# Voluntary running does not reduce neuroinflammation or improve non-cognitive behavior in the 5xFAD mouse model of Alzheimer's disease

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# **Supplementary Data**

## **Supplementary Table 1- Body weights**

	Sedentary (Mean±SD)	Running (Mean±SD)	T-test (unpaired) Sedentary vs.
Start Weight (g)	16.8±1.1	16.1±1.3	Exercised P=0.14
Final Weight (g)	23.4±1.3	22.6±3.0	P=0.35
Weight gain (%)	40±11	40±16	P=0.98

# **Supplementary Table 2- Corticosterone levels in feces (ELISA)**

	Sedentary	Running	Mann Whitney
	Median (IQR)	Median (IQR)	U-test
			Sedentary vs.
			Running
<b>Baseline levels</b>	2617 (1699-4455)	2167 (1644-4053)	P=0.66
(pg/ml)			
After 19 weeks	1523 (1331-2205)	1506 (1237-1722)	P=0.85
(pg/ml)			
Wilcoxon test	P=0.001	P=0.02	
Baseline vs.			
After 19 weeks			

# Supplementary Table 3- distance traveled in EPM and OF

	Sedentary	Running	Mann Whitney
	Median (IQR)	Median (IQR)	U-test
			Sedentary vs.
			Running
Distance moved	1048 (741-1357)	1278 (964-1616)	P=0.23
Elevated plus			
maze			
Distance moved	4569 (3115-5561)	4017 (3746-5568)	P=0.93
Open field			

# **Supplementary Table 4- Sucrose Preference**

	Sedentary	Running	Mann Whitney
	Median (IQR)	Median (IQR)	U-test
			P-value
Sucrose	79.6 (76.4-86.1)	79.7 (73.2-88.4)	1.0
Preference (%)			

# Supplementary Table 5- Aβ levels (ELISA)

	Different Aβ species	Sedentary concentration (ng Aβ/mg protein) Median (IQR)	Running concentration (ng Aβ/mg protein) Median (IQR)	Mann Whitney U- test p-values prior to Bonferroni correction
Insoluble fraction in	Αβ-38	164.5 (131.5- 322.4)	255.3 (147.8- 303.4)	0.57
hippocampus	Αβ-40	1048 (712.4- 1286)	1194 (989.7-1526)	0.35
	Αβ-42	8007 (5909- 10173)	9612 (4285- 11546)	0.78
<b>CSF</b> (n=7+7)	Αβ-38	0.59 (0.54-0.88)	0.44 (0.19-0.54)	0.21
	Αβ-40	3.87 (2.17-4.92)	2.28 (0.76-3.23)	0.32
	Αβ-42	1.81 (1.18-2.46)	1.25 (0.35-1.68)	0.32

# **Supplementary Table 6- Iba1 and gal-3 in hippocampus (immunohistochemistry)**

	Sedentary Running		Mann Whitney	
	Median (IQR)	Median (IQR)	U-test	
	Fold to actin %	Fold to actin %	P-value	
Galectin-3	116 (84-125)	114 (102-127)	0.94	
(n=6+6)				
NLRP3	22 (18-33)	17 (16-20)	0.63	
(n=3+4)				

# **Supplementary Table 7- Cytokine levels (ELISA)**

	Cytokine	Sedentary	Running	T-test
		Mean	Mean	P-values prior
		concentration	concentration	to Bonferroni
		$(pg/ml) \pm SD$	$(pg/ml) \pm SD$	correction
Hippocampus	IL-1 β	$1.29 \pm 0.7$	$1.34 \pm 0.8$	0.85
	IL-2	$0.022 \pm 0.01$	$0.029 \pm 0.02$	0.26
	IL-4	$0.045 \pm 0.02$	$0.052 \pm 0.04$	0.58
	IL-5	$0.008 \pm 0.003$	$0.010 \pm 0.005$	0.26
	IL-6	$0.45 \pm 0.3$	$0.68 \pm 0.7$	0.27
	IL-10	$0.15 \pm 0.09$	$0.15 \pm 0.07$	0.99
	IL-12p70	$1.57 \pm 0.8$	$1.78 \pm 0.9$	0.52
	IFNγ	Below	Below	
		detection	detection	
	TNFα	$0.067 \pm 0.02$	$0.082 \pm 0.06$	0.40
	KC/GRO	$2.75 \pm 1.9$	$2.94 \pm 2.6$	0.82
Serum	IL-1 β	$0.27 \pm 0.2$	$0.20 \pm 0.2$	0.37
	IL-2	$0.12 \pm 0.1$	$0.13 \pm 0.1$	0.84
	IL-4	Below	Below	
		detection	detection	
	IL-5	$1.48 \pm 1.3$	$1.68 \pm 1.0$	0.64
	IL-6	$3.25 \pm 2.6$	$3.59 \pm 4.2$	0.80
	IL-10	$7.14 \pm 3.9$	$6.14 \pm 3.6$	0.48
	IL-12p70	Below	Below	
		detection	detection	
	IFNγ	$0.33 \pm 0.1$	$0.20 \pm 0.1$	0.06
	TNFα	$3.66 \pm 1.0$	$4.17 \pm 1.8$	0.38
	KC/GRO	$34.14 \pm 29.8$	$23.52 \pm 10.2$	0.22

