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Supplemental Information

Pharmacological Activation of Pyruvate

Kinase M2 Inhibits CD4⁺ T Cell

Pathogenicity and Suppresses Autoimmunity

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SUPPLEMENTARY FIGURE TITLES AND LEGENDS



Figure S1. PKM1 expression in resting and activated murine and human CD4⁺ T cells. Related to Figure 1 and 7. (A-C) Murine $CD4^+CD62^+$ T cells were stimulated *in vitro* for 3 days with CD3/CD28 antibodies and collected at different time points of activation. (A) Left: western blot showing upregulation of PKM1 protein in murine CD4⁺ T cells following activation. Right: quantification of PKM1 expression by densitometry analysis (n=3 from two independent

experiments). (**B**) Quantification of *Pkm1* mRNA expression in resting versus activated murine CD4⁺ T cells by qRT-PCR (n=4 from three independent experiments). ****P*<0.001 compared to resting condition, by one-way Anova with Dunnett's post-hoc test. (**C**) Fold abundance of *Pkm2* over *Pkm1* mRNA in resting and activated murine CD4⁺ T cells (n=4-6 from three-four independent experiments). (**D**-**F**) Human naïve CD4⁺ T cells were stimulated *in vitro* for 4 days with anti-CD3/CD28 antibodies and collected at different time points of activation. (**E**) Left: western blot showing upregulation of PKM1 protein in human CD4⁺ T cells following activation. Right: quantification of PKM1 expression by densitometry analysis (n=3 from three independent experiments). (**F**) Quantification of *Pkm1* mRNA expression in human CD4⁺ T cells by qRT-PCR (n=5 from three independent experiments). ***P*<0.01 compared to resting condition, by one-way Anova with Dunnett's post-hoc test. (**C**) Fold abundance of *PKM2* versus *PKM1* expression levels in resting and activated human CD4⁺ T cells (n=5 from three independent experiments). For all panels, data are the mean \pm SD.



Figure S2. Effect of TEPP-46 on PKM2 tetramerisation, PKM2 nuclear translocation and T cell viability. Related to Figure 2. (A) Cells were collected after 3 days of stimulation, crosslinked with DSS and analysed for PKM2 expression. Left: Representative western blot showing induction of PKM2 tetrameric isoform by TEPP-46. Right: quantification of relative tetrameric and monomeric isoform expression over total PKM2 by densitometry analysis (n=4 from three independent experiments). (B) Cells were collected after 2 days of stimulation and PKM2 expression in nucleus and cytoplasm was analysed by western blot after cell fractionation. One representative experiment out of three showing a dose-dependent reduction of nuclear PKM2 expression in cells treated with TEPP-46 is displayed. (C) Murine CD4⁺ T cells were collected after

3 days of activation in the presence of TEPP-46. Cells were stained with PI and Annexin V (AV) to determine live (PI⁻AV⁻), apoptotic (PI⁻AV⁺) or necrotic/dead cells (PI⁺AV⁺) (n=6 from three independent experiments). For all panels, data are the mean \pm SD.



Figure S3. Effect of DASA-58 on CD4⁺ T cell activation and generation of Th17 and Th1 cells in vitro. Related to Figure 2, Figure 4 and Figure 5. (A-C) Murine CD4⁺CD62⁺ T cells were activated in vitro with CD3/CD28 antibodies in the presence of DMSO (CTRL condition) or DASA-58 25 μ M. (A) Cells were collected after 3 days of stimulation. Left: representative flow cytometry plot displaying T cell proliferation assessed as CellTraceTM Violet dilution. Right: quantification of division index by FlowJo software (n=3 from two independent experiments). (B) Percentage of IL-2-producing cells and IL-2 MFI in CTRL versus DASA-58-treated cells 24 hours upon activation (n=4 from two independent experiments). (C) MFI of surface CD62L and CD44 expression, evaluated by flow cytometry. (n=4 from two independent experiments). (**D**) Murine CD4⁺CD62⁺ T cells were activated *in vitro* under Th17-polarising conditions in the presence of DMSO (CTRL condition) or DASA-58 100 µM. Expression of Il17a, Il17f, Il21 and Tnfa mRNA in CTRL and DASA-treated Th17 cells was quantified by gRT-PCR (n=5 from two independent experiments). (E) Murine CD4⁺CD62⁺ T cells were activated *in vitro* under Th1-polarising conditions in the presence of DMSO (CTRL condition) or DASA-58 25 µM. Expression of Ifng and Tnfa mRNA in CTRL and DASA-treated Th1 cells was quantified by qRT-PCR (n=8 from three independent experiments). For all panels, data are the mean \pm SD. *P<0.05, ** P<0.01, ***P<0.001 or ****P<0.0001 compared to CTRL condition, by unpaired (A-C) or paired (D-E) Student's t test.



