

Nuclear bodies formed by polyQ-ataxin-1 protein are liquid RNA/protein droplets with tunable dynamics

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Supplementary Figure Legends

Supp figure 1. Following photobleaching of GFP-ataxin-1[85Q], fluorescence recovers from both within the ataxin-1 NB and the surrounding nucleoplasm.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24 h post-transfection, cells were used for FRAP. (A) Representative images are shown from 3 independent experiments for quantitative assessments of GFP-ataxin-1[85Q] exchange dynamics. White rectangles (in pre-bleach images) and open arrowheads (in post-bleach images) indicate the ataxin-1 NB analyzed. Different ROIs (ROI 1: across the NB, ROI 2: inside the NB, ROI 3: the entire NB; indicated by white open arrowheads) were photobleached and fluorescence recovery subsequently monitored post-bleach at 5 s intervals for 250 s. Scale bar = 10 μ m. (B) Plot of fluorescence recovery (%) from experiments as shown in (A). Each symbol represents fluorescence measured at the indicated time. (C-D) Pooled data from FRAP experiments measuring recovery of fluorescence for GFP-ataxin-1[85Q] over time. Each symbol represents a single data point. The results are also represented as the mean \pm SEM for (C) fluorescence maximum recovery percentage in the bleached area (where 100% represents full recovery) and (D) initial rates (average percentage recovery of fluorescence in the first 15 s) (Fn%/s). (Significance values calculated by ANOVA, $n > 10$, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, n.s. = not significant).

Supp figure 2: Fusion of ataxin-1[85Q] NBs observed during live cell imaging.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24 h post-transfection, live cell imaging reveals fusion events shown here for two NBs (white square in the top panel, shown as enlarged images in the zoom panels). Representative images are shown from 3 independent experiments. All scale bars = 10 μ m.

Supp figure 3: Arsenite treatment increases ataxin-1[85Q] NB average size but does not change the total NB size per nucleus.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24h post-transfection, cells were left untreated (Ctrl) or treated with arsenite (0.3 mM, 1 h) as indicated. Cells were then fixed and stained with DAPI before CLSM imaging. (A) Representative images from 3 independent experiments (~100 cells per condition) for quantitation of NB average size and the total NB size per nucleus are shown. White arrows indicate NBs with visibly increased size upon arsenite exposure. Scale bar = 50 μ m. (B-C) Quantitation of NB size. Each symbol represents a single data point. The results are also represented as the mean \pm SEM for (B) average size of ataxin-1 NBs or the (C) total size of the ataxin-1 NBs per nucleus. (Significance values calculated by unpaired two-tailed student's t-test, * $p < 0.05$, n.s. = not significant).

Supp figure 4: Ataxin-1[85Q] self association domain mutant (Δ SAD) forms smaller NBs that do not increase in size following arsenite stress.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q] or GFP-ataxin-1[85Q] Δ SAD. At 24h post-transfection, cells were left untreated or treated with arsenite (0.3 mM, 1 h) as indicated. Cells were then fixed and stained with DAPI before CLSM imaging. (A) Representative images from 3 independent experiments (~100 cells per condition) for quantitation of NB average size are shown. White squares in the right panels are shown as enlarged images in the zoom panels. Scale bars = 50 μ m (left panels without zoom) or 10 μ m (right panels with zoom). (B) Quantitation of NB size. Each symbol represents a single data point. The results are also represented as the mean \pm SEM. (Significance values calculated by ANOVA, *** p <0.001, n.s. = not significant). (C) Cell death percentage for the transfected cell population was assessed by staining with the SYTOX Red dead cell stain followed by flow cytometry analysis.

Supp figure 5: Ataxin-1[30Q] also forms NBs that show high dynamics and fusion

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24h post-transfection, cells were incubated in an imaging chamber equilibrated with 5% CO₂ at 37°C prior to FRAP using CLSM. (A) Representative images are shown from 3 independent experiments for quantitative assessments of GFP-ataxin-1[85Q] exchange dynamics. White rectangle indicates ataxin-1 NB analyzed. A small ROI (indicated by white open arrowhead for dynamic NB) was photobleached and fluorescence recovery was subsequently monitored post-bleach at 5 s intervals for 250 s. (B) Plot of fluorescence recovery (%) from experiments as shown in (A). Each symbol represents fluorescence measured at the indicated time. (C) Pooled data from FRAP experiments measuring recovery of fluorescence for GFP-ataxin-1[30Q] over time (average percentage recovery of fluorescence in the first 15 s) (Fn%/s). Each symbol represents a single data point. The results are also represented as the mean \pm SEM ($n > 7$). (D) Live cell imaging reveals fusion events shown here for two NBs (white square in the top panel, shown as enlarged images in the zoom panels). Representative images are shown from 3 independent experiments. All scale bars = 10 μ m.

