# **Supplementary Information**

Direct C-H Difluoromethylation of Heterocycles via Organic Photoredox Catalysis

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## **Supplementary Methods**

## **General information**

Commercial reagents were used as received, unless otherwise stated. <sup>1</sup>H and <sup>13</sup>C NMR were recorded on a Bruker-DPX 400 spectrometer. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance as the internal standard. The following abbreviations were used to designate chemical shift mutiplicities: s = singlet, d = doublet, t = triplet, q = quartet, h = heptet, m = multiplet, br = broad. All first-order splitting patterns were assigned on the basis of the appearance of the multiplet. Splitting patterns that could not be easily interpreted are designated as multiplet (m) or broad (br). <sup>19</sup>F NMR were recorded on a Varian NMR 400 spectrometer. Mass spectra were obtained using electrospray ionization (ESI) mass spectrometer. Substrates **1** was synthesized according to the literature method.<sup>5</sup> sodium difluoromethane sulfonate (CF<sub>2</sub>HSO<sub>2</sub>Na) was purchased from *J&K* without further purification.

## **Experimental procedure**

### General procedure for synthesis of quinoxalin-2(1H)-ones



To a 10 mL Schlenk tube equipped with a magnetic stir bar, added quinoxalin-2(1*H*)-ones **1** (0.2 mmol), CF<sub>2</sub>HSO<sub>2</sub>Na **2** (0.4 mmol) and rose bengal (0.004 mmol, 2 mol%) in DMSO (1.0 mL). Then the mixture was stirred and irradiated by the two 3W green LEDs at room temperature for 12 h. The residue was added water (10 mL) and extracted with ethyl acetate (5 mL  $\times$  3). The combined organic phase was dried over Na<sub>2</sub>SO<sub>4</sub>. The resulting crude residue was purified *via* column chromatography on silica gel to afford the desired products.

## General procedure for synthesis of heteroarenes



To a 10 mL Schlenk tube equipped with a magnetic stir bar, added heteroarenes **4** (0.1 mmol), CF<sub>2</sub>HSO<sub>2</sub>Na **2** (0.4 mmol) and rose bengal (0.002-0.005 mmol, 2-5 mol%) in DMSO (1.0 mL). Then the mixture was stirred and irradiated by the two 3W green LEDs at room temperature for 24 h. The residue was added water (10 mL) and extracted with ethyl acetate (5 mL  $\times$  3). The combined organic phase was dried over Na<sub>2</sub>SO<sub>4</sub>. The resulting crude residue was purified *via* column chromatography on silica gel to afford desired products.



Supplementary Figure 1. Reaction set-up

Analytical data of compounds quinoxalin-2(1H)-ones



**3a:** yellow solid, 30.3 mg, 72% yield;  $R_f$ =0.6 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.97-7.93 (m, 1H), 7.68-7.64 (m, 1H), 7.42-7.34 (m, 2H), 6.93 (t, *J*=53.6 Hz, 1H), 3.70 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.2, 148.51 (t, *J* = 22.6 Hz), 134.0, 132.7, 131.8, 131.3, 124.4, 114.0, 110.1 (t, *J* = 241.6 Hz), 29.0; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -125.1 (d, *J* = 58.8 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>9</sub>F<sub>2</sub>N<sub>2</sub>O [M+H<sup>+</sup>]: 211.0677, found: 211.0678.



**3b:** yellow solid, 30.5 mg, 68% yield;  $R_f$ =0.6 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.55 (t, *J* = 8.0 Hz, 1H), 7.28-7.25 (m, 1H), 7.20 (d, *J* = 8.4 Hz, 1H), 6.92 (t, *J* = 53.8 Hz, 1H), 3.72 (s, 3H), 2.71 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.1, 146.6 (t, *J* = 22.8 Hz), 140.7, 132.4 (t, *J* = 187.7 Hz), 125.6, 113.1, 111.7, 110.7, 108.3, 29.1, 17.4; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -123.6 (d, *J* = 54.1 Hz, 2F); HRMS (ESI) calcd for C<sub>11</sub>H<sub>10</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 247.0653, found: 247.0658.



**3c:** orange solid, 27.4mg, 60% yield;  $R_f$ =0.4 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.73-7.69 (m, 1H), 7.50-7.45 (m, 1H), 7.41-7.37 (m, 1H), 6.97 (t, J = 53.6 Hz, 1H), 3.76 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  158.9 (d, J = 245.7 Hz), 152.9, 150.0 (t, J = 22.8 Hz), 132.38 (d, J = 11.2 Hz), 130.8, 120.7(d, J = 24.1 Hz), 116.7(d, J = 22.4 Hz), 115.3(d, J = 8.7 Hz), 109.9 (t, J = 242.0 Hz), 29.3; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -117.2 (s, 1F), -124.6 (d, J = 53.5 Hz, 2F); HRMS (ESI) calcd

for C<sub>10</sub>H<sub>7</sub>F<sub>3</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 251.0403, found: 251.0406.



**3d:** yellow solid, 31.4mg, 70% yield;  $R_f$ =0.4 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.97 (d, J = 2.4 Hz, 1H), 7.63 (dd, J = 9.0, 2.4 Hz, 1H), 7.32 (d, J = 9.0 Hz, 1H), 6.92 (t, J = 53.6 Hz, 1H), 3.72 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$ 152.8, 151.4, 149.9 (t, J = 22.8 Hz), 132.7, 132.3, 131.1, 130.6, 129.8, 115.2, 109.9 (t, J = 242.5 Hz), 29.2; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.6 (d, J = 53.6 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>7</sub>ClF<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 267.0111, found: 267.0107.



**3e:** yellow solid, 47.4mg, 82% yield;  $R_f$ =0.2 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.14 (d, J = 2.2 Hz, 1H), 7.77 (dd, J = 8.9, 2.2 Hz, 1H), 7.28-7.26 (m, 1H), 6.93 (t, J = 53.6 Hz, 1H), 3.72 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  152.8, 149.8 (t, J = 22.8 Hz), 135.4, 133.7, 133.2, 132.6, 117.0, 115.4, 112.3, 109.9 (t, J = 243.0 Hz), 29.2; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.6 -124.55 (d, J = 53.6 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>7</sub>BrF<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 310.9602, found: 310.9605.



**3f:** white solid, 39.9mg, 83% yield;  $R_f$ =0.5 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.45 (s, 1H), 7.30 (d, J = 1.6 Hz, 2H), 6.98 (t, J = 53.8 Hz, 1H), 3.89 (s, 3H), 3.72 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  156.4, 153.0, 149.0 (t, J = 22.1 Hz), 132.7, 128.4, 122.5, 114.9, 112.1, 109.9 (t, J = 242.4 Hz), 55.9, 29.1; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.4 (d, J = 55.5Hz, 2F); HRMS (ESI) calcd for C<sub>11</sub>H<sub>10</sub>F<sub>2</sub>N<sub>2</sub>NaO<sub>2</sub> [M+Na<sup>+</sup>]: 263.0603, found: 263.0606.



