATGGCGCTAACAGCAACAG AGAGAATTTGATCATGATTTCCGATGG	<i>rfa1-K45E</i>
RFA1_R	CTGACAATAGCAGGTTCTGCAATTATCACGCGAATG ATATCACCCC	<i>rfa1-K45E</i>
POL3_D520V_template	AATGGCGATAGTGAAACAAGAAGAAGTTGGCCGTT TACTGTTTGAAAGTCGCATACCTGCCTTTAAGGCTTA TGAAAAACTAATGGCGTTAGTTAAC	<i>pol3-D520V (ssDNA template)</i>
For Sanger sequencing of the missense mutations		
DNA2_seq_R	CCTTGAACCTCGTATGAAACGCTTC	<i>dna2_H547A</i>
DNA2_seq_F	GCTAACGACAACCTGTTGGTGC	<i>dna2_H547A</i>
REV1_F	AATGTGTAGGGTCCGCATTG	<i>rev1-D467A-E468A (CD)</i>
REV1_R	TGTGCAACCATTCTGTTCCCTTGA	<i>rev1-D467A-E468A (CD)</i>
RFA1_seq_F2	CTCCAACCAGCTAAGCCCCA	<i>rfa1-S351P</i>
RFA1_seq_R2	GATGAAGTTTGCCTTGC	<i>rfa1-S351P</i>
RFA1_seq_F	ACACACCACAAATACCCATTTC	<i>rfa1-K45E</i>
RFA1_seq_R	CGACTGGACCAACTCAAAGTCATC	<i>rfa1-K45E</i>
POL3_seq_F	GTGGTTTTCTAACGGCAGACT	<i>pol3-D520V</i>
POL3_seq_R	CATATTGGTCATCAGAGGCCTGAG	<i>pol3-D520V</i>
RAD27_seq_F	TAACATCGCGCAAATGAAGG	
For knockout fragments amplification		
APN1_pRS_F	ATGCCTTCGACACCTAGCTTTGTTAGATCTGCTGTCT CGAAATACAAATTTGGTGTGCGGGCTGGCTTAA	<i>apn1::HIS3</i>
APN1_pRS_R	TTTGTTAACCTCAAACCTTGTCTAACTGTTCTTACG CGATTTAGCACCTGTTTACAATTTCTGATGCGGTA	<i>apn1::HIS3</i>
CHL1-pRS-F	CCAAAAATGGACAAAAAGGAATATTCGGAGACTTT CTATCATCCTTATAAGCCCGGTGTCGGGGCTGGCTTA A	<i>chl1::HIS3</i>

CHL1_pRS-R	ATCACAGTATACACGTAAACGTATTCCTTTTAGCGTG AATTCAGGCTGCGCATTGGTTTACAATTCCTGATGC GGTA	<i>chl1::HIS3</i>
EXO1_pRS_F	GAAAGGAATGGGTATCCAAGGTCTTCTCCTCAGTT AAAGCCCATACAGAGGTGTCGGGGCTGGCTTAA	<i>exo1::HIS3</i>
EXO1_pRS_R	GATTTTCATTTGAAAAATATACCTCCGATATGAAAC GTGCAGTACTTAACGTTTACAATTCCTGATGCGGTA	<i>exo1::HIS3</i>
MLH1_pAG-F	ATGTCTCTCAGAATAAAAGCACTTGATGCATCAGTG GTTAACAAAATTGCCAGCTGAAGCTTCGTACGC	<i>mlh1::HphMX4</i>
MLH1_pAG-R	TTAACACCTCTCAAAAACCTTGTATAGATCTGGAAG GTTGGCTATTTCCACATAGGCCACTAGTGGATCTG	<i>mlh1::HphMX4</i>
MLH2_pRS_F	CACATCCCATCATCTCGGTTTGAGGAACAGACGCCT TTTCATAGTTTTGGGGTGTCGGGGCTGGCTTAA	<i>mlh2::HIS3</i>
MLH2_pRS_R	TCTATTATGAAGTAATCTATTGTGCTGAGTGGTGATA GTGCACCCGATCAGTTTACAATTCCTGATGCGGTA	<i>mlh2::HIS3</i>
MLH3_pRS_F	CATAAACCCAGCGAGGCTTTCAAGGAAGAATGAACGT GAACTCGTCAACTCGGTTGTCGGGGCTGGCTTAA	<i>mlh3::HIS3</i>
MLH3_pRS_R	GCGCAATTTAAAATGCAGGCGACAAACCTTGTCCA GGATTAAGGTTCTCGTTTACAATTCCTGATGCGGTA	<i>mlh3::HIS3</i>
JK230_MRE11_SP	TAAAGAGAATGCAGACAATTGACGCAAGTTGTACCT GCTCACATCCGATAAACTCGACTCAGCTGAAGCTT CGTACGC	<i>mre11::HphMX4</i>