Supp figure 6: Arsenite or sorbitol stress treatment does not alter total GFP-ataxin-1[85Q] fluorescence intensity.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24h post-transfection, cells were left untreated or treated with arsenite (0.3 mM, 1 h) or sorbitol (0.5M, 1h) as indicated. Cells were then fixed and stained with DAPI before CLSM imaging using identical settings and laser intensity for all three conditions. (A) Representative images from 3 independent experiments (~100 cells per condition) for quantitation of total GFP-ataxin-1[85Q] fluorescence intensity per nucleus are shown. Scale bars = 50 μ m. (B) Quantitation of total GFP-ataxin-1[85Q] fluorescence intensity per nucleus. Each symbol represents a single data point. The results are also represented as the mean \pm SEM. (Significance values calculated by ANOVA, *** p <0.001, n.s. = not significant).

Supp figure 7: Intracellular ATP is depleted by CCCP+2DG treatment but recovers after inhibitor wash-out.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q] in a 96-well plate. At 24 h post-transfection, cells were treated for the times indicated with 2DG and CCCP (inhibitors of glycolysis and oxidative phosphorylation, respectively). For ATP recovery, inhibitors were removed by three washes with recovery for the times indicated. ATP levels were measured using a luminescent detection assay kit. (Significance values calculated by ANOVA, n=3 independent experiments, **p<0.01, ****p<0.0001, n.s. = not significant).

Supp figure 8: siRNA-mediated knockdown of RNA helicases.

Neuro-2a cells were co-transfected with GFP-ataxin-1[85Q] and different siRNAs: negative ctrl siRNA, siDDX21, siDDX19A, siDDX42, siDDX46, or siDHX15. Protein lysates were analysed by Western blotting using specific antibodies to the following proteins: DDX21, DDX19A, DDX42, DDX46 and DHX15; detection of tubulin was used as a loading control. Arrows indicate the protein bands of interest according to the size predicted for each protein. Representative results of full-length blots are shown from 3 independent experiments.

Supp figure 9: Endogenous RNA helicases DDX42, DDX46 and DHX15 show nuclear localization.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24 h post-transfection, cells were left untreated or treated with arsenite (0.3 mM, 1 h) as indicated. (A) Cells were then fixed and immunostained to visualize the RNA helicases DDX21, DHX19A, DDX42, DDX46 and DHX15, together with DAPI staining of DNA, before CLSM imaging. All scale bars = 10 μ m. Representative images are shown from 3 independent experiments. (B) Quantitative analysis for the fluorescence ratio nucleus to cytoplasm (Fn/c) for each of the RNA helicases detected with immunostaining. Each symbol represents a single data point. The results are also represented as the mean \pm SEM (Significance values calculated by ANOVA, n>10, *p<0.05, ****p<0.0001, n.s. = not significant).

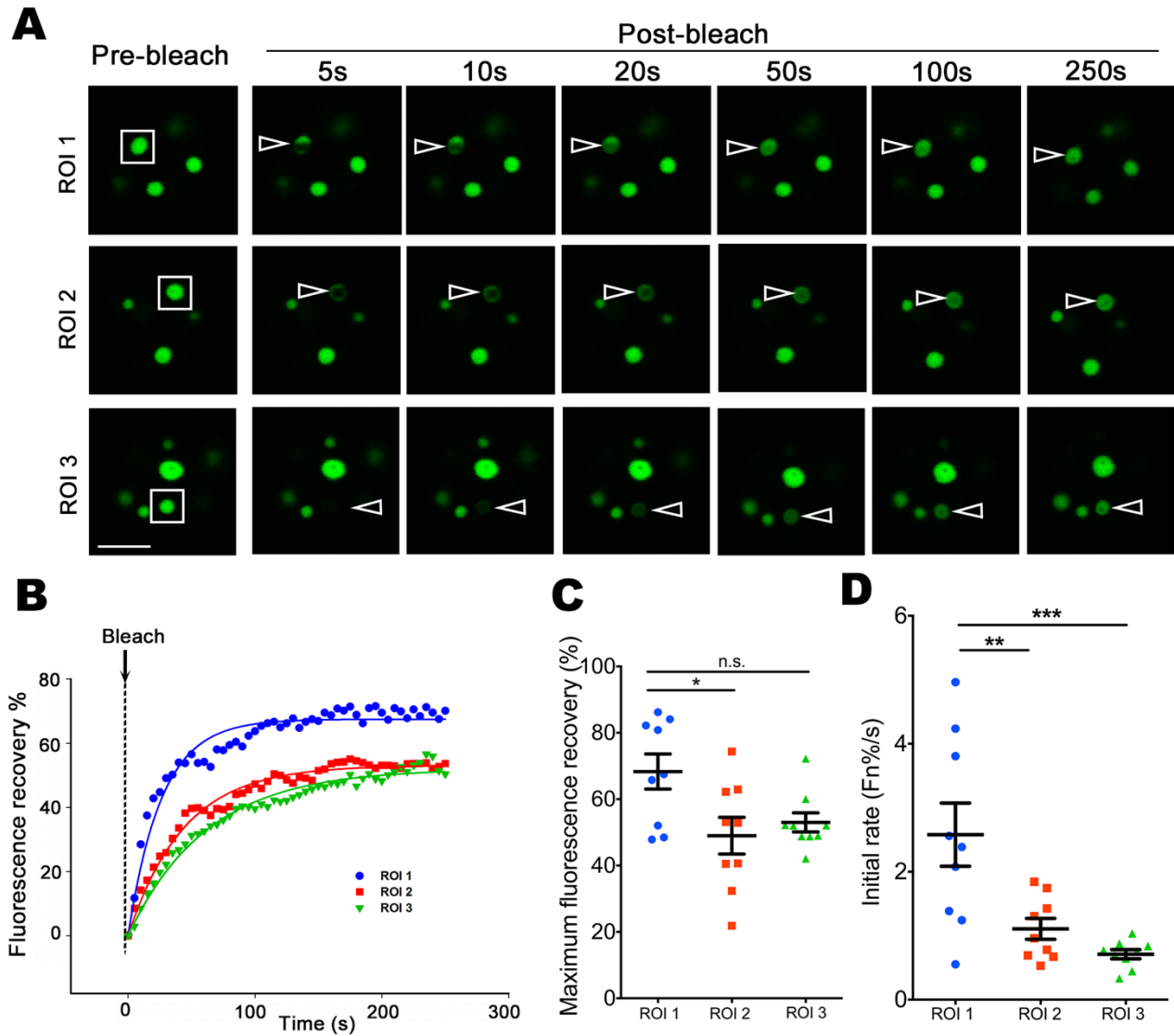
Supp figure 10: siRNA-mediated knockdown of RNA helicases DDX42, DDX46, or DHX15 does not increase cell death.