**3g:** yellow solid, 34.7mg, 60% yield;  $R_f$ =0.4 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.83 (d, *J* = 8.5 Hz, 1H), 7.54-7.51 (m, 2H), 6.91 (t, *J* = 53.6 Hz, 1H), 3.70 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  152.9, 148.8 (t, *J* = 23.0 Hz), 134.9, 132.6, 130.7, 127.8, 127.3, 117.1, 109.9 (t, *J* = 241.9 Hz), 29.1; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.4 (d, *J* = 53.6 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>7</sub>BrF<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 310.9602, found: 310.9606.



**3h:** yellow solid, 20.4mg, 40% yield;  $R_f$ =0.5 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.28-8.16 (m, 3H), 6.95 (t, *J* = 53.3 Hz, 1H), 3.81 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  152.6, 152.2 (t, *J* = 22.9 Hz), 149.3, 134.9, 134.5, 132.7, 118.7, 109.9, 109.5 (t, *J* = 244.0 Hz), 29.6; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -125.0 (d, *J* = 53.5 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>7</sub>F<sub>2</sub>N<sub>3</sub>NaO<sub>3</sub> [M+Na<sup>+</sup>]: 278.0348, found: 278.0350.



**3i:** yellow solid, 41.9mg, 75% yield;  $R_f$ =0.6 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.24 (s, 1H), 8.00 (s, 1H), 7.09 (t, *J* = 53.0 Hz, 1H), 3.63 (s, 3H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  152.3, 149.8 (t, *J* = 21.6 Hz), 134.9, 134.0, 130.9, 130.4, 126.1, 117.0, 110.1 (t, *J* = 239.9 Hz), 29.3; <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -119.9 (d, *J* = 52.9Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>6</sub>Cl<sub>2</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 300.9717, found: 300.9720.



**3j:** yellow solid, 21.9mg, 42% yield;  $R_f$ =0.3 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.52 (s, 1H), 7.97 (dd, J = 28.4, 8.3 Hz, 2H), 7.66-7.62 (m, 2H), 7.54 (t, J = 7.6 Hz, 1H), 6.99 (t, J = 53.7 Hz, 1H), 3.79 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.0, 149.2 (t, J = 22.3 Hz), 134.8, 131.5, 131.5, 131.0, 129.8, 129.2, 128.9, 127.3, 125.9, 110.5, 110.1 (t, J = 242.8 Hz), 28.9; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.3 (d, J = 53.9 Hz, 2F); HRMS (ESI) calcd for C<sub>14</sub>H<sub>10</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 283.0653, found: 283.0653.



**3k:** white solid,16.1mg, 34% yield;  $R_f$ =0.6 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.01 (dd, J = 8.1, 1.5 Hz, 1H), 7.67-7.63 (m, 1H), 7.44-7.40 (m, 1H), 7.36 (dd, J = 8.5, 1.1 Hz, 1H), 6.98 (t, J = 53.7 Hz, 1H), 5.98-5.89 (m, 1H), 5.31 (d, J = 8.7 Hz, 1H), 5.21 (d, J = 17.2 Hz, 1H), 4.95-4.92 (m, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  152.8, 148.7 (t, J = 22.3 Hz), 133.3, 132.6, 132.1, 131.5, 130.0, 124.4, 118.8, 114.5, 110.0 (t, J = 241.6 Hz), 44.5; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.2 (d, J = 53.6 Hz, 2F); HRMS (ESI) calcd for C<sub>12</sub>H<sub>10</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 259.0653, found: 259.0658.



**31:** yellow solid, 26.7mg, 56% yield;  $R_f$ =0.5 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.00 (dd, J = 8.0, 1.5 Hz, 1H), 7.69-7.65 (m, 1H), 7.43-7.36 (m, 2H), 6.97 (t, J = 53.7 Hz, 1H), 4.26-4.22 (m, 2H), 1.86-1.77 (m, 2H), 1.06 (t, J = 7.4 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.0, 148.6 (t, J = 22.4 Hz), 133.3, 132.5,

132.2, 131.7, 124.1, 114.0, 110.0 (t, J = 22.4 Hz) 43.8, 20.7, 11.4; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -129.0 (d, J = 53.6 Hz, 2F); HRMS (ESI) calcd for C<sub>12</sub>H<sub>12</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 261.0810, found: 261.0815.



**3m:** yellow solid, 29.8mg, 52% yield;  $R_f$ =0.6 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.01 (d, *J* = 8.0 Hz, 1H), 7.56 (t, *J* = 7.9 Hz, 1H), 7.40-7.25 (m, 7H), 7.03 (t, *J* = 53.7 Hz, 1H), 5.52 (s, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.4, 148.8 (t, *J* = 22.7 Hz), 134.5, 133.4, 132.6, 132.2, 131.6, 129.1, 128.0, 126.9, 124.5, 114.7, 110.0 (t, *J* = 241.7 Hz); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.1 (d, *J* = 53.6 Hz, 2F); HRMS (ESI) calcd for C<sub>16</sub>H<sub>12</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 309.0810, found: 309.0815.



**3n:** yellow solid, 38.9mg, 73% yield;  $R_f$ =0.6 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.00 (d, J = 7.7 Hz, 1H), 7.69-7.65 (m, 1H), 7.43-7.36 (m, 2H), 6.97 (t, J = 53.7 Hz, 1H), 4.28-4.24 (m, 2H), 1.81-1.73 (m, 2H), 1.49-1.35 (m, 4H), 0.93 (t, J = 6.9 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.0, 148.5 (d, J = 22.5 Hz), 133.3, 132.5, 132.2, 131.7, 124.2, 113.9, 110.0 (t, J = 241.5 Hz), 42.4, 29.0, 26.9, 22.3, 13.9; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.3 (d, J = 53.7 Hz, 2F); HRMS (ESI) calcd for C<sub>14</sub>H<sub>16</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 289.1123, found: 289.1126.



**30:** white solid, 34.1 mg, 87% yield;  $R_f=0.4$  (50% EtOAc/petroleum ether); <sup>1</sup>H NMR

(400 MHz, DMSO- $d_6$ )  $\delta$  12.90 (s, 1H), 7.93-7.85 (m, 1H), 7.71-7.64 (m, 1H), 7.44-7.34 (m, 2H), 7.08 (td, J = 53.1, 16.2 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  153.7, 150.2 (t, J = 22.2 Hz), 133.3, 132.8, 131.2, 130.0, 124.4, 116.3, 110.8 (t, J = 238.4 Hz); <sup>19</sup>F NMR (376 MHz, DMSO- $d_6$ )  $\delta$  -124.3 (d, J = 53.5 Hz, 2F); HRMS (ESI) calcd for C<sub>9</sub>H<sub>5</sub>F<sub>2</sub>N<sub>2</sub>O [M-H<sup>+</sup>]: 195.0370, found: 195.0373.

#### Analytical data of heteroarenes

Ph N CF<sub>2</sub>H Ph N OH

**5a:** yellow solid, 24.5mg, 82% yield;  $R_f$ =0.5 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  12.93 (s, 1H), 7.47-7.42 (m, 1H), 7.39-7.38 (m, 4H), 7.32-7.29 (m, 2H), 7.27-7.23 (m, 3H), 6.77 (t, J = 53.7 Hz, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  155.3, 144.7 (t, J = 24.9 Hz), 139.6, 136.1, 133.7, 131.4, 130.7, 129.5, 129.5, 128.9, 128.3, 128.1, 110.9 (t, J = 240.7 Hz); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -122.3 (d, J = 53.8 Hz, 2F); HRMS (ESI) calcd for C<sub>17</sub>H<sub>11</sub>F<sub>2</sub>N<sub>2</sub>O [M-H<sup>+</sup>]: 297.0839, found: 297.0842.