JK321_MRE11_ASP	TCGCGAAGGCAAGCCCTTGGTTATAAATAGGATATA ATATAATATAGGGATCAAGTACAACATAGGCCACTA GTGGATCTG	<i>mre11::HphMX4</i>
KMH2_F	GCACTCTACTCCAATATCAACTGTAAAAAACTCTTT ATCTGCTGACCTAACATCAAAATCCTCAGATCTGTTT AGCTTGCCTCGTC	<i>msh2::HphMX4</i>
KMS2_R	CCAAAAGACTGATCTGAAATACCAGGCTCAACTTTG TATAACAACGTGATGTCTCATCGATGAATTCGAGC TCGTTTTCG	<i>msh2::HphMX4</i>
JK215_MSH3_SP	GTACTIONTTGAGAGCCAAAAGCAGTGCAAATAGATTT ATTTTGTGTAATCTATTAACAATACAGCTGAAGCTTC GTACGC	<i>msh3::HphMX4</i>
JK216_MSH3_ASP	GCATAAGAAATTGCTATACCATCGTGCGTGCCAGTA CCTCTTCCCACTTCGTCTAATAATCATAGGCCACTAG TGGATCTG	<i>msh3::HphMX4</i>
JK217_MSH6_SP	GATAAGATTTTTTAATTGGAGCAACTAGTTAATTTTG ACAAAGCCAATTTGAACTCCAAACAGCTGAAGCTTC GTACGC	<i>msh6::HphMX4</i>
JK218_MSH6_ASP	TAAAGCATGGATGTCTTAATGATTTAAATTCAGAA AACCATTTAATTGAGTATTCGTTTCATAGGTTACTAG TGGATCTG	<i>msh6::HphMX4</i>
JK246_MUS81_SP	ACATTGGCGTAAACAAAGTTTCAAAGGATTGATACG AACACACATTCCTAGCATGAAAGCCAGCTGAAGCTT CGTACGC	<i>mus81::HphMX4</i>
JK247_MUS81_ASP	AAAGAATATCATCACTTTTTTCTTTATAAAACCTTGC AGGGATGACTATATTTCAAATTGCATAGGCCACTAG TGGATCTG	<i>mus81::HphMX4</i>
OGG1_F	ACTACTATTTCCAGCGGAAGAAGGCATTTGAAGCGT CCTGATTCATAATTGCGATCGGATCCCCGGGTAAATT AA	<i>ogg1::HphMX4</i>

OGG1_R	TTTGCTTCTTTGATGTGAAGATCAGACAATTCAACTT TCAGTTTCATTTGTTTCGCATAGGCCACTAGTGGATC TG	<i>ogg1::HphMX4</i>
PMS1_F	GATGTTTCATCGAATTACATCTGGACAAGTTATCACC GACTTAACAACCTGCCGGATCCCCGGGTTAATTA	<i>pms1::HphMX4</i>
PMS1_R	GGACAATTCCAAGGCTTATCAAGTTCAGTACTGAGATTA TGAACGACTCTGGTCATAGGCCACTAGTGGATCTG	<i>pms1::HphMX4</i>
JK219_POL32_SP	ATAATATTTTACATTAACATAACAACCAGAAATAGGC TTTAGTTAACTCAATCGGTAATTACAGCTGAAGCTTC GTACGC	<i>pol32::HphMX4</i>
JK220_POL32_ASP	CATTGTATTATACATTACATCACAATTAGTAATGGA AAGTGTGGAAAAAAGAAGACATAGGCCACTA GTGGATCTG	<i>pol32::HphMX4</i>
RAD14_F	AGAGTTTGGATCTTCGTAGTGAAGGTATCGAACGTA ACGCTATGACTCCCCGGATCCCCGGGTTAATTA	<i>rad14::HphMX4</i>
RAD14_R	TATACATAACCAACATTTAAATGTCAATTTCTTCAGT TTCTAGCCCCGAGCATAGGCCACTAGTGGATCTG	<i>rad14::HphMX4</i>
RAD18_pRS_F	ATGGACCACCAAATAACCACTGCAAGCGACTTCACG ACTACTTCAATACCGATCTTGGCCTCCTCTAGTACAC TC	<i>rad18::HIS3</i>
RAD18_pRS_R	TTAATTGTTACCGGGTGGGTCTTTACTATATTCATTC AAGTCCATTAATTCTGTTTACAATTTCTGATGCGGT A	<i>rad18::HIS3</i>
RAD2_Hyg_Int_F	CTAGTATAACCCATTCGAACCTCCGTGGAGGCATTA AAAGGGAGAGTGAAATTAGAACGCGGCTACAATTA	<i>rad2::HphMX4</i>
RAD2_Hyg_Int_R	AAGGACCGTATATATCTACTATTCCTGGATCGGTTGA CTTTGTAAACATGCAGAAACACCCTGATTCTGTGGAT AACC	<i>rad2::HphMX4</i>