Neuro-2a cells were co-transfected with GFP and different siRNAs: negative ctrl siRNA, siDDX42, siDDX46, or siDHX15. At 24 h post-transfection, cells were collected into round-bottom glass tubes, and added with SYTOX Red dead cell stain for 15 min prior to fluorescence detection by flow cytometry (BD LSRFortessa). (A) Representative gates are shown for (I) forward scattering and APC-A (SYTOX Red) gates to focus on the intact cell population, (II) side scattering by height (SSC-H) and width (SSC-W) focusing on the single cell population, (III) forward scattering by height (FSC-H) and width (FSC-W) focusing on the single cell population. (B) Representative images are shown for gates applied to each siRNA condition (Neg ctrl, siDDX42, siDDX46, siDHX15) with GFP transfected cells (GFP-A positive) or non-transfected cells (NT) (GFP-A negative), and dead cells (APC-A positive) or live cells (APC-A negative). (C) Cell death percentage among transfected cells in all conditions. Results are calculated from 3 independent experiments. (Significance values calculated by ANOVA, *p<0.05, ***p<0.001, ****p<0.0001, n.s. = not significant).

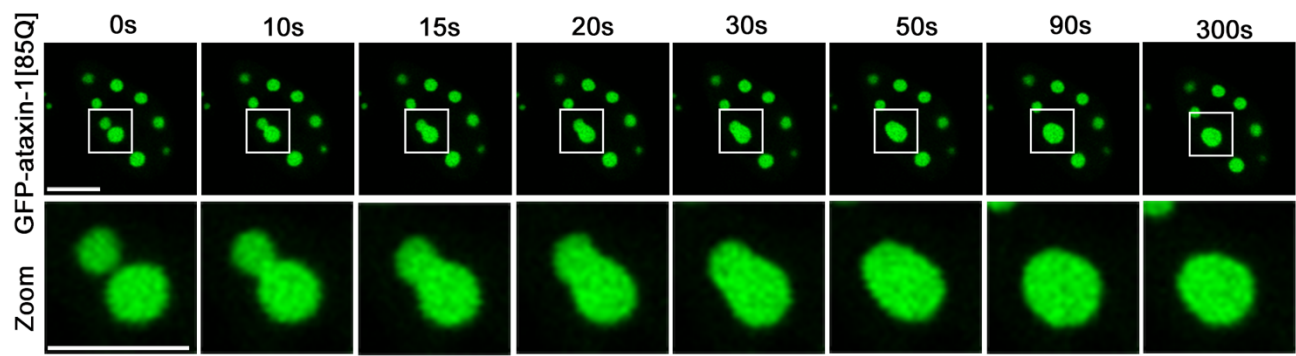
Supp movie 1: Fusion of ataxin-1[85Q] NBs visualized during live cell imaging.

Neuro-2a cells were transfected to express GFP-ataxin-1[85Q]. At 24 h post-transfection, images were recorded for 5 min (60 frames at 5 s intervals). Image stacks were compressed into a video (0.2 s per frame).