Figure S4. Induction of Foxp3⁺CD25⁺ T cells by TEPP-46 and effect of TEPP-46 on TGF-βinduced Tregs. Related to Figure 2. (A) Quantification of *Foxp3* mRNA levels by qRT-PCR in murine CD4⁺ T cells activated in the presence of DMSO (CTRL condition) or TEPP-46 (n=7 from six independent experiments). (B) Left: representative plot showing induction of Foxp3⁺CD25⁺ T cells by TEPP-46 treatment. Right: quantification of the percentage of Foxp3⁺CD25⁺ T cells in CTRL and TEPP-46-treated cells (n=6 from four independent experiments). (C) Left: representative flow cytometry plots showing Foxp3⁺CD25⁺ T cells in freshly-isolated CD4⁺CD62L⁺ resting T cells and naïve CD4⁺ T cells. Right: quantification of Foxp3⁺CD25⁺ T cell percentage in the two populations (n=3). (D) Naïve CD4⁺ T cells were activated with CD3/CD28 antibodies in the presence of TEPP-46. The percentage of Foxp3⁺CD25⁺ T cells was evaluated after 3 days of stimulation. (n=4 from 2 independent experiments). (E) Murine CD4⁺CD62⁺ T cells were activated

in vitro with CD3/CD28 antibodies under Treg-polarising conditions, in the presence of TEPP-46. The percentage of Foxp3⁺CD25⁺ T cells in CTRL and TEPP-46-treated cells was quantified by flow cytometry (n=8 from four independent experiments). (F) Analysis of Stat5 phosphorylation in TGF- β -induced Tregs. Left: western blot showing block of Stat5 phosphorylation by TEPP-46. Right: quantification of phospho-Stat5/Stat5 ratio by densitometry analysis (n=3 from three independent experiments). In all panels, data are the mean the mean ± SD. ***P*<0.01 or ****P*<0.001, compared to CTRL condition, by one-way Anova with Dunnett's post-hoc test (A, B and E) or unpaired Student's t test (D).



Figure S5. Global expression profile of resting CD4⁺ T cells and CD4⁺ T cells activated in the presence of DMSO (Th0 Ctrl) or TEPP-46 100 μ M (Th0 TEPP). Related to Figure 2 and Figure 3. (A) Plot showing Principal Component analysis of global gene expression in the three T cell populations. (B) Heat map showing an overview of global gene expression in resting and Th0 Ctrl or Th0 TEPP-46 cells. (C) Heat map showing expression of genes related to T cells activation in the three populations. (D) Heat map showing expression of regulatory T cell signature genes in the three populations.



Figure S6. Effect of TEPP-46 on Myc and Hif-1 α expression, mTORC1 activity and expression of glycolytic genes in activated T cells. Related to Figure 3. (A and B) Murine CD4⁺ T cells were collected after 24 hours of *in vitro* activation with CD3/CD28 antibodies in the presence of DMSO (CTRL condition) or TEPP-46. (A) Left: western blot image showing reduction of Myc expression by TEPP-46 treatment. Right: quantification of relative Myc expression in

CTRL and TEPP-46-treated cells by densitometry analysis (n=3 from two independent experiments). (**B**) Quantification of *Myc* mRNA levels in CTRL and TEPP-46-treated cells by qRT-PCR (n=8 from 5 independent experiments). (**C**) Left: image from one representative experiment showing reduction of p70 S6 and p85 S6 phosphorylation by TEPP-46. Right: quantification of relative P-p70 S6 and P-p85 S6 band intensity by densitometry analysis (n=4 from two independent experiments). (**D**-**F**) Murine CD4⁺ T cells were collected after 3 days of *in vitro* activation in the presence of DMSO or TEPP-46. (**D**) Left: western blot image showing Hif-1 α downregulation in TEPP-46-treated cells, compared to CTRL cells. Right: quantification of relative Hif-1 α expression by densitometry analysis (n=3 from two independent experiments). (**E**) *Hif1a* mRNA expression in CTRL and TEPP-46-treated cells quantified by qRT-PCR (n=5 from five independent experiments). (**F**) Expression of glycolytic genes in CTRL and TEPP-46-treated T cells by qRT-PCR (n=5-6 from 5 independent experiments). For all panels, data are the mean ± SD. *P<0.05, **P<0.01, ***P<0.001 or ****P<0.0001, compared to CTRL condition, by one-way Anova with Dunnett's post-hoc test.



Figure S7. Expression of PKM2 in murine Th17 and Th17 cells and induction of Tregs under Th17 and Th17 polarising-conditions *in vitro*. **Related to Figure 4 and 5.** Murine CD4⁺CD62⁺ T cells were activated *in vitro* for 3 days with CD3/CD28 antibodies under Th17- or Th1-polarising conditions. (**A**) Quantification of *Pkm2* mRNA expression levels in resting CD4⁺CD62L⁺ T cells versus Th17 and Th1 cells at different time points of activation by qRT-PCR (n=4-6 from four independent experiments). (**B**) Western blot showing upregulation of PKM2 protein in Th17 and Th1 cells following activation. A representative experiment out of two is shown. (**C**) *Foxp3* gene expression in CTRL versus TEPP-46-treated Th17 cells (n=7 from four independent experiments). (**D**) Left: representative plot showing induction of Foxp3⁺CD25⁺ T cells under Th17-polarising conditions by TEPP-46. Right: quantification of the percentage of Foxp3⁺CD25⁺ T cells in CTRL and TEPP-46-treated cell populations (n=6 from two independent experiments). (**E**) *Foxp3*

expression in CTRL versus TEPP-46-treated Th1 cells (n=5 from three independent experiments). (F) Left: representative plot showing induction of $Foxp3^+CD25^+$ T cells by TEPP-46 under Th1-polarising conditions. Right: quantification of the percentage of $Foxp3^+CD25^+$ T cells in CTRL and TEPP-46-treated cell populations (n=5 from two independent experiments). For all panels, data are the mean ± SD. **P*<0.05, ****P*<0.001 or *****P*<0.001, compared to CTRL condition, by one-way Anova with Dunnett's post-hoc test.