JK272_RAD27_SP	TATACATCGATGAAAAGCGTTGACAGCATAATTGG AAGAAATAGGAAACGGACACCGGAAGAAAAAATCA GCTGAAGCTTCGTACGC	<i>rad27::HphMX4</i>
JK273_RAD27_ASP	GTATACAAATACTATGTTACATATATGCCAAGGTGA AGGACCAAAAAGAAGAACTGGAAAAAGAACCCCC ATAGGCCACTAGTGGATCTG	<i>rad27::HphMX4</i>
RAD30-pRS-F	TTTTGAACGGCTTTGATAAAAACAAGACAAAGCATGT CAAATTTACTTGGGGTGTGCGGGCTGGCTTAA	<i>rad30::HIS3</i>
RAD30-pRS-R	ATTATCAGGACGTTTTAGTTGCTGAAGCCATATAATT GTCTATTTGGAATGTTTACAATTTCTGATGCGGTA	<i>rad30::HIS3</i>
RAD5del_Hyg_F	CCTTACTGCTAAGCGCATTGCTCACTTGAAAGTAAAT TATCTACAAAGTTACACATACGATTTAGGTGAC	<i>rad5::HphMX4</i>
RAD5del_Hyg_R	TCTATGCTATCTTGTATGATAAATCTCATAACTTTGA CGCTGTTTGTCTGTCTGTGGATAACCGTATTAC	<i>rad5::HphMX4</i>
RAD5_pRS_F	CCTTACTGCTAAGCGCATTGCTCACTTGAAAGTAAAT TATCTACAAAGTTACATTCGGTGATGACGGTGAAA	<i>rad5::HIS3</i>
RAD5_pRS_R	TCTATGCTATCTTGTATGATAAATCTCATAACTTTGA CGCTGTTTGTCTGGTTTACAATTTCTGATGCGGTA	<i>rad5::HIS3</i>
RAD51_Hyg_Int_F	AAATGTTGAAATGCACCACTACCGTTCTTCAACCA ATCTAGTTTAGCTATTTAGAACGCGGCTACAATTA	<i>rad51::HphMX4</i>
RAD51_Hyg_Int_R	AAAGAGGAGAATTGAAAGTAAACCTGTGTAAATAA ATAGAGACAAGAGACCAAATACCTACCCTGATTCTG TGGATAACC	<i>rad51::HphMX4</i>
RAD52_pAG_R	GGTTTCACGCGGTACTTGATTCCCAGCCCCTTCTAGC ATATGAGGCCCCAGTTCTTTATCATCGATGAATTCGA GCTCGTT	<i>rad52::HphMX4</i>

KR52t-F	CGAATGGCGTTTTTAAGCTATTTTGCCACTGAGAATC AACAAATGCAAACAAGGAGGTTGCCAGATCTGTTA GCTTGCCT	<i>rad52::HphMX4</i>
RAD59_pAG_F	GGTTACGTAGAGGAGAAGAGCATATTTTCAGGATAAA CAGACAAAATAATGCAGCTGAAGCTTCGTACGC	<i>rad59::HphMX4</i>
RAD59_pAG_R	GTGAAAATTATGACTTTTTATCAAGCAAATAAATTT GCTACTTGTGCCCATAGGCCACTAGTGGATCTG	<i>rad59::HphMX4</i>
RAD6S_HIS	CATGTCCACACCAGCTAGAAGAAGGTTGATGAGAGA TTTTAAACGTATGAAGGAAGGTGTCGGGGCTGGCTT AA	<i>rad6::HIS3</i>
RAD6A_HIS	TCAGTCTGCTTCGTGTCGTCGTCGTCGTCATCATCA TCATCATCATCGTCCATCTCCTTACGCATCTGTGCGG TA	<i>rad6::HIS3</i>
REV1_pRS_F	ACAGATTTTCTCAAAATAAATCGATACTGCATTTCTA GGCATATCCAGCGGGTGTCCGGGGCTGGCTTAA	<i>rev1::HIS3</i>
REV1_pRS_F	GATATTACAGGTAATGTTTCGAAACTGCGTGTTTACT GTATGCTGAAATGGTTTACAATTTCTGATGCGGTA	<i>rev1::HIS3</i>
REV3_pAG_F	ATGTCGAGGGAGTCAACGACACAATACAGAGCGA TACGGTTAGATCATCCGGATCCCCGGGTTAATTAA	<i>rev3::HphMX4</i>
REV3_pAG_R	GCGAGACATATCTGTGCTAGATTACCAATCATTTAG AGATATTAATGCTGCATAGGCCACTAGTGGATCTG	<i>rev3::HphMX4</i>
REV3_pRS_F	ATGTCGAGGGAGTCAACGACACAATACAGAGCGA TACGGTTAGATCATCGGTGTCGGGGCTGGCTTAA	<i>rev3::HIS3</i>