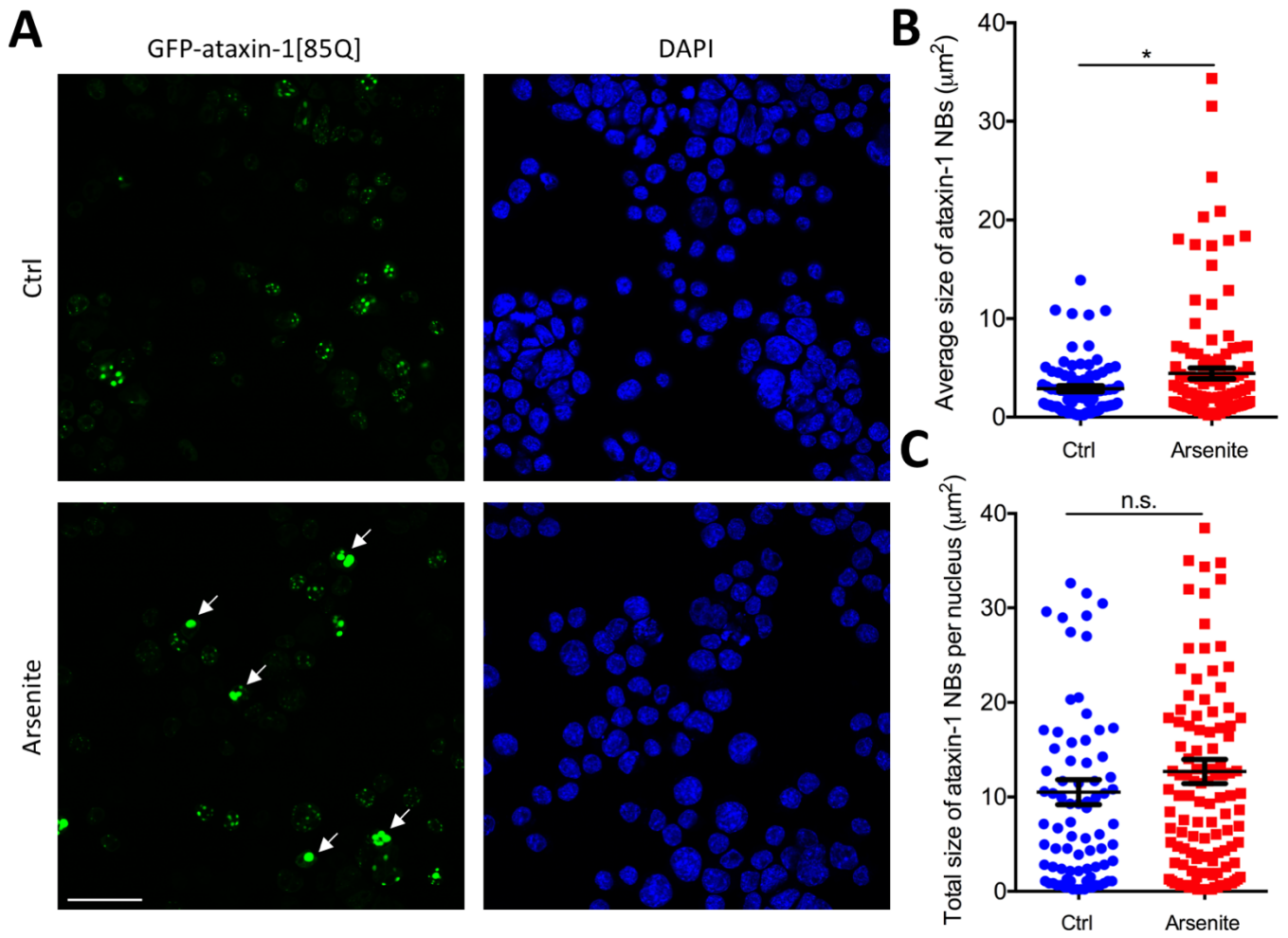
Supp Fig 1



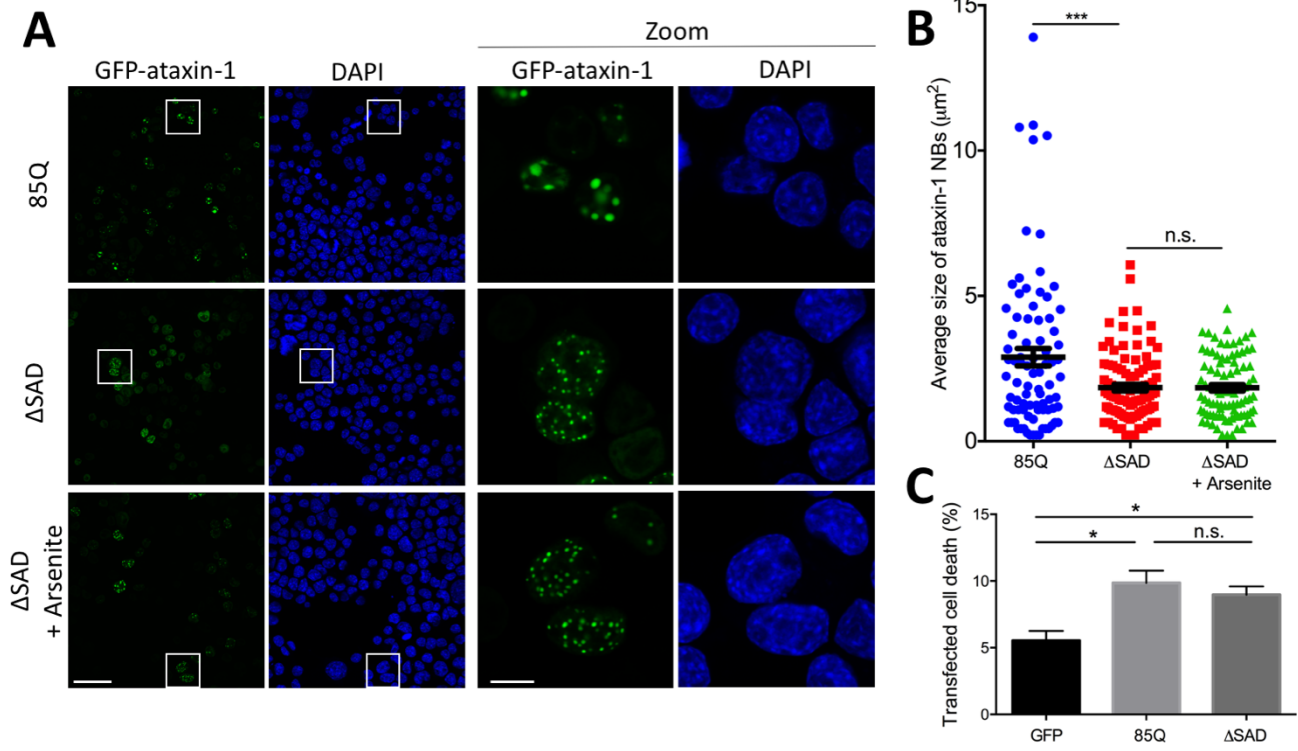
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