**5b:** colorless oil, 12.5mg, 65% yield;  $R_f$ =0.7 (5% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.67 (t, J = 54.1 Hz, 1H), 2.69 (s, 3H), 2.63 (s, 3H); Other spectral data of **5b** were consistent with previous reported data.<sup>3</sup>



**5c:** white solid, 12.1mg, 62% yield;  $R_f=0.8$  (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.10-8.06 (m, 2H), 7.85-7.75 (m, 2H), 6.83 (t, J = 54.3 Hz, 1H), 2.93 (s, 3H); Other spectral data of **5c** were consistent with previous reported data.<sup>1</sup>



**5d:** white solid, 17.6mg, 90% yield;  $R_f$ =0.5 (50% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 12.9 (s, 1H), 7.89 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.66 (td, *J* = 7.7, 1.4 Hz, 1H), 7.41-7.36 (m, 2H), 7.06 (t, *J* = 53.3 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 153.2, 149.7 (t, *J* = 21.4 Hz), 132.8, 132.3, 130.7, 129.4, 123.9, 115.7, 110.3 (t, *J* = 239.3 Hz); <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>) δ -119.5 (d, *J* = 53.1 Hz, 2F); HRMS (ESI) calcd for C<sub>9</sub>H<sub>5</sub>F<sub>2</sub>N<sub>2</sub>O [M-H<sup>+</sup>]: 195.0370, found: 195.0370.



**5e:** white solid, 9.2mg, 61% yield;  $R_f=0.7$  (35% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.71 (s, 1H), 6.87 (s, 1H), 6.71 (t, J = 55.3 Hz, 1H), 6.48-6.45 (m, 1H), 2.47 (s, 3H); Other spectral data of **5e** were consistent with previous reported data.<sup>1</sup>



**5f:** white solid, 12.3mg, 64% yield;  $R_f$ =0.2 (50% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.56 (t, *J* = 52.7 Hz, 1H), 7.51 (s, 1H), 3.92 (s, 3H), 3.86 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 162.7, 140.3, 133.7, 129.4 (t, *J* = 23.0 Hz), 108.7 (t, *J* = 233.9 Hz), 52.2, 33.6; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -114.4 (d, *J* = 51.8 Hz, 2F); HRMS (ESI) calcd for C<sub>12</sub>H<sub>12</sub>F<sub>2</sub>N<sub>2</sub>NaO [M+Na<sup>+</sup>]: 261.0810, found: 261.0815.



**5g:** white solid, 14.4mg, 81% yield;  $R_f$ =0.3 (35% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  13.96 (s, 1H), 7.46 (t, *J* = 53.1 Hz, 1H), 2.39 (s, 3H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  145.9, 130.1, 124.0, 108.6 (t, *J* = 235.6 Hz), 14.17; <sup>19</sup>F

NMR (376 MHz, DMSO- $d_6$ )  $\delta$  -113.2 (s, 2F); HRMS (ESI) calcd for C<sub>5</sub>H<sub>5</sub>F<sub>2</sub>N<sub>3</sub>NaO<sub>2</sub> [M+Na<sup>+</sup>]: 200.0242, found: 200.0245.



**5h:** colorless oil, 11.9 mg, 73% yield;  $R_f$ =0.2 (50% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  12.75 (s, 1H), 12.31 (s, 1H), 6.77 (t, *J* = 52.7 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  155.1, 149.0, 135.9 (t, *J* = 23.7 Hz), 110.7 (t, *J* = 237.9 Hz); <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -117.4 (d, *J* = 52.7 Hz, 2F); HRMS (ESI) calcd for C<sub>4</sub>H<sub>2</sub>F<sub>2</sub>N<sub>3</sub>O<sub>2</sub> [M-H<sup>+</sup>]: 162.0115, found: 162.0124.



**5i:** colorless oil, 7.8mg, 41% yield;  $R_f$ =0.3 (3% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.88 (t, J = 55.3, Hz), 6.50 (s, 1H), 4.30-4.23 (m, 4H); Other spectral data of **5i** were consistent with previous reported data.<sup>4</sup>



**5j:** colorless oil, 8.5mg, 42% yield;  $R_f = 0.4$  (25% dichloromethane/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.79 (d, J = 5.0 Hz, 1H), 8.17 (s, 1H), 7.96 (d, J = 5.0 Hz, 1H), 6.68 (t, J = 55.2 Hz, 2H), 1.41 (t, J = 7.1 Hz, 3H); Other spectral data of **5j** were consistent with previous reported data.<sup>1</sup>

**5k:** white solid, 5.6mg, 34% yield;  $R_f=0.5$  (80% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  6.94 (t, J = 53.7 Hz, 1H); Other spectral data of **5k** were consistent with previous reported data.<sup>1</sup>



**51:** white solid, 15.9mg, 38% yield;  $R_f$ =0.7 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  12.82 (s, 1H), 8.04 (d, *J* = 7.7 Hz, 1H), 7.73-7.46 (m, 2H), 7.59 (t, *J* = 53.6 Hz, 1H), 7.56 (d, *J* = 7.7 Hz, 1H) 2.67 (s, 3H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  193.6, 135.4, 134.7 (t, *J* = 22.9 Hz), 124.8, 123.9, 122.4, 121.3, 115.8 (t, *J* = 5.9 Hz), 113.0, 109.7 (t, *J* = 235.2 Hz), 30.9; <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -108.1 (d, *J* = 56.3 Hz, 2F); HRMS (ESI) calcd for C<sub>11</sub>H<sub>10</sub>F<sub>2</sub>NO [M+H<sup>+</sup>]: 210.0725, found: 210.0728.



**5m:** white solid, 16.3mg, 68% yield;  $R_f$ =0.3 (3% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.19 (d, *J* = 8.0 Hz, 1H), 7.93 (t, *J* = 52.0 Hz, 1H), 7.41-7.30 (m, 3H), 3.98 (s, 6H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  165.0, 137.5, 135.3 (t, *J* = 21.6 Hz), 125.2, 124.8, 122.9, 122.6, 109.9, 109.6 (t, *J* = 235.0 Hz), 108.0 (t, *J* = 6.4 Hz), 51.5, 31.8; <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -113.8 (d, *J* = 52.9 Hz, 2F); HRMS (ESI) calcd for C<sub>12</sub>H<sub>11</sub>F<sub>2</sub>NNaO<sub>2</sub> [M+Na<sup>+</sup>]: 262.0650, found: 262.0653.