REV3_pRS_R	GCGAGACATATCTGTGCTAGATTACCAATCATTTAG AGATATTAATGCTGTTTACAATTTCTGATGCGGTA	<i>rev3::HIS3</i>
Ksg1t-F	GGTGATCATTGGTGATACASTTTCCGATTTGTGGCTT TACCGTTTAGTTGTTTTATCAGCCAGATCTGTTTA GCTTGCCT	<i>sgs1::HphMX4</i>
Ksg1h-R	CGCACCAGTGATGGCTAATGCCTTAGTGACGGTAGT CGCAGTAGTACTTGTGAGGTTTGAATTCGAGCTCGTT TTCGACA	<i>sgs1::HphMX4</i>
JK180_SRS2_SP	TCTGCACCTTTGAGTATCATTCCAATTTGATCTTTCTTC TACCGGTACTIONTAGGGATAGCAACAGCTGAAGCTTCG TACGC	<i>srs2::HphMX4</i>
JK181_SRS2_ASP	CTTTGGCACCGTGAATTGTAGATATCGTGACAAACC CATTCTTCTCACGTTTTATTTTTGCATAGGCCACTAG TGGATCTG	<i>srs2::HphMX4</i>
STM1-pAG-F	CAAAGATGGCAAGTTAGAAACGTCAATGTTACGGTT CTTTTGAACGGTGTGCATAGGCCACTAGTGGATCTG	<i>stm1::HphMX4</i>
STM1-pAG-R	ATAGTGTCTTATCTAACAATCACTTCTGTGCAAAGTT CTAATTGTCTGGTCAGCTGAAGCTTCGTACGC	<i>stm1::HphMX4</i>
TOF1_SP	GTAAGTCGCCTCACATATGATAATACCATCTAGCTTG TGGGGTTTATGTATCTTCAGCTGAAGCTTCGTACGC	<i>tof1::HphMX4</i>
TOF1_ASP	TCTGTAGCTCTTATGCTTTCAATACTTGGTATGGATC CACCAAACAAGCTCGTATCATAGGCCACTAGTGGAT CTG	<i>tof1::HphMX4</i>
TOF1-pRS_F	GTAAGTCGCCTCACATATGATAATACCATCTAGCTTG TGGGTTCCGGTATGACGGTGAAA	<i>tof1::HIS3</i>
TOF1-pRS_F	TCTGTAGCTCTTATGCTTTCAATACTTGGTATGGATC CACCGTTTACAATTTCTGATGCGGTA	<i>tof1::His3</i>
YKU70_SP	GCGCTCAGTCACTAATGCATTTGGCAATAGTGGAGA ACTTAACGATCAAGTGGATCAGCTGAAGCTTCGTAC GC	<i>yku70::HphMX4</i>
YKU70_ASP	GCCTTTGGATGATTGGATCTTCTGACTTCTCCAGATT CTAAAATTTTATTTTCGCATAGGCCACTAGTGGATCTG	<i>yku70::HphMX4</i>

YKU70-pRS_F	GCGCTCAGTCACTAATGCATTTGGCAATAGTGGAGA ACTTAACGATCAAGTGGATTCGGTGATGACGGTGAA A	<i>yku70::HIS3</i>
YKU70-pRS_R	GCCTTTGGATGATTGGATCTTCTGACTTCTCAGATTC TAAAATTTTATTTTCGGTTTACAATTCCTGATGCGGT A	<i>yku70::HIS3</i>
For knockout verification		
APN1_chk_F	CTGGGAACCTGAACGTGGA	<i>APN1</i>
APN1_chk_R	GGGCAACAGCATCTTGGA	<i>APN1</i>
APN1_int_F	TGAAAGGAGACCATCAGTTGC	<i>APN1</i>
APN1_int_R	ATGTATCTATGCAAACGCCGA	<i>APN1</i>
CHL1-upstr-F	GGCACTACTGCAACTTCAGT	<i>CHL1</i>
CHL1-F	CCCTCGTTCCCATCATCCTT	<i>CHL1</i>
CHL1-R	CGTATCAGGACAATGACGCC	<i>CHL1</i>
EXO1_KOcheckFwd	GTATTACGTCCAAACTAAGTTCGCG	<i>EXO1</i>
EXO1_KOcheckRev	GACCGCTAGCGGCTTGATTAG	<i>EXO1</i>
EXO1_F	CAGCGGGAGGGAAAAGTCTGAT	<i>EXO1</i>
EXO1_R	CTCTGTTGGCTAGAGTTGGTG	<i>EXO1</i>
MLH1-upstr-F	GTAATCGCGCTAGCATGCTA	<i>MLH1</i>
MLH1-F	GCGTTGATGGAAAGGTGTGT	<i>MLH1</i>
MLH1-R2	CAATGGCAGATAATTCGGCG	<i>MLH1</i>
MLH2_upstr_F	CATATCCCTCATATACATGGCCC	<i>MLH2</i>