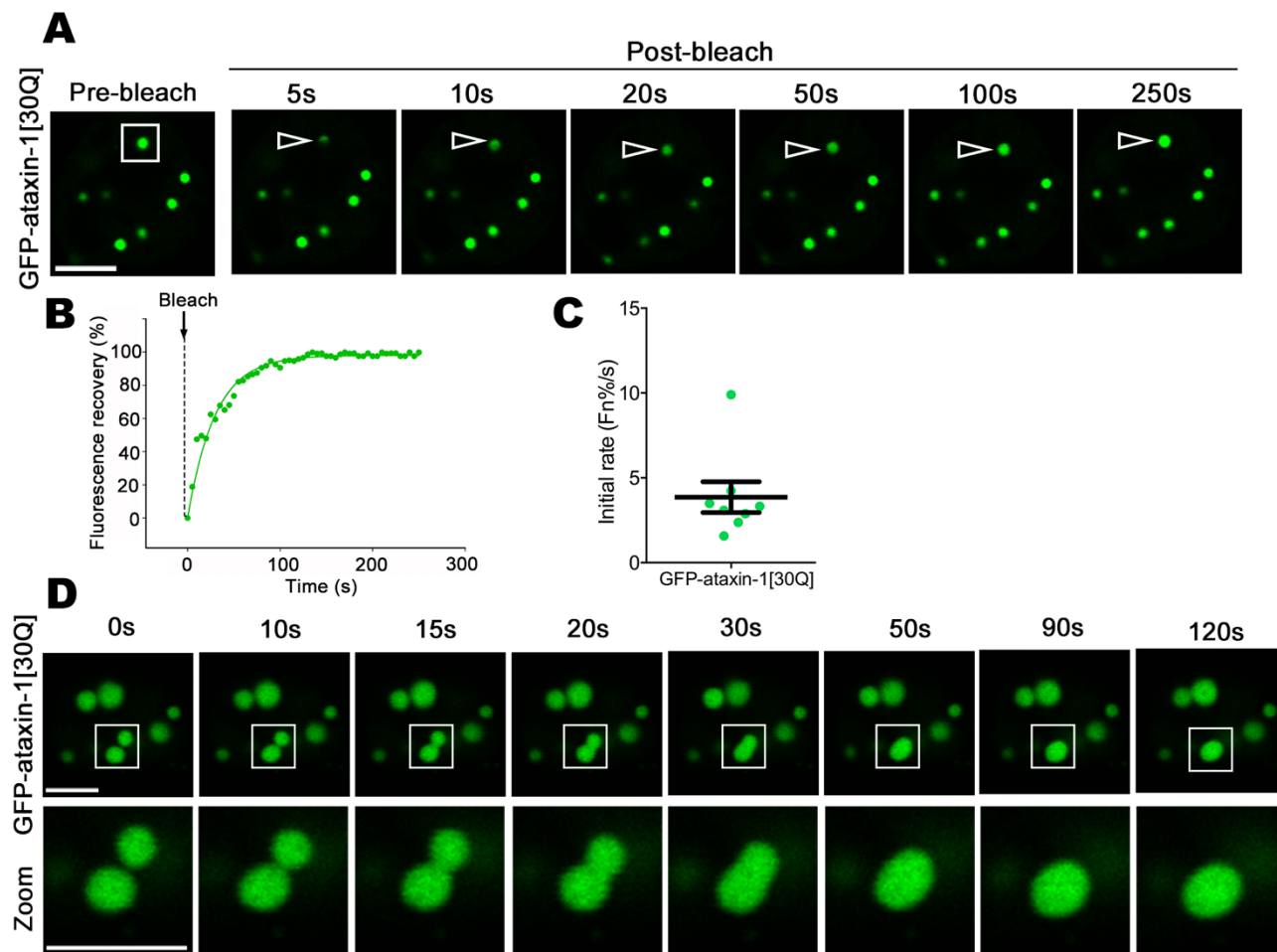
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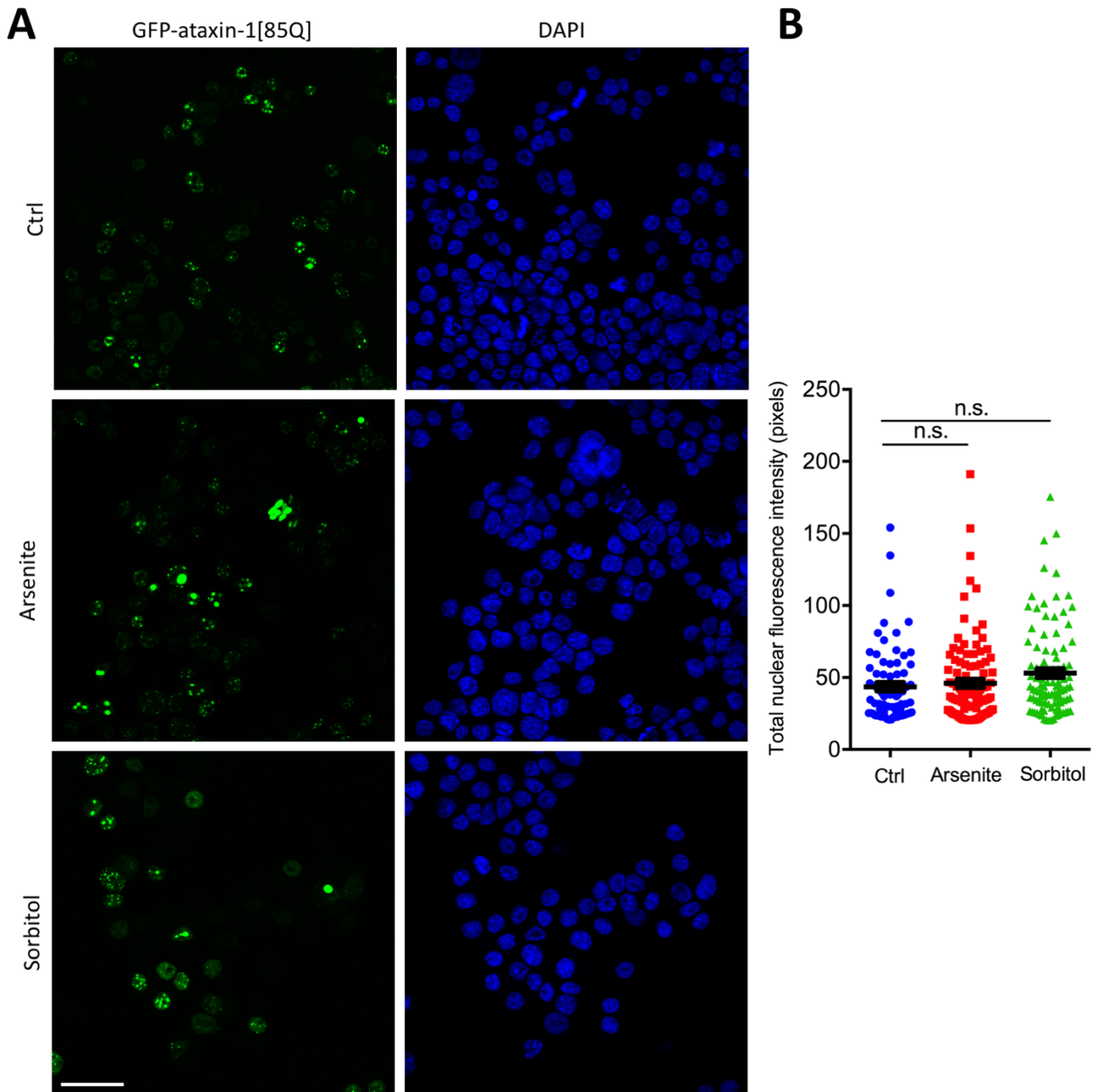