**5n:** white solid, 11.7mg, 52% yield;  $R_f$ =0.6 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.94 (t, J = 53.2 Hz, 1H), 3.65 (s, 3H), 3.37 (s, 3H). Other spectral data of **5n** were consistent with previous reported data.<sup>2</sup>



**50:** white solid, 8.0mg, 42% yield;  $R_f$ =0.2 (10% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.56 (s, 1H), 6.68 (t, *J* = 55.2 Hz, 1H), 3.47 (s, 3H), 3.36 (s, 3H). Other spectral data of **5n** were consistent with previous reported data.<sup>2</sup>



**5p:** white solid, 10.0mg, 51% yield;  $R_f$ =0.5 (30% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.60 (s, 1H), 8.67 (s, 1H), 8.01 (d, *J* = 8.2 Hz, 1H), 7.82 (t, *J* = 55.2 Hz, 1H), 7.78 (d, *J* = 7.3 Hz, 1H), 7.65-7.61 (m, 2H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ 151.5, 144.3, 139.1 (t, *J* = 5.0 Hz), 130.4 (t, *J* = 21.1 Hz), 129.6, 129.1, 126.3, 122.6 (t, *J* = 6.5 Hz), 115.7, 112.7 (t, *J* = 234.6 Hz); <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -114.1 (d, *J* = 56.8 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>8</sub>F<sub>2</sub>NO [M+Na<sup>+</sup>]: 196.0568, found: 196.0573.



**5q:** white solid, 17.6mg, 70% yield;  $R_f$ =0.8 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 6.92 (t, *J* = 51.9 Hz, 1H), 4.04 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 154.6, 154.3, 153.3, 147.8 (t, *J* = 27.8 Hz), 129.2, 110.2 (t, *J* = 240.2 Hz), 30.4; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -116.0 (d, *J* = 51.9 Hz, 2F); HRMS (ESI) calcd for C<sub>7</sub>H<sub>5</sub>Cl<sub>2</sub>F<sub>2</sub>N<sub>4</sub> [M+H<sup>+</sup>]: 252.9854, found: 252.9856.



**5r:** white solid, 18.5mg, 73% yield;  $R_f$ =0.4 (20% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.00 (t, *J* = 51.9 Hz, 1H), 4.30 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 161.0, 153.9, 151.2 (t, *J* = 27.7 Hz), 145.4, 123.7, 110.1 (t, *J* = 240.7 Hz); 33.6; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -115.9 (d, *J* = 51.9 Hz, 2F); HRMS (ESI) calcd for C<sub>7</sub>H<sub>5</sub>Cl<sub>2</sub>F<sub>2</sub>N<sub>4</sub> [M+H<sup>+</sup>]: 252.9854, found: 252.9857.



**5s:** colorless oil, 11.3mg, 54% yield;  $R_f$ =0.6 (2% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  6.70 (t, *J* = 54.3 Hz, 1H), 5.97 (s, 2H), 4.01 (s, 1H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  157.9, 144.4 (t, *J* = 25.6 Hz), 137.0, 129.5, 112.4 (t, *J* = 239.4 Hz), 55.2; <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -115.6 (d, *J* = 54.6 Hz, 2F); HRMS (ESI) calcd for C<sub>6</sub>H<sub>7</sub>ClF<sub>2</sub>N<sub>3</sub>O [M+H<sup>+</sup>]: 210.0240, found: 210.0242.



**5t:** colorless oil, 17.0mg, 65% yield;  $R_f$ =0.5 (10% EtOAc/petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.98 (t, *J* = 54.8 Hz, 1H), 6.72 (s, 1H), 3.94 (s, 3H), 3.90 (s, 3H), 3.89 (s, 3H), 2.57 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  202.1, 155.1, 152.9, 143.7, 136.3, 118.0 (t, *J* = 22.8 Hz), 112.0 (t, *J* = 236.4 Hz), 106.5, 61.9, 60.9, 56.2, 30.9; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -109.2 (d, *J* = 54.9 Hz, 2F); HRMS (ESI) calcd for C<sub>12</sub>H<sub>14</sub>F<sub>2</sub>NaO<sub>4</sub> [M+Na<sup>+</sup>]: 283.0752, found: 283.0754.



**6a:** white solid, 18.0mg, 74% yield;  $R_f = 0.6$  (60% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  7.13 (t, J = 52.9 Hz, 1H), 3.43 (s, 3H), 3.37 (s, 3H), 3.24 (s, 3H); Other spectral data of **6a** were consistent with previous reported data.<sup>1</sup>



**6b:** white solid, 15.1mg, 70% yield;  $R_f = 0.6$  (80% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  14.6 (s, 1H), 7.13 (t, *J* = 52.9 Hz, 1H), 3.44 (s, 3H), 3.24 (s, 3H); Other spectral data of **6b** were consistent with previous reported data.<sup>1</sup>



**6c:** white solid, 12.5mg, 38% yield;  $R_f = 0.3$  (30% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.73 (t, J = 52.2 Hz, 1H), 4.13 (s, 3H), 4.02-3.98 (m, 2H), 3.53 (s, 3H), 2.49 (t, J = 6.9 Hz, 2H), 2.13 (s, 3H), 1.69-1.58 (m, 4H); Other spectral data of **6c** were consistent with previous reported data.<sup>1</sup>



**6d:** grey solid, 22.5mg, 81% yield;  $R_f = 0.6$  (100% EtOAc); <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN) δ 9.30 (s, 1H), 8.15 (s, 1H), 6.38 (t, J = 54.8 Hz, 1H), 5.93 (t, J = 6.3 Hz, 1H), 4.16-4.11 (m, 1H), 3.67 (q, J = 3.3 Hz, 1H), 3.57-3.45 (m, 2H), 3.37-3.35 (m,1H), 3.19 (t, J = 4.8 Hz, 1H), 2.02-1.94 (m, 1H), 1.74-1.72 (m, 1H); <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>CN) δ 161.5, 150.5, 141.4, 112.41 (t, J = 234.5 Hz), 108.08 (t, J = 23.2 Hz), 88.2,

86.4, 70.9, 61.6, 41.2; <sup>19</sup>F NMR (376 MHz, CD<sub>3</sub>CN)  $\delta$  -117.2 (d, J = 161.2 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>12</sub>F<sub>2</sub>N<sub>2</sub>O<sub>5</sub> [M-H<sup>+</sup>]: 277.0636, found: 277.0640.



**6e:** yellow solid, 16.7mg, 57% yield;  $R_f = 0.5$  (5% MeOH/ CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN) δ 9.39 (s, 1H), 8.13 (s, 1H), 6.64 (t, *J* = 54.9 Hz, 1H), 6.08 (dd, *J* = 3.6, 1.6 Hz, 1H), 4.22 (d, *J* = 6.8 Hz, 1H), 4.11 (s, 2H), 3.94 (d, *J* = 3.7 Hz, 1H), 3.85-3.7 (m, 4H); <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>CN) δ 160.3 (t, *J* = 3.9 Hz), 149.3, 141.4 (t, *J* = 7.7 Hz), 111.5 (t, *J* = 234.5 Hz), 106.1 (t, *J* = 23.3 Hz), 85.8, 84.7, 75.6, 75.2, 60.6; <sup>19</sup>F NMR (376 MHz, CD<sub>3</sub>CN) δ -112.08 (dd, *J* = 54.1, 34.3 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>13</sub>F<sub>2</sub>N<sub>2</sub>O<sub>6</sub> [M+H<sup>+</sup>]: 295.0736, found: 295.0731.