MLH2_dnstr_R	GTGCGGTTACCATGAGTTAC	<i>MLH2</i>
MLH2_F	TGAACTTGGAGACGGGGAAGAC	<i>MLH2</i>
MLH2_R	CGGGTGTAGGTATCACCAGTGC	<i>MLH2</i>
MLH3_upstr_F	TTTGCCTTTATTTGCGAGCG	<i>MLH3</i>
MLH3_dnstr_R	GGTTTTGACAACGTGATGAGG	<i>MLH3</i>
MLH3_F	GTTTGGGGCAATAATACCACCGG	<i>MLH3</i>
MLH3_R	CGAACATCAAGGATGAAGACGGG	<i>MLH3</i>
JK232 MRE11_fwd	CCAATCATTTTCGACCGTCACTC	<i>MRE11</i>
JK254 MRE11_int_fwd	TGGATATACTTCATGCGACTGG	<i>MRE11</i>
JK255 MRE11_int_rev	ATGACCCCATATCACCATATCC	<i>MRE11</i>
MSH2_upstr_F	GCACTCCATCAAGTGAACCT	<i>MSH2</i>
MSH2_dnstr_R	TCGTTCCGACCTAACATCTC	<i>MSH2</i>
MSH2_F	CATCCCATGGATTCGGAAAG	<i>MSH2</i>
MSH2_R	CAGCTTCTTACAAGGTACG	<i>MSH2</i>
JK222-MSH3-rev	AAGGGGCAGTCACTTAACTCAG	<i>MSH3</i>
MSH3_F	GATTTACCACTCCCAGAACCCA	<i>MSH3</i>
JK221 MSH3_fwd	GTGTTCAAATCACGGTATGTGG	<i>MSH3</i>
MSH6-F	GGCTCGATAGTGTGACTCTTT	<i>MSH6</i>
JK223 MSH6_fwd	TGACATAATGAATGGCTTCTGG	<i>MSH6</i>
JK224 MSH6_rev	CCCGTTAACAATCCTAATCTGG	<i>MSH6</i>
JK248 MUS81_fwd	AGAGGTGGTGGTCAAATCATCC	<i>MUS81</i>
JK249 MUS81_rev	ACTGCCTCCAATTTGATTGCC	<i>MUS81</i>
JK260 MUS81_int_fwd	CACAGCAAATCTGACTGACCTC	<i>MUS81</i>
JK261 MUS81_int_rev	TTCGAAATCACCCTACACCAC	<i>MUS81</i>
OGG1_chk_F	CGCCTTTCTTAATGTAACGCC	<i>OGG1</i>
OGG1_chk_R	GCTCCTTTAAAGAATATGTATCGCC	<i>OGG1</i>
OGG1_in_chk_F	GCTCTAATTTCCGGAACTTAATCAC	<i>OGG1</i>
OGG1_in_chk_R	AGATACTTTGAAGGTATGTGGTGC	<i>OGG1</i>
PMS1_chk_F	GAACAGAGGTATATCCCTGTGAAA	<i>PMS1</i>
PMS1_chk_R	CCATCAAGCATCTTCAATGCAC	<i>PMS1</i>
PMS1_chk_in_F	CTCACAATCAGAACAACAAGCTC	<i>PMS1</i>

PMS1 chk in R	TTAGTCGGTGTACTTGAGTTGC	<i>PMS1</i>
JK225 POL32 fwd	TTTCCACTACGGTGTAAC TTTCC	<i>POL32</i>
POL32 in F	GACCACGCCAGAAGAAACAA	<i>POL32</i>
POL32 in R	GCTGTCGTTTCCAACAAGTC	<i>POL32</i>
RAD14 chk F	CGTTTGCTAAGTTGTAGGGAGA	<i>RAD14</i>
RAD14 chk R	GTACGAGTGACAAATGGGATATCA	<i>RAD14</i>
RAD14 chk in F	CCGATGACCAAGAATTTGAATCTG	<i>RAD14</i>
RAD14 chk in R	CTGGATGCTCCTTAGAACACTG	<i>RAD14</i>
RAD18 upstr F	GAGCAATGCCACATTAGAAG	<i>RAD18</i>
RAD18 dnstr R	GTGCACAAGCTAACAAACAG	<i>RAD18</i>
RAD18 F	CCACTGAGTTCCAAACCATC	<i>RAD18</i>
RAD18 R	GACTTCTGGAGTTCGTACCT	<i>RAD18</i>
RAD2 chk F	TTGATGTTTCCAGAGGATGTGA	<i>RAD2</i>
RAD2 F	ACCGTCTCAGCAGGAGGATA	<i>RAD2</i>
RAD2 R	GAGATTTAGTGGGAACGTCCTC	<i>RAD2</i>
JK274 RAD27 fwd	GACTAGTACCCGGCTGAATCAC	<i>RAD27</i>
JK275 RAD27 rev	CTGTGTATACTGCGGAGAAACC	<i>RAD27</i>