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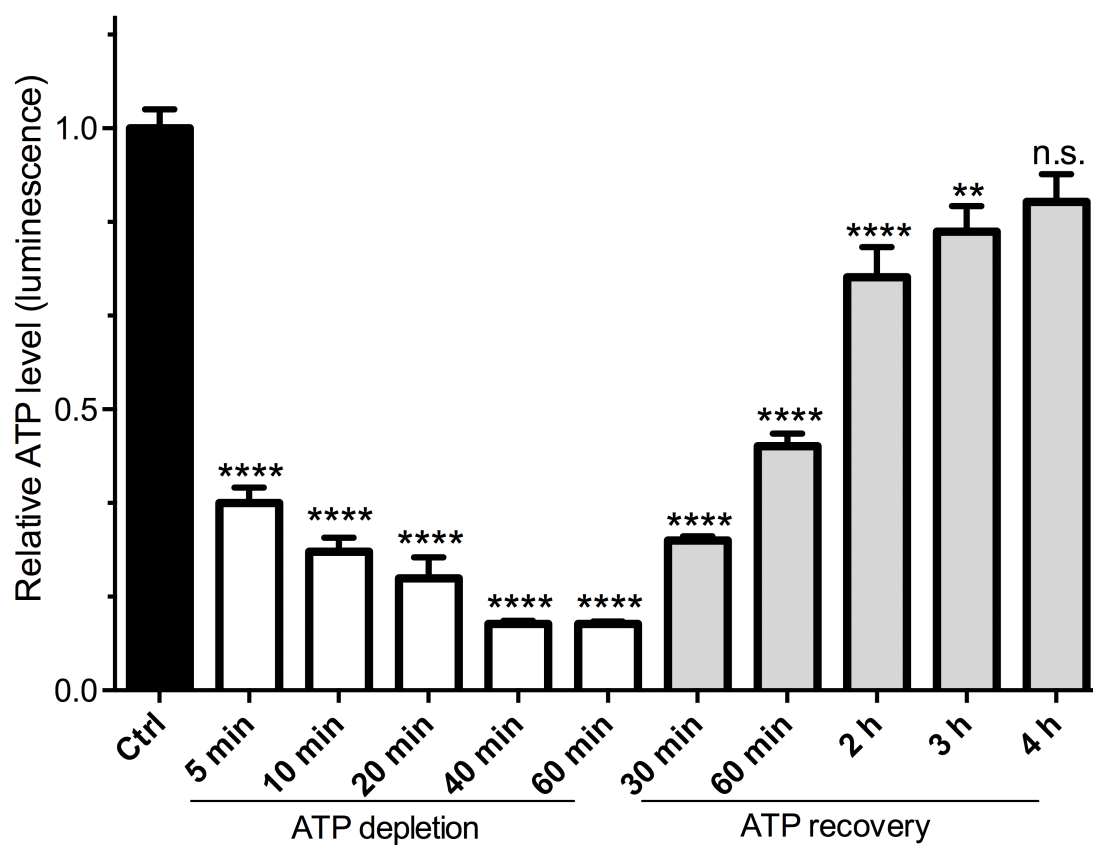