**6f:** colorless oil, 18.4mg, 62% yield;  $R_f = 0.7$  (100% EtOAc); <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN) δ 9.37 (s, 1H), 8.50 (s, 1H), 6.59 (t, J = 54.8 Hz, 1H), 5.92 (dd, J = 16.0, 1.2 Hz, 1H), 4.96 (dd, J = 52.9, 3.9 Hz, 1H), 4.31-4.20 (m, 1H), 4.02-3.93 (m, 1H), 3.97-3.93(m,1H), 3.76-3.66 (m, 2H), 3.48 (t, J = 4.7 Hz, 1H); <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>CN) δ 161.3 (t, J = 4.0 Hz), 150.2, 140.9 (t, J = 7.5 Hz), 112.2 (t, J = 235.6 Hz), 108.1 (t, J = 23.3 Hz), 94.5 (d, J = 185.4 Hz), 88.6 (d, J = 34.2 Hz), 83.7, 67.8 (d, J = 16.3 Hz), 59.3; <sup>19</sup>F NMR (376 MHz, CD<sub>3</sub>CN) δ -112.2 (dd, J = 236.0, 54.9 Hz, 2F), -199.1 (ddd, J = 52.7, 23.3, 16.1 Hz, 1F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>10</sub>F<sub>3</sub>N<sub>2</sub>O<sub>5</sub>



**6g:** yellow solid, 20.3 mg, 72% yield;  $R_f = 0.5$  (80% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  7.95 (s, 1H), 7.18-6.87 (m, 3H), 6.78-6.74 (m, 1H), 3.72 (s, 3H), 3.32-3.27 (m, 2H), 3.21-3.20 (m, 1H), 2,92-2.88 (m, 2H), 1.79 (s, 3H); Other spectral data of **6g** were consistent with previous reported data.<sup>4</sup>



**6h:** white solid, 8.0 mg, 43% yield;  $R_f = 0.4$  (80% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.11 (s, 1H), 7.14 (t, *J* = 53.4 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ 156.4, 154.5, 148.9, 141.3 (t, *J* = 27.9 Hz), 110.4 (t, *J* = 233.8 Hz), 102.8; <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -112.6 (d, *J* = 53.2 Hz, 2F); HRMS (ESI) calcd for C<sub>6</sub>H<sub>4</sub>F<sub>2</sub>N<sub>4</sub>O [M-H<sup>+</sup>]: 185.0275, found: 185.0278.



**6i:** colorless oil, 10.0 mg, 25% yield;  $R_f = 0.4$  (60% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.95 (s, 1H), 7.97 (s, 1H), 7.67-7.60 (m, 1H), 7.53 (s, 1H), 6.89-6.80 (m, 1H), 6.26 (s, 1H) 4.55 (dd, J = 120.9, 14.2 Hz, 2H), 4.20 (qd, J = 7.1, 1.1 Hz, 1H), 1.13 (d, J = 7.1 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  162.9 (dd, J = 251.5, 12.4Hz ), 161.9 (d, J = 13.0 Hz), 158.4 (dd, J = 246.4, 11.7 Hz), 153.3 (d, J = 270.1 Hz), 152.9 (d, J = 9.3 Hz), 151.1, 146.9 (td, J = 26.3, 11.7 Hz), 144.1, 130.6 (dd, J = 9.3, 5.5 Hz), 123.4 (dd, J = 12.2, 4.0 Hz), 111.8 (dd, J = 20.7, 3.3 Hz), 111.1 (t, J

= 244.3 Hz), 104.2 (t, J = 25.9Hz), 57.2 (d, J = 5.4 Hz), 37.4 (d, J = 5.0 Hz), 29.7, 16.1; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -113.7 (s, 1F), -114.4(s, 1F), -123.8(dd, J = 53.5, 10.2 Hz, 2F), -140.9 (s, 1F); HRMS (ESI) calcd for C<sub>17</sub>H<sub>15</sub>F<sub>5</sub>N<sub>5</sub>O [M+H<sup>+</sup>]: 400.1191, found: 400.1195.



**6j:** colorless oil, 6.1mg, 28% yield;  $R_f = 0.5$  (10% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.36 (s,1H), 7.87 (d, J = 8.7 Hz, 1H), 7.34 (t, J = 53.4 Hz, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 189.6 (t, J = 4.0 Hz), 157.1, 128.7 (t, J = 24.1 Hz), 127.9, 126.6, 113.6, 111.8(t, J = 4.0 Hz), 110.3, 62.0, 56.2; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ-104.3 (d, J = 54.6 Hz, 2F); HRMS (ESI) calcd for C<sub>10</sub>H<sub>11</sub>F<sub>2</sub>O<sub>3</sub> [M+H<sup>+</sup>]: 217.0671, found: 217.0667.



**6k:** colorless oil, 9.1mg, 33% yield; 3:1 r.r.;  $R_f = 0.6$  (50% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.68-6.58 (m, 3H), 7.96-7.76 (m, 1H), 7.67-7.56 (m, 2H), 7.36-7.28 (m, 1H), 6.57 (t, J = 55.2 Hz, 1H), 1.68 (d, J = 2.0 Hz, 6H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  200.5, 154.9 (t, J = 26.0 Hz), 152.6, 150.8, 147.4, 139.4, 138.4, 133.5, 132.5, 124.1, 119.8 (t, J = 3.0 Hz), 113.1 (t, J = 242.1 Hz), 50.5, 27.1; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) -121.6 (d, J = 55.3 Hz, 2F); HRMS (ESI) calcd for C<sub>15</sub>H<sub>15</sub>F<sub>2</sub>N<sub>2</sub>O [M+H<sup>+</sup>]: 277.1147, found: 277.1156.



**61:** white solid, 9.3mg, 57% yield;  $R_f = 0.3$  (50% EtOAc/ petroleum ether); <sup>1</sup>H NMR

(400 MHz, DMSO- $d_6$ )  $\delta$ 11.42 (s, 1H), 7.79 (s, 1H), 6.67 (t, J = 54.6 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  162.1 (t, J = 3.1 Hz), 151.3, 143.1 (t, J = 7.5 Hz), 112.7 (t, J = 233.4 Hz), 106.4 (t, J = 23.1 Hz); <sup>19</sup>F NMR (376 MHz, DMSO- $d_6$ )  $\delta$  -115.00 (d, J = 54.7 Hz, 2F); HRMS (ESI) calcd for C<sub>5</sub>H<sub>3</sub>F<sub>2</sub>N<sub>2</sub>O<sub>2</sub> [M-H<sup>+</sup>]: 161.0163, found: 161.0173.



**6m:** colorless oil, 6.0mg, 31% yield;  $R_f = 0.7$  (10% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.51 (s, 1H), 6.72 (t, J = 54.7 Hz, 1H), 2.58 (s, 3H), 2.56 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 174.9, 165.8, 154.4, 121.1 (t, J = 22.7 Hz), 113.1 (t, J = 238.4 Hz), 21.8, 14.2; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -118.9 (d, J = 54.1 Hz, 2F); HRMS (ESI) calcd for C<sub>7</sub>H<sub>9</sub>F<sub>2</sub>N<sub>2</sub>S [M+H<sup>+</sup>]: 191.0449, found: 191.0448.