JK276 RAD27 int fwd	CAAAGAAGGGAAAGGTGTATGC	<i>RAD27</i>
JK277 RAD27 int rev	CCCCAGATTCAATAAACTCCAC	<i>RAD27</i>
RAD30 upstr F	TTGTAGTCTTCTAGCGCAGG	<i>RAD30</i>
RAD30 dnstr R	GCGCCCGTGAATCATTTAGA	<i>RAD30</i>
RAD30 F	CGTGTGGTTTGTCTAGCACC	<i>RAD30</i>
RAD30 R	CAGACCGTACCGGCCTCTAC	<i>RAD30</i>
RAD5 del chk F	TTACGCGTCATAAACCCCTT	<i>RAD5</i>
RAD5 del chk R	ACAGCATCTGGATTTCTTCA	<i>RAD5</i>
RAD5 in F	CTGAAGTCCATAACAATCTCCGA	<i>RAD5</i>
RAD5 in R	GCCATTTGAACTGCTTCCATAA	<i>RAD5</i>
R51TF	AGTACGCGTGGTGGGACCATA	<i>RAD51</i>
R51TR	GGACCCTGCAGGAGGAAGTA	<i>RAD51</i>
RAD51 chk in F	AGATCGGAGCTGATTTGTTTGAC	<i>RAD51</i>
RAD51 chk in R	CTTCACCGCCACCAATATCC	<i>RAD51</i>
Ex52 R	ACGTCGCTAAAGATGGTATGGTA	<i>RAD52</i>
RAD52 chk in F	GCCAAGAAATCTGCCGTTAC	<i>RAD52</i>
RAD52 chk in R	TGAGCTTTTCGCTGATTTTCATCC	<i>RAD52</i>
RAD59 upstr F	GCAAGGGCAGATATGATAGG	<i>RAD59</i>
RAD59 dnstr R	CCTTCGTTACCTTGGAATGG	<i>RAD59</i>
RAD59 F	TTCGACTACATACGGCACAG	<i>RAD59</i>
RAD59 R	GCTTGCTATTAGTCGCTGAC	<i>RAD59</i>
RAD6 TS	GCCGGAGTAGAAAGCTGGAA	<i>RAD6</i>
RAD6 TA	AAAGATACGGGTATCGGCAGTT	<i>RAD6</i>
RAD6 F	GGGTGTATCTGCTTCACCAT	<i>RAD6</i>
RAD6 R	ATCGTCCATATCATCCTCCC	<i>RAD6</i>
REV1 upstr F	TACGGCAACCTTTAAGCACC	<i>REV1</i>
REV1 upstr R	GAGTCGGCCATTCCAATACC	<i>REV1</i>
REV1 F	AATGTGTAGGGTTCGGCATTG	<i>REV1</i>
REV1-R	TGTGCAACCATTTCCTTGA	<i>REV1</i>
REV3 upstr F	CGAGTGCAGTGCCTAGAAATAGTGT	<i>REV3</i>
REV3 F	GCATGCACACCCCTCATAGTAAGT	<i>REV3</i>
REV3 2400B	TGGCATTGACTCTGGCAAGTTCC	<i>REV3</i>
AS36 SGS1	CGTGCGTTTTCGAAGTGGATTG	<i>SGS1</i>
JK341 SGS1 int fwd	TGCAAAC TTTGTGCGAACGATAC	<i>SGS1</i>
JK342 SGS1-int rev	CGACAAGAGA ACTAGCCATGTG	<i>SGS1</i>
ESrsH F	CAGCTATCCTGATACTACTGCTT	<i>SRS2</i>

JK300 SRS2 int fwd	GCTTCCTCAATCACAAGGAAAG	<i>SRS2</i>
JK301 SRS2 int rev	TCTAGTCAGCAAACGAAAGGTG	<i>SRS2</i>
STM1 upstr F	CAAATTTCTCTTCCCCCAC	<i>STM1</i>
STM1 F	GCTGAAAAGGAAGCTCAAGC	<i>STM1</i>
STM1 R2	CAACCTTCTTAGCTTCTGGG	<i>STM1</i>
JK105 TOF1 fwd	TTCTGAAGACCACAGCAACG	<i>TOF1</i>
TOF1 in F	CAAGCATACATCTGCAAGACACT	<i>TOF1</i>
TOF1 in R	AAGTTACCATCCTATCCTGCTCA	<i>TOF1</i>
KU70 test SP	TTAATTGACTCTCGGTAGCCAAGTT	<i>YKU70</i>
KU70 int F	GCATGAAGATATCAGACAAGAAGC	<i>YKU70</i>
KU70 int R	TGCTCGATGAACGGAACC	<i>YKU70</i>
Hyg-R	GACTGTCAAGGAGGGTATTC	
JK183 hygRleft rev	ACAGTCACATCATGCCCTG	
HIS3H R	CCTGTGTGGACGTTAATCACTTGCGAT	
For amplification of <i>RAD27::rad27-HphMX4</i> fragments		