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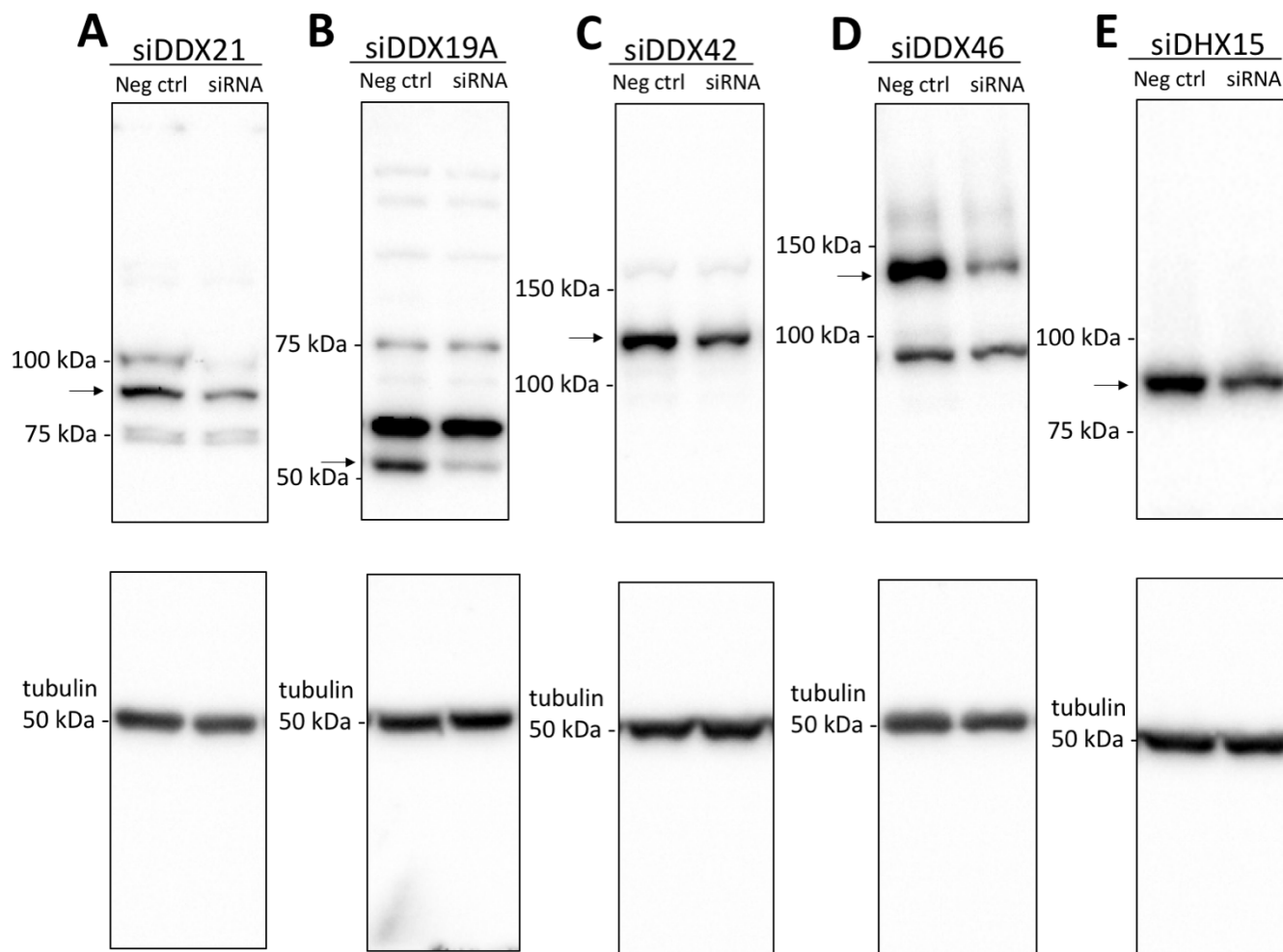
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