### **Supplementary Figure S1- Distance in running wheels**

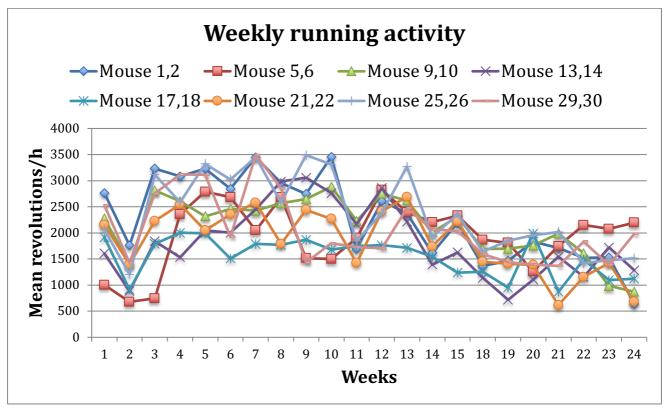


Figure S1. Running activity displayed as mean revolution per hour for each week and each couple sharing a running wheel in their home cage. Each revolution corresponds to a running distance of around 42 cm. ANOVA repeated measurements showed a change in the amount of running over time (p<0.001) and a paired T-test comparing the running during the first week with that of the last week of intervention revealed a trend towards decreased running over time (means $\pm$ SD were 2039 $\pm$ 549 revolutions/h during the first week compared to 1288 $\pm$ 578 revolutions/h during the last week, paired T-test, p=0.07).

#### **Supplementary Methods**

Sucrose preference test

To assess anhedonic behavior, a Sucrose preference test was performed during the night before sacrifice. Mice were introduced to a sucrose solution in their home cages one night before the test. A bottle containing 2% sucrose solution was put in the place where the regular bottle with tap water used to be during the night. The regular bottle with tap water was placed in the other corner of the cage, allowing the mice to choose. The day before the test, mice were deprived from drinking five hours prior to the test. Later, mice were individually caged with access to nesting material, food pellet, as well as two bottles, one tap water and one sucrose solution as described

before <sup>1</sup>. Bottles were weighed before and after the test and the volume consumed was calculated. A sucrose preference index was calculated using the following formula:

Sucrose preference index=weight of consumed sucrose/total weight consumed of both solutions

#### **Multiplex ELISA**

#### Cytokine ELISA

The concentrations of different cytokines in serum as well as in the pooled first and second fraction of homogenized hippocampus (25  $\mu$ l/sample) were measured with the MSD Mouse Proinflammatory V-Plex Plus Kit (IFN $\gamma$ , IL-1 $\beta$ , IL-2, IL-4, IL-5, IL-6, IL-10, IL-12p70, CXCL1, TNF $\alpha$ ; K15012C, Mesoscale) using a QuickPlex SQ120 (Mesoscale Discovery, Rockville, USA) Plate Reader according to the manufacturer's instructions. The recorded data was analyzed using MSD Discovery Workbench software. For the brain homogenate samples, the cytokine concentrations were normalized to the total protein concentrations measured in the BCA or Bradford assay.

#### *Aβ ELISA*

The concentration of different Aβ species in the insoluble fraction of homogenized hippocampus as well as in the CSF were measured with the MSD MULTI-SPOT Human (4G8) Aβ Triplex Assay (Aβ38, Aβ40 and Aβ42; K15199G-1, Mesoscale) using QuickPlex SQ120 (Mesoscale Discovery, Rockville, USA) Plate Reader according to the manufacturer's instructions. The recorded data was analyzed using MSD Discovery Workbench software. For the brain homogenate samples, Aβ concentrations were normalized to total protein concentrations measured in the BCA or Bradford assay.

#### Fecal corticosterone levels

Fecal samples were collected from the Open field arena after conducting the Open field test in order to measure the stress levels of the mice. The feces were stored at -80°C until use. Corticosterone was then extracted and analyzed with a corticosterone ELISA kit (Enzo Life Sciences) described by Touma et al.<sup>2</sup> except that feces was homogenized in 1 ml of 80% Methanol per 100 mg sample, as we have done before<sup>3</sup>.

#### References

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