RAD27_ASP	AAAAACTGGCAAAAAAAGAGAAGTATTAGATGAAA AAAGTTCGTGTATACAAATATCTATGTTACATATACA TAGGCCACTAGTGGATCTG	
RAD27_G240D_mut	TTGCATAATGCTTGGTTGTGACTACTGTGAAAGCATC AGAGGTGTTGATCCAGTGACAGCCTTAAAATTGATA	
RAD27_R104A_R105A_K13 0A_R127A	AGCGGTCTTCAGCTGCTGTGGAAACAGAAAAAAAAC TGGCAGAGGCAACAACAGAATTGGAAAAGATGAAG CAAGAAAGAGCTTTGGTGGCTGTCTCAA	
RAD27_G67S_mut	ACGGTGGGCAGTTGACCAATGAAGCCGGTGAACA ACGTCACACTTGATGTCCATGTTTTATAGGACACTGA	



**Table S3.** Yeast strains used in this project

Strain	Genotype	Notes
SMYU	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::P<sub>Gal1</sub>-UR-(GAA)<sub>124</sub>-A3-TRP1</i>	(1)
SMYU7/8	SMYU <i>mre11::HphMX4</i>	
SMYU14/16	SMYU <i>msh3::HphMX4</i>	
SMYU22/23	SMYU <i>rad52::HphMX4</i>	
SMYU28/31/32	SMYU <i>msh6::HphMX4</i>	
SMYU29/41	SMYU <i>rad51::HphMX4</i>	
SMYU30/89	SMYU <i>msh2::HphMX4</i>	
SMYU35/36	SMYU <i>rad59::HphMX4</i>	
SMYU39/40	SMYU <i>srs2::HphMX4</i>	
SMYU44/46	SMYU <i>rad27::HphMX4</i>	
SMYU42/43	SMYU <i>tof1::HphMX4</i>	
SMYU47/48	SMYU <i>stm1::HphMX4</i>	
SMYU51/52	SMYU <i>mus81::HphMX4</i>	
SMYU55/67	SMYU <i>yku70::HphMX4</i>	
SMYU60/61	SMYU <i>rad6::HIS3</i>	
SMYU63/64	SMYU <i>pol32::HphMX4</i>	
SMYU70/291	SMYU <i>rad5::HphMX4</i>	
SMYU71/72	SMYU <i>rad27::rad27-4A-HphMX4</i>	
SMYU73/73	SMYU <i>sgs2::HphMX4</i>	
SMYU76/77	SMYU <i>rad18::HIS3</i>	
SMYU81/82	SMYU71 <i>rad5::HIS3</i>	
SMYU83/84/85	SMYU71 <i>yku70::HIS3</i>	
SMYU86/87	SMYU71 <i>tof1::HIS3</i>	
SMYU90/93	SMYU <i>rev3::HIS3</i>	
SMYU95/96	SMYU <i>chl1::HIS3</i>	
SMYU101/102	SMYU <i>rad27::rad27-G67S-HphMX4</i>	
SMYU114/120	SMYU <i>rad27::rad27-G240D-HphMX4</i>	
SMYU116/117	SMYU <i>mlh1::HphMX4</i>	
SMYU125/127	SMYU <i>rev1::HIS3</i>	
SMYU129/131	SMYU <i>rad30::HIS3</i>	
SMYU133/134	SMYU <i>oggl::HphMX4</i>	
SMYU135/136	SMYU <i>rad2::HphMX4</i>	
SMYU137/138	SMYU <i>rad14::HphMX4</i>	
SMYU143/144	SMYU <i>exo1::HIS3</i>	
SMYU150	SMYU114 <i>tof1::HIS3</i>	
SMYU151	SMYU120 <i>tof1::HIS3</i>	
SMYU152/152	SMYU <i>apn1::HIS3</i>	
SMYU157/162	SMYU114 <i>rad6::HIS3</i>	
SMYU191/192	SMYU <i>pol3-D520V</i>	
SMYU195/196/197/198	SMYU <i>rfa1-S351P</i>	

SMYU201/202	SMYU <i>dna2-H547A</i>	
SMYU204/208	SMYU63 <i>rev1::HIS3</i>	
SMYU220/221/222	SMYU <i>pms1::HphMX4</i>	
SMYU225/226	SMYU129 <i>rev3::HphMX4</i>	
SMYU227	SMYU131 <i>rev3::HphMX4</i>	