8: white solid, 9mg, 54% yield; 10:1 r.r.;  $R_f = 0.5$  (15% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.40 (s, 1H), 7.67 (d, J = 7.9 Hz, 1H), 7.43 (d, J = 8.2 Hz, 1H), 7.30 (t, J = 7.6 Hz, 1H), 7.17 (t, J = 7.5 Hz, 1H), 6.84 (t, J = 54.8 Hz, 1H), 6.77 (s, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  136.3, 130.0 (t, J = 24.5 Hz), 127.0, 124.1, 121.7, 120.7, 111.6, 110.5 (t, J = 7.1 Hz), 104.0 (t, J = 7.1 Hz); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -109.9 (d, J = 53.0 Hz, 2F); HRMS (ESI) calcd for C<sub>9</sub>H<sub>6</sub>F<sub>2</sub>N [M-H<sup>+</sup>]: 166.0468, found: 166.0474.



**10**: yellow solid, 15mg, 71% yield;  $R_f = 0.4$  (40% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.61 (s, 1H), 7.96 (d, J = 8.9 Hz, 1H), 7.61 (t, J = 56.0 Hz, 1H), 7.56 (d, J = 8.9 Hz, 1H), 7.47 (d, J = 3.6 Hz, 1H), 7.12-7.09 (m, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.9, 138.7, 128.3, 125.3, 122.2 (t, J = 25.4 Hz), 118.8, 113.2, 112.3 (t, J = 239.2 Hz), 105.6 (t, J = 5.0 Hz); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -110.5 (dd, J = 54.2, 2.6Hz, 2F); HRMS (ESI) calcd for C<sub>9</sub>H<sub>5</sub>F<sub>2</sub>N<sub>2</sub>O<sub>2</sub> [M-H<sup>+</sup>]: 211.0319, found: 211.0326.



**12:** white solid, 7.8mg, 38% yield;  $R_f = 0.7$  (30% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  13.41 (s, 1H), 8.73 (s, 1H), 7.31 (t, *J* = 53.7 Hz, 1H), 7.01 (s, 1H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ 152.2, 152.2, 133.0 (t, *J* = 25.9 Hz), 129.6, 115.7, 110.1 (t, *J* = 234.2 Hz), 100.0 (t, *J* = 6.8 Hz); <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  -107.4 (d, J = 53.8 Hz, 2F); HRMS (ESI) calcd for C<sub>7</sub>H<sub>3</sub>ClF<sub>2</sub>N<sub>3</sub> [M-H<sup>+</sup>]: 201.9984, found: 201.9993.

**14:** colorless oil, 16.7mg, 92% yield;  $R_f = 0.7$  (2% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.58 (d, *J* = 7.8 Hz, 1H), 7.50 (d, *J* = 8.3 Hz, 1H), 7.39 (t, *J* = 7.7 Hz, 1H), 7.30 (t, *J* = 7.4 Hz, 1H), 6.81 (t, *J* = 53.4 Hz, 1H), 2.38 (t, *J* = 2.6 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ154.3, 143.0 (t, *J* = 26.7 Hz), 128.7, 126.2, 123.0, 120.3, 117.0 (t, *J* = 4.5Hz) ,111.8, 109.2 (t, *J* = 234.7 Hz), 7.5; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -115.1 (d, *J* = 53.3 Hz, 2F); HRMS (EI) calcd for C<sub>10</sub>H<sub>8</sub>F<sub>2</sub>O (M<sup>+</sup>): 182.0543, found: 182.0536.



**16:** colorless oil, 12.3mg, 65% yield;  $R_f = 0.7$  (2% EtOAc/ petroleum ether); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.89-7.82 (m, 1H), 7.77-7.74 (m, 1H), 7.45-7.40 (m, 2H), 7.04 (t, J = 55.6 Hz, 1H), 2.49 (t, J = 2.0 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  139.6, 139.1, 133.0 (t, J = 7.6 Hz), 130.1 (t, J = 24.3 Hz), 125.9, 124.5, 122.8, 122.6, 111.4 (t, J = 235.7 Hz), 11.8; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -104.4 (d, J = 55.4 Hz, 2F); HRMS (EI) calcd for C<sub>10</sub>H<sub>8</sub>F<sub>2</sub>S (M<sup>+</sup>): 198.0315, found: 198.0306.



A many products form and most of starting materials remain



B no desired products formation







C no reaction





**D** trace products formation

CO<sub>2</sub>Et CO<sub>2</sub>Me

**Supplementary Figure 2.** 

Unsuccessful substrates



Supplementary Figure 3. The x-ray single crystal structure of 3a.

<b>Supplementary Table 1.</b> Crystal data and structure refinement for shelxl. <b>X-ray crystallography data of 3a.</b>		
Identification code	shelxl	
Empirical formula	$C_{10}H_8F_2N_2O$	
Formula weight	210.18	
Temperature	113(2) K	
Wavelength	0.71073 A	
Crystal system, space group	Monoclinic, P2(1)/c	
Unit cell dimensions	a = 7.1342(14) A alpha = 90 deg.	
	b = 10.658(2) A beta = 106.21(3) deg	
	c = 12.383(3) A gamma = 90 deg.	
Volume	904.1(3) A <sup>3</sup>	
Z, Calculated density	4, 1.544 $Mg/m^3$	
Absorption coefficient	0.130 mm <sup>-1</sup>	
F(000)	432	
Crystal size	0.200 x 0.180 x 0.120 mm	
Theta range for data collection	2.566 to 27.766 deg.	
Limiting indices	-9<=h<=9, -13<=k<=13, -16<=l<=16	
Reflections collected / unique	10502 / 2123 [R(int) = 0.0739]	
Completeness to theta $= 25.242$	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1 and 0.5621	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	2123 / 0 / 137 23	

Goodness-of-fit on F^2	0.974
Final R indices [I>2sigma(I)]	R1 = 0.0472, wR2 = 0.1104
R indices (all data)	R1 = 0.0705, wR2 = 0.1232
Extinction coefficient	n/a
Largest diff. peak and hole	0.333 and -0.447 e.A <sup>-3</sup>

#### **Evaluation of anti-tumor activity**

The in vitro anti-tumor activity of 6d against all cell lines was assessed using the CCK-8, according to the manufacturer's instructions. To test 6d, MCF-7 and HepG-2 cells were seeded into 96-well plates at a density of 5000 cells/well in 100 µL of RPMI 1640 medium containing 10% FBS, 1% penicillin, and 1% streptomycin and cultured for 24 h in 5% CO<sub>2</sub> at 37 °C. HCT116 cells were seeded into 96-well plates at a density of 5000 cells/well in 100 µL of McCoy's 5A medium supplemented with 10% FBS, 1% penicillin, and 1% streptomycin and cultured for 24 h in 5% CO<sub>2</sub> at 37 °C. Likewise, Hela cells were seeded into 96-well plates at a density of 5000 cells/well in 100 µL of MEM supplemented with 10% FBS, 1% penicillin, and 1% streptomycin and cultured for 24 h in 5% CO<sub>2</sub> at 37 °C. 6d was dissolved in PBS contain 10% DMSO and then diluted to the required concentration. It was then added to the cell-containing wells and further incubated at 37  $\,^{\circ}$ C under 5% CO<sub>2</sub> for 72 h. Subsequently, 10 µL of CCK-8 was added into each well and incubated for another 1 h. The plates were then measured at 450 nm using a SpectraMax ® M5 plate reader (Molecular Devices, San Jose, CA, USA). All experiments were carried out five times. For trifluridine, the same procedures were performed by varying the concentration of the specie in question to determine the cytotoxicity.