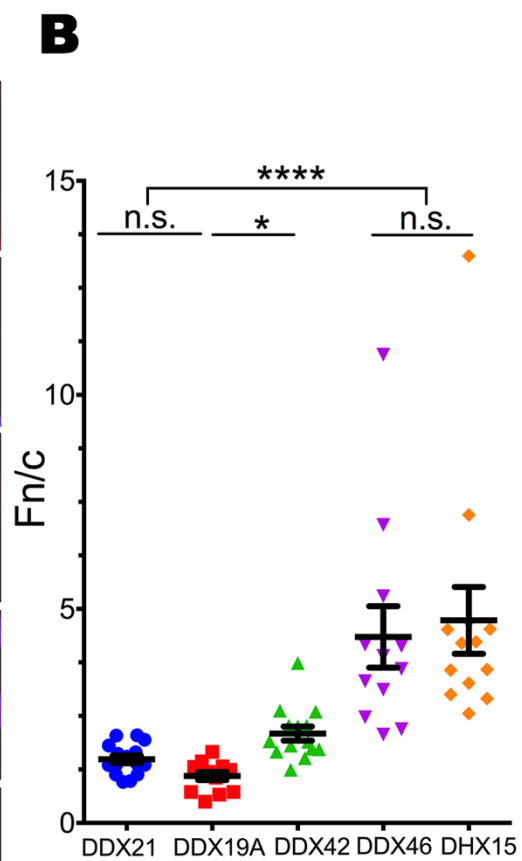
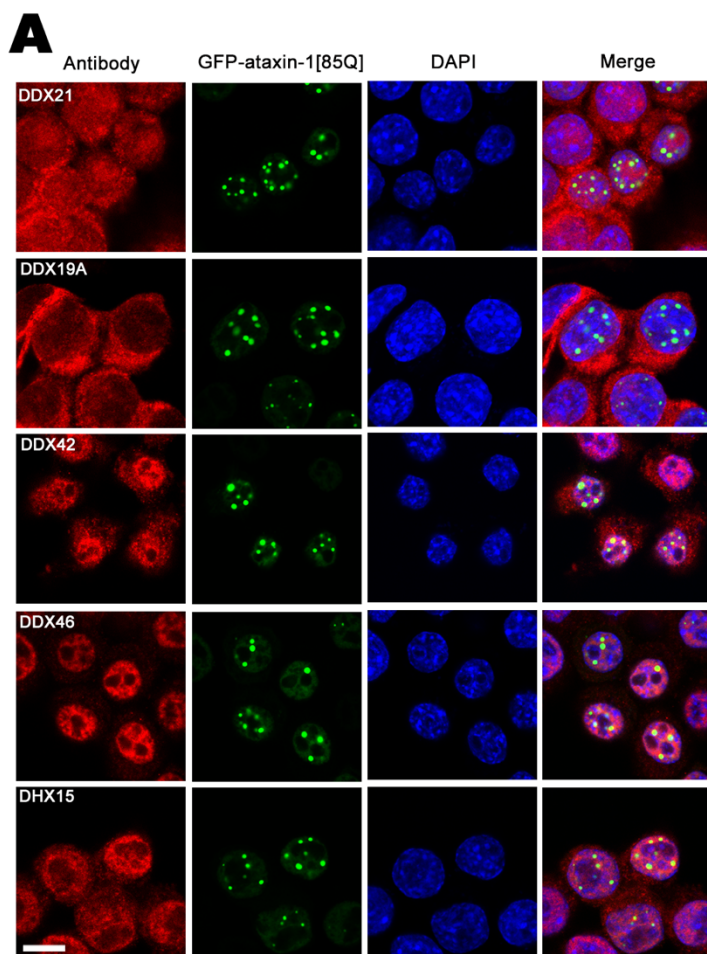
Supp Fig 7



Supp Fig 8



Supp Fig 9



Supp Fig 10