SMYU233/234	SMYU <i>rfa1-K45E</i>	
SMYU244/245	SMYU <i>mlh2::HIS3</i>	
SMYU255/256	SMYU <i>mlh3::HIS3</i>	
SMYU298/299	SMYU <i>rev1-CD</i>	
SMYUI2/4	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::A3-(GAA)<sub>124</sub>-UR-P<sub>Gall</sub>-TRP1</i>	
SMYUI10/11	SMYUI2 <i>rev1::HIS3</i>	
SMYUI15/16	SMYUI <i>yku70::HIS3</i>	
SMYUI22/23	SMYUI2 <i>pol32::HphMX4</i>	
SMYUI30/31	SMYUI2 <i>rad52::HphMX4</i>	
SMYUI34/35/36	SMYUI2 <i>chl1::HIS3</i>	
SMYUD14/15/16	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::P<sub>Gall</sub>-UR-(GAGAA)<sub>74</sub>GA-A3-TRP1</i>	
SMYUID13/14/15	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::A3-(GAGAA)<sub>74</sub>GA-UR-P<sub>Gall</sub>-TRP1</i>	
SMYUD23/24/25	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::P<sub>Gall</sub>-UR-(GAGAAGAAA)<sub>41</sub>GAG-A3-TRP1</i>	
SMYUID16/17	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::A3-(GAAGAAGAAA)<sub>41</sub>GAG-UR-P<sub>Gall</sub>-TRP1</i>	
SMYUD21/22	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::P<sub>Gall</sub>-UR-(GAGAAGGAAA)<sub>37</sub>GA-A3-TRP1</i>	
SMYUID25/26	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::A3-(GAAGAAGGAAA)<sub>37</sub>GA-UR-P<sub>Gall</sub>-TRP1</i>	
SMYUD4/5/6	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::P<sub>Gall</sub>-UR-(GAAGAA)<sub>64</sub>-A3-TRP1</i>	
SMYUID1/2	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ade2Δ::KanMX4, HAP1-wt, ChrIII(75594-75641)::A3-(GAAGAA)<sub>64</sub>-UR-P<sub>Gall</sub>-TRP1</i>	
SMYP1/3	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ChrIII(75594-75641)::P<sub>Gall</sub>-UR-GAA<sub>124</sub>-A3-TRP1</i>	
SMYP4/7	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, pol1-Y868F, ChrIII(75594-75641)::P<sub>Gall</sub>-UR-(GAA)<sub>124</sub>-A3-TRP1</i>	(3)

SMYP7/15	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, poll-Y869A, ChrIII(75594-75641)::P<sub>Gal1</sub>-UR-(GAA)<sub>124</sub>-A3-TRP1</i>	(3)
SMYPI9	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, ChrIII(75594-75641)::A3-(GAA)<sub>124</sub>-UR-P<sub>Gal1</sub>-TRP1</i>	
SMYPI4/5/6	<i>MATa, leu2-Δ1, trp1-Δ63, ura3-52, his3-200, poll-Y869A, ChrIII(75594-75641)::A3-(GAA)<sub>124</sub>-UR-P<sub>Gal1</sub>-TRP1</i>	(3)

## Supplemental references

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