Eight standard reaction mixtures in 10 mL schlenk tube were equipped with a magnetic stir bar, added 1a (0.2 mmol, 1.0 equiv), CF<sub>2</sub>HSO<sub>2</sub>Na (0.4 mmol, 2.0 equiv) and rose bengal (0.004 mmol, 2 mol%) in DMSO (1.0 mL). Then the mixture was stirred and irradiated by two 3W green LEDs at room temperature. After 1.5 h, the green LEDs were turned off, and one schlenk tube was removed from the irradiation setup for analysis. The remaining seven schlenk tubes were stirred in the absence of light for an additional 1.5 h. Then, one schlenk tube was removed for analysis, and the green LEDs were turned back on to irradiate the remaining six reaction mixtures. After an additional 1.5 h of irradiation, the green LEDs were turned off, and one schlenk tube was removed for analysis. The remaining five schlenk tubes were stirred in the absence of light for an additional 1.5 h. Then, schlenk tube was removed for analysis, and the green LED s were turned back on to irradiate the remaining four reaction mixtures. After 1.5 h, the green LEDs were turned off, and one schlenk tube was removed for analysis. The remaining three schlenk tubes were stirred in the absence of light for an additional 1.5 h, then, a schlenk tube was removed for analysis and the green LEDs were turned back on to 8 irradiate the remaining two reaction mixtures. After 1.5 h, the green LEDs were turned off, and one schlenk tube was removed for analysis. The last schlenk tube was stirred in the absence of light for an additional 1.5 h, and then it was analyzed. The yield was determined by <sup>19</sup>F NMR spectroscopy using benzotrifluoride as the internal standard.



Supplementary Figure 4. Light on/ off experiment

Investigation on the effect of TEMPO and Oxygen.



Reaction conditions: a mixture of **1a** (0.1 mmol),  $NaSO_2CF_2H$  (0.2 mmol), rose bengal (2 mol %) and 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO, 0.2 mmol) in DMSO (1 mL) irradiated with two 3 W green LEDs for 12 hours at room temperature in air.



Reaction conditions: a mixture of **1a** (0.1 mmol), NaSO<sub>2</sub>CF<sub>2</sub>H (0.2 mmol), rose bengal (2 mol %) and 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO, 0.2 mmol) in DMSO (1 mL) irradiated with two 3 W green LEDs for 12 hours at room temperature in argon.

The radical trapping experiments were conducted with and  $CF_2HSO_2Na$  under the standard conditions with a trapping agent 1,1-diphenylethylene (2.0 equiv) to capture the radical intermediate expected in our system, and the products were detected by HRMS techniques. Supplementary Figure 5 showed that 1,1-diphenylethylene, the most common trapping agent, captured diarylmethane radical with 1,1-diphenylethylene-trapped compound **17** observed. HRMS (ESI): compound **17**, HRMS (ESI) calcd for  $C_{15}H_{13}F_2 [M+H]^+$ : 231.0980, found: 231.0985.



**Supplementary Figure 5.** Radical trapping experiment for 1a and  $CF_2HSO_2Na$  under standard conditions with ethene-1,1-diyldibenzene (2.0 equiv)



Supplementary Figure 6. Detection of hydrogen peroxide



**Supplementary Figure 7.** <sup>1</sup> H NMR spectrum of detection of hydrogen peroxide

## **Supplementary Discussions**

We did not observed the peak of hydrogen peroxide  $(H_2O_2)$  by *in situ* <sup>1</sup>H NMR analysis after the reaction mixture in DMSO-*d*<sub>6</sub> was irradiated with two 3W green LEDs for 12 hours in air at room temperature (Supplementary Figure 7, eq 1). However, the peak of water  $(H_2O)$  increased possibly due to the oxygen was finally converted to H<sub>2</sub>O rather than H<sub>2</sub>O<sub>2</sub>. This is because (I) When the reaction was conducted in the H<sub>2</sub>O<sub>2</sub> in the absence of rose bengal under the irradiation of two 3W green LEDs for 12h, no formation of desired product **3a** was observed by <sup>1</sup>H NMR analysis but the peak of hydrogen peroxide  $(H_2O_2)$  remained after the reaction (Supplementary Figure 7, eq 2). (II) When the reaction was conducted in the H<sub>2</sub>O<sub>2</sub> (35 wt % aqueous solution) in the presence of rose bengal under the irradiation of two 3W green LEDs for 12h, a yield of 20% of **3a** was obtained and the peak of H<sub>2</sub>O<sub>2</sub> in <sup>1</sup>H NMR spectrum disappeared after the reaction (Supplementary Figure 7, eq 3). Therefore, the generated H<sub>2</sub>O<sub>2</sub> could participate in the catalytic cycle and ultimately convert to H<sub>2</sub>O as the byproduct and and the result was similar to the previous report by Wu group.<sup>6</sup>



To a 10 mL Schlenk tube equipped with a magnetic stir bar, was added quinoxalin-2(1*H*)-ones **1a** (0.2mmol), CF<sub>2</sub>HSO<sub>2</sub>Na (0.4 mmol) and rose bengal (0.004 mmol, 2 mol%) in DMSO (1.0 mL) Then the mixture was stirred and irradiated by sunlight at room temperature for 12 h (Location:  $39^{\circ}6'2''$  N,  $117^{\circ}9'51''$  E). Afterward, the residue was added water (10 mL) and extracted with ethyl acetate (5 mL × 3). The combined organic phase was dried over Na<sub>2</sub>SO<sub>4</sub>. The resulting crude residue was purified *via* column chromatography on silica gel to afford **3a** in 68% yield.



Supplementary Figure 8. Sunlight-driven experiment



- 3.70

Supplementary Figure 10. <sup>13</sup>C NMR Spectrum of 3a





240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

# Supplementary Figure 11. <sup>19</sup>F NMR Spectrum of 3a



Supplementary Figure 13. <sup>13</sup>C NMR Spectrum of 3b





260 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

# Supplementary Figure 14. <sup>19</sup>F NMR Spectrum of 3b



Supplementary Figure 16.<sup>13</sup>C NMR Spectrum of 3c





Supplementary Figure 17.<sup>19</sup>F NMR Spectrum of 3c


Supplementary Figure 19. <sup>13</sup>C NMR Spectrum of 3d





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260 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

## Supplementary Figure 20.<sup>19</sup>FNMR Spectrum of 3d

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Supplementary Figure 22.<sup>13</sup>C NMR Spectrum of 3e



Supplementary Figure 23.<sup>19</sup>F NMR Spectrum of 3e







Supplementary Figure 26.<sup>19</sup>F NMR Spectrum of 3f



Supplementary Figure 28.<sup>13</sup>C NMR Spectrum of 3g



Supplementary Figure 29. <sup>19</sup>F NMR Spectrum of 3g



Supplementary Figure 31. <sup>13</sup>C NMR Spectrum of 3h



Supplementary Figure 32. <sup>19</sup>F NMR Spectrum of 3h



Supplementary Figure 34. <sup>13</sup>C NMR Spectrum of 3i



260 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

## Supplementary Figure 35. <sup>19</sup>F NMR Spectrum of 3i



Supplementary Figure 37. <sup>13</sup>C NMR Spectrum of 3j



Supplementary Figure 38. <sup>19</sup>F NMR Spectrum of 3j

#### $\begin{array}{c} 8 & 8 & 0 & 0 \\ 8 & 0$



Supplementary Figure 40. <sup>13</sup>C NMR Spectrum of 3k



Supplementary Figure 41. <sup>19</sup>F NMR Spectrum of 3k





Supplementary Figure 43. <sup>13</sup>C NMR Spectrum of 31



Supplementary Figure 44. <sup>19</sup>F NMR Spectrum of 31

### 



Supplementary Figure 46.<sup>13</sup>C NMR Spectrum of 3m



Supplementary Figure 47.<sup>19</sup> F NMR Spectrum of 3m



Supplementary Figure 49.<sup>13</sup>C NMR Spectrum of 3n



Supplementary Figure 50.<sup>19</sup> F NMR Spectrum of 3n



Supplementary Figure 52. <sup>13</sup>C NMR Spectrum of 30



Supplementary Figure 53. <sup>19</sup>F NMR Spectrum of 30



— 12.93



Supplementary Figure 55. <sup>13</sup>C NMR Spectrum of 5a



200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 -260 -280 -300 f1 (ppm)

## Supplementary Figure 56. <sup>19</sup>F NMR Spectrum of 5a



Supplementary Figure 58.<sup>1</sup>H NMR Spectrum of 5c

# 



#### Supplementary Figure 59. <sup>1</sup>H NMR Spectrum of 5d

<ul> <li>153.16</li> <li>149.91</li> <li>149.70</li> <li>149.49</li> </ul>	∠ 132.81 ∠ 132.26 ∖ 130.65 ∩ 129.45 - 123.87	115.73 112.69 110.31 107.93
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Supplementary Figure 60. <sup>13</sup>C NMR Spectrum of 5d



Supplementary Figure 61. <sup>19</sup>F NMR Spectrum of 5d



Supplementary Figure 62.<sup>1</sup>H NMR Spectrum of 5e



Supplementary Figure 64.<sup>13</sup>C NMR Spectrum of 5f



Supplementary Figure 65. <sup>19</sup>F NMR Spectrum of 5f



Supplementary Figure 67.<sup>13</sup>C NMR Spectrum of 5g



Supplementary Figure 68. <sup>19</sup>F NMR Spectrum of 5g



Supplementary Figure 70. <sup>13</sup>C NMR Spectrum of 5h



260 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

## Supplementary Figure 71. <sup>19</sup>F NMR Spectrum of 5h




Supplementary Figure 73.<sup>1</sup>H NMR Spectrum of 5j



Supplementary Figure 73. <sup>1</sup>H NMR Spectrum of 5k



Supplementary Figure 75.<sup>13</sup>C NMR Spectrum of 51



Supplementary Figure 76. <sup>19</sup>F NMR Spectrum of 51



Supplementary Figure 78.<sup>13</sup>C NMR Spectrum of 5m



60 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

Supplementary Figure 79. <sup>19</sup>F NMR Spectrum of 5m



Supplementary Figure 81. <sup>1</sup>H NMR Spectrum of 50





Supplementary Figure 83. <sup>13</sup>C NMR Spectrum of 5p



Supplementary Figure 84. <sup>19</sup>F NMR Spectrum of 5p



Supplementary Figure 86. <sup>13</sup>C NMR Spectrum of 5q



200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 -260 -280 -300 f1 (ppm)

## Supplementary Figure 87. <sup>19</sup>F NMR Spectrum of 5q



Supplementary Figure 89. <sup>13</sup> C NMR Spectrum of 5r

- -115.86 - -116.00



260 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

Supplementary Figure 90. <sup>19</sup>F NMR Spectrum of 5r



Supplementary Figure 92. <sup>13</sup>C NMR Spectrum of 5s



Supplementary Figure 93. <sup>19</sup>F NMR Spectrum of 5s



**Supplementary Figure 95.** <sup>13</sup>C NMR Spectrum of **5t** 



## Supplementary Figure 96. <sup>19</sup>F NMR Spectrum of 5t



## Supplementary Figure 98. <sup>1</sup>H NMR Spectrum of 6b



Supplementary Figure 99. <sup>1</sup>H NMR Spectrum of 6c





Supplementary Figure 101. <sup>13</sup>C NMR Spectrum of 6d



Supplementary Figure 102. <sup>19</sup>F NMR Spectrum of 6d



Supplementary Figure 103. <sup>13</sup> C NMR Spectrum of 6e



Supplementary Figure 104.<sup>19</sup> F NMR Spectrum of 6e



Supplementary Figure 106. <sup>13</sup>C NMR Spectrum of 6f



Supplementary Figure 107. <sup>19</sup>F NMR Spectrum of 6f



Supplementary Figure 108. <sup>1</sup>H NMR Spectrum of 6g



Supplementary Figure 110. <sup>13</sup>C NMR Spectrum of 6h



Supplementary Figure 111. <sup>19</sup>F NMR Spectrum of 6h

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## Supplementary Figure 113. <sup>13</sup>C NMR Spectrum of 6i



Supplementary Figure 114. <sup>13</sup>F NMR Spectrum of 6i



Supplementary Figure 116. <sup>13</sup>C NMR Spectrum of 6j



Supplementary Figure 117. <sup>19</sup>F NMR Spectrum of 6j



Supplementary Figure 119. <sup>13</sup>C NMR Spectrum of 6k



Supplementary Figure 120.<sup>19</sup>F NMR Spectrum of 6k



Supplementary Figure 122. <sup>13</sup>C NMR Spectrum of 6l



Supplementary Figure 123.<sup>19</sup>F NMR Spectrum of 61


Supplementary Figure 125. <sup>13</sup>C NMR Spectrum of 6m



Supplementary Figure 126. <sup>19</sup>F NMR Spectrum of 6m

#### - 8.40 7.68 7.66 7.42 7.42 7.42 7.23 7.23 7.23 7.23 7.23 7.23 7.23 6.53 6.73 6.73 6.73 6.73 6.73





135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 15 10 5 f1(ppm)

### Supplementary Figure 128. <sup>13</sup>C NMR Spectrum of 8



# 7.97 7.95 7.75 7.61 7.57 7.56 7.55 7.55 7.54 7.48 7.48 7.48 7.48 7.48 7.47 7.11 7.11 7.11 7.11 7.11 7.11 7.10





Supplementary Figure 130. <sup>1</sup>H NMR Spectrum of 10







145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 15 10 5 f1 (ppm)

### Supplementary Figure 131. <sup>13</sup>C NMR Spectrum of 10



Supplementary Figure 132. <sup>19</sup>F NMR Spectrum of 10



## Supplementary Figure 134. <sup>13</sup>C NMR Spectrum of 12



Supplementary Figure 135. <sup>19</sup>F NMR Spectrum of 12

2.38 2.38 2.37



Supplementary Figure 137. <sup>13</sup>C NMR Spectrum of 14



Supplementary Figure 138. <sup>19</sup>F NMR Spectrum of 14

2.51
2.50
2.50
2.50



155 150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 20 15 10 5 f1 (ppm)

#### Supplementary Figure 140. <sup>13</sup>C NMR Spectrum of 16





260 240 220 200 180 160 140 120 100 80 60 40 20 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240 f1 (ppm)

#### **Supplementary Figure 140.** <sup>19</sup>F NMR Spectrum of **16 Supplementary References**

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